

Agilent VnmrJ 4

Command and Parameter Reference Guide



Notices

© Agilent Technologies, Inc. 2013

No part of this manual may be reproduced in any form or by any means (including electronic storage and retrieval or translation into a foreign language) without prior agreement and written consent from Agilent Technologies, Inc. as governed by United States and international copyright laws.

Manual Part Number

G7446-90519

Software Revision

This guide is valid for 4.0 and 4.0 revisions of the Agilent VnmrJ software, until super-seded.

Edition

Rev. A, March 2013

Printed in USA

Agilent Technologies, Inc. 5301 Stevens Creek Boulevard Santa Clara, CA 95051 USA

Warranty

The material contained in this document is provided "as is," and is subject to being changed, without notice, in future editions. Further, to the maximum extent permitted by applicable law, Agilent disclaims all warranties, either express or implied, with regard to this manual and any information contained herein, including but not limited to the implied warranties of merchantability and fitness for a particular purpose. Agilent shall not be liable for errors or for incidental or consequential damages in connection with the furnishing, use, or performance of this document or of any information contained herein. Should Agilent and the user have a separate written agreement with warranty terms covering the material in this document that conflict with these terms, the warranty terms in the separate agreement shall control.

Technology Licenses

The hardware and/or software described in this document are furnished under a license and may be used or copied only in accordance with the terms of such license.

Restricted Rights Legend

If software is for use in the performance of a U.S. Government prime contract or subcontract. Software is delivered and licensed as "Commercial computer software" as defined in DFAR 252.227-7014 (June 1995), or as a "commercial item" as defined in FAR 2.101(a) or as "Restricted computer software" as defined in FAR 52.227-19 (June 1987) or any equivalent agency regulation or contract clause. Use, duplication or disclosure of Software is subject to Agilent Technologies' standard commercial license terms, and non-DOD Departments and Agencies of the U.S. Government will receive no greater than Restricted Rights as defined in FAR 52.227-19(c)(1-2) (June 1987). U.S. Government users will receive no greater than Limited Rights as defined in FAR 52.227-14

(June 1987) or DFAR 252.227-7015 (b)(2) (November 1995), as applicable in any technical data.

Safety Notices

CAUTION

A **CAUTION** notice denotes a hazard. It calls attention to an operating procedure, practice, or the like that, if not correctly performed or adhered to, could result in damage to the product or loss of important data. Do not proceed beyond a **CAUTION** notice until the indicated conditions are fully understood and met.

WARNING

A WARNING notice denotes a hazard. It calls attention to an operating procedure, practice, or the like that, if not correctly performed or adhered to, could result in personal injury or death. Do not proceed beyond a WARNING notice until the indicated conditions are fully understood and met.

Contents

1 Notational Conventions

Title line codes63Applicability63Command and macro syntax64Parameter Syntax65Notational Conventions65Other Sources of Information65

2 A

aaAbort acquisition with error (C) 71
abort Terminate action of calling macro and all higher macros (C) 72
abortallacgs Reset acquisition computer in a drastic situation (C) 72
abortoff Terminate normal functioning of abort in a macro (C) 73
aborton Restore normal functioning of abort in a macro (C) 73
abs Find absolute value of a number (C) 73
AC1S-AC11S . Autocalibration macros (M) 73
ACbackup Make backup copy of current probe file (M) 74
acctWrites records for operator login and logoff (M) 74
ACreport Print copy of probe file after autocalibration (M) 74
acos Find arc cosine of number (C) 75
acosy Automatic analysis of COSY data (C) 75
acosyold Automatic analysis of COSY data, old algorithm (C) 75
acq_errors Acquisition Done and Error Codes 76
acgdequeue Dequeue an acquisition 81
acqdisp Display message on the acquisition status line (C) 81
acgiInteractive acquisition display process (C) 82
acqmeter Open Acqmeter window (M) 83
Acqmeter Open Acqmeter window (U) 84
acgmode Acquisition mode (P) 85
acgreserve Reserve the acquisition console for the current owner 85
acgstat Open Acquisition Status window (M) 86
Acgstat Open Acquisition Status window (U) 87
acgstatus Acquisition status (P) 87
acmire Acquire data (M) 90
actionid Current study queue node id (P) 90

add Add current FID to add/subtract experiment (C) 91 addi...... Start interactive add/subtract mode (C) 93 addnucleus.... Add new nucleus to existing probe file (M) 94 addpar..... Add selected parameters to current experiment (M) 95 addparams Add parameter to current probe file (M) 96 addprobe...... Create new probe directory and probe file (M) 97 adept Automatic DEPT analysis and spectrum editing (C) 97 aexpp1..... Automatic plot of spectral expansion (M) 98 ai..... Select absolute-intensity mode (C) 99 aig Absolute-intensity group (P) 99 aipDisplay.... Display images (C) 99 100 aipLoadSpec . Load fdf spectra (C) aipMakeMaps ... Make csi map (C) 100 aipOverlayFramesOverlay images in selected frames (C) 101 aippars..... Create parameters for imaging browser (M) 101 aipRemoveSpec Remove all or specified spectral data (C) 106 aipRQcommand. Load and display images (C) 106 aipSaveColormapSave color map for selected images (C) 108 aipSetColormap Load color map for selected images (C) 109 aipSetTransparencySet transparency for images or text (C) 109 aipShow3PCursorsTurn on/off cursors for 3-plane extraction 109 aipShowCSIDataDisplay CSI spectral data in grid format 110 aipShowSpec Display spectra specified by key(s), in specified layout (C) 111 aipViewLayers... Get information for overlaid images (C) 112 alfa..... Set alfa delay before acquisition (P) 112 alock Automatic lock control (P) 113 **ampmode** Independent control of amplifier mode (P) 114 amptype Amplifier type (P) 114 analyz..... Calculate standard peak height (M) 115 **analyze** Generalized curve fitting (C) 115 annotation.... Display annotation specified by the parameter "template" or the default 118 **ap**..... Print out "all" parameters (C) 118 ap..... "All" parameters display control (P) 119 apa Plot parameters automatically (M) 119 **aph** Automatic phase adjustment of spectra (C) 119 **aph0**..... Automatic phase of zero-order term (C) 120 **aphb**...... Auto phasing for Bruker data (C) 120

aphx...... Perform optimized automatic phasing (M) 120 appdir..... Application directory information 122 appdirs Starts Applications Directory Editor (M) 123 appmode Application mode (P) 123 124 **apptype** Application type (P) 124 aptaph...... Automatic processing for APT spectra (M) 124 array Easy entry of linearly spaced array values (M) 125 array Parameter order and precedence (P) 125 **arraydim**...... Dimension of experiment (P) 126 array2csv Formats Array into Comma Separate Variable 126 array2stringFormats Array into String 126 array2strsv. Formats Array into String Separated Variable 127 **asin**...... Find arc sine of number (C) 127 **asize** Make plot resolution along f₁ and f₂ the same (M) 128 assign...... Assign transitions to experimental lines (M) 128 at Acquisition time (P) 129 atan..... Find arc tangent of a number (C) 129 atan2 Find arc tangent of two numbers (C) 129 atcmd Call a macro at a specified time (M) 130 atext Append string to current experiment text file (M) 131 attva1...... Calculate pulse width (M) 131 atune ProTune Present (P) 131 (M) 132 AuCALch3i Set up autocalibration with CH3I sample (M) 133 AuCALch3i1.... Get autocalibration with CH₃I sample (M) 133 **AuCALch3oh**.... Set up autocalibration with Autotest sample (M) 133 **AuCALch3oh1**. Get autocalibration with Autotest sample (M) 134 Aucalibz0 Automatic Hz to DAC calibration for Z0 (M) 134 **AuCdec**...... Carbon decoupler calibration macro (M) 134 AuCgrad Carbon/proton gradient ratio calibration macro (M) 134 AuCobs...... Carbon observe calibration macro (M) 135 audiofilter. Audio filter board type (P) 135 Aufindz0...... Automatic adjustment of Z0 (M) 135 Augca1..... Probe gcal calibration macro (M) 135 Augmap Automated gradient map generation (M) 136 Augmapz0...... Automatic lock gradient map generation and z0 calibration (M) 136

AuHdec	Proton decoupler calibration (M) 136
AuHobs	Proton observe calibration macro (M) 137
Aumakegmap	Auto lock gradient map generation (M) 137
AuNuc	Get parameters for a given nucleus (M) 137
auto	Prepare for an automation run (C) 137
auto	Automation mode active (P) 138
autoaa	Abort an automation run with no error 138
auto_au	Controlling macro for automation (M) 139
autog	Utility commands for the automation queue 139
Autobackup	Back up current probe file (M) 140
autodept	Automated complete analysis of DEPT data (M) 140
autodir	Automation directory absolute path (P) 141
autogo	Start automation run (C) 141
autolist	Set up and start chained acquisition (M) 142
automerge	Merges overniteQ with daytimeQ 142
Automkdir	Creates Data Directory from Template 143
autoname	Create path for data storage (C) 143
autoname	Prefix for automation data file (P) 146
autora	Resume suspended automation run (C) 146
autosa	Suspend current automation run (C) 146
autoscale	Resume autoscaling after limits set by scalelimits macro (M) 147
autostack	Automatic stacking for processing and plotting arrays (M) 147
autotest	Open Auto Test Window (C) 147
autotime	Displays approximate time for automation (M) 147
av	Set abs. value mode in directly detected dimension (C) 148
av1	Set abs. value mode in 1st indirectly detected dimension (C) 148
av2	Set abs. value mode in 2nd indirectly detected dimension (C) 149
averag	Calculate average and standard deviation of input (C) 150
awc	Additive weighting const. in directly detected dimension (P) 150
awc1	Additive weighting const. in 1st indirectly detected dimension (P) 151
awc2	Additive weighting const. in 2nd indirectly detected dimension (P) 151
axis	Provide axis labels and scaling factors (C) 151

axis Axis label for displays and plots (P) 152		52
axisf	Axis label for FID displays and plots (P)	153

3 B

bandinfo Shaped pulse information for calibration (M) 1	55
bannerDisplay message with large characters (C) 156	
ъс 1D and 2D baseline correction (C) 156	
beepoffTurn beeper off (C) 158	
beeponTurn beeper on (C) 158	
bigendian Determine system byte order (C) 158	
binom Set up parameters for BINOM pulse sequence (M	l) 159
biorefBio-NMR Referencing (P) 159	
bootup Macro executed automatically (M) 159	
ьож Draw a box on a plotter or graphics display (C)	160
boxes Draw boxes selected by the mark command (M)	161
bpa Plot boxed parameters (M) 161	
bph Individually phase each trace of arrayed 1D data	162
br24Set up parameters for BR24 pulse sequence (M)	163
ъs Block size (Р) 163	

4 C

c13 Automated carbon acquisition (M) 169
c13p Process 1D carbon spectra (M) 169
calcdim Calculate dimension of experiment (C) 170
calcECC Calculate ECC corrections (C) 170
$\verb calfa Recalculate alfa so that first-order phase is zero (M) 170$
calibflag Correct systematic errors in DOSY experiments (P) 171
calibrate Start a dialog for autocalibration routines (M) 171
callacg Utility macro to call Acq command (M) 171
captAutomated carbon and APT acquisition (M) 172
Carbon Set up parameters for 13C experiment (M) 172
cat Display one or more text files in text window (C) 172
cattn Coarse attenuator type (P) 173
ca Change working directory (C) 173
cac Cancel drift correction (C) 173
cdept Automated carbon and DEPT acquisition (M) 174
cdump Prints the current graphics screen (M) 174
celem Completed FID elements (P) 175
center

centerprobe . Calculates probe position relative to the ISO-Center 1	75
centersw Move cursor to center of spectrum (M) 176	
centersw1 Move cursor to center of spectrum in 1st indirect dimens (M) 176	ion
centersw2 Move cursor to center of spectrum in 2nd indirect dimension (M) 176	
сехр Create aVnmr experiment (M) 176	
cfCurrent FID (P) 177	
cfpmult Calculate first-point multiplier for 2D experiments (M) 178	
change Submit a change sample experiment to acquisition (M) 178	
checkstring Find and replace unwanted characters (C) 179	
<pre>chiliConf Control flag set by ecc_on and ecc_off (P) 179</pre>	
chkname Parse the template and return substituted strings and lis of parameters defined by the template 179	sts
Cigar2j3j Convert the parameter to a CIGAR2j3j experiment (M) 182	
ckresloc Macro to Reserve Specific Locations 182	
ckstring Utility to Check String Variables for Illegal Characters	182
claClear all line assignments (M) 183	
claCalculated transition number (P) 183	
clamp Calculated transition amplitude (P) 183	
cleanexp Remove old files and directories from an experiment (M) 184	
clear Clear a window (C) 184	
<pre>cleardosy Delete temporarily saved data in current sub experiment</pre>	:
clfreg Calculated transition frequency (P) 185	
clindex Index of experimental frequency of a transition (P) 18	5
clradd Clear add/subtract experiment (C) 185	
cmdlineox Determine if an operator has a command line 185	
coldprobe Tells system a coldprobe is present 186	
color Select plotting colors from a graphical interface (M) 1	86
combiplate View a color map for visual analysis of VAST microtiter plate (U) 186	
combishow Display regions (red, green, and blue) in CombiPlate window (M) 187	
compressfid . Compress double-precision FID data (M,U) 187	
config Display current configuration and possibly change it (M) 188	

confirm Confirm message using the mouse (C) 189
Console System console type (P) 189
contact_timeMAS cross-polarization spin-lock contact time (M) 190
continflag The command ddif creates a CONTIN display if continflag='y'. 190
continprepareCalled by the macro dosy to prepare the input file for the CONTIN programme. 191
continread Called by the macro dosy to take the output of the CONTIN programme and create an input file for ddif. 191
continueMovie Continue movie in either forward or backward direction (C) 191
convert Convert data set from a VXR-style system (M,U) 192
convertbru Convert Bruker data (M,U) 192
сору Copy a file (C) 195
cos Find cosine value of an angle (C) 196
Cosy Convert the parameter to a COSY experiment (M) 196
совуря Set up parameters for phase-sensitive COSY pulse sequence (M) 196
cp Copy a file (C) 197
cp Cycle phase (P) 197
cpdone Macro called upon study completion (M) 197
cpgo Macro called when acquisition is started (M) 198
cpmgt2 Set up parameters for CPMGT2 pulse sequence (M) 198
cpos_cvt Convert data set from a VXR-style system (M,U) 198
cptmp Copy experiment data into experiment subfile (M) 199
cptmpltdefaultsDefaults for Save Data Template 199
срх Create pbox shape file (M) 199
cqexp Load experiment from protocol (M) 200
cqfindz0 Run an experiment to find the value of z0 (M) 200
cggmap Perform gradient shimming utility functions (M) 200
cqinit Initialize liquids study queue (M) 200
cqpars Create study queue parameters for liquids (M) 201
cgplot Macro to perform generic 2D plot (M) 201
cgprotocol Macro to create protocols (M) 201
cqprotocol Macro to create protocols (M) 201 cqreset
cqprotocolMacro to create protocols (M)201cqresetReset study queue parameters (M)201cqsavestudy .Macro to save study queue parameters (M)201
cqprotocol Macro to create protocols (M)201cqreset
cqprotocol Macro to create protocols (M)201cqreset

cr2	Cursor position in 2nd indirectly detected dimension (P) 203
crcom	Create user macro without using text editor (M) 203
create	Create new parameter in a parameter tree (C) 203
create (P)	Parameter used for RF transmitter board temperature compensation 205
createqcomp	Create qcomp parameter (M) 205
crf	Current time-domain cursor position (P) 205
crl	Clear reference line in directly detected dimension (M) 206
crl1	Clear reference line in 1st indirectly detected dimension (M) 206
cr12	Clear reference line in 2nd indirectly detected dimension (M) $$206$$
crmode	Current state of the cursors in df, ds, or dconi programs (P) 207
crof2	Recalculate rof2 so that $Ip = 0$ (M) 207
cryo_noisete	estRun Cold Probe conditioning experiments (M) 207
cryoclient	Start the CryoBay Monitor program (M, U) 208
csv2cpQ	Imports CSV data (M) 209
ct	Completed transients (P) 210
ctext	Clear the text of the current experiment (C) 211
curexp	Current experiment directory (P) 211
curscan	Scan currently in progress (P) 211
curwin	Current window (P) 211
cutoff	Data truncation limit (P) 212
cyclence	Set up parameters for CYCLENOE pulse sequence (M) 212
cy1br24	Set up parameters for cycled BR24 pulse sequence (M) 212
cylmrev	Set up parameters for cycled MREV8 pulse sequence (M) 213
CZ	Clear integral reset points (C) 213
d0	Overhead delay between FIDs (P) 222
d1	First delay (P) 223
d2	Incremented delay in 1st indirectly detected dimension (P) 223

d2pul Set up parameters for D2PUL pulse sequence (M) 223

5 D

d3	Incremented delay for 2nd indirectly detected dimension (P) 224
d4	Incremented delay for 3rd indirectly detected dimension (P) 224
DAC_to_G	Store gradient calibration value in DOSY sequences (P) 225
da	Display acquisition parameter arrays (C) 225
daslp	Increment for t1 dependent first-order phase correction (P) 225
date	Date (P) 226
daxis	Display horizontal LC axis (M) 226
Dbppste	Set up parameters for Dbppste pulse sequence (M) 226
Dbppsteinept	Set up parameters for Dbppsteinept pulse sequence: (M) 227
dbsetup	. Set up VnmrJ database (U) 227
dbupdate	Update the VnmrJ database (U) 228
dc	Calculate spectral drift correction (C) 228
dc2d	Apply drift correction to 2D spectra (C) 229
dcg	Drift correction group (P) 229
dcon	Display noninteractive color intensity map (C) 229
dconi	Interactive 2D data display (C) 231
dconi	Control display selection for the dconi program (P) 232
dconn	Display color intensity map without screen erase (C) 233
dcrmv	Remove dc offsets from FIDs in special cases (P) 234
ddf	Display data file in current experiment (C) 234
ddff	Display FID file in current experiment (C) 234
ddfp	Display phase file in current experiment (C) 235
ddif	Synthesize and show DOSY plot (C) 235
ddrcr	Direct digital receiver coefficient ratio (P) 236
ddrpm	Set ddr precession mode (P) 236
ddrtc	Set ddr time constant (P) 237
dds	Default display (M) 237
dds_seqfil	Sequence-specific default display (M) 237
debug	Trace order of macro and command execution (C) 237
decasynctype	Decoupler asynchronous scheme (P) 238
decay_gen	Calculates the form of diffusional attenuation expected for the measured gradient and signal maps in non-uniform gradient calibration. 238
deccwarnings	Control reporting of DECC warnings from PSG (P) 239
decomp	Decompose a VXR-style directory (M) 239

def_osfilt	Default value of osfilt parameter (P) 240
defaultdir	Default directory for Files menu system (P) 240
delcom	Delete a user macro (M) 240
delete	Delete a file, parameter directory, or FID directory (C) 241
delexp	Delete an experiment (M) 241
delexpdata	Delete data from the current experiment 242
deletenucleu	s Removes nucleus entry from current probe file (M) 242
dels	Delete spectra from T_1 or T_2 analysis (C) 243
delta	Cursor difference in directly detected dimension (P) 243
delta1	Cursor difference in 1st indirectly detected dimension (P) 243
delta2	Cursor difference in 2nd indirectly detected dimension (P) 244
deltaf	Difference of two time-domain cursors (P) 244
dept	Set up parameters for DEPT experiment (M) 244
deptgl	Set up parameters for DEPTGL pulse sequence (M) 244
deptproc	Process array of DEPT spectra (M) 245
destroy	Destroy a parameter (C) 245
destroygroup	Destroy parameters of a group in a tree (C) 246
df	Display a single FID (C) 246
df2d	Display FIDs of 2D experiment (C) 247
dfid	Display a single FID (C) 247
dfmode	Current state of display of imaginary part of a FID (P) 248
dfrq	Transmitter frequency of first decoupler (P) 248
dfrq2	Transmitter frequency of second decoupler (P) 248
dfrq3	Transmitter frequency of third decoupler (P) 249
dfrq4	Transmitter frequency of fourth decoupler (P) 249
dfs	Display stacked FIDs (C) 249
dfsa	Display stacked FIDs automatically (C) 250
dfsan	Display stacked FIDs automatically without screen erase (C) 251
dfsh	Display stacked FIDs horizontally (C) 251
dfshn	Display stacked FIDs horizontally without screen erase (C) 251
dfsn	Display stacked FIDs without screen erase (C) 252
dfww	Display FIDs in whitewash mode (C) 252
dg	Display group of acquisition/processing parameters (C) 252
dg	Control dg parameter group display (P) 253
dg1	Display group of display parameters (M) 253

dg1	Control dg1 parameter group display (P) 254
dg2	Display group of 3rd and 4th rf channel/3D parameters (M) 254
dg2	Control dg2 parameter group display (P) 254
dga	Display group of spin simulation parameters (M) 255
dgcsteSL	Set up parameters for DgcsteSL pulse sequence (M) 255
dgcstecosy	Set up parameters for Dgcstecosy pulse sequence (M) 255
dgcstehmqc	Set up parameters for Dgcstehmqc pulse sequence (M) 255
dglc	Display group of LC-NMR parameters (M) 256
dglc	Control dglc parameter group display (P) 256
dglp	Display group of linear prediction parameters (C) 256
dgs	Display group of shims and automation parameters (M) 256
dgs	Control dgs parameter group display (P) 257
dhp	Decoupler high-power control with class C amplifier (P) 257
diagth2d	Exclude diagonal peaks when peak picking 257
dialog	Display a dialog box from a macro (C) 258
diffparam	Report differences between parameter sets (UNIX) 258
diffparams	Report differences between two parameter sets (U) 259
diffshims	Compare two sets of shims (M,U) 260
digfilt	Write digitally filtered FIDs to another experiment (M) 260
dir	List files in directory (C) 260
display	Display parameters and their attributes (C) 261
dla	Display spin simulation parameter arrays (M) 261
dlalong	Long display of spin simulation parameter arrays (C) 262
dLC	Display LC detector trace(s) in a horizontal format 262
dlcnmr	Displays all forms of LC-NMR data 262
dli	Display list of integrals (C) 263
dlivast	Produce text file and process wells (M) 263
d11	Display listed line frequencies and intensities (C) 264
dlni	Display list of normalized integrals (M) 265
dlp	Decoupler low-power control with class C amplifier (P) 265
dm	Decoupler mode for first decoupler (P) 265
dm2	Decoupler mode for second decoupler (P) 266
dm3	Decoupler mode for third decoupler (P) 267

dm4	Decoupler mode for fourth decoupler (P) 267
dmf	Decoupler modulation frequency for first decoupler (P) 267
dmf2	Decoupler modulation frequency for second decoupler (P) 268
dmf3	Decoupler modulation frequency for third decoupler (P) 268
dmf4	Decoupler modulation frequency for fourth decoupler (P) 269
dmfadj	Adjusts the parameter 'dmf' 269
dmf2adj	Adjust tip-angle resolution time for second decoupler (M) 270
dmf3adj	Adjust tip-angle resolution time for third decoupler (M) 270
dmf4adj	Adjust tip-angle resolution time for fourth decoupler (M) 270
dmg	Data display mode in directly detected dimension (P) 271
dmg1	Data display mode in 1st indirectly detected dimension (P) 272
dmg2	Data display mode in 2nd indirectly detected dimension (P) 272
dmgf	Absolute-value display of FID data or spectrum in acqi (P) 273
dmm	Decoupler modulation mode for first decoupler (P) 273
dmm2	Decoupler modulation mode for second decoupler (P) 274
dmm3	Decoupler modulation mode for third decoupler (P) 274
dmm4	Decoupler modulation mode for fourth decoupler (P) 275
dn	Nucleus for first decoupler (P) 275
dn2	Nucleus for second decoupler (P) 276
dn3	Nucleus for third decoupler (P) 276
dn4	Nucleus for fourth decoupler (P) 276
dndfid	Retrieve and process fid data from the locator (M) 277
dndjoin	Join a work space from the locator (M) 277
dndpar	Retrieve a parameter set from the locator (M) 277
dndshims	Retrieve a shimset set from the locator (M) 278
doautodialog	אָלאָד a dialog window using def file (M) 278
dodialog	Start a dialog window with dialoglib file (M) 278
dof	Frequency offset for first decoupler (P) 278
dof2	Frequency offset for second decoupler (P) 279
dof3	Frequency offset for third decoupler (P) 279

dof4	Frequency offset for fourth decoupler (P) 279
doneshot	Set up parameters for Doneshot pulse sequence (M) 280
dopardialog.	Start a dialog with dialoglib/experiment def file (M) 280
do_pcss	Calculate proton chemical shifts spectrum (C) 280
dosy	Process DOSY experiments (M) 281
dosy2d	Apptype macro for dosy 2D experiments (M) 281
dosy3Dflag	Used by the dosy macro to determine whether to use 2D or 3D DOSY processing $$282$$
dosy3Dproc	Used by the dosy macro to determine whether to use 2D or 3D processing 282
dosybypoints	Determines whether peak picking is used by the dosy macro 282
dosyfit	fits 2D or 3D DOSY data to obtain diffusion coefficients, amplitudes and statistics 283
dosyfrq	Larmor frequency of phase encoded nucleus in DOSY (P) 283
dosygamma	Gyromagnetic constant of phase encoded nucleus in DOSY (P) $$283$$
dosypeaks	Determines whether peak picking is used by the dosy macro 283
dosyproc	Determines the type of processing performed by the dosy macro 284
dosytimecube	aGyromagnetic constant of phase encoded nucleus in DOSY (P) 284
dot1	Set up a <i>T</i> ₁ experiment (M) 284
dotflag	Display FID as connected dots (P) 285
dousermacro.	Mechanism to provide customization to VnmrJ operations 285
downsamp	Downsampling factor applied after digital filtering (P) 286
dp	Double precision (P) 287
dpcon	Display plotted contours (C) 287
dpconn	Display plotted contours without screen erase (C) 288
dpf	Display peak frequencies over spectrum (C) 288
dpir	Display integral amplitudes below spectrum (C) 289
dpirn	Display normalized integral amplitudes below spectrum (M) 289
dpiv	Display integral values below spectrum (M) 290
dpivn	Display normalized integral values below spectrum (M) 290
dp1	Default plot (M) 291

dp1_seqfil	Sequence-specific default plot (M) 291
dplane	. Display a 3D plane (M) 291
dpr	. Default process (M) 292
dpr_seqfil	. Sequence-specific default process (M) 292
dprofile	Display pulse excitation profile (M) 292
dproj	Display a 3D plane projection (M) 293
dps	Display pulse sequence (C) 294
dpwr	Power level for first decoupler with linear amplifier (P) 294
dpwr2	Power level for second decoupler with linear amplifier (P) 295
dpwr3	Power level for third decoupler with linear amplifier (P) 296
dpwr4	Power level for fourth decoupler amplifier (P) 296
dpwrf	First decoupler fine power (P) 297
dpwrf2	Second decoupler fine power (P) 297
dpwrf3	Third decoupler fine power (P) 298
dpwrm	First decoupler linear modulator power (P) 298
dpwrm2	Second decoupler linear modulator power (P) 298
dpwrm3	Third decoupler linear modulator power (P) 299
Dqcosy	Convert the parameter to a DQCOSY experiment (M) 299
draw	Draw line from current location to another location (C) 299
dres	Measure linewidth and digital resolution (C) 300
dres	. Tip-angle resolution for first decoupler (P) 300
dres2	. Tip-angle resolution for second decoupler (P) 301
dres3	Tip-angle resolution for third decoupler (P) 301
dres4	Tip-angle resolution for fourth decoupler (P) 302
ds	Display a spectrum (C) 302
ds2d	Display 2D spectra in whitewash mode (C) 304
ds2dn	Display 2D spectra in whitewash mode without screen erase (C) 304
dscale	Display scale below spectrum or FID (C) 305
dsnarray	Report statistical signal-to-noise for Cold Probes (M) 305
dscoef	Digital filter coefficients for downsampling (P) 306
dseq	Decoupler sequence for first decoupler (P) 306
dseq2	Decoupler sequence for second decoupler (P) 306
dseq3	Decoupler sequence for third decoupler (P) 307
dseq4	Decoupler sequence for fourth decoupler (P) 307
dsfb	Digital filter bandwidth for downsampling (P) 307

dshape	Display pulse shape or modulation pattern (M) 308
dshapef	Display last generated pulse shape (M) 308
dshapei	Display pulse shape or modulation pattern interactively (M) 308
dshim	. Display a shim "method" string (M) 309
dslsfrq	. Bandpass filter offset for downsampling (P) 310
dsn	. Measure signal-to-noise (C) 310
dsnmax	. Calculate maximum signal-to-noise (M) 311
dsp	Display calculated spectrum (C) 311
dsp	. Type of DSP for data acquisition (P) 312
dsplanes	Display a series of 3D planes (M) 313
dsptype	. Type of DSP (P) 314
dss	Display stacked spectra (C) 314
dssa	Display stacked spectra automatically (C) 316
dssan	Display stacked spectra automatically without erasing (C) 318
dssh	Display stacked spectra horizontally (C) 318
dsshn	Display stacked spectra horizontally without erasing (C) 320
dssl	Label a display of stacked spectra (M) 320
dssn	Display stacked spectra without screen erase (C) 321
dsvast	Display VAST Data in a stacked 1D-NMR matrix format 322
dsvast2d	Display VAST Data in a pseudo-2D format 322
dsww	. Display spectra in whitewash mode (C) 323
dtext	Display a text file in graphics window (M) 323
dtrig	Delay to wait for another trigger or acquire a spectrum (P) 324
dutyc	Duty cycle for homodecoupling (optional) (P) 324
e	. Eject sample (macro) 328

e Eject sample (macro) 328
ecc_on Turns on eddy current compensation for Cold Probes (M) 328
ecc_off Turns off eddy current compensation for Cold Probes (M) 328
echo Simple echo command similar to unix echo 329
edit Edit file or a macro with user-selectable editor 329
editht Create and edit a Hadamard frequency list. 329
editLog Customize the log details 332

6 E

eject Eject sample (M) 334 email Tool to Send Email 335 335 **enter** Enter sample information for automation run (M,U) **enterdialog**. Start a dialog window using enterexp file (M) 336 336 epage Emails Output eplot Emails PostScript 336 ernst Calculate the ernst angle 336 errlog...... Display recent Vnmr error messages 336 errloglen Number of lines in error message display (P) 337 exec..... Execute a VNMR command 337 **execpars**......... Set up the exec parameters (M) 338 338 **execplot**...... Execute plotting macro (P) **execprep**...... Execute prepare macro (P) 339 339 **execprescan**. Execute prescan macro (P) **execproc**...... Execute processing macro (P) 339 339 **execprocess**. Execute processing macro (P) 339 **execsetup** Execute setup macro (P) exists...... Checks if parameter, file, or macro exists and file type (C) 340 exit...... Macro to call vnmrexit 342 expactive Determine if the experiment has an active acquisition 342 expfit...... Unix program for making a least squares fit to a polynomial or exponential curve. 344 exp1..... Display data on the screen 345 expladd Add another diffusion analysis to current display (M) 346 explib..... Display experiment library (M) 347 explist Display current experiment chain and approx. time for each (M) 347 explog...... Display an experiment's log file 347 exptime Display experiment time 347

7 F

f	Set display parameters to full spectrum	352	
£19	Automated fluorine acquisition (M)	352	
f19p	Process 1D fluorine spectra (M) 352		
f1coef	Coefficient to construct F1 interferogram	n (P)	353
f2coef	Coefficient to construct F2 interferogram	n (P)	353
fastuserlogi	n Gateway macro for fastuserlogin fun (M) 354	ction	

fattn Fine attenuator (P) 354	
£ъ Filter bandwidth (P) 354	
fbc Applies 'bc' type baseline correction to all the spectra in array 355	n an
fdm1 Set, write 1D FDM parameters, run FDM (M) 355	
fid_scan Start up the interactive acquisition display process 3	56
fiddc3d Flag for 3D time-domain DC correction 357	
fiddle Perform reference deconvolution 357	
fiddle_examplesIllustrates some of the simple ways that fiddle can used to extract worthwhile results from poor quality data 360	be
fiddled Perform reference deconvolution subtracting alternate (C) 361	FIDs
fiddleu Perform reference deconvolution subtracting successiv FIDs (C) 361	е
fiddle2a Perform 2D reference deconvolution (C) 361	
fiddle2D Perform 2D reference deconvolution (C) 361	
fiddle2Dd 2D reference deconvolution subtracting alternate FIDs (C) 362	
fidmax Find the maximum point in an FID 362	
fidpar Add parameters for FID display in the current experiment 362	
fidsave Save data (M) 363	
fifolpsize FIFO loop size (P) 363	
file File name of parameter set (P) 363	
files Interactively handle files (C) 363	
filesinfo Return file information for files display (C) 364	
filtfile File of FIR digital filter coefficients (P) 364	
findxmlmenu. Find an xml menu (M) 365	
fitspec Spectrum deconvolution 365	
fixgrd Convert gauss/cm value to DAC (M) 366	
fixpar Correct parameter characteristics in experiment (M)	366
fixpar3rf Create parameters for third rf channel (M) 367	
fixpar4rf Create parameters for fourth rf channel (M) 367	
fixpar5rf Create parameters for fifth rf channel (M) 367	
fixgrdR Converts Gradient Strength to DAC values 368	
fixup Adjust parameter values selected by setup macros (M) 368	
fixpsgUpdate psg libraries (M) 368	
flashc Convert compressed 2D data to standard 2D format	368

flipflop	Set up parameters for FLIPFLOP pulse sequence (M) 370
Fluorine	Set up parameters for 19F experiment (M) 370
flush	Write out data in VNMR memory 371
fn	Fourier number in directly detected dimension (P) 371
fn1	Fourier number in 1st indirectly detected dimension (P) 371
fn2	Fourier number in 2nd indirectly detected dimension (P) 372
fn2D	Fourier number to build up 2D DOSY display in freq. domain (P) $$372$$
focus	Send keyboard focus to input window (C) 372
foldcc	Fold INADEQUATE data about 2-quantum axis 372
foldj	Fold J-resolved 2D spectrum about the F1=0 axis 373
foldt	Fold COSY-like spectrum along diagonal axis 373
fontselect	Open FontSelect window (C) 373
format	374
fp	Find peak heights or phases (C) 375
fpi	Report integral values from arrayed spectra. 375
fpmult	First point multiplier for np FID data (P) 376
fpmult	First point multiplier for "np" FID data 377
fpmult1	First point multiplier for "ni" interferogram data 378
fpmult2	First point multiplier for "ni2" interferogram data 378
fr	Recall all display parameters from set #n, n=19 (n) 378
framecmd	Create a new frame 378
fread	Read in variables from a file and load them in a tree 379
fsave	Save parameters from a tree to a file (C) 380
fsq	Frequency-shifted quadrature detection (P) 380
ft	Fourier transform 1D data (C) 381
ft1d	Fourier transform along f ₂ dimension (C) 386
wft1d(coeffi	cients) Weight and Fourier transform F2 of 2D data 388
ft1da	Fourier transform phase-sensitive data (M) 388
ftldac	Combine arrayed 2D FID matrices (M) 388
ftldac and w	vftldac Help file for wft1dc macro used to combine arrayed 2D FID matrices 389
ft2d	Fourier transform 2D data (C) 389
wft2d(coeffi	cients) Weight and Fourier transform 2D data 392
ft2da	Fourier transform phase-sensitive data (M) 395
ft2dac	Combine arrayed 2D FID matrices (M) 396

ft2dac and w	2D FID matrices 397
ft3d	Perform a 3D FT on a 3D FID data set 397
ftargs	Macro to create parameters 399
full	Set display limits for a full screen (C) 399
fullsq	Display largest square 2D display (M) 399
fullt	Set display limits for a full screen with room for traces (C) 400

8 G

g2pul_ecc	Setup macro for eddy current compensation parameters (M) 405
ga	Submit experiment to acquisition and FT the result (M) 405
gain	Receiver gain (P) 406
gap	Find gap in the current spectrum (M) 407
gaussian	Set up unshifted Gaussian window function (M) 407
gcal	Local value of the conversion factor between gradient in DAC points and gradient in G/cm 408
gcal	Gradient calibration constant (P) 408
gcoil	Current gradient coil (P) 408
Gcosy	Convert the parameter to a gradient COSY experiment (M) 409
gdiff	Diffusion gradient level (P) 410
Gdqcosy	Convert the parameter to a gradient DQCOSY experiment (M) 410
get1d	Select a 1D experiment for processing (M) 410
get2d	Select a 2D experiment for processing (M) 411
getdim	Return dimensionality of experiment (M) 411
getemailaddr	e Get email addresses from a file 412
geterror	Return or display an acquisition error 412
getfile	Get information about directories and files (C) 412
getgamma	Retrieves Gamma from /vnmr/nuctabref 413
getht	Retrieve/Save a Hadamard frequency list from a file. 413
getlcdata	An LC-NMR communications macro 415
getlimit	get the limits of a variable in a tree (C) 415
get11	Get intensity and line frequency of line (C) 416
getmodule	Gets module (C) 416
getoffset	Sets offset based on current reference parameters 417
getparam	Retrieve parameter from probe file (M) 417

getplane	Extract planes from a 3D spectral data set (M) 417
getplotterty	קי e The getplottertype command retrieves plotter information. 418
getppm	Returns Cursor Value in ppm 419
getreg	Get frequency limits of a specified region (C) 419
getsampgloba	Loads sample global parameters 420
getshimmetho	as Get proshim methods list (M) 420
getsn	Get signal-to-noise estimate of a spectrum (M) 420
gettoken	Utility macro to separate a string into tokens (M) 421
gettxt	Get text file from VnmrJ data file (C) 421
gettype	Get the type of a variable (C) 422
getvalue	Get value of parameter in a tree (C) 422
gf	Prepare parameters for FID/spectrum display in acqi (M) 423
gf	Gaussian function in directly detected dimension (P) 424
gf1	Gaussian function in 1st indirectly detected dimension (P) 424
gf2	Gaussian function in 2nd indirectly detected dimension (P) 424
gflow	Flow encoding gradient level (P) 425
gfs	Gaussian shift const. in directly detected dimension (P) 425
gfs1	Gaussian shift const. in 1st indirectly detected dimension (P) 425
gfs2	Gaussian shift const. in 2nd indirectly detected dimension (P) 425
Ghmbc	Convert the parameter to a gradient HMBC experiment (M) 426
ghmqc	Set up a PFG HMQC pulse sequence (M) 426
Ghmqc	Convert the parameter to a gradient HMQC experiment (M) 426
gHMQC15	Set up parameters for ¹⁵ N gHMQC experiment (M) 426
gHMQC_d2	. Set up parameters for ^{15}N gHMQC experiment using dec. 2 (M) $$426$$
gHMQC_d213	Set up parameters for ${}^{13}C$ gHMQC experiment using dec. 2 (M) 427
ghmqcps	Set up a PFG HMQC phase-sensitive pulse sequence (M) 427
ghsqc	Set up a PFG HSQC pulse sequence (M) 427
Ghsqc	Convert the parameter to a gradient HSQC experiment (M) 427

gHSQC15	Set up parameters for ¹⁵ N gHSQC experiment (M) 427
gHSQC_d2	Set up parameters for 15 N gHSQC experiment using dec. 2 (M) 428
gHSQC_d213	Set up parameters for ¹³ C gHSQC experiment using dec. 2 (M) 428
Ghsqctoxy	Convert parameters for gradient HSQCTOXY experiment (M) 428
gilson	Open the Gilson Liquid Handler window (C) 428
gilson	Allow starting the Gilson Liquid Handler GUI 428
gin	Return current mouse position and button values (C) 429
globalauto	Automation directory name (P) 430
glue	Create a pseudo-2D dataset (M) 430
gmapshim	Start gradient autoshimming (M) 430
gmapshim_au.	Start acquisition with gradient shimming (M) 431
gmapspin	Enable or disable spinning during gradient shimming (P) 431
gmapsys	Run gradient autoshimming, set parameters, map shims (M) 431
gmapz	Get parameters and files for gmapz pulse sequence (M) 432
gmap_findtof	Gradient shimming flag to first find tof (P) 433
gmap_z1z4	Gradient shimming flag to first shim z1-z4 (P) 433
gmax	Maximum gradient strength (P) 434
gmqcosy	Set up PFG absolute-value MQF COSY parameter set (M) 434
gnoesy	Set up a PFG NOESY parameter set (M) 434
go_ <pslabel></pslabel>	Experiment-Specific Runtime Macro 434
go	Submit experiment to acquisition (M) 434
go	Pulse sequence setup macro called by go, ga, and au (M) 436
gpat-gpat3	Gradient shape (P) 437
gplan	Start interactive image planning (C) 437
х	Multiplier for gradient pulses on alternating scans (P) 437
gradfit	calculates fit coefficients describing the variation of gradient strength with position in calibration of non-uniform pulsed field gradients 437
gradientdisa	ызе Disable PFG gradients (P) 438
gradientsha <u>r</u>	pingActivate shaping on the gradient pulses (P) 438
gradstepsz	Gradient step size (P) 439
gradtype	Gradients for X, Y, and Z axes (P) 439

graphis	Return the current graphics display status (C) 439
grayctr	Gray level window adjustment (P) 440
grays1	Gray level slope (contrast) adjustment (P) 440
grecovery	Eddy current testing (M) 441
grid	Draw a grid on a 2D display (M) 441
groupcopy	Copy parameters of group from one tree to another (C) 442
gspoil	Spoiler gradient level (P) 442
gsspat	Slice-select gradient shape (P) 442
gtnnoesy	Set up a PFG TNNOESY parameter set (M) 442
gtnroesy	Set up a PFG absolute-value ROESY parameter set (M) 443
gtotlimit	Gradient total limit (P) 443
gtrim	Trim gradient level (P) 443
gxmax,gymax,	gzmaxMaximum gradient strength for each axis (P) 443
gzlvl	Pulsed field gradient strength (P) 444
gzsize	Number of z-axis shims used by gradient shimming (P) 444
gzwin	Spectral width percentage used for gradient shimming (P) 444

9 H

h1	Automated proton acquisition (M) 450
hlfreq	Proton frequency of spectrometer (P) 450
h1p	Process 1D proton spectra (M) 450
h2cal	Calculate strength of the decoupler field (C) 451
halt	Abort acquisition with no error (C) 452
hc	Automated proton and carbon acquisition (M) 452
hcapt	Automated proton, carbon, and APT acquisition (M) 453
hcchtocsy	Set up parameters for HCCHTOCSY pulse sequence (M) 453
hccorr	Automated proton, carbon, and HETCOR acquisition (M) 453
hcdept	Automated proton, carbon, and DEPT acquisition (M) 454
hcosy	Automated proton and COSY acquisition (M) 454
hdmf	Modulation frequency for homonuclear decoupling (P) 455
hcmult	Execute protocol actions of apptype hcmult (M) 455
hdof	Frequency offset for homodecoupling (P) 455
hdpwr	Power level for homodecoupling (P) 456

457 hdres Sets the tip angle resolution (P) 457 hdseq Waveform filename for band selective decoupling (P) 458 hdwshim...... Hardware shimming (P) 459 hdwshimlist. List of shims for hardware shimming (P) 459 help..... Display current help file 459 **HELP**...... Help File for this Tool 460 460 **HETCOR**...... Change parameters for HETCOR experiment (M) 460 460 hetcorcp1 Set up parameters for solids HETCOR pulse sequence (M) 461 hetcorps....... Set up parameters for HETCORPS pulse sequence (M) 461 hetero2d...... Execute protocol actions of apptype hetero2d (M) 461 hidecommand. Execute macro instead of command with same name 461 (C) **hipwrampenable** High Power Amplifier Enable (P) 462 Hmbc...... Convert the parameter to a HMBC experiment (M) 462 462 463 **HMQC** d2 Set up parameters for ¹⁵N HMQC experiment using dec. 2 (M) 463 **HMQC d213** Set up parameters for ¹³C HMQC experiment using dec. 2 (M) 463 463 Hmgctoxy....... Convert the parameter to a HMQCTOXY experiment 463 (M) **HMOCTOXY15**.... Set up parameters for ¹⁵N HMQCTOXY experiment 463 (M) **HMOCTOXY d2**. Set up parameters for ¹⁵N HMQCTOXY using decoupler 2 (M) 464 HMQCTOXY_d213Set up parameters for ¹³C HMQCTOXY using decoupler 2 (M) 464 hmgctoxy3d.... Set up parameters for HMQC-TOCSY 3D pulse sequence (M) 464 **ho**..... Horizontal offset (P) 464 hom2dj Set up parameters for HOM2DJ pulse sequence (M) 464 homo...... Homodecoupling control for the observe channel (P) 464 **HOMODEC** Change parameters for HOMODEC experiment (M) 465 **homo2d**..... Execute protocol actions of apptype homo2d (M) 465

homorof1	Delay before turning on homo decoupling rf (P) 466
homorof2	Delay after blanking the amp and setting T/R switch to recv (P) $-\!$
homorof3	Delay between setting T/R to receive and gating the recvr on (P) 467
hoult	Set parameters alfa and rof2 according to Hoult (M) 467
hpa	Plot parameters on special preprinted chart paper (C) 468
Hprescan	Proton prescan (P)) 468
hregions	Select integral regions in proton spectrum (M) 468
hs	Homospoil pulses (P) 468
Hsqc	Convert the parameter to a HSQC experiment (M) 469
HSQC15	Set up parameters for ¹⁵ N HSQC experiment (M) 469
HSQC_d2	Set up parameters for ¹⁵ N HSQC experiment using dec. 2 (M) 469
HSQC_d213	Set up parameters for ¹³ C HSQC experiment using dec. 2 (M) 469
hsqcHT	Set up the hsqcHT experiment (M) 469
Hsqctoxy	Convert parameters to a HSQCTOXY experiment (M) 470
нѕостоху15	Set up parameters for ¹⁵ N HSQCTOXY experiment (M) 470
HSQCTOXY_d2	Set up parameters for ¹⁵ N HSQCTOXY using decoupler 2 (M) 470
HSQCTOXY_d21	L3Set up parameters for ¹³ C HSQCTOXY using decoupler 2 (M) 470
hsqctoxySE	Set up parameters for HSQC-TOCSY 3D pulse sequence (M) 470
hsrotor	Display rotor speed for solids operation (P) 471
hst	Homospoil time (P) 471
ht	Setting up and processing Hadamard experiments. 471
htbitrev	Hadamard bit reversal flag (P) 473
htbw1	Hadamard pulse excitation bandwidth in ni (P) 473
htcal1	. RF calibration flag for Hadamard waveforms in ni (P) 473
htfrq1	Hadamard frequency list in ni (P) 474
htfrqdisp	Read, write, and display Hadamard frequencies. 474
htofs1	Hadamard offset in ni (P) 475
htpwr1	Power level for RF calibration of Hadamard waveforms in ni (P) 475
htss1	Stepsize for Hadamard waveforms in ni (P) 475
hzmm	Scaling factor for plots (P) 476
hztomm	Convert locations from Hz or ppm to plotter units (C) 476

iInsert sample (M) 480
ihwinfo Hardware status of console (U) 480
i1Interleave arrayed and 2D experiments (P) 481
ilfid Interleave FIDs during data processing (C) 481
<pre>imagefile Display an image file (M) 482</pre>
imagemath Fit images to an specified function (M) 483
<pre>imageprint Plot non interactive gray scale image (M) 484</pre>
imconi Display 2D data in interactive grayscale mode (M) 484
<pre>import1DspecImport ASCII Spectrum into VnmrJ / VNMR (M) 484</pre>
import1Dspec Create phasefile and data from ASCII spectrum (U) 485
in Lock and spin interlock (P) 486
<pre>inadgtSet up parameters for INADEQUATE pulse sequence</pre>
index2 Projection or 3D plane index selected (P) 487
inept Set up parameters for INEPT pulse sequence (M) 488
<pre>initialize_iterateSet iterate string to contain relevant parameters</pre>
input Receive input from keyboard (C) 488
ins Integral normalization scale (P) 489
ins22D volume value (P) 489
insref Fourier number scaled value of an integral (P) 489
<pre>ins2ref Fourier number scaled volume of a peak (P) 490</pre>
insertInsert sample (M) 490
inset Display an inset spectrum (C) 490
integ Find largest integral in a specified region (C) 490
integrate Automatically integrate 1D spectrum (M) 491
int_flg 491
intmod Integral display mode (P) 492
intvast Produce a text file of integral regions (M) 492
intvast Produce a text file containing the integral of the partial regions 492
ioIntegral offset (P) 493
isIntegral scale (P) 493
isadj Automatic integral scale adjustment (M) 493
isadj2Automatic integral scale adjustment by powers of two (M) 494
isCSIMode Determine if graphics area is split for CSI mode 494
isiin System global parameter for ISI interlock 495

<pre>isrea1Utility macr</pre>	o to determine a pa	arameter type (M)	495
isstring Utility macr	o to determine a pa	arameter type (M)	496
isvnmrj Identifies th	ne interface is use,	either Vnmr or Vnmr	J 496
iterate Parameters	to be iterated (P)	497	

11 J

jaddsub	Join the add/subtract experiment 499
jcurwin	Work space numbers of all viewports (P) 500
jdesignS	Start Plot Designer Program (M) 501
jexp	Join existing experiment (C) 501
jexp1-jexp999 (99 Join existing experiment and display new parameters M) 502
jexpn	Join experiment n, where n is a number between 1 and 502
jnewexp (Creates and Joins a New Experiment 502
jplot l	Plot from Plot Designer program (C) 503
jplotscaleS	Scale plot parameters (M) 503
jplotunscale	Restore current experiment parameters (M) 503
jprint	Prints the selected images to a printer or file (M) 504
jpublish	Macro to archive and/or copy to system a local protocol M) 504
jumpretS	Set up parameters for JUMPRET pulse sequence (M) 504
jviewport	Work space numbers of the current viewports (P) 504
jviewportlabe	1 Work space labels for all viewport buttons (P) 505
jviewports	Viewport layout (P) 505
jwinA	Activate and record activity in current window (M) 505

12 K

killft3d	. Terminate any ft3d process started in an experiment (M,U) 507	
killplot	. Stop plot jobs and remove from plot queue (M) 50	8
killprint	. Stop print jobs and remove from print queue (M)	508
kind	Kinetics analysis, decreasing intensity (M) 509	
kinds	Kinetics analysis, decreasing intensity, short form (M) 509	
kini	Kinetics analysis, increasing intensity (M) 509	
kinis	Kinetics analysis, increasing intensity, short form (M) 510	

13 L

laser SVS adiabatic localization 515

lastlk	Last lock solvent used (P) 515
lastmenu	Menu to display when Return button is selected (P) 515
latch	Frequency synthesizer latching (P) 516
1b	Line broadening in directly detected dimension (P) 516
1b1	Line broadening in 1st indirectly detected dimension (P) 517
1b2	Line broadening in 2nd indirectly detected dimension (P) 517
lc1d	Pulse sequence for LC-NMR (M) 517
lcdatast	An LC-NMR plotting and display macro 518
lcpar2d	Create 2D LC-NMR acquisition parameters (M) 518
lcpeak	Peak number (P) 518
lcplot	Plot LC-NMR data (M) 519
lcpsgset	Set up parameters for various LC-NMR pulse sequences (M) 519
lcset2d	General setup for 2D LC-NMR experiments (M) 519
left	Set display limits to left half of screen (C) 520
legrelay	Independent control of magnet leg relay (P) 520
length	Determine length of a string (C) 520
1f	List files in directory (C) 521
lgcp	X Lee-Goldburg cross polarization (CP) between protons and X with a choice of SPINAL or TPPM decoupling 521
liamp	Amplitudes of integral reset points (P) 524
lifrq	Frequencies of integral reset points (P) 524
liMMap	Calculate csi map of integrals for a specified peak (C) 524
liqbear	Liquids Bearing Air Level (P) 525
listenoff	Disable receipt of messages from send2Vnmr (M) 525
listenon	Enable receipt of messages from send2Vnmr (M) 525
listparam	List parameters in simple format (UNIX) 526
lkof	Track changes in lock frequency (P) 527
112d	Automatic and interactive 2D peak picking (C) 527
112dbackup	Copy current II2d peak file to another file (M) 530
112dmode	Control display of peaks picked by II2d (P) 530
11amp	List of line amplitudes (P) 531
11frg	List of line frequencies (P) 531
11MMap	Calculate csi map of peak height for a peak defined by cs (C) 531
ln	Find natural logarithm of a number (C) 532
load	Load status of displayed shims (P) 532
loadcolors	Load colors for graphics window and plotters (M) 533

loaduserprefsLoad Operator Preferences 533 loc Location of sample in tray (P) 534 locaction Locator action (M) 534 534 535 **lockacgtc** Lock loop time constant during acquisition (P) **lockfreg**...... Lock frequency (P) 535 lockgain..... Lock gain (P) 536 lockphase Lock phase (P) 536 lockpower Lock power (P) 537 locktc..... Lock time constant (P) 537 log 537 logate..... Transmitter local oscillator gate (P) 538 **lookup**..... Look up words and lines from a text file (C) 538 **locprotoexec** Execute a protocol from the locator (M) 546 1p..... First-order phase in directly detected dimension (P) 546 1p1 First-order phase in 1st indirectly detected dimension (P) 546 1p2 First-order phase in 2nd indirectly detected dimension (P) 547 **1pa1g** LP algorithm in np dimension (P) 547 lpalg1..... LP algorithm in ni dimension (P) 548 **1pa1g2**..... LP algorithm in ni2 dimension (P) 548 **1pext** LP data extension in np dimension (P) 549 **1pext1**..... LP data extension in ni dimension (P) 549 1pext2..... LP data extension in ni2 dimension (P) 549 1pfilt..... LP coefficients to calculate in np dimension (P) 550 lpfilt1 LP coefficients to calculate in ni dimension (P) 550 1pfilt2 LP coefficients to calculate in ni2 dimension (P) 550 **1pnupts** LP number of data points in np dimension (P) 551 **1pnupts1**...... LP number of data points in ni dimension (P) 551 1pnupts2...... LP number of data points in ni2 dimension (P) 551 **1popt** LP algorithm data extension in np dimension (P) 552 **1popt1**...... LP algorithm data extension in ni dimension (P) 552 1popt2..... LP algorithm data extension in ni2 dimension (P) 553 **1pprint** LP print output for np dimension (P) 553 **1pprint1**...... LP print output for ni dimension (P) 554 554 **1pprint2**...... LP print output for ni2 dimension (P) **1ptrace** LP output spectrum in np dimension (P) 555 **1ptrace1**...... LP output spectrum in ni dimension (P) 555

lptrace2	_P output spectrum in ni2 dimension (P) 555	
ls	_ist files in directory (C) 556	
lsfid	Number of complex points to left-shift the np FID (P) 55	6
lsfid1	Number of complex points to left-shift ni interferogram P) 557	
lsfid2	Number of complex points to left-shift ni2 interferogram P) 557	
lsfrq	Frequency shift of the fn spectrum (P) 558	
lsfrq1	Frequency shift of the fn1 spectrum (P) 558	
lsfrq2	Frequency shift of the fn2 spectrum (P) 559	
lvl	Zero-order baseline correction (P) 559	
lvltlt	Control sensitivity of IvI and tlt adjustments (P) 560	

14 M

macro Macro name (P) 563
macrocat Display a user macro file in text window (C) 563
macrocp Copy a user macro file (C) 564
macrodir List user macro files (C) 564
macroedit Edit a macro with user-selectable editor (M) 564
macrold Load a macro into memory (C) 565
macrorm
macrosyscat . Display a system macro file in text window (C) 566
macrosyscp Copy a system macro to become a user macro (C) 566
macrosysdir. List system macros (C) 567
macrosysrm Remove a system macro (C) 567
macrovi Edit a user macro with the vi text editor (M) 567
make3dcoef Make a 3D coefficients file from 2D coefficients (M) 568
makedosyparamsCreate parameters for DOSY processing (M) 569
makefid Make a FID element using numeric text input (C) 570
makeeccglobals Create global parameters for ECC control (M) 571
makeslice Synthesize 2D projection of 3D DOSY experiment (C) 571
makeStudy Create and manage Study Clones. 571
makeuser Add a new Vnmr user account or update an existing Vnmr user account (U) 572
makeuserpsg. Compiles the user PSG sources and constructs the user PSG object library 573
man Display online description of command or macro (M) 574
managedb Update user files (U) 574
manualpath Path to user's manual directory (P) 574
manvi Edit online description of a command or macro (M) 575

mapwin	. List of experiment numbers (P) 575
mark	. Determine intensity of spectrum at a point (C) 575
masvt	. Type of variable temperature system (P) 578
maxattench1	 -4 Maximum limit for attenuator setting for rf channel 1-4 (P) 578
maxpen	. Maximum number of pens to use (P) 578
md	. Move display parameters between experiments (C) 579
menu	. Change status of menu system (C) 579
menuvi	. Edit a menu with vi text editor (M) 580
method	. Autoshim method (P) 580
mf	. Move FIDs between experiments (C) 580
mfblk	. Copy FID block (C) 581
mfclose	. Close memory map FID (C) 582
mfdata	. Move FID data (C) 582
mfopen	. Memory map open FID file (C) 583
mftrace	. Move FID trace (C) 584
mht	. Move Hadamard parameters from one workspace to another 585
minsw	. Reduce spectral width to minimum required (M) 586
mkchsums	Make checksum(s) for a given directory or file 586
mkCPprotocol	. Make Protocol 586
mkdir	. Create new directory (C) 587
mlabel	. Menu label (P) 587
move	. Move to an absolute location to start a line (C) 587
movedssw	. Set downsampling parameters for selected spectral region (M) 588
moveossw	. Set oversampling parameters for selected spectral region (M) 588
movesw	. Move spectral window according to cursors (M) 589
movetof	. Move transmitter offset (M) 589
mp	. Move parameters between experiments (C) 590
mparval	. Moves a Paramter Value Between Experiments 590
mqcosy	. Set up parameters for MQCOSY pulse sequence (M) 590
mref	. Set referencing based on a existing spectrum of the sample (M) 590
mrev8	. Set up parameters for MREV8 pulse sequence (M) 592
mrfb	. Set the filter bandwidths for multiple receivers (P) 592
mrgain	. Set the gain for multiple receivers (P) 593
mspec	. Select multiple spectra to display (C) 593

mstat Display memory usage statistics (C) 595	
mstring Menu string (P) 595	
mtune Tune probe using swept-tune graphical display (M)	596
mv Move and/or rename a file (C) 596	
<pre>mvsampglobalMoves sample global parameters 597</pre>	
mxconst Maximum scaling constant (P) 597	
mz Move Integral Reset Points to specified experiment	597

15 N

n1,n2,n3	Name storage for macros (P) 600
ncomp	The number of components to be used in discrete DOSYfitting601
newexp	Create a new VNMR experiment (M) 601
newmenu	Select a menu without immediate activation (C) 601
newshm	Interactively create a shim method with options (M) $\qquad 602$
nextexp	Value of Next Experiment 603
nextlocQ	Next Available Location 603
nextpl	Display the next 3D plane (M) 603
nfni	Number of increments in 1st indirectly detected dimension (P) $$604$$
ni2	Number of increments in 2nd indirectly detected dimension (P) $$604$$
ni3	Number of increments in 3rd indirectly detected dimension (P) $$604$$
niter	Number of iterations (P) 605
nimax	Maximum limit of ni (P) 605
nimax	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605
nimax nl nli	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606
nimax nl nli nlivast	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606
nimax nl nli nlivast nlivast2	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606
nimax nl nli nlivast nlivast2 nlivast3	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606 Produces a text file with normalized integral regions (M) 607
nimax nl nli nlivast nlivast2 nlivast3 nli	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606 Produces a text file with normalized integral regions (M) 607 Find line frequencies and intensities (C) 607
nimax nl nli nlivast nlivast2 nlivast3 nli	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606 Produces a text file with normalized integral regions (M) 607 Find line frequencies and intensities (C) 607 Find normalized integral values 608
nimax nl nli nlivast nlivast2 nlivast3 nli nli	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606 Produces a text file with normalized integral regions (M) 607 Find line frequencies and intensities (C) 607 Find normalized integral values 608 Select normalized intensity mode (C) 608
nimax nl nlivast nlivast 2 nlivast 3 nlivast 3 nli nli nli nm1	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606 Produces a text file with normalized integral regions (M) 607 Find line frequencies and intensities (C) 607 Find normalized integral values 608 Select normalized intensity mode (C) 608 Returns the current transmitter corresponding to the nucleus in argument 1. 608
nimax nl nlivast nlivast 2 nlivast 3 nlivast 3 nli nm nm nm1 nm2d	Maximum limit of ni (P) 605 Position cursor at the nearest line (C) 605 Find integral values (C) 606 Produces a text file of integral regions without a sum region (M) 606 Produces a text file with normalized integral regions (M) 606 Produces a text file with normalized integral regions (M) 607 Find line frequencies and intensities (C) 607 Find normalized integral values 608 Select normalized intensity mode (C) 608 Returns the current transmitter corresponding to the nucleus in argument 1. 608 Select Automatic 2D normalization (M) 608

Noesy1dC (1	convert the parameter set to a Noesy1d experiment M) 609
noiseN	Aleasure noise level of FID (C) 610
noisemultC (1	Control noise multiplier for automatic 2D processing M) 610
noislmL	imit noise in spectrum (M) 611
notebookN	lotebook name (P) 611
np N	Iumber of data points (P) 612
npointN	lumber of points for fp peak search (P) 612
nrecordsD	letermine number of lines in a file (M) 612
nt N	Iumber of transients (P) 613
ntrigN (F	lumber of trigger signals to wait before acquisition P) 613
ntype3dS (f	pecify whether f ₁ or f ₂ display expected to be N-type P) 613
nuctable [([Display VNMR style nucleus table for a given H1 frequency M) 614
nugcalA	parameter array containing calibration information from alibration of non-uniform field gradients 614
nugcalA	parameter array containing calibration information from alibration of non-uniform field gradients 614
nugcalib T s tł g	he nugcalib macro calculates the probe/pulse sequence pecific coefficients from an experiment designed to map he non-uniformity (NUG) of the pulsed field radients. 615
nugflag To N	ells the macro dosy to use processing with correction for on-uniform field gradients 616
numrcvrsN	lumber of receivers in the system (P) 616
numregR	leturn the number of regions in a spectrum (C) 617
numrfchN	lumber of rf channels (P) 617

16 0

: Make a parameter inactive (C) 619
Make a parameter active or test its state (C) 620
ancel Specify special functions and labels for the "Cancel" Command" button 621
erator Operator name (P) 623
ratorloginSets workspace and parameters for the operator (M) 623
: Open shape definition file for Pbox (M) 623
:oefDigital filter coefficients for over sampling (P) 624
ته Digital filter bandwidth for oversampling (P) 624

osfilt	Oversampling filter for real-time DSP (P) 62	25
oslsfrq	Bandpass filter offset for oversampling	(P)	625
overrange	Frequency synthesizer overrange (P)	626	
oversamp	Oversampling factor for acquisition (P)	626	;
owner	Operating system account owner (P)	628	

17 P

p1 Enter pu	Ilse width for p1 in degrees (C) 637
p1 First pul	se width (P) 638
p1pat Shape o	f excitation pulse (P) 638
p2pu1 Set up s	equence for PFG testing (M) 638
p31 Automa	ted phosphorus acquisition (M) 639
p31p Process	1D phosphorus spectra (M) 639
ра Set phas (C) 64	se angle mode in directly detected dimension 40
pa1 Set phas (C) 64	se angle mode in 1st indirectly detected dimension 40
pacosy Plot aut	omatic COSY analysis (C) 641
pad Preacqu	isition delay (P) 641
padept Perform	adept analysis and plot resulting spectra (C) 642
pageSubmit	plot and change plotter page (C) 642
pageName o	f page (P) 643
<pre>panellevelDisplay</pre>	level for VnmrJ interface pages (P) 643
pap Plot out	"all" parameters (C) 644
par2dCreate 2 (M) 6	D acquisition, processing, and display parameters
par3dCreate 3 (M) 6	D acquisition, processing, and display parameters
par3rfGet disp (M) 6	lay templates for 3rd rf channel parameters 45
par4d Create 4	D acquisition parameters (M) 646
paramedit Editapa (C) 64	arameter and its attributes with user-selected editor 46
paramgroup Create a optional paramet	a set of new parameters in a workspace and ly add a display string to the dg and ap ters. 647
paramvi Edit a pa	arameter and its attributes with vi editor (M) 650
pardiff Report of	lifferences between parameter sets (M) 650
pards Create a	The table of t

parfidss	Create parameters for time-domain solvent subtraction (M) 652
parfix	Update parameter sets (M) 653
parlc	Create parameters for LC-NMR experiments (M) 653
parlist	List complete parameters in simple format (M) 654
par112d	Create parameters for 2D peak picking (M) 654
parlp	Create parameters for linear prediction (M) 655
parmax	Parameter maximum values (P) 655
parmin	Parameter minimum values (P) 656
paros	Create additional parameters used by oversampling (M) 656
parside	Sets Up Parameters for Plotting Reference on Side 656
parstep	Parameter step size values (P) 657
partop	Sets Up Parameters for Plotting Reference on Top 657
parversion	Version of parameter set (P) 657
patchinstall	Install a VnmrJ patch 658
patchmake	Build a custom Vnmr patch 659
patchuninsta	11 Uninstall a VnmrJ patch 660
path3d	Path to currently displayed 2D planes from a 3D data set (P) 660
paxis	Plot horizontal LC axis (M) 661
РЬож	Pulse shaping software (U) 661
pbox_bw	Define excitation band (M) 663
pbox_bws	Define excitation band for solvent suppression (notch) pulses (M) 663
pbox_dmf	Extract dmf value from pbox.cal or Pbox shape file (M) 664
pbox_dres	Extract dres value from pbox.cal or Pbox shape file (M) 664
pbox_name	Extract name of last shape generated by Pbox from pbox.cal (M) 664
pbox_pw	Extract pulse length from pbox.cal or Pbox shape file (M) 665
pbox_pwr	Extract power level from Pbox.cal or Pbox shape file (M) 665
pbox_pwrf	Extract fine power level from pbox.cal or Pbox shape file (M) 665
pbox_rst	Reset temporary Pbox/Vnmr variables (M) 666
pbox_shapeir	foReturns Pbox Shape Information 666
pboxget	Extract Pbox calibration data (M) 666
pboxget Extract Pbox calibration data from pbox.cal or Pbox shapefile (M) 667	
--	-----
pboxpar	
pboxrst	
pboxunits Converts to Pbox default units (M) 668	
pcmapapply Apply Phase Correction Map to Data (C) 668	
pcmapgen Generate Phase Correction Map (C) 669	
pcmapclose Phase Correction Map Close (C) 670	
pcon Plot contours on a plotter (C) 671	
pcssCalculate and show proton chemical shifts spectrum (M) 672	
peak Find tallest peak in specified region (C) 672	
peak2đ Return information about maximum in 2D data (C) 67	3
peakmin Find the minimum point 673	
pen Select a pen or color for drawing (C) 674	
pexp1 Plot exponential or polynomial curves (C) 674	
pexpladd Add another diffusion analysis to current plot (M) 675	ō
pfgon Pulsed field gradient amplifiers on/off control (P) 676	\$
pfwwPlot FIDs in whitewash mode (C) 676	
pge Convert parameter set to PGE pulse sequence (M) 67	7
pge_calib Calibrate gradient strengths for PGE pulse sequence (M) 677	
pge_data Extract data from single element of PGE pulse sequence (M) 677	;
pge_output Output results from PGE pulse sequence (M) 678	
pge_process . Automated processing of data from PGE pulse sequence (M) 678	3
$pge_results$. Calculate diffusion constant for integral region (M) 63	78
pge_setup Set up gradient control parameters for PGE pulse seque (M) 679	nce
phSet phased mode in directly detected dimension (C)	579
ph1 Set phased mode in 1st indirectly detected dimension (C) 680	
בתק Set phased mode in 2nd indirectly detected dimension (C) 681	
phase Change frequency-independent phase rp (M) 681	
phase Phase selection (P) 682	
phase1 Phase of first pulse (P) 682	
phase2 Phase selection for 3D acquisition (P) 682	
phase3 Phase selection for 4D acquisition (P) 683	

phasing	Control update region during interactive phasing (P) 683
phfid	Zero-order phasing constant for the np FID (P) 683
phfid1	Zero-order phasing constant for ni interferogram (P) 684
phfid2	Zero-order phasing constant for ni2 interferogram (P) 685
Phosphorus	Set up parameters for ³¹ P experiment (M) 685
pi3ssbsq	Set up pi/3 shifted sinebell-squared window function (M) 685
pi4ssbsq	Set up pi/4 shifted sinebell-squared window function (M) 686
pin	Pneumatics Router Interlock ((P) 686
pintvast	Plot VAST Intergral Data in a stacked 1D-NMR matrix format 687
pir	Plot integral amplitudes below spectrum (C) 687
pirn	Plot normalized integral amplitudes below spectrum (M) 687
piv	Plot integral values below spectrum (M) 688
pivn	Plot normalized integral values below spectrum (M) 688
p1	Plot spectra (C) 688
p12d	Plot 2D spectra in whitewash mode (C) 690
plane	Currently displayed 3D plane type (P) 690
plapt	Plot APT-type spectra automatically (M) 691
plarray	Plotting macro for arrayed 1D spectra (M) 691
plate_glue	Define a glue order for plotting and display (U) 692
plc	Plot a carbon spectrum (M) 692
pLCNMR	Plot all forms of LC-NMR data (M) 693
plcosy	Plot COSY- and NOESY-type spectra automatically (M) 693
pldept	Plot DEPT data, edited or unedited (M) 694
plexpinfo	Plots Experiment Information 694
plfid	Plot FIDs (C) 694
plfit	Plot deconvolution analysis (M) 695
plgrid	Plot a grid on a 2D plot (M) 695
plh	Plot proton spectrum (M) 696
plhet2dj	Plot heteronuclear J-resolved 2D spectra automatically (M) 696
plhom2dj	Plot homonuclear J-resolved 2D spectra automatically (M) 697
plhxcor	Plot X,H-correlation 2D spectrum (M) 698
p11	Plot a line list (M) 699

pllogo	Plots Logo 700
p112d	Plot results of 2D peak picking (C) 700
Plock	Sets Protection Bit for a Parameter 700
plockport	Port number to use to lock out multiple ProTune processes (P) $$700^{\circ}$$
plot	Automatically plot spectra (M) 701
plot_ <pslabe< td=""><td>21> 702</td></pslabe<>	21> 702
plot1d	Plotting macro for simple (non-arrayed) 1D spectra (M) 702
plot2D	Plot 2D spectra (M) 702
plotfile	Plot to a file (M) 703
plothirespre	\mathbf{p} High resolution plot output preparation (M) 703
plotlcnmr	An LC-NMR plotting macro (M) 703
plotmanual	Plot manually (M) 704
plotlogo	Plots a logo (M) 704
plotpreview.	Creates temporary plots of the current plot output (M) 704
plotside	Plot spectrum on side (M) 704
plotter	Plotter device (P) 704
plottop	Plot spectrum on top (M) 705
plottopside.	Plot spectrum on top and side (M) 705
plp	Plot phosphorus spectrum (M) 705
plplanes	Plot a series of 3D planes (M) 706
plt2Darg	Plot 2D arguments (P) 706
pltext	Plot text file (M) 707
pltmod	Plotter display mode (P) 707
plvast	Plot VAST Data in a stacked 1D-NMR matrix format 708
plvastget	Plot VAST spectral data in a vertical stacked plot mode 708
plvast_replo	Replot VAST spectral data one spectrum per page of paper (M) 709
plvast2d	Plot VAST data in a stacked pseudo-2D format (M) 709
plww	Plot spectra in whitewash mode (C) 710
pmode	Processing mode for 2D data (P) 710
poly0	Display mean of the data in regression.inp file (M) 712
pow	Find the value of a number raised to a power 712
powerfit	Fits the diffusional attenuation calcuated by decay_gen to the exponential of a power series in the calibration of the non-uniformity of pulsed field gradients. 712
pp	Decoupler pulse length (P) 713

ppa Plot a parameter list in plain English (M) 713 ppca1 Proton decoupler pulse calibration (M) 714 ppf Plot peak frequencies over spectrum (C) 714 pph Print pulse header (M) 715 715 715 **pps** Plot pulse sequence (C) 716 prealfa Specify a delay for longer ring down (P) 716 preAmpConfigSet the band of the preamp, high or low, connected to each transmitter channel. 717 prep...... Run prepare acquisition macro (M) 718 **Presat**...... Set up parameters for presat ¹H experiment (M) 718 prevp1..... Display the previous 3D plane (M) 718 719 prescan CoilTableRead or update the CoilTable File (M) 719 prescan_tn.... Return tn string for a given atomic number (M) 719 presig...... Preamp Signal Level Selection Parameter (parameter) 720 printer Printer device (P) 720 printfile Path to the print-to-file image (P) 720 721 printformat. Format of saved-to-file image (P) **printlayout**. Layout of printed image (P) 721 printoff......... Stop sending text to printer and start print operation (C) 721 printon Direct text output to printer (C) 721 **printregion**. Screen region to be printed (P) 722 printsize Size of printed image (P) 722 **printsend** Defines where image will print (P) 722 probe Probe type (P) 722 probeConnectSpecify which nucleus can be acquired on each RF channel (P) 723 **Probe_edit**.... Edit probe for specific nucleus (U) 723 probe edit.... Edit probe for specific nucleus (M) 723 probe_protectionProbe protection control (P) 724 proc...... Type of processing on np FID (P) 724 proc1 Type of processing on ni interferogram (P) 725 proc1d...... Processing macro for simple (non-arrayed) 1D spectra (M) 725 proc2 Type of processing on ni2 interferogram (P) 726 proc2d..... Process 2D spectra (M) 726

procarray Process arrayed 1D spectra (M) 727
process
procplot Automatically process FIDs (M) 728
profile Set up pulse sequence for gradient calibration (M) 729
profile_int . Normalise the experimental signal profile during calibration of non-uniform pulsed gradients. 729
proj Project 2D data (C) 730
proshimhelp. Proshim help (C) 730
Proton Set up parameters for ¹ H experiment (M) 731
protune Macro to start ProTune (M) 731
protune
protunegui Macro to start ProTune in graphical user interface (M) 732
prune Prune extra parameters from current tree (C) 732
pscale Plot scale below spectrum or FID (C) 733
pseudo Set default parameters for pseudo-echo weighting (M) 734
psg Display pulse sequence generation errors (M) 734
psggen Compile a user PSG object library (M,U) 734
psgset
psgupdateon. Enable update of acquisition parameters (C) 735
psgupdateoffPrevent update of acquisition parameters (C) 735
pshape Plot pulse shape or modulation pattern (M) 736
pshapef Plot the last created pulse shape (M) 736
pshr PostScript High Resolution plotting control (P) 736
pslabel Pulse sequence label (P) 737
<pre>satup Experiment-Specific Setup Macro (M) 737</pre>
pslw PostScript Line Width control (P) 737
psMain Prescan controlling macro 737
pssl Plot Arrayed Numbers (C) 738
ptext Print out a text file (M) 739
ptspec3d Region-selective 3D processing (P) 739
ptsval PTS frequency synthesizer value (P) 740
pulseinfo Shaped pulse information for calibration (M) 741
pulsetool RF pulse shape analysis (U) 741
purge Remove macro from memory (C) 742
puttxt Put text file into a data file (C) 742
putwave Write a wave into Pbox.inp file (M) 742
pw Enter pulse width pw in degrees (C) 743

pw	. Pulse width (P) 743
pw90	. 90° pulse width (P) 744
pwd	. Display current working directory (C) 744
pwpat	. Shape of refocusing pulse (P) 744
pwr	. Set power mode in directly detected dimension (C) 745
pwr1	. Set power mode in 1st indirectly detected dimension (C) 745
pwr2	. Set power mode in 2nd indirectly detected dimension (C) 746
pwsadj	. Adjust pulse interval time (M) 747
pwxcal	. Decoupler pulse calibration (M) 747
pxbss	. Bloch-Siegert shift correction during Pbox pulse generation (P) 748
pxrep	. Flag to set the level of Pbox reports (P) 748
pxset	. Assign Pbox calibration data to experimental parameters (M) 748
pxshape	. Generates a single-band shape file (M) 749
Pxsim	. Simulate Bloch profile for a shaped pulse (U) 749
Pxspy	. Create shape definition using Fourier coefficients (U) 750
<pslabel>_p</pslabel>	Lot Experiment-Specific Plot Macro 750
<pslabel>_p:</pslabel>	rocessExperiment-Specific Processing Macro 751
<pslabel>_s</pslabel>	Experiment-Specific Setup Macro 751

18 Q

qcomp	Longer dead time for longer rin	g down (P) 7	53
QKexp	Set up quick experiment (M)	753	
qtune	Tune probe using swept-tune g	raphical tool (C) 754
?	Display the value of an individu	al parameter (C	;) 754
quadtt	Prints differences in wideline re	eceiver channel	ls 755

19 R

$m{r}$ Recall display parameter set (M) 760
r (n) Recall some display parameters (C) 761
r1-r7 Real-value storage for macros (P) 761
ra Resume acquisition stopped with sa command (C) 761
random Return a random number 762
rcvrwt Weighting for different receivers (P) 763
react Recover from error conditions during werr processing (M) 763
readallshimsRead all shims from hardware (M) 763

readbrutape.	Read Bruker data files from 9-track tape (U) 764
readfile	Read the contents of a text file into two parameters (C) 764
readhw	Read current values of acquisition hardware (C) 766
readlk	Read current lock level (C) 767
readparam	Read one or more parameters from a file (C) 767
readultra	Read shim coil setting for Ultra•nmr shim system (M) $$770$$
real	Create a real variable without a value (C) 770
recon_all	Reconstruct images from 2D MRI fid data (C) 770
record	Record keyboard entries as a macro (M) 773
redor1	Set up parameters for REDOR1 pulse sequence (M) 774
redosy	Restore 2D DOSY display from sub experiment (M) 774
reff1	Reference f1 Indirect Dimension from Observe Dimension (M) 774
reff2	Reference f2 Indirect Dimension from Observe Dimension (M) 775
reffrq	Reference frequency of reference line (P) 776
reffrq1	Reference freq. of reference line in 1st indirect dimension (P) 777
reffrq2	Reference freq. of reference line in 2nd indirect dimension (P) 777
refpos	Position of reference frequency (P) 778
refpos1	Position of reference frequency in 1st indirect dimension (P) 778
refpos2	Position of reference frequency in 2nd indirect dimension (P) 779
refsource1	Center frequency in 1st indirect dimension (P) 779
refsource2	Center frequency in 2nd indirect dimension (P) 779
region	Divide spectrum into regions (C) 780
relayh	Set up parameters for RELAYH pulse sequence (M) 781
rename	Move and/or rename a file (C) 781
reorder3D	Reorders array elements in arrayed phase sensitive 2D experiment 781
reqparcheck.	Flag which enables/disables required parameters (P) 782
reqparclear.	Clears the parameters in required parameter list (M) 782
reqparlist	List of required parameters (P) 783
reqpartest	Tests whether required parameters are set (M) 783
resetf3	Reset parameters after a partial 3D Fourier transform (M) 784
resetplotter	Reset plotter to system plotter (M) 785

Roesy Convert the parameter to a ROESY experiment (M) 800
Roesy1d Convert the parameter set to a Roesy1d experiment (M) 801
rof1 Receiver gating time preceding pulse (P) 801
rof2 Receiver gating time following pulse (P) 801
rof3 Receiver gating time following T/R switch (P) 801
rotate Rotate 2D data (C) 802
rotorsync Rotor synchronization (P) 802
rp Zero-order phase in directly detected dimension (P) 802
عتZero-order phase in 1st indirectly detected dimension (P) 803
ع ت 2Zero-order phase in 2nd indirectly detected dimension (P) 803
rt Retrieve FIDs (M) 803
rtcmx Return Spinsight data into current experiment (C) 804
rtp Retrieve parameters (M) 804
rts Retrieve shim coil settings (C) 805
rttmp Retrieve experiment data from experiment subfile (M) 805
rty Retrieve individual parameters (C) 806
rtx Retrieve parameters based on rtx rules (C) 807
s Save display parameters as a set (M) 817
s (n) Save display parameters (C) 817
s2pul Set up parameters for standard two-pulse sequence (M) 818
sa Stop acquisition (C) 818
<pre>sampleSubmit change sample, Autoshim experiment to acquisition (M) 819</pre>
<pre>sample ChangeAutomation utility 819</pre>
<pre>samplename Sample name (P) 819</pre>
<pre>sampling Parameter to control elliptical k-space sampling 820</pre>
save Save data (M) 820
savefdfspec Save 1D or arrayed 1D spectra as fdf file (C) 821
savefid Save fid 822

savefile...... Base file name for saving files (P) 822

saveglobal.... Save selected parameters from global tree (P) 822

savemodule. Save module 823

savesampglobalSaves Sample Global Parameters 823

20 S

sb	Sinebell constant in directly detected dimension (P) 823
sb1	Sinebell constant in 1st indirectly detected dimension (P) 824
sb2	Sinebell constant in 2nd indirectly detected dimension (P) 824
sbs	Sinebell shift in directly detected dimension (P) 825
sbs1	Sinebell shift in 1st indirectly detected dimension (P) 825
sbs2	Sinebell shift in 2nd indirectly detected dimension (P) 826
sc	Start of chart (P) 826
sc2	Start of chart in second direction (P) 826
scalelimits	Set limits for scales in regression (M) 826
scalesw	Set scaling factor for multipulse experiments (M) 827
scalesw	Scale spectral width in directly detected dimension (P) 827
scalesw1	Set f ₁ scaling factor for 2D multipulse experiments (M) 828
scalesw1	Scale spectral width in 1st indirectly detected dimension (P) 828
scalesw2	Scale spectral width in 2nd indirectly detected dimension (P) 828
schedulerhel	ع Proshim Maintenance Scheduler help(C) 829
sd	Set first decoupler frequency to cursor position (M) 829
sd2	Set second decoupler frequency to cursor position (M) 829
sd2a	Set second decoupler frequency array (M) 829
sd3	Set third decoupler frequency to cursor position (M) 830
sda	Set first decoupler frequency array (M) 830
sd3a	Set third decoupler frequency array (M) 830
sdp	Show diffusion projection (M) 831
selld	Apptype macro for Selective 1D experiments (M) 831
select	Select spectrum, FID, trace, or 2D plane without display (C) 831
selex	Defines excitation band (M) 832
selexcit	Set up PFG selective excitation pulse sequence (M) 833
selexHT	Set up a selective Hadamard experiment (M) 833
send2vnmr	. Send a command to VnmrJ (U) 833
seqfil	Pulse sequence name (P) 834
seqgen	Initiate compilation of user's pulse sequence (M,U) 834
seqgenupdate	∍Update compilation of user's pulse sequence 835

serverport.... Returns the VnmrJ network listening port value (C) 835 set2D...... General setup for 2D experiments (M) 835 836 set2d General setup for 2D experiments (M) **set3dproc** Set 3D processing (C) 836 setallshims . Set all shims into hardware (M) 837 setcolor........ Set colors for graphics window and for plotters (C) 837 **setDECpars**.... Sets Decoupler Parameters 839 setdec2pars. Set decoupler 2 parameter values from probe file (M) 839 setdgroup Set the Dgroup of a parameter in a tree (C) 839 **setenumeral**. Set values of a string parameter in a tree (C) 840 setether....... Connect or reconnect host computer to Ethernet (U) 840 **setexport** Set parameter bits for use with protocols (M) 840 setfrg.....Set frequency of rf channels (C) 841 setgauss........ Set a Gaussian fraction for lineshape (M) 841 setgcal Set the gradient calibration constant (M) 842 **setgcoil**...... Assign sysgcoil configuration parameter (M) 842 setgrid Divide graphics window into rows and columns (C) 842 setgroup...... Set group of a parameter in a tree (C) 843 sethtfrg1 Set a Hadamard frequency list from a line list ((M) 844 844 **sethwshim**..... Special case of sethw for setting shims (C) 846 846 setlimit...... Set limits of a parameter in a tree (C) 847 set1k Set up lock parameters (M) 848 setlockfreq.Set lock frequency (M) 849 setLP Set up linear prediction in the direct dimension (M) 849 setLP1...... Set F1 linear prediction parameters (M) 849 set1p0..... Set parameters for zero linear phase (M) 850 **setnoether**.... Disconnect host computer from Ethernet (U) 850 **setobspars**.... Sets Observe Parameters 850 setoffset Calculate offset frequency for given nucleus and ppm (M) 850 setparams Write parameter to current probe file (M) 851 setpen...... Set maximum number of HP plotter pens (M) 851 **setplotdev**.... Return characteristics of a named plotter (C) 852 setpower........ Set power and pulsewidth for a given $\gamma B1$ value (M) 852 **setprotect**.... Set the protection bits of a variable in a tree (C) 853 setpw180ad.... Creates and sets observe adiabatic pulse shapes (M) 855

setpwx180ad	Creates and sets decoupler adiabatic pulse shapes (M) 855
setrc	Set receiver constants (M) 856
setref	Set frequency referencing (M) 856
setref1	Set freq. referencing for 1st indirectly detected dimension (M) 858
setref2	Set freq. referencing for 2nd indirect detected dimension (M) 858
setscout	Set up a scout run (M) 859
setssfilter	Set sslsfrq to the frequencies of each suppressed solvents (M) 860
setsw	Set spectral width (M) 860
setsw1	Set spectral width in evolution dimension (M) 860
setsw2	Set spectral width in 2nd evolution dimension (M) 861
setselfrqc	Set selective frequency and width (M) 861
setselinv	Set up selective inversion (M) 861
settcldefaul	LtSelect default display templates for pulse sequence (M) 862
settune	Opens the Auto Tune Setup dialog (M) 862
settype	Set the type of a parameter (C) 862
setup	Set up parameters for basic experiments (M) 863
setup_dosy	Set up gradient levels for DOSY experiments (M) 864
setuserpsg	Creates/initializes user PSG directory 864
setvalue	Set value of any parameter in a tree (C) 864
setwave	Write a wave definition string into Pbox.inp file (M) 865
setwell	Adjust the label of the "t1" axis for VAST contour maps 865
setwin	Activate selected window (C) 866
sf	Start of FID (P) 866
sf1	Start of interferogram in 1st indirectly detected dimension (P) 867
sf2	Start of interferogram in 2nd indirectly detected dimension (P) 867
sfrq	Transmitter frequency of observe nucleus (P) 867
sh2pu1	Set up for a shaped observe excitation sequence (M) 868
shdec	Set up for shaped observe excitation sequence (M) 868
shell	Start a UNIX shell (C) 868
shelli	Start an interactive UNIX shell (C) 869
shim	Submit an Autoshim experiment to acquisition (C) 869
shimmult	Multiple the shim dacs of the current shimset 870

shimnames	Returns shim names 870
shimset	Type of shim set (P) 870
showconfig	Show system configuration settings (M) 872
showconsole.	Show system configuration settings (U) 872
showdosy	Show DOSY Plot (M) 873
showdosyfit.	Plots the experimental signal attenuation, fitted attenuation and residual for one peak from a 2D or 3D DOSY experiment 873
showdosyresi	.dual Plots the residual for one peak from a 2D or 3D DOSY experiment 873
showgradfit.	Plots the experimental gradient variation with position and the power series fit in non-uniform gradient calibration. 874
showfit	Display numerical results of deconvolution (M) 874
showloginbox	Shows operator login dialog (M) 874
shownugfit	Plots the logarithm of the calculated diffusional attenuation and of the power series fit in non-uniform gradient calibration 875
shownumx	Show x position of number (P) 875
shownumy	Show y position of number (P) 875
showoriginal	Restore first 2D spectrum in 3D DOSY experiment (M) 875
showplotter.	Show list of currently defined plotters and printers (M) 876
showplotq	Display plot jobs in plot queue (M) 876
showprintq	Display print jobs in print queue (M) 876
showprotuneg	ruishow the graphical interface while tuning (P) 876
showrfmon	Show RF Monitor Button in Hardware Bar (P) 877
showsampglob	alShows sample global parameters 877
showstat	Display information about status of acquisition (M,U) 877
sim	Sample in magnet (For systems equipped with a robot) 877
sin	Find sine value of an angle (C) 878
sine	Find values for a sine window function (M) 878
sinebell	Select default parameters for sinebell weighting (M) 879
sinesq	Find values for a sine-squared window function (M) 879
size	Returns the number of elements in an arrayed parameter (0) 880
slfreq	Measured line frequencies (P) 880
slw	Spin simulation linewidth (P) 880
smaxf	Maximum frequency of any transition (P) 881

sminf	Minimum frequency of any transition (P) 881
smsport	Sample Management System serial port connection (P) 881
sn	Signal-to-noise ratio (P) 882
solppm	Return ppm and peak width of solvent resonances (M) 882
solvent	Lock solvent (P) 882
solvinfo	Retrieve information from solvent table (C) 883
sort	Sort real values of a parameter (M) 883
sp	Start of plot in directly detected dimension (P) 884
sp1	Start of plot in 1st indirectly detected dimension (P) 884
sp2	Start of plot in 2nd indirectly detected dimension (P) 884
spadd	Add current spectrum to add/subtract experiment (C) 885
spcfrq	Display frequencies of rf channels (M) 887
specdc3d	3D spectral drift correction (P) 887
spin	Submit a spin setup experiment to acquisition (C) 888
spin	Sample spin rate (P) 888
spinll	Set up a slfreq array (M) 889
spinner	Open the Spinner Control window (C) 889
spins	Perform spin simulation calculation (C) 890
split	Split difference between two cursors (M) 892
spintype	Spinner Type ((P) 893
splmodprepar	eUsed by the dosy macro to prepare data for the program SPLMOD 893
splmodread	Used by the dosy macro to convert the output of the SPLMOD program into a form suitable for ddif 893
spmax	Take the maximum of two spectra (C) 894
spmin	Take minimum of two spectra in add/subtract experiment(C)894
spsm	Enter spin system (M) 894
spsub	Subtract current spectrum from add/subtract experiment (C) 895
sącosine	Set up unshifted cosine-squared window function (M) 896
sqdir	Study queue directory (P) 896
sqend	End a study queue (M) 897
sqexp	Load experiment from protocol (M) 897
sqfilemenu	. Study queue file menu commands (M) 897
sqLog	Records specific events from a study queue 898

sqmode	. Study queue mode (P) 900
sqname	. Study queue parameter template (P) 900
sqpars	. Create study queue parameters for imaging (M) 900
sqprotocol	. Macro to create protocols (M) 901
sqreset	. Reset study queue parameters for imaging (M) 901
sqrt	. Return square root of a real number (0) 901
sqsavestudy	. Macro to save study parameters for imaging (M) 901
sqsinebell	. Set up unshifted sinebell-squared window function (M) 902
srate	. Spinning rate for magic angle spinning (P) 902
sread	. Read converted data into VnmrJ (C) 902
srof2	. Calculate exact rof2 value for Cold Probes (M) 903
SS	. Steady-state transients (P) 903
ssecho	. Set up solid-state echo pulse sequence (M) 903
ssechol	. Set up parameters for SSECH01 pulse sequence (M) 904
ssfilter	. Full bandwidth of digital filter to yield a filtered FID (P) 904
sslsfrq	. Center of solvent-suppressed region of spectrum (P) 904
ssntaps	Number of coefficients in digital filter (P) 905
ssorder	. Order of polynomial to fit digitally filtered FID (P) 905
stack	. Stacking mode for processing and plotting arrayed spectra (M) 906
stackmode	. Stacking control for processing arrayed 1D spectra (P) 907
startq	. Start a chained study queue (M) 907
status	. Display status of sample changer (C,U) 907
std1d	Apptype macro for Standard 1D experiments (M) 908
stdshm	. Interactively create a method string for autoshimming (M) 908
sth	. Minimum intensity threshold (P) 909
string	. Create a string variable (C) 909
string2array	Formats a String Variable into an Array 909
strstr	. Sets ret to the starting position of the first occurrence of string2 in string1 909
strsv2array	. Formats a String Separated Variable into an Array 911
strtext	. Starting point for LP data extension in np dimension (P) 911
strtext1	. Starting point for LP data extension in ni dimension (P) 911

strtext2	Starting point for LP data extension in ni2 dimension (P) 912
strtlp	Starting point for LP calculation in np dimension (P) \qquad 912
strtlp1	Starting point for LP calculation in ni dimension (P) 913
strtlp2	Starting point for LP calculation in ni2 dimension (P) 913
studyid	Study identification (P) 913
studypar	Study parameters (P) 914
studystatus.	Study status (P) 914
studytime	Determine start and end times for studies (P) 914
su	Submit a setup experiment to acquisition (M) 915
sub	Subtract current FID from add/subtract experiment (C) 915
substr	Select a substring from a string (C) 916
suselfrq	Select peak, continue selective excitation experiment (M) 923
svdat	Save data (C) 923
svf	Save FIDs in current experiment (M) 924
svfdf	Save FID data in FDF format (M) 925
svfdir	Directory for non-study data (P) 925
svfj	Save FID in JCAMP-DX format (M) 926
Svfname	Create path for data storage (C) 927
svfname	Filename parameter template for non-study data (P) 929
svimg	Generate and Save images as FDF files. (macro) 929
svllj	Save peak listing in JCAMP-DX X,Y or X,Y,M format (M) 930
svlsj	Save large dynamic range spectrum in JCAMP-DX format (M) 931
svp	Save parameters from current experiment (M) 932
svpdp	Compare workspace parameters to parameter file 932
svr	Save secured REC data for VnmrJ SE 932
svs	Save shim coil settings (C) 933
svs	Spin simulation vertical scale (P) 934
svsis	Generate and Save images as FDF files. (macro) 934
svsj	Save spectrum in JCAMP-DX format (M) 935
svtmp	Move experiment data into experiment subfile (M) 936
svxyj	Save spectrum in JCAMP-DX X,Y format (M) 937
sw	Spectral width in directly detected dimension (P) 938
sw1	Spectral width in 1st indirectly detected dimension (P) 938

sw2	Specti (P)	al width in 2 939	2nd indire	ctly detected dimension
sw3	Specti (P)	al width in 3 939	Brd indired	tly detected dimension:
sysgcoil	Syster	n gradient c	oil (P)	939
system	Syster	n type (P)	940	
systemdir	Vnmr	system dire	ectory (P)	940

21 T

t1	T_1 exponential analysis (M) 943
t1s	T_1 exponential analysis with short output table (M) 944
t2	T ₂ exponential analysis (M) 944
t2s	T_2 exponential analysis with short output table (M) 945
tabc	Convert data in table order to linear order (M) 945
tan	Find tangent value of an angle (C) 946
tape	Read tapes from VXR-style system (M,U) 947
tape	Control tape options of files program (P) 947
target_bval.	Adjust gdiff to achieve target b-value (M) 948
tcapply	Apply Table Conversion Reformatting to Data (C) 948
tchan	RF channel number used for tuning (P) 949
tcl	Send Tcl script to Tcl version of dg window (C) 949
tcclose	Table Convert Close (C) 949
temp	Open the Temperature Control window (C) 950
temp	Sample temperature (P) 950
tempcal	Temperature calculation (C) 951
tempcalc	Measure approximate sample temperature in Cold Probes (M) 951
testacquire	Test acquire mode (P) 952
testct	Check ct for resuming signal-to-noise testing (M) 952
testsn	Test signal-to-noise of a spectrum (M) 952
teststr	Find which array matches a string M) 953
text	Display text or set new text for current experiment (C) 954
textis	Return the current text display status (C) 954
textvi	Edit text file of current experiment (M) 955
th	Threshold (P) 955
th2d	Threshold for integrating peaks in 2D spectra (P) 955
thadj	Adjust threshold for peak printout (M) 955
time	Display experiment time or recalculate number of transients (M) 956

tin	Temperature interlock (P) 957
tlt	First-order baseline correction (P) 957
tmove	Left-shift FID to time-domain cursor (M) 958
tmsref	Reference 1D proton or carbon spectrum to TMS (M) 958
tn	Nucleus for observe transmitter (P) 958
tncosyps	Set up parameters for TNCOSYPS pulse sequence (M) 959
tndqcosy	Set up parameters for TNDQCOSY pulse sequence (M) 959
tnmqcosy	Set up parameters for TNMQCOSY pulse sequence (M) 959
tnnoesy	Set up parameters for TNNOESY pulse sequence (M) 959
tnroesy	Set up parameters for TNROESY pulse sequence (M) 960
tntocsy	Set up parameters for TNTOCSY pulse sequence (M) 960
Тосзу	Convert the parameters to a TOCSY experiment (M) 960
Tocsy1d	Convert the parameter set to a Tocsy1d experiment (M) 960
tocsyHT	Set up the tocsyHT experiment (M) 960
tof	Frequency offset for observe transmitter (P) 961
tpwr	Observe transmitter power level with linear amplifiers (P) 961
tpwrf	Observe transmitter fine power (P) 962
tpwrm	Observe transmitter linear modulator power (P) 962
trace	Mode for <i>n</i> -dimensional data display (P) 963
traymax	Sample changer tray slots (P) 963
trfunc	Translates screen co-ordinates 963
trfuncd	Translates a screen distance 964
troesy	Set up parameters for TROESY pulse sequence (M) 964
trunc	Truncate real numbers (0) 964
trtune	Allows the user to view multiple tuning traces apparently simultaneously 964
tshift	Adjust tau2 to current cursor position (M) 965
tugain	Receiver gain used in tuning (P) 965
tune	Assign a frequency to a channel for probe tuning (C) 965
tunehf	Tune both H1 and F19 on an HFX probe (M) 967
tunematch	Default match target, in percent of optimum (P) 967
tunemethod	Method to use for tuning (P) 967
tuneResult	Message indicating how well the tuning succeeded (P) 968

tunerp A pulse sequence for pulse tuning thro couplers in the VNMRJ display 968	ough the directional
tunesw Width of the tuning sweep in Hz (P)	971
tupwr Transmitter power used in tuning (P)	972
typeof Return identifier for argument type (0)	972

22 U

ultra8selects the Ultra 8 shim configuration (M) 973
ultra18 Select 18 shim configuration for Ultra 18 shim power supply (M) 974
undospins Restore spin system as before last iterative run (M) 974
undosy Restore original 1D NMR data from sub experiment (M) 974
unitDefine conversion units (C) 975
unixtime
unlock Remove inactive lock and join experiment (C) 977
updatepars Update all parameter sets saved in a directory (M) 977
updateprobe. Update probe file (М) 978
updaterev Update after installing new VnmrJ version (M) 978
updtgcoil Update gradient coil (M) 978
updtparam Update specified acquisition parameters (C) 979
usemark Use "mark" output as deconvolution starting point (M) 979
userdir VnmrJ user directory (P) 980
usergoExperiment setup macro called by go, ga, and au (M) 980
userfixpar Macro called by fixpar (M) 980

23 V

vast1d	Set up initial parameters for VAST experiments (M) 982
vastget	Selects and displays VAST spectra (M) 983
vastglue	Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M) 983
vastglue2	Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M) 984
vastgo	Turn off LC stop flow automation, start VAST automation (M) 984
vbg	Run VNMR processing in background (U) 984
vf	Vertical scale of FID (P) 985
v i	Edit text file with vi text editor (M) 985
vibradd	Display relative amplitudes of Cold Probe vibrations (M) 988

989 vjhelp..... Display VnmrJ help (U) 989 vn..... Start VNMR directly (U) vnmr...... Starts VnmrJ (U) 989 vnmr_accountingOpen Accounting window (U) 989 vnmremail Utility to Send Files via Email 990 **vnmrexit**...... Exit from the VNMR system (C) 990 vnmrj Start VnmrJ (U) 990 **vnmrjcmd()** ... Commands to invoke the GUI popup (C) 991 **vnmrjOptions**Installer for passworded VnmrJ options (C) 991 vnmrplot...... Plot files (U) 992 992 **vnmrprint** Print text files (U) vo..... Vertical offset (P) 993 vp..... Vertical position of spectrum (P) 993 vpaction...... Set initial state for multiple viewports (M) 993 vpf Current vertical position of FID (P) 993 vpfi..... Current vertical position of imaginary FID (P) 994 vpset3def Set the viewport state to three default viewports (M) 994 vpsetup Set new viewports (M) 995 **vs**..... Vertical scale (P) 995 vs2d..... Vertical scale for 2D displays (P) 995 vsadj Automatic vertical scale adjustment (M) 996 vsadj2..... Automatic vertical scale adjustment by powers of 2 (M) 996 vsadjc...... Automatic vertical scale adjustment for 13C spectra (M) 997 vsadjh...... Automatic vertical scale adjustment for ¹H spectra (M) 997 vsproj...... Vertical scale for projections and traces (P) 998 vtairflow...... Variable Temperature Air Flow (P) 998 **vtairlimits**. Variable Temperature Air Flow Limits (P) 998 **vtc** Variable temperature cutoff point (P) 999 vtcomplv1 Variable temperature compensation for gradient shimming 999 (P) vttype...... Variable temperature controller present (P) 1000 vtwait...... Variable temperature wait time (P) 1000 vxr_unix....... Convert VXR-style text files to UNIX format (M, U) 1001

24 W

w..... Who is using system (C) 1005

walkup	Walkup automation (M) 1006
walkupQ_runt	imeMacro to Control Study Queue 1006
waltz	WALTZ decoupling present (P) 1008
warmprobe	Tells the system a warm probe is present 1008
wbs	Specify action when bs transients accumulate (C) 1008
wbs	When block size (P) 1009
wc	Counts Words in a String 1009
wc2	Width of chart in second direction (P) 1009
wcmax	Maximum width of chart (P) 1010
wc2max	Maximum width of chart in second direction (P) 1010
wdone	Specify action when experiment is done (C) 1010
wdone	Specify action when experiment is done (P) 1011
werr	Specify action when error occurs (C) 1011
werr	When error (P) 1012
wet	Flag to turn on or off wet solvent suppression ((P) 1012
Wet1d	Set up parameters for wet ¹ H experiment (M) 1012
wetdqcosy	Set up parameters for a WETDQCOSY pulse sequence (M) 1012
wetgcosy	Set up parameters for a WETGCOSY pulse sequence (M) 1013
wetghmqcps	Set up parameters for a WETGHMQCPS pulse sequence (M) 1013
wetghsqc	Set up parameters for a WETGHSQC pulse sequence (M) 1013
wetgmącosy	Set up parameters for a WETGHSQC pulse sequence (M) 1013
wetit	Set up and create pulse shapes for Wet1d experiment (M) 1013
wetnoesy	Set up parameters for a WETNOESY pulse sequence (M) 1014
wetpeaks	Number of peaks for wet solvent suppression (P) 1014
wetpwxcal	Set up parameters for a WETPWXCAL pulse sequence (M) 1014
wettntocsy	Set up parameters for a WETTNTOCSY pulse sequence (M) 1014
wetshape	Shape for pwwet pulses (P) 1014
wexp	Specify action when experiment completes (C) 1015
wexp	When experiment completes (P) 1015
wf	Width of FID (P) 1016
wf1	Width of interferogram in 1st indirectly detected dimension (P) 1016

wf2	. Width of interferogram in 2nd indirectly detected dimension (P) 1017
wfgtest	. Waveform generator test (M) 1017
wft	. Weight and Fourier transform 1D data (C) 1017
wft1d	. Weight and Fourier transform f_2 for 2D data (C) 1018
wft1da	. Weight and Fourier transform phase-sensitive data (M) 1018
wft1dac	. Combine arrayed 2D FID matrices (M) 1018
wft2d	. Weight and Fourier transform 2D data (C) 1019
wft2da	. Weight and Fourier transform phase-sensitive data (M) 1020
wft2dac	. Combine arrayed 2D FID matrices (M) 1020
wftt3	. Process f ₃ dimension during 3D acquisition (M) 1020
which	. Display which command or macro is used (M) 1021
wnt	. Specify action when nt transients accumulate (C) 1021
wnt	. When number of transients (P) 1022
wp	. Width of plot in directly detected dimension (P) 1022
wp1	. Width of plot in 1st indirectly detected dimension (P) 1023
wp2	. Width of plot in 2nd indirectly detected dimension (P) 1023
write	. Write formatted text to a device (C) 1023
writefid	. Write numeric text file using a FID element (C) 1025
writejxy	. Create x,y ascii file from phasefile for JCAMP-DX conversion (M) 1026
writeparam	. Write one of more parameters to a file (C) 1026
writespectr	um write a spectrum to a binary file (C) 1027
writetrace	. Create ascii file from phasefile (f1 or f2) trace (M) 1028
writexy	. Create x,y ascii file from phasefile (f1 or f2) trace (M) 1028
wrtp	. Command string executed after rtp command (P) 1029
wsram	. Send hardware configuration to acquisition console (C) 1029
wshim	. Conditions when shimming is performed (P) 1029
wtfile	. User-defined weighting in directly detected dimension (P) 1030
wtfile1	. User-defined weighting in 1st indirectly detected dimension (P) 1031
wtfile2	. User-defined weighting in 2nd indirectly detected dimension (P) 1031
wtgen	. Compile user-written weighting functions (M,U) 1031

wti Interactive weighting (C) 1	032
wtia Interactive weighting for 2D a	absorptive data (M) 1033
wtune Specify when to tune (P)	033
wtunedone What to do after ProTune tuni	ing is done (P) 1034
wysiwyg Set plot display or full display	r (P) 1034

25 X

ж0	X-zero position of HP pen plotter or Postscript device (P) 1037
x1	X1 shim gradient (P) 1037
x2y2	X2Y2 shim gradient (P) 1037
х3	X3 shim gradient (P) 1037
x4	X4 shim gradient (P) 1038
xdiag	Threshold for excluding diagonal peaks when peak picking (P) 1038
xgate	Load time counter (M) 1038
xm1	Utility macro for study queue experiment manager (M) 1039
xmaction	Perform study queue action (M) 1039
xmactionw	Perform study queue action for walkup (M) 1039
xmaddreq	Add a required protocol before the main protocol (M) 1039
xmcheckreq	Check required protocol name (M) 1039
xmconvert	Convert a temporarily stored study into a submitted study (M) 1040
жисору	Copy protocols in a study queue (M) 1040
xmdelete	Delete nodes in a study queue (M) 1040
xmenablepane	1 Enable or disable a parameter panel (M) 1041
xmendq	End a chained study queue (M) 1041
xmgetatts	Get study queue attributes (M) 1041
xmHprescan	Set up and process Proton prescans (M) 1041
xminit	Initialize an imaging study queue (M) 1041
xmlockup	Move a study queue node up and lock it (M) 1042
xmmakenode	Make a new study queue node (M) 1042
xmnext	Find next prescan or next experiment in study queue (M) 1042
xmprescan	Run prescans in study queue (M) 1042
xmreact	Recover from error conditions during automation study (M) 1043
xmreadnode	Read attributes from a study queue node (M) 1043

xmrtpar	Retrieve parameters from a study queue node (M) 1043
xmsample	Write enterQ entry for a sample for study queue – liquids (M) 1044
xmsara	Write enterQ entry for a sample for study queue – imaging (M) $$1044$$
xmsatfrq	Processing for Presat experiment (M) 1044
xmselect	Action when study queue node is selected (M) 1044
xmsetatts	Set an attribute for a study queue node (M) 1045
xmsetattr	Set an attribute for a study queue node (M) 1045
xmshowdata	Show data from a study queue node (M) 1045
xmstartnight	ε φ Start the night queue (M) 1045
xmsubmit	Submit sample(s) to the study queue (M) 1046
xmtime	Update the study queue time (M) 1046
xmtune	Check tune parameter during automation (M) 1046
xmwerr	Recover from acquisition error in study queue (M) 1046
xmwexp	Processing macro for end of acquisition in study queue (M) 1047
xmwritenode	Write study queue node attributes (M) 1047
xmwritesq	Write study queue node order (M) 1047
xpol	Cross-polarization (P) 1048
xpolar1	Set up parameters for XPOLAR1 pulse sequence (M) 1048
ху	XY shim gradient (P) 1048
xz	XZ shim gradient (P) 1048
xz2	XZ2 shim gradient (P) 1049

26 Y

y0 Y-zero position of HP p (P) 1051	en plotter or Postscript device
у1 Y1 shim gradient (Р)	1051
${f y}$ 3Y3 shim gradient (P)	1052
y4 Y4 shim gradient (P)	1052
yzYZ shim gradient (P)	1052
yz2 YZ2 shim gradient (P)	1052
yz2 YZ2 shim gradient (P)	1052

27 Z

z Add integral reset poi	nt at cursor position (C)	1056
zoZ0 field position (P)	1056	
z1Z1 shim gradient (P)	1057	
z1cZ1C shim gradient (P)	1057	

z2	Z2 shim gradient (P) 1057
z2c	Z2C shim gradient (P) 1057
z2x2y2	Z2X2Y2 shim gradient (P) 1058
z2x3	Z2X3 shim gradient (P) 1058
z2xy	Z2XY shim gradient (P) 1058
z2y3	Z2Y3 shim gradient (P) 1058
z3	Z3 shim gradient (P) 1058
z3c	Z3C shim gradient (P) 1058
z3x	Z3X shim gradient (P) 1059
z3x2y2	Z3X2Y2 shim gradient (P) 1059
z3x3	Z3X3 shim gradient (P) 1059
z3xy	Z3XY shim gradient (P) 1059
z3y	Z3Y shim gradient (P) 1059
z3y3	Z3Y3 shim gradient (P) 1060
z4	Z4 shim gradient (P) 1060
z4c	Z4C shim gradient (P) 1060
z4x	Z4X shim gradient (P) 1060
z4x2y2	Z4X2Y2 shim gradient (P) 1060
z4xy	Z4XY shim gradient (P) 1060
z4y	Z4Y shim gradient (P) 1061
z5	Z5 shim gradient (P) 1061
z5x	Z5X shim gradient (P) 1061
z5y	Z5Y shim gradient (P) 1061
z6	Z6 shim gradient (P) 1061
z7	Z7 shim gradient (P) 1062
z8	Z8 shim gradient (P) 1062
zeroneg	Set all negative intensities of 2D spectra to zero (C) 1062
zoom	Adjust display to given width (M) 1062
zx2y2	ZX2Y2 shim gradient (P) 1062
zx3	ZX3 shim gradient (P) 1063
zxy	ZXY shim gradient (P) 1063
zy3	ZY3 shim gradient (P) 1063



Agilent VnmrJ 4 Command and Parameter Reference Guide

Notational Conventions

The *VnmrJ Command and Parameter Reference* describes in detail the commands, macros, and parameters in VnmrJ software. Information new to VnmrJ in this version is shown by a change bar (as shown to the left of this paragraph).

Title line codes

Each entry has a letter in parentheses in the title line that identifies the type of entry:

- (C) VnmrJ command
- (M) VnmrJ macro command (from the maclib directory)
- (0) MAGICAL programming operator
- (P) VnmrJ parameter
- (U) UNIX command (not executable within VnmrJ)
- (C,U) (M,U) Executable from UNIX or VnmrJ (note that syntax is different)

Applicability

An entry with applicability information applies only to the system or accessory listed. If the entry does not include applicability information, the entry applies to all systems.



Command and macro syntax

	Each command and macro entry includes the syntax used when entering it into the system. The following examples illustrate this syntax:
halt	If no parentheses are shown, enter the command or macro exactly as shown, e.g., enter halt.
delexp(exp_num)	If parentheses are shown, enter the command or macro name as shown, but replace arguments with a value, e.g., if exp_num is 5, enter $delexp(5)$.
rttmp(file)	Arguments can be a string (e.g., name of file or solvent), number, variable, or parameter (e.g., pw),. If a string, enclose it with single quote marks, <i>e.g.</i> , <i>if</i> file <i>is</i> samp02, <i>enter</i> rttmp('samp02'). If number, variable, or parameter, do <i>not</i> use marks.
rl<(frequency)>	Angle brackets (< and >) indicate optional input, e.g., if frequency not needed or the default value of frequency is acceptable, enter rl, but if frequency has a value such as 10, enter rl(10).
<pre>md(<from_exp,>to_exp)</from_exp,></pre>	Arguments can also be optional. Use a comma to separate arguments, e.g., $md(2,3)$. Unless stated otherwise, the order of arguments is often important.
nll<('pos')>	A keyword is frequently used as an argument. In the syntax, keywords are shown in single quotes and are entered exactly as shown, e.g., to use the optional keyword 'pos' for nll, enter nll('pos').
dc2d('f1' 'f2')	A vertical bar indicates an OR condition, e.g., either 'f1' or 'f2' can be an argument to dc2d.
<pre>sin(angle)<:n></pre>	Some commands return values to a calling macro. This is shown by a colon followed by one or more variables, e.g., if angle is variable x and n is variable rt, then $sin(x):rt$ returns the value of $sin(x)$ to the calling macro via the variable rt.
z(reset1,reset2,)	Three dots indicate the sequence of arguments continues. Unless a limit is given, you can enter one argument, two, three, or as many as needed.

Parameter Syntax

Parameter syntax is always in the form parameter_name=value. If value is a string, enclose it in single quote marks; otherwise, no marks are used, e.g., auto='y', plotter='ThinkJet', spin=5. Note that some parameters are not user-enterable.

Notational Conventions

Throughout all Agilent NMR manuals, typewriter-like characters identify commands, parameters, directories, file names, and text displayed on the screen.

Because pressing the Return key is required at the end of almost every command or line of text you type on the keyboard, assume this use of the Return key unless stated otherwise.

Other Sources of Information

For further information about an entry, refer to the manual listed under "See also." For general coverage on VnmrJ, refer to the following manuals (each manual is also online):

Automation User Guide Spectroscopy User Guide VnmrJ Installation and Administration VnmrJ Imaging User Guide

Notational Conventions



Agilent VnmrJ 4 Command and Parameter Reference Guide

aa	Abort acquisition with error (C)
abort	Terminate action of calling macro and all higher macros (C)
abortallacqs	Reset acquisition computer in a drastic situation (C)
abortoff	Terminate normal functioning of abort in a macro (C)
aborton	Restore normal functioning of abort in a macro (C)
abs	Find absolute value of a number (C)
AC1S-AC11S	Autocalibration macros (M)
ACbackup	Make backup copy of current probe file (M)
acct	Writes records for operator login and logoff (M)
ACreport	Print copy of probe file after autocalibration (M)
acos	Find arc cosine of number (C)
acosy	Automatic analysis of COSY data (C)
acosyold	Automatic analysis of COSY data, old algorithm (C)
acq_errors	Acquisition Done and Error Codes
acqdequeue	Dequeue an acquisition
acqdisp	Display message on the acquisition status line (C)
acqi	Interactive acquisition display process (C)
acqmeter	Open Acqmeter window (M)
Acqmeter	Open Acqmeter window (U)
acqmode	Acquisition mode (P)
acqreserve	Reserve the acquisition console for the current owner



acqstat	Open Acquisition Status window (M)
Acqstat	Open Acquisition Status window (U)
acqstatus	Acquisition status (P)
acquire	Acquire data (M)
actionid	Current study queue node id (P)
activestudy	Active study name (P)
add	Add current FID to add/subtract experiment (C)
addi	Start interactive add/subtract mode (C)
addnucleus	Add new nucleus to existing probe file (M)
addpar	Add selected parameters to current experiment (M)
addparams	Add parameter to current probe file (M)
addprobe	Create new probe directory and probe file (M)
adept	Automatic DEPT analysis and spectrum editing (C)
aexppl	Automatic plot of spectral expansion (M)
ai	Select absolute-intensity mode (C)
aig	Absolute-intensity group (P)
aipDisplay	Display images (C)
aipLoadSpec	Load fdf spectra (C)
aipMakeMaps	Make csi map (C)
aipOverlayFrames	Overlay images in selected frames (C)
aippars	Create parameters for imaging browser (M)
aipRemoveSpec	Remove all or specified spectral data (C)
aipRQcommand	Load and display images (C)
aipSaveColormap	Save color map for selected images (C)
aipSetColormap	Load color map for selected images (C)
aipSetTransparency	Set transparency for images or text (C)
aipShow3PCursors	Turn on/off cursors for 3-plane extraction (C)
aipShowCSIData	Display CSI spectral data in grid layout (C)
aipShowSpec	Display spectra specified by key(s), in specified layout (C)
aipViewLayers	Get information for overlaid images (C)
alfa	Set alfa delay before acquisition (P)

alock	Automatic lock control (P)
ampmode	Independent control of amplifier mode (P)
amptype	Amplifier type (P)
analyz	Calculate standard peak height (M)
analyze	Generalized curve fitting (C)
annotation	Display annotation specified by the parameter "template" or the default.
ap	Print out "all" parameters (C)
ap	"All" parameters display control (P)
ара	Plot parameters automatically (M)
aph	Automatic phase adjustment of spectra (C)
aph0	Automatic phase of zero-order term (C)
aphb	Auto phasing for Bruker data (C)
aphx	Perform optimized automatic phasing (M)
appdir	Application directory information
appdirs	Starts Applications Directory Editor (M)
appmode	Application mode (P)
apptype	Application type (P)
Apt	Set up parameters for APT experiment (M)
aptaph	Automatic processing for APT spectra (M)
array	Easy entry of linearly spaced array values (M)
array	Parameter order and precedence (P)
arraydim	Dimension of experiment (P)
array2csv	Formats Array into Comma Separate Variable
array2string	Formats Array into String
array2strsv	Formats Array into String Separated Variable
asin	Find arc sine of number (C)
asize	Make plot resolution along f_1 and f_2 the same (M) $% f_1$
assign	Assign transitions to experimental lines (M)
at	Acquisition time (P)
atan	Find arc tangent of a number (C)
atan2	Find arc tangent of two numbers (C)

atcmd	Call a macro at a specified time (M)
atext	Append string to current experiment text file (M)
attval	Calculate pulse width (M)
atune	ProTune Present (P)
au	Submit experiment to acquisition and process data (M)
AuCALch3i	Set up autocalibration with CH3I sample (M)
AuCALch3i1	Get autocalibration with $\ensuremath{CH_3}\ensuremath{I}$ sample (M)
AuCALch3oh	Set up autocalibration with Autotest sample (M)
AuCALch3oh1	Get autocalibration with Autotest sample (M)
Aucalibz0	Automatic Hz to DAC calibration for Z0 (M)
AuCdec	Carbon decoupler calibration macro (M)
AuCgrad	Carbon/proton gradient ratio calibration macro (M)
AuCobs	Carbon observe calibration macro (M)
audiofilter	Audio filter board type (P)
Aufindz0	Automatic adjustment of Z0 (M)
Augcal	Probe gcal calibration macro (M)
Augmap	Automated gradient map generation (M)
Augmapz0	Automatic lock gradient map generation and z0 calibration (M)
AuHdec	Proton decoupler calibration (M)
AuHobs	Proton observe calibration macro (M)
Aumakegmap	Auto lock gradient map generation (M)
AuNuc	Get parameters for a given nucleus (M)
auto	Prepare for an automation run (C)
auto	Automation mode active (P)
auto_au	Controlling macro for automation (M)
autoaa	Abort an automation run with no error
Autobackup	Back up current probe file (M)
autodept	Automated complete analysis of DEPT data (M)
autodir	Automation directory absolute path (P)
autogo	Start automation run (C)

autolist	Set up and start chained acquisition (M)
automerge	Merges overniteQ with daytimeQ
Automkdir	Creates Data Directory from Template
autoname	Create path for data storage (C)
autoname	Prefix for automation data file (P)
autoq	Utility commands for the automation queue
autora	Resume suspended automation run (C)
autosa	Suspend current automation run (C)
autoscale	Resume autoscaling after limits set by scalelimits macro (M)
autostack	Automatic stacking for processing and plotting arrays (M)
autotest	Open Auto Test Window (C)
autotime	Displays approximate time for automation (M)
av	Set abs. value mode in directly detected dimension (C)
av1	Set abs. value mode in 1st indirectly detected dimension (C)
av2	Set abs. value mode in 2nd indirectly detected dimension (C)
averag	Calculate average and standard deviation of input (C)
awc	Additive weighting const. in directly detected dimension (P)
awc1	Additive weighting const. in 1st indirectly detected dimension (P)
awc2	Additive weighting const. in 2nd indirectly detected dimension (P)
axis	Provide axis labels and scaling factors (C)
axis	Axis label for displays and plots (P)
axisf	Axis label for FID displays and plots (P)

Abort acquisition with error (C)

Syntax aa

In some circumstances, there is a delay between the time g_0 is entered and the acquisition is started. During this time, instructions based on the selected pulse sequence are being generated. This is signified by the letters "PSG" appearing in the upper left corner of the status window. An aa command issued under these circumstances reports that no acquisition is active but it instead stops the instruction generation process and the message "PSG aborted" appears.

See also NMR Spectroscopy User Guide

Related	file	File name of a parameter set (P)
	go	Submit experiment to acquisition (C)
	halt	Abort acquisition with no error (C)
	werr	Specify action when error occurs (C)
	werr	When error (P)

abort Terminate action of calling macro and all higher macros (C)

Syntax	abort	
Description	Terminates the action of the calling macro and all higher levels of nested macros. abort is used only in macros and not entered from the keyboard. It generates an error condition, which is the reason why the calling macro and any parent (nested) macros above will also be aborted. To exit from the execution of a macro without generating an error, use return.	
See also	VnmrJ User Programming	
Related	abortoff	Terminate normal functioning of abort in a macro (C)
	aborton	Restore normal functioning of abort in a macro (C)
	return	Terminate execution of a macro (C)

abortallacqs Reset acquisition computer in a drastic situation (C)

Syntax	abortallacqs
Description	Reboots the acquisition system from the host computer. Wait at least 30 seconds before attempting new acquisitions.
See also	NMR Spectroscopy User Guide
abortoff Terminate normal functioning of abort in a macro (C)

Syntax abortoff

- Description Changes the action of an abort command in a macro. Normally, abort (or any command aborting with an error condition) terminates the action of the calling macro and all higher levels of nested macros; however if the abortoff command is executed prior to a macro containing the abort command, only the macro containing abort terminates and execution continues to the next macro. The operation of the abortoff command is nullified by the aborton command. abortoff is used only in macros and not entered from the keyboard. See also VnmrJ User Programming
 - Related abort Terminate action of calling macro and all higher macros (C) aborton Restore normal functioning of abort in a macro (C)

aborton Restore normal functioning of abort in a macro (C)

Syntax	aborton	
Description	Nullifies the operation of a abortoff command and restores the normal functioning of the abort command. aborton is used only in macros and not entered from the keyboard.	
See also	VnmrJ User Programming	
Related	abortoff Terminate normal functioning of abort in a macro (C)	

abs Find absolute value of a number (C)

Syntax	abs(number)<:value>	
Description	Finds the absolute value of a number. Absolute value is a nonnegative number equal in numerical value to the given number (e.g., $abs(-6.5)$) is 6.5).	
Arguments	number is the given real number.	
	value is the return value with the absolute value of the given number. The default is to display the value in the status window.	
Examples	abs(-25) abs(n):abs val	
See also	VnmrJ User Programming	

AC1S-AC11S Autocalibration macros (M)

Syntax ACnS, where n is a number from 1 to 11.

- Description Performs automatic system calibration. When finished with the calibration routines, the current probe file is updated. If the probe is new to the system (i.e., all values in the probe file are zero), system power levels are determined followed by calibration. If power levels are listed in the current probe file, these values are used. The macro AC1S determines ¹H pw90, AC5S begins ¹³C calibration, including decoupler power calibrations. AC10S performs ¹⁹F calibration, and AC11S performs ³¹P calibration.
 - See also NMR Spectroscopy User Guide

ACbackup Make backup copy of current probe file (M)

Syntax	ACbackup		
Description	Called by the autocalibration macros AC1S-AC11S to back up the probe file after calibration ends. This macro is not usually called by the user.		
See also	NMR Spectroscopy User Guide		
Related	AC1S-AC11S Autocalibration macros (M)		

acct Writes records for operator login and logoff (M)

Applicability	VnmrJ		
Syntax	acct('start' 'done')		
Description	acct writes operator login and logoff records to the system adm/tmp/macrorecords.txt file used by the accounting package.		
See also	VnmrJ Installation and Administration manual		
Related	operator operator name (P)		
	operatorlogin	Sets work space and parameters for the operator	
		(M)	
	vnmr_accounting	Open Accounting window (U)	

ACreport Print copy of probe file after autocalibration (M)

Syntax	ACreport	
Description	Called by the autocalibration macros <u>AC1S-AC11S</u> to print a copy of the probe file before beginning a new autocalibration run.	
See also	NMR Spectroscopy User Guide	
Related	AC1S-AC11S Autocalibration macros (M)	

acos Find arc cosine of number (C)

Syntax	<pre>acos(value)<:n></pre>		
Description	Finds the arc cosine (also called the inverse cosine) of a number.		
Arguments	value is a number in the range of ± -1.0 to ± 1.0 .		
	n is a return argument giving the arc cosine, in radians, of value. The default is to display the arc cosine value in the status window.		
Examples	acos(.5) acos(value):acos_val		
See also	VnmrJ User Programming		
Related	sin Find sine value of an angle (C)		

acosy Automatic analysis of COSY data (C)

Syntax	acosy	
Description	Automatically analyzes a 2D COSY data set with fn=fn1 and sw=sw1. In this algorithm, a fuzzy pattern recognition technique is used to detect peaks and cluster the cross peaks into groups. Symmetry measures and chemical shifts for all cross peaks are calculated. Connectivities and the correlation table are displayed on the computer screen. This method is less sensitive to the threshold and rejects most artifacts in the peak list.	
See also	NMR Spect	roscopy User Guide
Related	acosyold	Automatic analysis of COSY data (C)
	fn	Fourier number in 1st indirectly detected dimension (P)
	fn1	Fourier number in directly detected dimension (P)
	112d	Automatic and interactive 2D peak picking (C)
	SW	Spectral width in directly detected dimension (P)
	sw1	Spectral width in 1st indirectly detected dimension (P)

acosyold Automatic analysis of COSY data, old algorithm (C)

Syntax	acosyold		
Description	Analyzes COSY data using an old algorithm.		
See also	NMR Spectroscopy User Guide		
Related	acosy	Automatic analysis of COSY data (C)	
	fn	Fourier number in 1st indirectly detected dimension (P)	
	fn1	Fourier number in directly detected dimension (P)	
	112d	Automatic and interactive 2D peak picking (C)	

SW	Spectral width in directly detected dimension (P)
sw1	Spectral width in 1st indirectly detected dimension (P)

Acquisition Done and Error Codes acq errors

Applicability VnmrJ 3.1

Description Whenever wbs, wnt, wexp, or werr processing occurs, the acquisition condition which initiated that processing is available from the parameter acqstatus. This acquisition condition is represented by two numbers, a "done" code and an "error" code. The done code is set in acgstatus[1] and the error code is set in acgstatus[2]. Macros may take different action depending on the acquisition condition. The done codes and error codes are listed below. As an example, a

> werr macro could specify special processing if the maximum number of transients of accumulated. The appropriate test in the macro would be:

if (acgstatus[2] = 200) then "do special processing, e.g. dp='y' au" endif

The acquisition error messages printed by Vnmr may be modified by creating an "acqerrmsgs" file with substitute messages. Each line in the file contains an error code followed by the text of the desired message. Error codes that do not occur in the acgerrmsgs file will continue to produce the standard messages. Vnmr first searches for

the acqerrmsgs file in the user's "\$vnmruser/templates" directory; if the file is not there Vnmr looks the the system directory

"\$vnmrsystem/user_templates". Entries are taken only from one file or the other, their contents are not "merged". A typical entry in the file would be:

301Can't spin the spinner

Error codes marked with an asterisk (*) are not used on Mercury and GEMINI 2000

Done Codes:

- 11. FID Complete.
- 12. Block Size Complete. (error code indicates BS # completed)
- 13. Soft Error.
- 14. Warning.
- 15. Hard Error.
- 16. Experiment Aborted.

17. Setup Completed. (error code indicates type of setup completed)

101. Experiment Complete.

102. Experiment Started.

Error codes:

Note: WARNINGS - Experiment acquisition continues.

SOFTERRORS - Experiment acquisition is stopped.

WARNINGS:

101. Low Noise Signal.

102. High Noise Signal.

103. ADC overflow occurred.

104. Receiver overflow occurred.*

SOFTERROR:

200. Maximum Transient Completed for Single Precision Data. WARNINGS or SOFTERRORS: (User selectable)

201. Lost Lock during experiment. (LOCKLOST)

Spinner Errors:

301. Sample failing to spin after three attempts of repositioning. (BUMPFAIL)

302. Spinner did not regulate in the allowed time period. $({\tt RSPINFAIL})^*$

303. Spinner went out of regulation during the experiment. $({\tt SPINOUT})^*$

395. Unknown Spinner device specified. (SPINUNKNOWN)*

396. Spinner device is not powered up. (SPINNOPOWER)*

 $397.\ \mathrm{RS}232$ Cable not connected between console and Spinner device.

(SPINRS232)*

398. Spinner does not acknowledge commands. (SPINTIMEOUT)* VT Errors:

400. VT did not regulate in the given time 'vttime' after being set.

401. VT went out of regulation during the experiment. (VTOUT)

402. VT is in manual mode after the automatic command given. (see oxford manual)*

403. VT Safety Sensor has reached limit. (see oxford manual)*

404. VT can not turn on cooling gas. (see oxford manual)*

405. VT main sensor on bottom limit. (see oxford manual)*

406. VT main sensor on top limit. (see oxford manual)*

407. VT sc/ss error. (see oxford manual)*

408. VT oc/ss error. (see oxford manual)*

495. Unknown VT device specified. (VTUNKNOWN)*

496. VT device is not powered up. (VTNOPOWER)*

497. RS232 Cable not connected between console and VT device. $(VTRS232)^*$

498. VT does not acknowledge commands. (VTTIMEOUT) SOFTERROR:

Sample Changer Errors:

- 501. Sample changer has no sample to retrieve.
- 502. Sample changer arm unable to move up during retrieve.
- 503. Sample changer arm unable to move down during retrieve.
- 504. Sample changer arm unable to move sideways during retrieve.
 - 505. Invalid sample number during retrieve.

 - 510. Robot arm failed to find home position during retrieve.*
 - 511. Sample tray size is not consistent.*
 - 512. Sample changer power failure during retrieve.*
 - 513. Illegal sample changer command during retrieve.*
 - 514. Gripper failed to open during retrieve.*
 - 515. Air supply to sample changer failed during retrieve.*
 - 525. Tried to insert invalid sample number.*
 - 526. Invalid temperature during sample changer insert.*
 - 527. Gripper abort during insert.*
 - 528. Sample out of range during automatic insert.
 - 529. Illegal command character during insert.*
 - 530. Robot arm failed to find home position during insert.*
 - 531. Sample tray size is not consistent.*
 - 532. Sample changer power failure during insert.*
 - 533. Illegal sample changer command during insert.*
 - 534. Gripper failed to open during insert.*
 - 535. Air supply to sample changer failed during insert.*
 - 593. Failed to remove sample from the magnet.*
 - 594. Sample failed to spin after automatic insert.
 - 595. Sample failed to insert properly.
 - 596. Sample changer not turned on.
 - 597. Sample changer not connected to RS-232 interface.
 - 598. Sample changer not responding.*

Shimming Errors:

- 601. Shimming User Aborted.*
- 602. Lost Lock while Shimming.*
- 604. Lock Saturation while Shimming.*

- 506. Invalid temperature during retrieve.
- 507. Gripper abort during retrieve.
- 508. Sample out of range during automatic retrieve.
- 509. Illegal command character during retrieve.*

608. A Shim Coil DAC limit hit while Shimming.*

Auto-Lock Errors:

701. User Aborted.(ALKABORT)*

702. Auto Lock Failure in finding resonance of sample. (ALKRESFAIL)

703. Auto Lock Failure in lock power adjustment. $(ALKPOWERFAIL)^*$

704. Auto Lock Failure in lock phase adjustment. (ALKPHASFAIL)*

705. Auto Lock Failure, lock lost in finial gain adjustment.

(ALKGAINFAIL)*

Auto-Gain Errors:

801. Auto-Gain failure, gain driven to zero, reduce pulse width (pw).

(AGAINFAIL)

HARDERRORS:

901. Incorrect PSG version for Acquisition.

 $902.\ \mbox{Sum-to-Memory Error}, \ \mbox{Number of points acquired not equal to np.}$

903. Fifo Underflow Error. (A delay too small?). *

904. Requested number of data points (np) to acquire is too large for acquisition.*

905. Acquisition Bus Trap (Experiment maybe lost). *

SCSI Errors

1001. Recoverable SCSI read transfer from Console Occurred. *

1002. Recoverable SCSI write transfer from Console Occurred. *

1003. Unrecoverable SCSI read transfer Error. *

1004. Unrecoverable SCSI write transfer Error. *

Host disk errors

1101. Error opening disk file. (most likely a Unix premission problem.)*

1102. Error on closing disk file.*

1103. Error on reading from disk file.*

1104. Error on writing to disk file.*

RF Monitor errors (only on Inova systems with RF monitor)

1400. An RF monitor trip occurred but the error status is OK

1401. Reserved RF monitor trip A occurred

1402. Reserved RF monitor trip B occurred

1404. Excessive reflected power at quad hybrid

1405. STOP button pressed at operator station

1406. Power for RF Monitor board (RFM) failed

Α

1407. Attenuator control or readback failed

1408. Quad reflected power monitor bypassed (Warning)

1409. Power supply monitor for RF Monitor board (RFM) bypassed (Warning)

1410. Ran out of memory to report RF monitor errors

1411. No communication with RF monitor system

1421. Reserved RF monitor trip A1 occurred on observe channel

1422. Reserved RF monitor trip B1 occurred on observe channel

1423. Reserved RF monitor trip C1 occurred on observe channel

1424. RF Monitor board (PALI/TUSUPI) missing on observe channel

1425. Excessive reflected power on observe channel

1426. RF amplifier gating disconnected on observe channel

1427. Excessive power detected by PALI on observe channel

1428. RF Monitor system (TUSUPI) heartbeat stopped on observe channel

1429. Power supply for PALI/TUSUPI failed on observe channel

1430. PALI asserted REQ_ERROR on observe channel (should never occur)

1431. Excessive power detected by TUSUPI on observe channel

1432. RF power amp: overdrive on observe channel

1433. RF power amp: excessive pulse width on observe channel

1434. RF power amp: maximum duty cycle exceeded on observe channel

1435. RF power amp: overheated on observe channel

1436. RF power amp: power supply failed on observe channel

1437. RF power monitoring disabled on observe channel (Warning)

1438. Reflected power monitoring disabled on observe channel (Warning)

1439. RF power amp monitoring disabled on observe channel (Warning)

1461. Reserved RF monitor trip A2 occurred on decouple channel 1462. Reserved RF monitor trip B2 occurred on decouple channel

1463. Reserved RF monitor trip C2 occurred on decouple channel

1464. RF Monitor board (PALI/TUSUPI) missing on decouple channel

1465. Excessive reflected power on decouple channel

1466. RF amplifier gating disconnected on decouple channel

1467. Excessive power detected by PALI on decouple channel

1468. RF Monitor system (TUSUPI) heartbeat stopped on decouple channel

1469. Power supply for PALI/TUSUPI failed on decouple channel

1470. PALI asserted REQ_ERROR on decouple channel (should never occur)

1471. Excessive power detected by TUSUPI on decouple channel

1472. RF power amp: overdrive on decouple channel

1473. RF power amp: excessive pulse width on decouple channel

1474. RF power amp: maximum duty cycle exceeded on decouple channel

1475. RF power amp: overheated on decouple channel

1476. RF power amp: power supply failed on decouple channel

1477. RF power monitoring disabled on decouple channel (Warning)

1478. Reflected power monitoring disabled on decouple channel (Warning)

1479. RF power amp monitoring disabled on decouple channel (Warning)

acqdequeue Dequeue an acquisition

Syntax	acqdequeue<:\$ret> - dequeue acquisition from current experiment		
	<pre>acqdequeue<('go_id')><:\$ret> - dequeue an acquisition</pre>		
Applicability	VnmrJ 3.1		
Description	When a go, ga, or au command is issued, instructions are sent to t		
	acquisition system to run that experiment. If another experiment is		

acquisition system to run that experiment. If another experiment is already running, the request is queued. When the prior experiment finishes, the queued acquisition will start. The acqdequeue command will remove an experiment from this queue. The acqdequeue command will not stop an experiment that is already started. An optional return argument will be set to 1 if the experiment is successfully dequeued; otherwise it will be set to 0.

Arguments When a go, ga, or au command is issued, a unique identifier is added to the parameter set, in the processed tree. This parameter is named 'go_id'. This parameter can be used as an argument for the acqdequeue command. If no argument is given, the value of this parameter in the current experiment's processed tree is used.

acqdisp Display message on the acquisition status line (C)

Syntax	acqdisp(message)		
Description	Displays the message specified on the acquisition status line. acqdisp		
	is used primarily by the acquisition process to update the screen.		
Arguments	message is a text string, up to 8 characters long.		

acgi Interactive acquisition display process (C)

Syntax acgi<('par'|'disconnect'|'exit'|'standby')><:\$ret> Description Opens the Acquisition window for interactive locking and shimming on the lock signal, FID, or spectrum. When using a spectrometer, acqi normally automatically starts. On all systems, if the console has been recently rebooted, enter su before running acqi. If acgi is connected to the console and you start an acquisition (su/go/au), acqi automatically disconnects. The pulse sequence and parameter set for the FID/spectrum display can be selected by entering gf. Note that if clicking the FID button in acqi causes acqi to "disconnect," the common cause is that gf had not been executed. The FID display is controlled by the parameters lsfid, phfid, and dmgf. These display parameters are automatically sent to acgi when acqi is first invoked. These parameters may subsequently be changed and sent again to acqi with the command acqi ('par'). If phfid is not set to "Not Used" for the FID display in acqi, a slide control will be available in acqi for the interactive adjustment of the phfid parameter. The slide will be in the IPA set of adjustments. If the parameter dmgf exists and is set to 'av', the FID display in acqi displays the square root of the sum of the squares of the real and imaginary channels. The spectrum display is controlled by parameters sp, wp, dmg, rp, lp, rfl, rfp, vs, vp, sw, and fn. These parameters are automatically sent to acqi when acqi is first invoked. These parameters can subsequently be changed and sent again to acqi with the command acqi('par'). The preparation macro gf also calls acqi('par'), thereby causing these parameters to be sent to acqi. If fn is greater than 64K, it is lowered to 64K. A convenient method of setting these parameters is to acquire a spectrum with go, then ft and adjust the display with the ds command options. Once the display is set the way you want, enter gf. The same display should then appear when the spectrum display is selected from acqi. Note that weighting parameters are not used in the *acqi* spectrum display.

The manual NMR Spectroscopy User Guide has a step-by-step description of using acqi.

Arguments 'par' causes the current values of parameters lsfid, phfid, dmgf, sp, wp, dmg, rp, lp, rfl, rfp, vs, sw, and fn to be sent to acqi.

'disconnect' causes acqi to be disconnected. Clicking the Close button in acqi is equivalent, and puts acqi in the standby mode. Lock parameters, the spin parameter, and the shim values are sent back to the current experiment when acqi is "disconnected." If the experiment

has the load parameter set to 'y', then the shim values are not delivered to the experiment. 'exit' causes an exit from acqi. Clicking the exit button in the Acquisition window is equivalent. \$ret is a return value with the success or failure of running acqi. The default is a warning displayed in the status window if acqi fails. 'standby' starts acgi and puts it into the standby mode. Examples acgi acqi('par') acqi('disconnect') acqi('exit') acqi:\$ok See also NMR Spectroscopy User Guide Related Acqstat Bring up the acquisition status display (U) Display mode in directly detected dimension (P) Absolute-value display of FID data or spectrum in acqi dmgf (P)

- dsDisplay a spectrum (C)fnFourier number in directly detected dimension (P)
- ft Fourier transform 1D data (C)
- gf Prepare parameters for FID/spectrum display in acqi (M)
- go Submit an experiment to acquisition (C)
- load Load status of displayed shims (P)
- lkof Track changes in lock frequency (P)
- 1p First-order phase in directly detected dimension (P)
- lsfid Number of complex points to left-shift the np FID (P)
- phfid Zero-order phasing constant for np FID (P)
- rfl Ref. peak position in 1st indirectly detected dimension (P)
- rfp Ref. peak frequency in directly detected dimension (P)
- rp Zero-order phase in directly detected dimension (P)
- sp Start of plot in directly detected dimension (P)
- spin Sample spin rate (P)
- sw Spectral width in directly detected dimension (P)
- vp Vertical position of the spectrum (P)
- vs Vertical scale (P)
- WP Width of plot in directly detected dimension (P)

acqmeter Open Acqmeter window (M)

Syntax acqmeter<(remote_system)>

Description Opens the Acqmeter window and shows a time line of lock level, temperature (VT), and/or spinner speed. When first opened, only lock level is displayed. By clicking anywhere in the lock level window with

 Arguments remote_system is the host name of a remote machine on the same network. The default is the local machine. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file). Examples acqmeter acqueter ('nmr500') See also NMR Spectroscopy User Guide Related acqi Interactive acquisition display (C) Acqmeter Open Acqmeter window (U) 		the right mou level window, open Click on the mouse button graphical mo window with or close window	use button, a menu pops up with choices to close the lock c, show a temperature (VT) window, show a spinner n a properties window, or close the Acqmeter window. choice desired in the menu with either the left or right n. In the properties window, the host, font, color, and ode can be changed. Continue to click in any Acqmeter the right mouse button to open the menu and then open dows, or close the Acqmeter window, as desired.	
Examples acqmeter acqmeter('nmr500') See also NMR Spectroscopy User Guide Related acqi Interactive acquisition display (C) Acqmeter Open Acqmeter window (U)	Arguments	remote_system is the host name of a remote machine on the same network. The default is the local machine. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file).		
See alsoNMR Spectroscopy User GuideRelatedacqiAcqmeterInteractive acquisition display (C)AcqmeterOpen Acqmeter window (U)	Examples	acqmeter acqmeter('nmr500')		
RelatedacqiInteractive acquisition display (C)AcqmeterOpen Acqmeter window (U)	See also	NMR Spectroscopy User Guide		
	Related	acqi Acqmeter	Interactive acquisition display (C) Open Acqmeter window (U)	

Acqmeter Open Acqmeter window (U)

Syntax Acqmeter <remote_system> <-f file> <&>

- Description Opens the Acqmeter window and shows a time line of lock level, temperature (VT), and/or spinner speed. When first opened, only lock level is displayed. By clicking anywhere in the lock level window with the right mouse button, a menu pops up with choices to close the lock level window, show a temperature (VT) window, show a spinner window, open a properties window, or close the Acqmeter window. Click on the choice desired in the menu with either the left or right mouse button. In the properties window, the host, font, color, and graphical mode can be changed. Continue to click in any Acqmeter window with the right mouse button to open the menu and then open or close windows, or close the Acqmeter window, as desired.
- Arguments remote_system is the host name of a remote machine on the same network. The default is the local machine. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file).

-f file is the name of a template file in the directory \$vnmruser/vnmrsys/templates/acqstat used to set the attributes of the Acqmeter window when it opens. This allows customizing the Acqmeter window for different users and experiments. The default name of the file is default.

& (ampersand) character added to the command makes Acqmeter into a background process. For example, if "lab" is the remote machine host

name, entering the command Acqmeter lab & displays the acquisition status of the "lab" remote machine as a background process. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file).

Examples	Acqmeter &			
	Acqmeter nmr400 &			
	Acqmeter gem300 -f inova500.lisa &			
See also	NMR Spectroscopy User Guide			
Related	acqi Interactive acquisition display (C)			
	acqmeter Open Acqmeter window (M)			

acqmode Acquisition mode (P)

Description	A global parameter specifying the normal acquisition mode for acquiring, locking, fid shimming, and prescan in VnmrJ.	
Values	' ' (empty string) normal acquisition	
	'lock' lock acquisition	
	'fidscan' fid shimming acquisition	
	'prescan' prescan acquisition	
See also	VnmrJ Imaging, User Guide, NMR Spectroscopy User Guide	

acgreserve Reserve the acquisition console for the current owner

Syntax	acqreserve
Applicability	VnmrJ 3.1
Description	acqreserve controls reservation of the NMR acquisition console, allowing a user sole access. It reserves the console for the current user, as specified by the owner parameter. This user / owner will have access to the acquisition commands to acquire data, lock, shim, set temperature, etc. If the console is reserved, any VnmrJ session with a different user / owner will be forced into a datastation mode. Access to acquisition related commands and acquisition related panels will be prevented in this datastation mode.
	By default, a console reservation will be removed when the user / owner exits from the current VnmrJ session. Using the acqreserve('on','noAutoOff') option causes the reservation to remain intact, even after the user / owner exits. For example, they will maintain the console reservation while they travel and start a new session at a remote site. If they do not exit from the first VnmrJ session and start a second session somewhere else, that second session will share the reservation. The reservation does not need to be turned

Α

off from the VnmrJ that started it. The capitalization in the second argument is ignored. The argument 'noautooff' also works. The invocation acqreserve('autooff') is used when the user exits. This will turn off the reservation, as long as it was not turned on with the 'noAutoOff' option.

A force option will turn the reservation on or off, even if the current user / owner is not the one that made the original reservation. A record will be kept of forced reservation events.

If acqreserve is never used, or after acqreserve('off') is issued, access to the console will be available on a first come first served basis. As soon as the console becomes "Idle", any user not in datastation mode will be able to access it.

Arguments acqreserve takes up to three optional arguments. The first argument is 'on', 'off', or 'autooff'. If no argument is given, the default is 'on'. acqreserve('on') makes the reservation. acqreserve('off') removes the reservation. acqreserve('autooff') is described below. The other optional arguments are 'noAutoOff' and 'force'. They can be provided in either order, following the 'on', 'off', or 'autooff' argument.

acqstat Open Acquisition Status window (M)

Syntax acqstat<(remote_system)>

- Description Opens the Acquisition Status window, which displays acquisition information such as the current acquisition task, experiment number, spinner status, and temperature status. When the host computer is attached to a spectrometer, this window should open automatically when VnmrJ is started. In the properties window, the host, font, color, and graphical mode can be changed. For a complete description of these windows, refer to the manual *NMR Spectroscopy User Guide*.
- Arguments remote_system is the host name of a remote machine on the same network. The default is the local machine. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file).

Examples acqstat acqstat('u500') See also NMR Spectroscopy User Guide

Related Acqstat Open the Acquisition Status window (U) showstat Display information about status of acquisition (C,U)

Acqstat Open Acquisition Status window (U)

Syntax Acqstat <remote_system> <-f file> <&>
Description Opens the Acquisition Status window, which displays acquisition
information such as the current acquisition task, experiment number,
spinner status, and temperature status. When the host computer is
attached to a spectrometer, this window should open automatically
when VnmrJ is started. In the properties window, the host, font, color,
and graphical mode can be changed. For a complete description of
these windows, refer to the manual NMR Spectroscopy User Guide.

Arguments remote_system is the host name of a remote machine on the same network. The default is the local machine. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file).

> -f file is the name of a template file in the directory \$vnmruser/vnmrsys/templates/acqstat used to set the attributes of the Acquisition Status window when it opens. This allows customizing the Acquisition Status window for different users and experiments. The default name of the file is default.

> & (ampersand) character added to the command makes Acqstat into a background process. For example, if "lab" is the remote machine host name, entering the command Acqstat lab & displays the acquisition status of the "lab" remote machine as a background process. To activate the remote feature, the local and remote machines must be on the same Ethernet LAN (local area network) and the local machine must be able to get the Internet address of the remote machine (usually in the /etc/hosts file).

Examples Acqstat & Acqstat nmr400 & Acqstat gem300 -f inova500.lisa & See also NMR Spectroscopy User Guide

Related AcqstatOpen the Acquisition Status window (U)showstatDisplay information about status of acquisition
(C,U)

acqstatus Acquisition status (P)

Description Whenever wbs, wnt, wexp, or werr processing occurs, the acquisition condition that initiated that processing is available from the parameter acqstatus. This acquisition condition is represented by two numbers, a "done" code and an "error" code. The done code is set in acqstatus[1] and the error code is set in acqstatus[2]. Macros can take different actions depending on the acquisition condition. The done codes and error codes are listed below and in the file acq_errors in /vnmr/manual. For example, a werr macro could specify special processing if the maximum number of transients is accumulated. The appropriate test in the macro would be:

if (acgstatus[2] = 200) then "do special processing, e.g. dp='y' au" endif

Done codes:

- 11. FID complete
- 12. Block size complete (error code indicates bs number completed)
- 13. Soft error
- 14. Warning
- 15. Hard error
- 16. Experiment aborted
- 17. Setup completed (error code indicates type of setup completed)
- 101. Experiment complete
- 102. Experiment started

Error codes:

Warnings

- 101. Low-noise signal
- 102. High-noise signal
- 103. ADC overflow occurred
- 104. Receiver overflow occurred*
- Soft errors
- 200. Maximum transient completed for single-precision data
- 201. Lost lock during experiment (LOCKLOST)

300. Spinner errors:

- 301. Sample fails to spin after three attempts at repositioning
- 302. Spinner did not regulate in the allowed time period (RSPINFAIL)*
- 303. Spinner went out of regulation during the experiment (SPINOUT)*
- 395. Unknown spinner device specified (SPINUNKNOWN)*
- 396. Spinner device is not powered up (SPINNOPOWER)*
- 397. RS-232 cable not connected from console to spinner (SPINRS232)*
- 398. Spinner does not acknowledge commands (SPINTIMEOUT)*
- 400. VT (variable temperature) errors:
- 400. VT did not regulate in the given time vttime after being set
- 401. VT went out of regulation during the experiment (VTOUT)

402. VT in manual mode after automatic command (see Oxford manual)*

- 403. VT safety sensor has reached limit (see Oxford manual)*
- 404. VT cannot turn on cooling gas (see Oxford manual)*
- 405. VT main sensor on bottom limit (see Oxford manual)*

406. VT main sensor on top limit (see Oxford manual)*

407. VT sc/ss error (see Oxford manual)*

- 408. VT oc/ss error (see Oxford manual)*
- 495. Unknown VT device specified (VTUNKNOWN)*
- 496. VT device not powered up (VTNOPOWER)*
- 497. RS-232 cable not connected between console and VT (VTRS232)*
- 498. VT does not acknowledge commands (VTTIMEOUT)

- 500. Sample changer errors:
- 501. Sample changer has no sample to retrieve
- 502. Sample changer arm unable to move up during retrieve
- 503. Sample changer arm unable to move down during retrieve
- 504. Sample changer arm unable to move sideways during retrieve
- 505. Invalid sample number during retrieve
- 506. Invalid temperature during retrieve

507. Gripper abort during retrieve

- 508. Sample out of range during automatic retrieve
- 509. Illegal command character during retrieve*
- 510. Robot arm failed to find home position during retrieve*
- 511. Sample tray size is not consistent*
- 512. Sample changer power failure during retrieve*
- 513. Illegal sample changer command during retrieve*
- 514. Gripper failed to open during retrieve*
- 515. Air supply to sample changer failed during retrieve*
- 525. Tried to insert invalid sample number*
- 526. Invalid temperature during sample changer insert*
- 527. Gripper abort during insert*
- 528. Sample out of range during automatic insert
- 529. Illegal command character during insert*
- 530. Robot arm failed to find home position during insert*
- 531. Sample tray size is not consistent*
- 532. Sample changer power failure during insert*
- 533. Illegal sample changer command during insert*
- 534. Gripper failed to open during insert*
- 535. Air supply to sample changer failed during insert*
- 593. Failed to remove sample from magnet*
- 594. Sample failed to spin after automatic insert
- 595. Sample failed to insert properly
- 596. Sample changer not turned on
- 597. Sample changer not connected to RS-232 interface
- 598. Sample changer not responding*
- 600. Shimming errors:
- 601. Shimming user aborted*
- 602. Lost lock while shimming*
- 604. Lock saturation while shimming*
 - 608. A shim coil DAC limit hit while shimming*
 - 700. Autolock errors:
- 701. User aborted (ALKABORT)*
- 702. Autolock failure in finding resonance of sample (ALKRESFAIL)
- 703. Autolock failure in lock power adjustment (ALKPOWERFAIL)*
- 704. Autolock failure in lock phase adjustment (ALKPHASFAIL)*
- 705. Autolock failure, lock lost in final gain adjustment (ALKGAINFAIL)*
- 800. Autogain errors.
- 801. Autogain failure, gain driven to 0, reduce pw (AGAINFAIL)

Hard errors

- 901. Incorrect PSG version for acquisition
- 902. Sum-to-memory error, number of points acquired not equal to np 903. FIFO underflow error (a delay too small?)*

	904. Requested number of data points (np) too large for acquisition* 905. Acquisition bus trap (experiment may be lost)*
	 1000. SCSI errors: 1001. Recoverable SCSI read transfer from console* 1002. Recoverable SCSI write transfer from console** 1003. Unrecoverable SCSI read transfer error* 1004. Unrecoverable SCSI write transfer error*
	 1100. Host disk errors: 1101. Error opening disk file (most likely a UNIX permission problem)* 1102. Error on closing disk file* 1103. Error on reading from disk file* 1104. Error on writing to disk file*
See also	NMR Spectroscopy User Guide
Related	<pre>react Recover from error conditions during werr processing (M) werr Specify action when error occurs (C) werr When error (P)</pre>

acquire Acquire data (M)

See

Description	Macro to acquire data. It uses execpars to select the prep and prescan method, executes them, and then begins acquisition.		
See also	NMR Spectroscopy User Guide		
Related	execpars	Set up the exec parameters (M)	
	execprescan	Execute prescan macro (P)	
	xmnext	Find next prescan or next experiment in study queue (M)	
	xmwexp	Processing macro for end of acquisition in study queue (M)	

actionid Current study queue node id (P)

Applicability	Liquids, Imaging	
Description	Specifies the currently selected study queue node id.	
See also	VnmrJ Imaging, User Guide, NMR Spectroscopy User Guide	
Related	xmactionPerform study queue action (M)xmnextFind next prescan or next experiment in study qu (M)	
	xmselect	Action when study queue node is selected (M)

activestudy Active study name (P)

Applicability	Liquids, Imaging		
Description	A global parameter that specifies the currently active study name. In the Walkup interface, it specifies the currently active automation run.		
Values	's_20050601' active study name		
	'auto_2005.06.01	' active automation run name	
	'null' no active s	tudy or automation run	
See also	VnmrJ Imaging, Use	er Guide and NMR Spectroscopy User Guide	
Related	acquire	Acquire data (M)	
	autodir	Automation directory absolute pathname (P)	
	cqinit	Initialize liquids study queue (M)	
	studyid	Study identification (P)	
	xmaction	Perform study queue action (M)	
	xmselect	Action when study queue node is selected (M)	

add Add current FID to add/subtract experiment (C)

Syntax	clradd<:\$stat,\$message> – clear the add/subtract experiment
	jaddsub – join the add/subtract experiment
	add<(multiplier <,'new'>)> $-$ add the current FID to the add/subtract experiment
	sub<(multiplier <,'new'>)> — subtract the current FID from the add/subtract experiment
	add<(multiplier <,'trace',index>)> $-$ add the current FID to the "index" element in the add/subtract experiment
	sub<(multiplier <,'trace',index>)> — subtract the current FID from the "index" element in the add/subtract
	add subexp $-$ global integer identifying the add/subtract experiment. The default is 5
Description	Adds the last displayed or selected FID to the current contents of the add/subtract experiment (exp5). An optional argument allows the FID to be first multiplied by a 'multiplier'.
	The FID data are divided by the number of time averages of the data, reflected in the parameter ct. To get unscaled data, use a multiplier of ct.
	The add and sub commands use the cexp command to create the add-subtract experiment. They take the same return values as the cexp command. These can be used to suppress messages. See "cexp Create aVnmr experiment (M)" on page 176 for a description of the return values.

The parameters lsfid and phfid can be used to shift or phase rotate the selected FID before it is combined with the data in the add/subtract experiment. A multi-FID add/subtract experiment can be created by using the 'new' keyword. Individual FIDs in a multi-FID add/subtract experiment can subsequently be added to using the 'trace' keyword followed by the index number of the FID.

"clradd" deletes the add-subtract experiment. The add-subtract experiment number is defined by the global addsubexp parameter. The clradd program uses the delexp command to delete the add-subtract experiment. It takes the same return values as the delexp command. These are used to suppress messages. See "delexp Delete an experiment (M)" on page 241 for a description of the return values.

"jaddsub" joins the add-subtract experiment, as defined by the global addsubexp parameter. jaddsub creates this parameter if it does not exist, and sets it to a default value of 5. jaddsub with an argument, as in jaddsub('silent') will not clear the graphics, text window, or menu system. It does not matter what the argument is.

A multi-fid add/subtract experiment can be created with the add or sub command. The optional argument 'new' will create a new FID element in the add/subtract experiment. For example, the commands clradd select(1) add from some experiment will create the add/subtract experiment with a single FID in it.

If the next commands typed are select(2) add, then a single FID which is the sum of the original FIDs one and two will be made in the add/subtract experiment. If the commands select(2) add('new') were typed, then the add/subtract experiment will contain an array of two FIDs corresponding to the original FIDs one and two, respectively.

The arraydim parameter may need to be updated after constructing a multi-fid add/subtract experiment. To do this, join the add/subtract experiment (jaddsub) and enter

setvalue('arraydim', <num>, 'processed')

where <num> is replaced by the number of FIDs in that experiment.

For example, if twelve FIDs were put into the add/subtract experiment, enter

setvalue('arraydim',12,'processed')

Individual FIDs in a multi-fid add/subtract experiment may subsequently be added to and subtracted from. The add and sub command without a 'trace' argument will add or subtract from the first FID in the add/subtract experiment.

Adding the 'trace' argument followed by a required index number will select another FID to be the target of the add/subtract.For example, select(4) add('trace',6) will take the fourth FID from the current experiment and add it to the sixth FID in the add/subtract experiment When using the 'trace' argument, that FID must already exist in the add/subtract experiment by using an appropriate number of add('new') or sub('new') commands.

Arguments	multiplier is a value that the FID is to be multiplied by before being added to the add/subtract experiment (exp5). The default is 1.0.		
	'new' is a keyword to create a new FID element in a add/subtract experiment.		
	'trace' is a keyword to use the next argument (index) as the number of the FID to add to in an add/subtract experiment. The default is to add to the first FID in a multi-FID add/subtract experiment.		
	index is the index number of the FID to be used as a target in a multi-FID add/subtract experiment.		
Examples	add clradd:\$stat,\$message		
	add:\$stat,\$message		
	add(0.75) add('new') add('trace',2)		
See also	NMR Spectroscopy User Guide		
Related	<pre>clradd Clear add/subtract experiment (C) lsfid Number of complex points to left-shift ni interferogram (P) phfid Zero-order phasing constant for np FID (P) select Select a spectrum without displaying it (C) spadd Add current spectrum to add/subtract experiment (C) sub Subtract current FID from add/subtract experiment (C)</pre>		

addi Start interactive add/subtract mode (C)

Syntax addi

Description Starts the interactive add/subtract mode. Before entering addi, start the process with clradd and spadd, then display a second spectrum on the screen. This may involve changing experiments, selecting a second member of an array of spectra, a different trace of a 2D spectrum, or displaying a spin simulated spectrum. The Fourier numbers (fn) *must* be the same in the two spectra to be manipulated. The width (sw) of the two spectra need *not* be identical, although adding spectra of different widths will probably not be meaningful. Having selected the second spectrum and ensuring it is in nm mode, enter addi to begin the interactive process.

> After addi is invoked, spectrum 1, the spectrum selected by the spadd command, appears in the center of the display. Spectrum 2, the spectrum that was active when addi was entered, appears on the bottom. The sum or difference of these spectra appears on top of the screen. When addi is first entered, this spectrum will be the sum (1 + 2) by default. The spectra is manipulated using the mouse.

The select button toggles between different modes of control.

- When the label at the screen bottom reads "active: current", all of the parameters (except wp) control spectrum 2, and spectrum 2 can be phased, scaled, or shifted relative to spectrum 1.
- After clicking on select, the label at the screen bottom reads "active: addsub", and now all of the parameters except wp control spectrum 1.
- Clicking select again toggles the label to read "active: result", and now parameter changes affect only the sum or difference spectrum.

Note that wp always controls all spectra, because differential expansions of the two spectra are not supported. Note also that the colors of the labels change to match the colors of the different spectra.

The sum/difference spectrum displayed on the screen while addi is active is strictly a temporary display. Once all manipulations have been performed, and assuming the sum/difference is something you wish to perform further operations with (such as plotting), it must be saved into the add/subtract experiment (exp5) by clicking on save. At this point, spectrum 1, which was in the add/subtract experiment, is overwritten by the sum or difference spectrum, and addi ceases operation. In most cases, you will next want to enter jexp5 ds to display the difference spectrum on the screen, ready for further manipulation (expansion, line listing, etc.) and plotting. If you wish to continue with the add/subtract process by adding in a third spectrum, display that spectrum in the usual way and enter addi again.

See also NMR Spectroscopy User Guide

Related	clradd	Clear add/subtract experiment (C)	
	jexp	Join existing experiment (C)	
	nm	Select normalized intensity mode (C)	
	spadd	Add current spectrum to add/subtract experiment (C)	
	spmin	Take minimum of two spectra in add/subtract experiment	
		(C)	
	spsub	Subtract current spectrum from add/subtract experiment	
		(C)	
	wp	Width of plot in directly detected dimension (P)	

addnucleus Add new nucleus to existing probe file (M)

Applicability	ALL	
Description	Entries for nuclei not in the default probe file are appended to the end of the file. The argument should correspond to a nucleus in the nuctable.	
Syntax	addnucleus('nucleus')	
Arguments	nucleus - name followed by atomic number, e.g. C13 not 13C.	
Examples	addnucleus('Si29')	

See also	oNMR Spectroscopy User GuideedaddprobeCreate new probe directory and probe file (
Related			
	deletenucleus	Removes nucleus entry to probe file (M)	
	getparam	Receive parameter from probe file (M)	
	probe	Probe type (P)	
	setparams	Write parameter to current probe file (M)	

addpar Add selected parameters to current experiment (M)

Syntax	addpar<('2d' '3d' '3rf' '4d' 'downsamp' 'fid' 'image' '112d' '1p'<,dim> 'oversamp' 'ss')>	
Applicability	The '3d', '3rf', '4d', 'fid', and 'image' arguments work on all systems but are only useful if system has the proper hardware.	
Description	Creates selected parameters in the current experiment.	
Arguments	If no argument is entered, addpar displays instructions for its use.	
	'2d', '3d', '3rf', '4d', 'downsamp', 'fid', 'image', 'll2d', 'lp', 'oversamp', and 'ss' are keywords (only one keyword is used at a time) specifying the parameters to be created:	
	• '2d' specifies creating ni, phase, and sw1, which can be used to acquire a 2D data set (functions the same as macro par2d).	
	• '3d' specifies creating d3, ni2, phase2, and sw2, which can used to acquire a 3D data set (functions the same as macro par3d).	
	• '3rf' specifies retrieving the ap and dg2 display templates for third rf channel and 3D parameters (functions the same as macro par3rf).	
	• '4d' specifies creating the acquisition parameters d4, ni3, phase3, and sw3, which can be used to acquire a 4D data set (functions the same as macro par4d).	
	• 'downsamp' specifies creating the parameters downsamp, dscoef, dslsfrq, dsfb, and filtfile for digital filtering and downsampling (functions the same as macro pards).	
	• 'fid' specifies creating FID display parameters axisf, crf, deltaf, dotflag, vpf, and vpfi if the parameter set is older and lacks these parameters (functions the same as macro fidpar).	
	• 'll2d' specifies creating th2d and xdiag for the ll2d 2D peak picking program (functions the same as macro parll2d).	
	• 'lp' specifies creating lpalg, lpopt, lpfilt, lpnupts, strtlp, lpext, strtext, lptrace, and lpprint for linear prediction in the acquisition dimension (functions the same as macro parlp). The display template for the dglp macro is also created if necessary.	
	• 'oversamp' specifies creating parameters def_osfilt, filtfile, oscoef, osfb, osfilt, oslsfrq, and oversamp for oversampling and digital filtering (functions the same as macro paros).	

	•'ss' spect and sslsf	fies adding parameters ssorder, ssfilter, ssntaps, irg for time-domain solvent subtraction (functions the	
	same as macro partidss).		
	dim specifi 1 for the f dimension. addpar('1 creates lpo strtext1, lpalg2, lp lptrace2, multidimer	es the dimension when adding linear prediction parameters: irst implicit dimension or 2 for the second implicit Default is the acquisition dimension. Therefore, lp') creates the parameters listed above; addpar('lp',1) alg1, lpopt1, lpfilt1, lpnupts1, strtlp1, lpext1, lptrace1, and lpprint1; and addpar('lp',2) creates popt2, lpfilt2, lpnupts2, strtlp2, lpext2, strtext2, and lpprint2. Each separate dimension of a usional data set can have its own unique parameters.	
Examples	addpar	addpar	
	addpar('3	3d')	
	addpar('	lp',1)	
See also	NMR Spectroscopy User Guide; VnmrJ Imaging NMR		
Related	def_osfi Default value of osfilt (P)		
	lt		
	fidpar	Add parameters for FID display in current experiment (M)	
	ostilt	Oversampling filter for real-time DSP (P)	
	par2d	Create 2D acquisition parameters (M)	
	par3d	Create 3D acquisition parameters (M)	
	par3rf	Get display templates for 3rd rf channel parameters (M)	
	par4d	Create 4D acquisition parameters (M)	
	pards	Create digital filtering and downsampling parameters (M)	
	parfidss	Set up parameters for time-domain solvent subtraction (M)	
	paros	Create oversampling and digital filtering parameters (M)	
	parll2d	Create parameters for 2D peak picking (M)	
	parlp	Create parameters for linear prediction (M)	

addparams Add parameter to current probe file (M)

Syntax	<pre>addparams(param,value,nucleus<,'tmplt'><,'system'>)</pre>	
Description	Adds a new parameter and its value for a specified nucleus to the probe file or to the probe template.	
Arguments	param is the name of the parameter to be added.	
	value is a string with the value to be written for the parameter.	
	nucleus is the nucleus to add in the probe file.	
	'tmplt' is a keyword to add the parameter to the local template. Th default is the probe file.	
	'system' is a keyword to add the parameter to the system-level template or probe file, provided that you have write permission to that file. The default is to add the parameter to the local template or probe file.	

addprobe Create new probe directory and probe file (M)

Syntax	addprobe(probe_name<,'stdar' 'system'><,'stdpar'>)	
Description	Creates a new probe directory and a probe file. Default nuclei included in this file are 1 H, 19 F, 13 C, and 15 N. The information is saved in the user's directory vnmrsys/probes.	
Arguments	probe_name is the name to be given to the probe directory and probe file.	
	'stdpar' and 'system' are keywords for the second and third arguments:	
	• If the second argument is 'stdpar', calibration values from the standard parameter sets (stdpar/H1.par, stdpar/C13.par, etc.) will be read and written into the probe file.	
	• If the second argument is 'system' and the user has write permission into the VnmrJ system probes directory (typically /vnmr/probes), then a system-level probe directory will be made.	
	• If the second argument is 'system' and the third argument is 'stdpar', then both actions in the preceding bullets will occur.	
	• The default is the probe file is created with all parameters initialized to zero.	
Examples	addprobe('idpfg') addprobe('idpfg','stdpar') addprobe('idpfg','system','stdpar')	
See also	NMR Spectroscopy User Guide; VnmrJ Automation User Guide	
Related	addnucleusAdd new nucleus to existing probe file (M)deletenucleusRemoves nucleus entry to probe file (M)getparamReceive parameter from probe file (M)probeProbe type (P)setparamsWrite parameter to current probe file (M)	

adept Automatic DEPT analysis and spectrum editing (C)

Syntax adept<(<'noll'><,'coef'><,'theory'>)>

Description	Automatically analyzes a set of four DEPT spectra and edits the spectra
	so that the spectra is arrayed as follows:

- #4 is CH₃ carbons only
- #3 is CH₂ carbons only
- #2 is CH carbons only
- #1 is all protonated carbons

Because adept modifies the transformed data, it should not be repeated without retransforming the data between calls. adept produces a text file dept.out in the current experiment directory, which contains the result of the analysis.

Arguments The following keyword arguments can be supplied in any order:

'noll' causes the line listing to be skipped. If 'noll' is not supplied as an argument, adept first performs a line listing. In that case, the threshold parameter th must be set properly before starting adept.

'coef' causes the combination coefficients to be printed.

'theory' causes theoretical coefficients to be used. The default is optimized coefficients.

- Examples adept
 adept('coef')
 adept('theory','noll')
- See also NMR Spectroscopy User Guide

RelatedautodeptAutomated complete analysis of DEPT data (M)DeptSet up parameters for DEPT experimentdeptprocProcess DEPT data (M)padeptPerform adept analysis and plot resulting spectra
(C)pldeptPlot DEPT data, edited or unedited (M)thThreshold (P)

aexpp1 Automatic plot of spectral expansion (M)

Syntax	aexppl<(expansion_factor)>	
Description	Plots automa defined first in ds.	tically expansions of given regions. Regions have to be by using the region command or by using the cursors
Arguments	expansion_ The default i	factor is a spectral expansion factor in units of Hz/mm. s 2 Hz/mm.
Examples	aexppl aexppl(20)	
See also	NMR Spectroscopy User Guide	
Related	ds region	Display a spectrum (C) Divide spectrum into regions (C)

ai Select absolute-intensity mode (C)

Syntax ai

Description	Selects the <i>absolute-intensity display mode</i> in which the scale is kept constant from spectrum to spectrum to allow comparison of peak heights from one spectrum to another. The alternative is the normalized-intensity display mode (nm) in which spectra are scaled so that the largest peak in the spectrum is vs mm high. The modes are mutually exclusive—the system is always in either nm or ai mode. Enter aic ² to determine which mode is currently active
	arg: to determine which mode is currently active.

See also NMR S	Spectroscopy	User	Guide
----------------	--------------	------	-------

Related	aig	Absolute intensity group (P)
	nm	Select normalized-intensity mode (C)
	VS	Vertical scale (P)

aig Absolute-intensity group (P)

Description	Contains the result of the ai or nm command. aig is not set in the usual way but can be queried (aig?) to determine which display mode is active.	
Values	'ai' indicates the absolute-intensity display mode is active.	
	'nm' indicate	es the normalized-intensity display mode is active.
See also	NMR Spectroscopy User Guide	
Related	ai	Select absolute intensity mode (C)
	dmg	Display mode in directly detected dimension (P)
	nm	Select normalized-intensity mode (C)
	? Display individual parameter value (C)	

aipDisplay Display images (C)

Syntax	aipDisplay<('all'/'sel'/'redisplay'/'reset')>
	<pre>aipDisplay('batch','first'/'next'/'previous'/'last')</pre>
	If first argument is not given, 'redisplay' is used.
Arguments	'redisplay' - display/redisplay currently displayed or selected images.
	'all' - display all images currently loaded (in Review viewport, display all images selected in RQ).
	'sel' - display images in selected frames.
	'reset' - clear frame cache (without clear graphics area). This will cause next display use fresh frames.

'batch' - When more images are loaded than current frame layout, images are displayed as batches. This argument selects a batch to display ('first', 'next', 'previous', or 'last').

aipLoadSpec Load fdf spectra (C)

```
Syntax aipLoadSpec(fullpath<,key>)
    path - full path of fdf file (ends with ".fdf").
    key - a name to identify the data. The following
    are special keys reserved for data already in
    vnmrbg memory:
        'FID' - data in vnmrbg fid buffer,
        'SPEC' - data in vnmrbg phasefile buffer,
        'BASE' - data in vnmrbg baseline buffer.
```

If key is not specified, fdf file base name (not including ".fdf") will be the key.

aipMakeMaps Make csi map (C)

Syntax	aipMakeMaps<('li'/'ll'<,mapName<,specKey>>)>
	If first argument is not given, 'li' is used.
Arguments	'li' - make map of peak integrals. Peak is selected by cursor (cr). Integral region is defined by lifrq parameter.
	'll' - make map of peak height. Peak is selected by cursor (cr).
	<pre>mapName - is a full path including file name, but not suffix .fdf. Default is xxx.csi/maps/li_<chem_shift_ppm>.</chem_shift_ppm></pre>
	specKey - is the key of spectral data. If not specified, 'spec' is assumed. If 'spec' data is not loaded, phasefile buffer 'SPEC' is used.
	The major difference between aipMakeMaps and liMMap or llMMap is that aipMakeMaps can make maps for ROIs. If ROI of types box, oval, polygon or polyline exists (graphically displayed), aipMakeMaps will make CSI map for the ROI(s). The FOV and position of a csi map for a ROI will be calculated properly.
	This command exclusively uses <i>Cr</i> and lifrq parameters to determine peak position and integral region. Whereas liMMap and llMMap allow user to pass frequencies as arguments explicitly.
	This command allows selecting spectral data explicitly if one or more fdf spectral data are loaded.

aipOverlayFrames Overlay images in selected frames (C)

Syntax aipOverlayFrames('overlay'<, frameID1, frameID2,...>) overlay images in selected frames.

aipOverlayFrames('unoverlay') unoverlay images.

aipOverlayFrames('overlaid'):\$ret - return whether multiple
images are overlaid

aipOverlayFrames('canOverlay'):\$ret - return whether more
than one frame are selected

Arguments 'overlay' - overlay images in selected frames. Frames can be selected by mouse, or passing as arguments. Frame layout will be changed to 1x1. The first selected image will be displayed as the base image. Other selected image(s) will be overlaid on the base image. If the overlaid image has the same orientation as the base image (determined by Euler angles), images will be overlaid with proper scaling and translation to match the physical size and position. Otherwise the image will be overlaid without the consideration of FOV, position and data size.

'unoverlay' - unoverlay the images and return to the original frame layout.

Note, aipOverlayFrames('unoverlay') is different from aipViewLayers('remove'), which simply removes overlaid images (without changing frame layout).

'overlaid' - query whether images are overlaid as a result of aipOverlayFrames('overlay') command. Note, frame layout is changed to 1x1 temporarily and can be recoved by aipOverlayFrames('unoverlay') command.

Note, images can be overlaid by aipRQcommand command (loading image to a frame where an image already displayed). That does not change the layout.

aipOverlayFrames('overlaid'):\$n is true only if images are overlaid by aipOverlayFrames('overlay') command. Whereas aipViewLayers('hasOverlay') is true as long as any frame has an overlaid image.

'canOverlay' - query whether images are selected to overlay. More than one image frame should be selected. Multiple frames can be selected by clicking near to border of the frame while holding ctrl key.

aippars Create parameters for imaging browser (M)

Description aippars is a macro that creates global parameters used by imaging browser. These parameters may have different values in different viewports (parameter value is not shared by viewports, but is shared by all data in a viewport).

Parameters:

```
aipLayoutPolicy - 0 no effect, 1 rows=array_dim,
cols=slices, 2 rows=slices, cols=array_dim.
```

where array_dim and slices are from fdf header

aipStatUserVar - user selected variable for X-axis of ROI scatterplot. Default is "

aipVsHistFile=''

aipVsFunctionFile='/tmp/VjCltFunction'

aipVsFunction=''

aipWriteFmtConvert='FDF'

aipProfileLengthMsg=''

aipProfileFile=''

aipProfileMaxMsg=''

aipPointIntensityMsg=''

aipPointLabCoordsMsg=''

aipPointSeparationMsg=''

aipPointProjSeparationMsg=''

aipProfileDataCoordsStartMsg=''

aipProfileDataCoordsEndMsg=''

aipStatMinMsg=''

aipStatMaxMsg=''

aipStatAreaMsg=''

aipStatMeanMsg=''

aipStatMedianMsg=''

aipStatSdvMsg=''

```
aipStatVolumeMsg=''
```

- aipStatClipped=''
- aipStatGraphFile=''

```
aipFrameDefaultMax=3
```

```
aipVsBind=0
```

```
aipVsMode='individual'
```

- aipZoomBind=0
- aipZoomBindOnDrag=0
- aipPrintScale=100
- aipInfoUpdateOnMove=0
- aipProfileMIP=0
- aipZoomFactor=1.41421356
- aipRoiMaxActiveSlaves=-1
- aipRoiSelectOnCreation=1
- aipStatNumBins=100

aipStatHistRangeType=0 aipStatUpdateOnMove=0 aipStatCursMin=0 aipStatCursMax=1 aipStatHistMin=0 aipStatHistMax=1 aipStatOrdinate=0 aipStatAbscissa=0 aipVsDataMin=0 aipVsDataMax=0.01 aipVsTailPercentile=0.1 aipWriteFmtBits=32 aipWriteFmtFloat=1 aipMode=1 aipMovieRate=2 aipMselect='' aip2JExp='' aip2CExp='' aip2JCaret=0 aip2CCaret=0 aipProfileDataCoordsEndY=0 aipProfileDataCoordsEndX=0 enableExtractPanel=0 showObliquePlanesPanel=0 aipOrient='' aipXYfirst aipXZfirst aipYZfirst aipXYlast aipXZlast aipYZlast aipXYincr aipXZincr aipYZincr aipStatNumRois=0 aipPrevMode=1 aipRoiBind=1 aipRoiSelectSlavesOnCreation=1 aipRotationPolicy='neurological'

```
aipDisplay[1]=1 aipDisplay[2]=0 aipDisplay[3]=1
aipWindowSplit[1]=0 aipWindowSplit[2]=0
aipFrameResplitOnResize=0
aipPointDataCoordsMsg=''
aipStatUpdateCount=0
aipUserProfileFile='lineProfile'
aipUserStatGraphFile='statGraph'
aipUserStatListFile='statList'
aipInterpolationQuality=0
aipVsBindOnLoad=1
aipGrayFilename='/tmp/VjComm'
aipVsDynamicBinding=1
aipVsViewportBinding=1
aipZoomViewportBinding=1
aipProfileShowZero='y'
aipStatCursEnable=0
aipSegmentationType='r'
aipNumberFrames=0
aipNameFrames=2
aipNumberRois=0
aipPointInfoNumber=0,0
aipLineInfoNumber=0
aipAnnotation=''
aipClickedFrame=''
aipCurrentKey=''
aipMoviePath=userdir+'/data/images.mov'
aipMovieSpec='50'
aipMovieSettings[1]=1 aipMovieSettings[2]=1
aipMovieSettings[3]=1 aipMovieSettings[4]=100
aipMovieSettings[5]=100
aipAutoLayout = 1
aipDisplayMode = 1
aipMovieMode = 1
framelayoutName
framelayout
aipBatch = 1
aipBatches = 0
rqsort = 0
rqsort2 = 1
rqfull = 0
```

```
userselection=''
aipIOMode = 1
aipROIname_sel=''
aipROIname=''
aipROIpath=''
reconIn=''
reconOut=''
reconMode=1
rgImageNodes=100
aipShowFOV = 0 not to show border of FOV when image is displayed,
1 show border of FOV
aipLayerSel = 0 base image is selected, 1 first overlaid image is
selected, ...
aipShowPosition = 0 not to display position, distance, or intensity
1 show display position, distance or intensity according to
aipShowROIOpt.
aipShowROIPos= 0 show position, distance, or intensity when cursor
is over ROI. = 1 show position, distance, or intensity statically.
aipShowROIOpt = 0 show position or distance
                 1 show mean intensity.
                 2 show sum of intensity.
aipShowCenter= 0 hide center lines
         1 show center of magnet
         2 show center of FOV
aipAxis = 0,1,1,1,1,0,1,1 (default)
   aipAxis[1] = 0 or 1, hide or show axes.
         If 0, the rest of aipAxis values are ignored.
   aipAxis[2] = 0 or 1, hide or show axis on west
   aipAxis[3] = 0 or 1, hide or show axis on east
   aipAxis[4] = 0 or 1, hide or show axis on south
   aipAxis[5] = 0 or 1, hide or show axis on north
   aipAxis[6] = 0 or 1, ticks in or out
   aipAxis[7] = 0 or 1, show or hide crosshair in Point mode
   aipAxis[8] = 0 use frame borders as boundary, 1 use image FOV
as boundary
aipAxisGeom = 1, 1, 0, 0, 0, 0, 0, 0, 0, 0 (default)
   aipAxisGeom[1] = 0.0 to 1.0, fraction of image or frame
width as size of horizontal axis
   aipAxisGeom[2] = 0.0 to 1.0, fraction of image or frame
height as size of vertical axis
```

Α

<code>aipAxisGeom[3] = 0.0</code> to 1.0, fraction of image or frame width as X offset of west axis

aipAxisGeom[4] = 0.0 to 1.0, fraction of image or frame height as Y offset of west axis

<code>aipAxisGeom[5] = 0.0</code> to 1.0, fraction of image or frame width as X offset of east axis

aipAxisGeom[6] = 0.0 to 1.0, fraction of image or frame height as Y offset of east axis

aipAxisGeom[7] = 0.0 to 1.0, fraction of image or frame width as X offset of south axis

aipAxisGeom[8] = 0.0 to 1.0, fraction of image or frame height as Y offset of south axis

aipAxisGeom[9] = 0.0 to 1.0, fraction of image or frame width as X offset of north axis

aipAxisGeom[10] = 0.0 to 1.0, fraction of image or frame height as Y offset of north axis

aipAxisTransparency=0.0 total opaque for axis, and other text display, 1.0 total transparent.

```
aipUnits='mm','cm', or 'pix'
aipDefaultWindowSplit=1,1
aipAxisRef='logical'
planDecimal = 3
csiSpecVS - csi map scaling
```

aipRemoveSpec Remove all or specified spectral data (C)

Syntax aipRemoveSpec('all'/key)

'all' - remove all spectral data loaded with aipLoadSpec command key - remove spectral data specified by the key (given when loading the spectra, see aipLoadSpec details)

aipRQcommand Load and display images (C)

Syntax	aipRQcommand('read'/'loadImage'/'load'/)
Examples	aipRQcommand('load', fullpath) - load images in a given fullpath (a directory or a fdf file). Do not display images.
	aipRQcommand('load', fullpath, frameID) - load images in a given fullpath, and display images according to frameID. If
	fullpath is an image file, display the image in the frame specified
	by frameID. If there is already an image in the frame, the new image
	will be overlaid on existing image. If fullpath is a directory and

frameID < 0, then currently displayed images will be cleared, and new images will be display starting from frame 1. If frameID > 0, then new images will be appended to currently displayed images (enough frames will be laid out to display all images).

aipRQcommand('load', fullpath, '', x, y) - same as aipRQcommand('load', fullpath, frameID), but frameID will be determined by dropping location x,y (dropping to the graphics area). This command appends images because frameID is always > 0.

aipRQcommand('loadImage', fullpath<, frameID>) - same as aipRQcommand('load', fullpath<, frameID>), but fullpath is an image file, and images will be displayed in frame 1 if frameID is not specified.

aipRQcommand('display',path<,frameID>) - same as aipRQcommand('load', fullpath<, frameID>), but images will be displayed even when frameID is not supplied.

aipRQcommand('display'<, selection<, sortOption<, layoutOp tion>>>) - display selected images. selection=1,2,3,4,or 5 for "all","group","selected in RQ","selected frames", or "select...". Or selection='all', 'group', 'rq', 'frames', or 'images'. If selection is not given, parameter aipDisplayMode will be used. sortOption is meaningful only if images of multiple scans are loaded. Images will be sorted by scan if sortOption=0 or 1, and sorted by slices if sortOption=2. layoutOption = 0 to use current layout, 1 to do auto layout.

aipRQcommand('display',key<,sortOption<,layoutOption>>)
- similar to

aipRQcommand('display'<, selection<, sortOption<, layoutOp tion>>>), but images are selected by key (key=parentDir+' '+name+'0', where name can be a directory name or file name).

aipRQcommand('displayBatch'<,'first'/'next'/'prev'/'las
t') - similar to</pre>

aipDisplay('batch','first'/'next'/'previous'/'last').
Display current batch if the second argument is not given.

aipRQcommand('unselectDisplay') - unselect groups in RQ.

aipRQcommand('delete','all'/\$path/\$key) - delete image data
specified by the second argument. This command does not remove
images from Review Queue.

aipRQcommand('remove','all'/\$path/\$key) - delete image data
specified by the second argument. This command also removes images
from Review Queue.

aipRQcommand('reload'<,selection>) - reload selected images. selection can be 1,2,3,4,5 for "all", "group","selected in RQ","selected frames","select...". If selection is not supplied, parameter reconMode will be used.

aipRQcommand('read',xmlfile) - build "Review Queue" by reading a xml file (see userdir+'/persistence/RQtree3.xml' for example).

aipRQcommand('set',key,columnName,value) - set value for a RQ node specified by the key. aipRQcommand('get',key,columnName):value - get value for a RQ node specified by the key.

aipRQcommand('setvalue',key,columnName,value) - similar to 'set', but will not notify UI (java code) to update.

aipRQcommand('move',key1,key2) - move node1 (key1) above node2 (key2).

aipRQcommand('copy',key1,key2) - copy node1 (key1) and insert
above node2 (key2).

aipSaveColormap Save color map for selected images (C)

Syntax	aipSaveColormap<('all'/'dis'/'sel')>					
Description	Color map(s) are saved in image directory. If color map file(s) exist for individual image(s) or for a group of images (all images in a directory), they will be used to display the images, otherwise default color map will be used.					
Arguments	'all' - save a single color map file named "image.cmap", to be used for all images in the directory (all images in a directory is a "group").					
	'dis' - save a color map file for each displayed image. Color map files have the same name as image file, but the suffix .fdf is replaced by .cmap.					
	'sel' - the same as 'dis', but for images selected by mouse click.					
	If no argument is given, aipColormapOpt parameter will be used (value 0 is 'all', 1 is 'dis', 2 is 'sel'). Default value is 0.					
	Normally a color map is saved in a directory. The directory name is used to uniquely identify the color map. There is a color map file named image.cmap in this directory.					
	color map has the following format:					
	# Number of Colors					
	size 64 # Erronglugongu, rongo from 0.0 to 1.0					
	# Translucency, range from 0.0 to 1.0 translucency 0.0					
	# Color 0 is the color of below minimum					
	scale threshold					
	# Color 65 is the color of above maximum					
	# index Red Green Blue Transparent					
	Translucent					
	begin					
	2 0.0157 0.0 0.9804 0 1					
	3 0.0314 0.0 0.9608 0 1					
	4 0.0471 0.0 0.9451 0 1					
	5 0.0549 0.0 0.9255 0 1					
	0 0.0700 0.0 0.9098 0 I 7 0.0902 0.0 0.902 0 1					
	8 0.1059 0.0 0.8863 0 1					
9	0.1255	0.0	0.870	6	0	1
----	--------	------	-------	----	---	---
10	0.1412	0.0	0.85	1	0	1
11	0.1608	0.0	0.83	53	0	1
12	0.1765	0.0	0.81	57	0	1
13	0.1922	0.0	0.8	0	1	
14	0.2 0.	.0 0	.7804	0	1	

aipSetColormap Load color map for selected images (C)

Syntax aipSetColormap(colorMapName<, 'all'/'dis'/'sel'>)
Arguments ColorMapName is the name of the color map directory, such as
'default', 'blue.purple.red', etc...
'all' - apply color map to all images in the directory (a group of
images).
'dis' - apply color map all displayed images.
'sel' - apply color map all selected images.
If the second argument is not given, aipColormapOpt parameter will
be used (value 0 is 'all', 1 is 'dis', 2 is 'sel'). Default value is 0.

aipSetTransparency Set transparency for images or text (C)

Syntax	<pre>aipSetTransparency(transparency<, 'all'/'dis'/'sel'/'text'>)</pre>
Arguments	transparency = 0 to 1.0 for text, 0 to 100 for images. Default is 0 .
	Total opaque is 0, and total transparent is 1.0 or 100.
	'all' - set transparency for all images in the directory (a group of images).
	'dis' - set transparency for all displayed images.
	'sel' - set transparency for all selected images.
	'text' - set transparency for annotation, axis, text, etc
	If the second argument is not given, $aipColormapOpt$ parameter will be used (value 0 is 'all', 1 is 'dis', 2 is 'sel'). Default value is 0.

aipShow3PCursors Turn on/off cursors for 3-plane extraction

Syntax	aipShow3PCursors<(1/0)>
Description	This command turns on/off cursors for 3-plane extraction. Without the
	argument, this command toggles on/off cursor display.

aipShowCSIData Display CSI spectral data in grid format

Arguments key - a name given when loading the spectra. The following are special keys reserved for data already in vnmrbg memory:

'FID' - data in vnmrbg fid buffer,

'SPEC' - data in vnmrbg phasefile buffer,

'BASE' - data in vnmrbg baseline buffer.

comboKey - a string to combine (add/subtract/scale) spectra.

Examples key1='SPEC'

key2='spec'

comboKey1='spec1-spec2'

comboKey2='spec1*0.5'

'grid' - keyword to display grid

'3dgrid' - keyword to display 3D grid

'num' - keyword to display index

'frame:n' - display spectra in frame n. Default is to display in first frame or frame(s) selected by mouse.

'frame:all' - display spectra in all frames.

'slice:i' - display ith slice or ith xy-plane of 3D data. Default is to display first slice (index starts from 1).

This command is implement for visualizing CSI data with the following features:

1. Always in grid layout defined by fnv, fnv2, fnv3 parameters (see savefdfspec command for more info about fnv, fnv2, fnv3).

2. If base image is loaded, the position, size, orientation are properly adjusted based on the base image. Grid and data will be displayed only if CSI data intersects with the base image. The slice selection (by 'slice:n') is ignored if base image is displayed.

3. '3dgrid' option can be used to show grid when CSI data not intersecting base image.

4. If base image is not displayed, nv, nv2 plane will be displayed, grid will occupy max area while maintaining aspect ratio defined by lpe and lpe2.

5. If ROIs of type box, oval, polygon, polyline are defined (displayed), spectral data n ROIs will be shown. But spectral data outside the ROIs will not be shown.

To enter interactive mode, use jFunc(88,1) aipSetSate(11) commands.

To exit interactive mode, use jFunc(88,0) aipSetSate(1).

Α

aipShowSpec Display spectra specified by key(s), in specified layout (C)

Syntax	<pre>aipShowSpec(key/comboKey<, 'grid/vert/horiz'<, 'rows:n', 'cols:n '<, 'num'<, 'frame:n'<, 'sel'>>>>) - display spectra specified by key(s), in specified layout.</pre>
Description	aipShowSpec is an "alias" of aipShowCSIData command (or vice versa). It differs from aipShowCSIData by not imposing features specific for CSI data (see aipShowCSIData), and it supports horizontal and vertical layout in addition to grid layout. It also has an option to select traces with "dsSelect" parameter.
	This command requires to run aippars macro to create necessary parameters.
	To enter interactive mode, use jFunc(88,1) aipSetSate(11) commands.
	To exit interactive mode, use jFunc(88,0) aipSetSate(1).
Arguments	key - a name given when loading the spectra. The following are special keys reserved for data already in vnmrbg memory:
	'FID' - data in vnmrbg fid buffer,
	'SPEC' - data in vnmrbg phasefile buffer,
	'BASE' - data in vnmrbg baseline buffer.
	comboKey - a string to combine (add/subtract/scale) spectra.
	For example:
	key1='SPEC'
	key2='spec'
	comboKey1='spec1-spec2'
	<pre>comboKey2='spec1*0.5'</pre>
	'grid' - keyword to display grid
	'vert' - keyword for vertical layout
	'horiz' - keyword for horizontal layout
	'rows:n' - rows for grid layout
	'cols:n' - columns for grid layout
	'num' - keyword to display index
	'frame:n' - display spectra in frame n. Default is to display in first frame or frame(s) selected by mouse.
	'frame:all' - display spectra in all frames.
	'sel' - to display traces specified by "dsSelect" parameter.
Examples	<pre>aipShowSpec('SPEC','vert')</pre>
	aipLoadSpec('','spec') aipShowSpec('spec','rows:3','cols:4')

Syntax	<pre>aipViewLayers('names'<,frameID>):\$n,\$names - get names for overlaid images</pre>
	aipViewLayers('keys'<,frameID>):\$n,\$keys - get keys for overlaid images.
	<pre>aipViewLayers('hasOverlay'<,frameID>):\$n - get number of layers for overlaid images</pre>
	<pre>aipViewLayers('remove'<,frameID>) - remove overlaid images</pre>
Arguments	'names' - keyword to query image names.
	'keys' - keyword to query image keys
	'hasOverlay' - keyword to query number of layers over the base image. If no overlaid image, 0 is returned.
	'remove' - remove all overlaid images.

frameID - an integer to select a frame. If not specified, first selected frame (with mouse) is used. If no frame is selected, then first frame is used.

alfa Set alfa delay before acquisition (P)

- Description After the final event in the pulse sequence, including any receiver gate times occurring following the final pulse, acquisition occurs after a delay. This delay includes a fixed part, alfa, and a variable part, 1/(beta*fb).
 - On systems with 4-pole Butterworth filters, beta is 2.
 - On systems with 8-pole Butterworth (200-kHz) filters, beta is 3.8.
 - On systems with 8-pole elliptical filters, beta is 1.29.
 - On Systems with 4-pole Bessel filters, beta is 2.3 (only systems with 2-MHz and 5-MHz Analog-to-Digital Converter boards use this filter).

Because the total delay before acquisition is the sum of alfa and 1/(beta*fb), it is possible to shorten the delay beyond "normal" values by setting alfa negative (to a maximum of 1/(beta*fb)). The macros hoult and calfa frequently result in such negative values of alfa.

To set alfa to a negative number, use either the setvalue command to enter a specific value of alfa, or use the setlimit command to allow entry of negative values of alfa directly from the keyboard.

Values 0 to 100,000,000; in µs.

See also	NMR Spectroscopy User Guide		
Related	calfa	Recalculate alfa so that first-order phase is zero (M)	
	fb	Filter bandwidth (P)	
	hoult	Set parameters alfa and rof2 according to Hoult (M)	
	rof2	Receiver gating time following pulse (P)	
	setlimit	Set limits of a parameter in a tree (C)	
	setlp0	Set parameters for zero linear phase (M)	
	setvalue	Set value of any parameter in a tree (C)	

alock Automatic lock control (P)

Description	Governs Autolock control following the insertion of a sample with change or sample, and following initiation of an acquisition with the go, ga, or au. Manual adjustment of lock power, gain, and phase is possible using the acqi command.			
Values	Possible values are 'a', 'auto', 'n', 's', 'samp', 'u', or 'y', where:			
	'a' or 'auto' selects the optimizing Autolock function, which performs a lock capture and an automatic lock power and gain adjustment before data acquisition begins (lock phase is <i>not</i> optimized).			
	'n' leaves the lock in its current state.			
	's' or 'samp' selects the optimizing Autolock function, which performs a lock capture and an automatic lock power and gain adjustment before data acquisition begins (lock phase is <i>not</i> optimized) but only if the sample has just been changed.			
	'u' turns lock off so that the experiment runs unlocked.			
	$^{\prime}\mathrm{y}^{\prime}$ turns on the software Autolock function, which searches for the correct Z0 value only.			
See also	NMR Spectroscopy User Guide			
Related	acqiInteractive acquisition display process (C)auSubmit experiment to acquisition and process data (C)changeSubmit a change sample experiment to acquisition (M)gaSubmit experiment to acquisition and FT the result (C)gfPrepare parameters for FID/spectrum display in acqi (M)goSubmit experiment to acquisition (C)lockSubmit an Autolock experiment to acquisition (C)sampleSubmit change sample, Autoshim experiment to acquisition (M)			

ampmode Independent control of amplifier mode (P)

Description Gives override capability over the default selection of amplifier modes. Unless overridden, the usage of rf channels determines whether the amplifier for a channel is in pulse, CW (continuous wave), or idle mode:

- Observe channel is set to the pulse mode.
- Other used channels are set to the CW mode.
- Any unused channels are set to the idle mode.

The ampmode parameter can be used to override this selection.

ampmode does not normally exist but can be created by the user with the command create('ampmode','flag').

- Values List of characters in which the mode of the first amplifier is determined by the first character, the mode of the second amplifier by the second character, and so on. For each amplifier, one of the following characters is used:
 - 'c' selects CW mode.
 - 'i' selects idle mode.
 - 'p' selects pulse mode.
 - 'd' selects default behavior.

For example, ampmode='ddp' selects default behavior for the first two amplifiers and forces the third channel amplifier into pulse mode. Additional filtering is usually required when an amplifier in the same band as the observe amplifier is placed in the CW mode.

See also VnmrJ User Programming

RelatedcreateCreate new parameter in a parameter tree (C)dnNucleus for the first decoupler (P)tnNucleus for observe transmitter (P)

amptype Amplifier type (P)

Description Specifies the type of amplifier on each rf channel of the spectrometer. The value is set in the Spectrometer Configuration window (opened from config) using the label Type of Amplifier.

For each channel, the types are Class C, Linear Full Band, Linear Low Band, Linear Broadband, or, for the fourth channel only, Shared. Selecting Shared means that the amplifier is fully configured for the third channel, and that the fourth channel shares this amplifier with the third channel.

When a type is selected for a channel, a letter (one of the values described below) is added to the value of amptype. For example, a system already set to Linear Full Band on the observe transmitter channel and the first decoupler channel would have amptype='aa'.

	Selecting the third channel as Linear Low Band would set amptype='aal'. Finally, selecting Shared for the fourth channel would set amptype='aaln'.
Values	'a' indicates the channel uses a linear full-band amplifier. A full-band amplifier has two outputs: 12 MHz to $^{31}\rm{P},$ and $^{19}\rm{F}'^{1}\rm{H}.$
	'b' indicates the system uses a linear broadband amplifier.
	'c' indicates the system uses a class C amplifier.
	'l' indicates the channel uses a linear low-band amplifier. A low-band amplifier has one output from 12 MHz to $^{31}{\rm P}$ only.
	'n' indicates the fourth channel shares a linear amplifier with the third.
See also	NMR Spectroscopy User Guide, VnmrJ User Programming
Related	config Display current configuration and possibly change it (M)

analyz Calculate standard peak height (M)

Syntax analyz(\$option,\$title)

Description Macro to calculate average peak height and standard deviation and/or average phase and standard deviation.

Arguments \$option ='n' for amplitude and phase, 'a' for amplitude only, and 'p' for phase only. The \$title option puts a title on the plot.

Examples analyz - Does analysis for both amplitude and phase analyz('p') - Does analysis for phase only analyz('n','Stability') - Does analysis for amplitude and phase and puts title "Stability" on the plot.

analyze Generalized curve fitting (C)

Syntax (curve fitting) analyze('expfit', xarray<, options>)
 (regression) analyze('expfit', 'regression'<, options>)

Description Provides interface to curve fitting program expfit (using the curve fitting syntax), supplying expfit with input data in the form of the text file analyze.inp in the current experiment. expfit can be called from UNIX with the syntax:

expfit options <analyze.inp >analyze.list

expfit does a least-squares curve fitting to the data supplied in analyze.inp. Macros are available for the specialized uses of analyze, such as the 'T1' and 'kinetics' options. These macros avoid the need to select options and get the correct file format.

In the regression mode (using the regression syntax above), the type of curve fitting, ('poly1',...) must be selected. The regression section in the manual *NMR Spectroscopy User Guide* gives the input

file format and describes the menus that permit choices indirectly through menu buttons.

The text file analyze.inp for the options 'T1', 'T2', 'kinetics', 'contact_time', and 'regression' contains the following lines (note that (1), (2), (3), etc. do not appear in the file but are used to identify lines in the explanation):

- (1) <text line>
- (2) <text line>
- (3) npeaks npairs <xscale> <yscale>
 - (4) <NEXT npairs1>
- (5) peaks
- (6) x y
- (6) x y
- . . .
- (4) <NEXT npairs2>
- (5) peaks
- (6) x y
- (6) x y
 - . . .

Line-by-line explanation:

(1) Optional descriptive text line, for regression only. Omit line otherwise.

(2) Optional y-axis title, for regression only. Omit line otherwise.

(3) Line containing an integer for the number of peaks (npeaks) followed by another integer for the number of (x, y) pairs per peak (npairs). If regression, the *x*-scale type and *y*-scale type are also listed.

(4) In the regression mode, a line beginning with the keyword NEXT is inserted at the start of each data set when the number of pairs per peak is variable. In this case, the number of (x, y) pairs for the peak (npair1, npair2, etc.) is also given on the line.

(5) Peak index.

(6) Data pairs, one to a line, are listed by peak in the following order:

x y (first peak, first pair)

x y (first peak, second pair)

. . .

x y (second peak, first pair)

x y (second peak, second pair)

• • •

In the regression mode, the line beginning with NEXT is inserted at the start of the data for each peak when the number of pairs per peak is variable. In this case, the header contains the maximum number of pairs for any peak.

For 'T1', 'T2', 'kinetics', and 'contact_time', information from the file fp.out and values of the arrayed parameter xarray are used to construct the file; thus, it is necessary to run fp prior to analyze. For regression, analyze.inp is made by running
expl('regression'). If the regression mode is not selected,
analyze.inp may be slightly different.

In addition to output to the standard output, which is usually directed to analyze.list, expfit makes a file analyze.out, which is used by expl to display the results of the analysis.

User-supplied analysis programs can be called by analyze in place of expfit. Such programs should read their input from stdin and write the output listing to stdout. No analyze.out file needs to be generated unless display by expl is desired. Use the program expfit as a model.

Arguments 'expfit' is a required first argument.

xarray is the name of the parameter array holding x-values in 'T1', 'T2', 'kinetics', and 'contact_time', and is used only with these options.

'regression' sets regression mode and signifies generalized curve fitting with choices 'poly1', 'poly2', 'poly3', and 'exp'.

options are any of the following keywords:

- 'T1' sets T_1 analysis (the default).
- 'T2 ' sets T_2 analysis.
- 'kinetics' sets kinetics analysis, with decreasing peak height.
- 'increment' sets kinetics analysis, with increasing peak height.
- 'list' makes an extended listing for each peak.
- 'diffusion' sets a special analysis for diffusion experiments.
- 'contact_time' sets a special analysis for solids cross-polarization spin-lock experiments.
- 'poly1' sets a linear fitting. It is used in regression mode only.
- 'poly2' sets a quadratic fitting. It is used in regression mode only.
- 'poly3' sets a cubic fitting It is used in regression mode only.
- 'exp' sets exponential curve fitting. It is used in regression mode only.

Examples analyze('expfit','d2','T1','list')
analyze('expfit','pad',kinetics','list')
analyze('expfit','p2','contact_time','list')
analyze('expfit','regression','poly1','list')

See also NMR Spectroscopy User Guide

 Related contact_time
 MAS cross-polarization spin-lock contact time (M)

 expl
 Display exponential or polynomial curves (C)

 pexpl
 Plot exponential or polynomial curves (C)

 kini
 Kinetics analysis, increasing intensity (M)

t1 T_1 exponential analysis (M)

annotation Display annotation specified by the parameter "template" or the default

Syntax	annotation(<template>, <x, height="" width,="" y,="">)</x,></template>		
	pannotation(<template>, <x, height="" width,="" y,="">)</x,></template>		
Applicability	VnmrJ 3.1		
Description	"annotation" and "pannotation" will display or plot annotation specified by the parameter "template" or the default.		
Arguments	template: The name of template of annotation to be displayed. The default name is 'default'.		
	x, y: The origin point on the screen or plotter, in mm.		
	width: The width on the screen or plotter, in mm.		
	height: The height on the screen or plotter, in mm.		

ap Print out "all" parameters (C)

Applicability	VnmrJ			
Syntax	ap('template_name',<'filename'>)			
Description	Print a parameter list. The User Programming Manual describes the rules for building a template for the ap commands. The string parameter ap normally controls how the command, ap, displays the parameters. Use command paramvi('ap') to modify the ap parameter. The ap command writes the parameter list to a file if filename is provided as the second argument.			
Arguments	ments template_name template name must be the first argument. filename optional, name of file to which the parameters are v			
Examples	s ap('ap', 'apout') — writes the parameter list using defined by a parameter to the file apout.			
	ap('newa	.p')		
See also	NMR Spee	troscopy User Guide; VnmrJ User Programming		
Related	addpar ap	Add selected parameters to the current experiment (M) "All" parameters display control (P)		
	dg	Display group of acquisition/processing parameters (C)		
	hpa	Plot parameters on special preprinted chart paper (C)		
	pap	Plot out "all" parameters (C)		
	paramvi	Edit a variable and its attributes with vi text editor (C)		
	ppa	Plot a parameter list in "English" (M)		

ap "All" parameters display control (P)

Description	Controls the display of the ap and pap commands to print and plot a parameter list. Use paramvi('ap') to modify the string value of ap.		
See also	NMR Spectroscopy User Guide; VnmrJ User Programming		
Related	ap Print out "all" parameters (C)		
	dg Display group of acquisition/processing parameters (C)		
	pap Plot out "all" parameters (C)		
	paramvi	Edit a variable and its attributes with vi text editor (C)	

apa Plot parameters automatically (M)

Syntax	apa		
Description	Selects automatically the appropriate command on different plotter devices to plot the parameter list.		
See also	VnmrJ User Programming		
Related	hpa	Plot parameters on special preprinted chart paper (C)	
	ppa	Plot a parameter list in "English" (M)	

aph Automatic phase adjustment of spectra (C)

Syntax	aph<:\$ok,\$r	p,\$1p>
Description	Automatically calculates the phase parameters lp and rp required to produce an absorption mode spectrum and applies these parameters to the current spectrum. Values calculated do <i>not</i> depend on the initial values of lp and rp.	
Arguments	\$\\$ \$\\$ \$\\$ \$\\$ \$\\$ \$\\$ \$\\$ \$\\$ \$\\$ \$\\$	
	lp is the calculated value of lp . If lp is requested as a return val lp is returned but not applied to the current spectrum.	
See also	NMR Spectroscopy User Guide	
Related	aph0	Automatic phase of zero-order term (C)
	aphx	Perform optimized automatic phasing (M)
	lp	First-order phase in directly detected dimension (P)
	rp	Zero-order phase in directly detected dimension (P)

aph0 Automatic phase of zero-order term (C)

Syntax	aph0<:\$ok,\$rp,\$lp>	
Description	Automatically adjusts only the zero-order frequency-independent term rp and does not rely on the frequency-dependent term lp being previously adjusted. In favorable circumstances, spectra may be obtained in such a way that only rp is expected to change. In these cases, if lp has been determined for one spectrum, then rp only can be computer-adjusted for subsequent spectra by aph0 ("aph-zero"). Note that aph0 does not correctly phase an exactly on-resonance peak.	
Arguments	sok is 1 if the phase adjustment succeeds, or 0 if the adjustment fails.	
	\$rp is the calculated value of rp.	
	lp is the current value of lp .	
See also	NMR Spectroscopy User Guide	
Related	aph Automatic phase adjustment of spectra (C)	
	aphx Perform optimized automatic phasing (M)	
	1pFirst-order phase in directly detected dimension (P)	
	rp Zero-order phase in directly detected dimension (P)	

aphb Auto phasing for Bruker data (C)

Syntax	aphb<(threshold)>	
Description	Phases Bruker data using the autophasing program.	
Arguments	threshold determines if a data point is large enough to qualify it as part of a peak. If no argument is given, or if the value is equal to or less than 0, the threshold is calculated from the spectrum.	
Examples	aphb aphb(2)	
See also	NMR Spectroscopy User Guide	
Related	aphAutomatic phase adjustment of spectra (C)aph0Automatic phase of zero-order term only (C)	

aphx Perform optimized automatic phasing (M)

Syntax	aphx
Description	Optimizes parameters and arguments for the aph command. aphx first performs on aph then colculates a theoretical value for $\ln z$. If $\ln z$ set
	by the aph is different from the calculated value by 10 per cent, the
	calculated value is used and an aph0 is performed.

See also	NMR Spectroscopy User Guide	
Related	aph	Automatic phase adjustment of spectra (C)
	aph0	Automatic phase of zero-order term only (C)
	lp	First order phase along directly detected dimension (P)

appdir Application directory information

Syntax	appdir('info'):\$num - Applications directories information		
	<pre>appdir('info',n):\$label,\$path - Application directory information</pre>		
Applicability	VnmrJ 3.1		
Description	An application directory is a directory where VnmrJ can look for templates, maclib, manual, menujlib, parlib, probes, psg, psglib, seqlib, shims, tablib, shapelib, gshimlib, and mollib directories. It will not look for expN directories, global, or other files or directories. The exists command has been enhanced to search for other files and directories in the applications directories, allowing users flexibility to customize their applications. The appdirs macro starts an editor to set applications directories.		
Arguments	The appdir('info') command will tell you the number of application directories that are currently enabled. This value can be returned to a parameter as in appdir('info'):\$num		
	The label and path of a specific application directory can be returned		
	by supplying a number after the 'info' keyword. The number must be between		
	1 and the total number of applications directories (\$num from above).		
Examples	The following macro lists the current applications directories: clear		
	write('alpha','Applications Directories')		
	appdir('info'):\$num		
	\$i = 0		
	while (\$i < \$num) do		
	\$i=\$i+1		
	appdir('info',\$i):\$label,\$path		
	if (\$label = '') then		
	write('alpha','%d: "appdir %d" has path "%s"',\$i,\$i,\$path)		
	else		
	write('alpha','%d: "%s" has path "%s"',\$i,\$label,\$path)		
	endif		
	endwhile		
See also	The "which" macro for another example of the use of the appdir command		

appdirs Starts Applications Directory Editor (M)

Applicability	ALL		
Syntax	appdirs		
Description	The appdirs macro brings up an editor to set the applications directories. The top section of the editor has rows consisting of a menu and two entry boxes.		
Values	Menu selections:		
	Enabled – enable an application directory.		
	Disabled – disable an application directory.		
	$\operatorname{Remove}(d)$ – initial setting for other row and the and empty entry boxes.		
	Set an application directory menu to Remove(d) to completely remove it.		
	Fields in each row:		
	Applications directory path.		
	A comment can be added to the second entry box.		
	Radio-button choices:		
	Save as private applications directories – sets the applications directories for the current operator only.		
	Reset to system default applications directories – removes any private applications directories and return to the standard default set.		
	Save the applications directories for global use – available only to users with write permission for VnmrJ system files. A name must be provided for this choice. This will affect all users the administrator has set that name as their appdirs setting. The Agilent default names are Experimental, Walkup, Imaging, and LcNmrMs.		
	Buttons:		
	OK – exit the editor and apply the selections made in the editor.		
	Cancel – exit the editor and abort the editor session, making no changes to the applications directories.		
See also	VnmrJ Installation and Administration		
Related	exists Checks if parameter, file, or macro exists and file type (C)		

appmode Application mode (P)

Description A global parameter that allows selection of specialized system applications modes, such as imaging, by setting the global parameters sysmaclibpath, sysmenulipath, and syshelppath. For example, in /vnmr/maclib is a subdirectory maclib.imaging that contains macros used primarily with imaging applications. Similarly, in /vnmr/menulib is a subdirectory menulib.imaging for Α

imaging- related menus. By separating the imaging macros and menus into subdirectories, access to imaging-specific macros and menus is more convenient. This separation also allows minor modifications to some macros and menus while retaining the names that are in common use or required by other VnmrJ commands.

The value of appmode are set from either the System settings dialog in the Utilities menu or the VnmrJ Admin interface.

Values 'standard' sets standard application mode. 'imaging' sets imaging application mode. 'autotest' sets autotest application mode

apptype Application type (P)

Description	Specifies the application type, the group of pulse sequences to which
	a pulse sequence belongs. It is used by the execpars macros to specify
	the actions executed by the protocol for a pulse sequence. The actions
	are common to the group of pulse sequences specified by the apptype.

Values See the execpars directory in /vnmr.

See also VnmrJ Imaging, User Guide and NMR Spectroscopy User Guide

cqexp	Load experiment from protocol (M)
execpars	Set up the exec parameters (M)
execsetup	Execute setup macro (P)
execprep	Execute prepare macro (P)
execprescan	Execute prescan macro (P)
execpreocess	Execute processing macro (P)
execplot	Execute plotting macro (P)
sqexp	Load experiment from protocol (M)
	cqexp execpars execsetup execprep execprescan execpreocess execplot sqexp

Apt Set up parameters for APT experiment (M)

Description	Converts a parameter set to the APT (attached proton test) experiment.	
See also	NMR Spectroscopy User Guide	
Related	aptaph Automatic processing for APT spectra (M)	
	capt	Automated carbon and APT acquisition (M)
	hcapt	Automated proton, carbon, and APT acquisition (M)

aptaph Automatic processing for APT spectra (M)

SyntaxaptaphDescriptionAutomatically phases APT spectra.

See also NMR Spectroscopy User Guide

Related Apt Set up parameters for APT pulse sequence (M)

array Easy entry of linearly spaced array values (M)

Syntax	array<(parameter<,number_steps,start,step_size)>
Description	Arrays a parameter to the number of steps, starting value and step size given by the user. All values of the array will satisfy the limits of the parameter.
	If array is typed with none or only some of its arguments, you enter an interactive mode in which you are asked for the missing values.
Arguments	parameter is the name of the parameter to be arrayed. The default is an interactive mode in which you are prompted for the parameter. Only numeric parameters can be arrayed.
	number_steps is the number of values of the parameter. The default is an interactive mode in which you are prompted for the number of steps.
	start is the starting value of the parameter array. The default is an interactive mode in which you are prompted for the starting value.
	step_size is the magnitude of the difference between elements in the array. The default is an interactive mode in which you are prompted for the step size.
Examples	array array('pw') array('tof',40,1400,-50)
See also	NMR Spectroscopy User Guide

array Parameter order and precedence (P)

Description Whenever an array of one or more parameters is set up, the string parameter array tells the system the name of the parameter or parameters that are arrayed and the order and precedence in which the arraying is to take place. The parameter array is automatically updated when acquisition parameters are set. "Diagonal arrays" (those corresponding to using parentheses in the parameter array) must be entered by hand.

Values '' (two single quotes with no space between) indicates no parameter is arrayed.

'x' indicates the parameter x is arrayed.

'x,y' indicates the parameters x and y are arrayed, with y taking precedence. That is, the order of the experiments is x_1y_1 , x_1y_2 ,... x_1y_n , x_2y_1 , x_2y_2 ,... x_2y_n ,... x_my_n , with a total of $m \times n$ experiments being performed.

'y,x' indicates the parameters x and y are arrayed, with x taking precedence. That is, the order of the experiments is x_1y_1 , x_2y_1 ,... x_ny_1 , x_1y_2 , x_2y_2 ,... x_my_2 ,... x_my_n , with total of $m \times n$ experiments being performed.

'(x, y)' indicates the parameters x and y are jointly arrayed. The number of elements of the parameters x and y must be identical, and the order of experiments is x_1y_1 , x_2y_2 ,... x_ny_n , with n experiments being performed.

Joint arrays can have up to 10 parameters. Regular multiple arrays can have up to 20 parameters, with each parameter being either a simple parameter or a diagonal array. The total number of elements in all arrays can be 2^{32} -1.

See also NMR Spectroscopy User Guide

Related array Easy entry of linearly spaced array values (M)

arraydim Dimension of experiment (P)

DescriptionAfter calcdim calculates the dimension of an experiment, the result
is put into the parameter arraydim. If an experiment is arrayed,
arraydim is the product of the size of the arrays.See alsoNMR Spectroscopy User GuideRelatedcalcdimcalcdimCalculate dimension of experiment (C)
celemcelemCompleted FID elements (P)

array2csv Formats Array into Comma Separate Variable

Description	This macro converts an array into a comma separated variable.	
Syntax	array2csv('parameter'):\$csv	
Examples	array2csv('ni'):\$increments	
Related	array2strsv, array2string, string2array, strsv2array, csv2array	

array2stringFormats Array into String

Description	This macro converts an array into a string variable.
Syntax	array2string('parameter'):\$string
Examples	array2string ('d3'):\$delay

array2strsv Formats Array into String Separated Variable

Description This macro formats an array into a string separated variable. Syntax array2strsv('parameter'):\$strsv Examples array2strsv ('ni'):\$increments

asin Find arc sine of number (C)

Syntax	asin(value)<:n>	
Description	Finds the arc sine (also called the inverse sine) of a number.	
Arguments	value is a number in the range of ± 1.0 .	
	n is a return argument giving the arc sine, in radians, of value. The default is to display the arc sine value in the status window.	
Examples	asin(.5) asin(val):asin_val	
See also	VnmrJ User Programming	
Related	sin Find sine value of an angle (C)	

Syntax	asize		
Description	Adjusts the 2 displayed res suggested for spread of one	Adjusts the 2D display parameters (sc, wc, sc2, and wc2) so that the displayed resolution along both f_1 and f_2 is the same. It is not suggested for heteronuclear experiments where the chemical shift spread of one nucleus is much greater than that of the other.	
See also	NMR Spectroscopy User Guide		
Related	SC	Start of chart (P)	
	sc2	Start of chart in second direction (P)	
	WC	Width of chart (P)	
	wc2	Width of chart in second direction (P)	

assign Assign transitions to experimental lines (M)

Syntax	<pre>(1) assign< (2) aggign(</pre>	('mark')>	
Description	Assigns the n nll listing a not be assign spins ('ite differences in	hearest calculated transition to the lines from a dll or fter spinll has placed them in slfreq. All lines may ned and transitions must be greater than sth. The next rate') determines new parameters to minimize the n position of the assigned pairs.	
Arguments	'mark' makes assign use the lines selected with the mark button in place of dll. The results of the mark operation are stored in the file markld.out, which is cleared by the command mark('reset').		
	transition_number is a single calculated transition number that is assigned to a line from the dll listing.		
	line_numbe line_numbe	r is the index of the line from the dll listing. Setting r=0 removes an assignment from a calculated transition.	
Examples	assign assign('mark') assign(4,0)		
See also	NMR Spectro	scopy User Guide	
Related	d11	Display listed line frequencies and intensities (C)	
	mark	Determine intensity of the spectrum at a point (C)	
	nll	Find line frequencies and intensities (C)	
	slfreq	Measured line frequencies (P)	
	spinll	Set up slfreq array (M)	
	spins	Perform spin simulation calculation (C)	
	sth	Minimum intensity threshold (P)	

at Acquisition time (P)

Description	Length of time during which each FID is acquired. Since the sampling rate is determined by the spectral width sw , the total number of data points to be acquired $(2*sw^*at)$ is automatically determined and displayed as the parameter np. at can be entered indirectly by using the parameter np.	
Values	Number, in seconds. A value that gives a number of data points that is not a multiple of 2 is readjusted automatically to be a multiple of 2.	
See also	NMR Spectroscopy User Guide; VnmrJ User Programming	
Related	npNumber of data points (P)swSpectral width in directly detected dimension (P)	

atan Find arc tangent of a number (C)

Syntax	atan(value)<:n>	
Description	Finds the arc tangent (also called the inverse tangent) of a number.	
Arguments	value is a number between $\pi/2$ and $-\pi/2$.	
	n is a return argument giving the arc tangent, in radians, of value. The default is to display the arc tangent value in the status window.	
Examples	atan(.5) atan(val):atan_val	
See also	VnmrJ User Programming	
Related	sin Find sine value of an angle (C)	

atan2 Find arc tangent of two numbers (C)

Syntax	atan2(y,x)<:n>
Description	Finds the arc tangent (also called the inverse tangent) of the quotient of two numbers.
Arguments	$_Y$ and $_x$ are two numbers, where the quotient $_{y/x}$ is between $\pi/2$ and $-\pi/2$ and $_x$ is not equal to zero.
	n is a return argument giving the arc tangent, in radians, of $_{\rm Y}/x.$ The default is to display the arc tangent value in the status window.
Examples	atan2(1,2) atan2(val):atan2_val
See also	VnmrJ User Programming
Related	sin Find sine value of an angle (C)

atcmd Call a macro at a specified time (M)

```
Syntax atcmd('macro','timespec')
atcmd('macro','timespec','active')
atcmd('macro','timespec','start')
atcmd('macro','timespec','active','start')
atcmd('macro','cancel')
atcmd('macro','list')
atcmd
```

Description atcmd will call a macro at the specified time. It only functions on a spectrometer. If the 'active' argument is given, the macro will be executed by the Vnmr process that specified atcmd. If that process is no longer active, the macro will be removed from the database. If the 'active' argument is not give, then a background Vnmr will be started to execute the macro. This background Vnmr will not be started in an experiment. Therefore, the macro will need to execute jexp or run commands or macros which do not need experiment parameters. It will have access to global and systemglobal parameters. The bootup macro will not be executed automatically. It can be called from the atcmd macro.

Arguments When called with arguments, atcmd updates the database with the supplied information. It does not start the process that calls the macros at the specified times. atcmd with no arguments starts the program that calls the macros at the specified times.

timespec -- has the format hh:mm <mon tue wed thur fri sat sun> A 24 hour clock is used -- midnight is 0:0, noon is 12:00.

day -- If the optional day field is used, the command will be repeated on that day at the appointed time. The day fields are case insensitive. For monday, wednesday, and friday only a single character is needed. More can be used. For tuesday, thursday, saturday, and sunday, at least two characters must be given.

cancel -- If the cancel argument is given, it will cancel all the commands that match the supplied macro. For example, if you specify cmda to be run at 8:00 on mon and 9:00 on tue, then atcmd('cancel', 'cmda') will cancel both of them. If the macro is '', the cancel option will cancel all atcmd macros.

list -- The list argument lists the timespec for all the atcmds that match the supplied macro. If the macro is '', the list option lists all of the atcmd macros and their timespecs. Optional arguments can be returned. The first is the number of atcmds. The macro and timespec for each atcmd can be returned.

When the command specified by atcmd is executed in background, it will be executed using the environment of the user who requested the atcmd. Also, the background VnmrJ will initially not be joined to a specific experiment.

Examples atcmd('echo(`good morning`)','8:00 mon tue wed thu fri') Displays a welcome message every weekday at 8:00 am. atcmd('echo(`What are you doing here on a weekend?`)','8:00 Sat Sun') Questions your intentions on the weekend. atcmd('startNightQueue','22:00') Runs the macro startNightQueue at 22 hr. (10:00pm). atcmd('startNightQueue','cancel') Cancels the scheduled startNightQueue cmd atcmd('','cancel') Cancels all scheduled commands atcmd('','list') Lists all scheduled commands

atext Append string to current experiment text file (M)

Syntax	atext(string)		
Description	Adds a line of text to the current experiment text file.		
Arguments	string is a single line of text.		
Examples	atext('T1 Experiment')		
See also	NMR Spectro	oscopy User Guide	
Related	ctext	Clear the text of the current experiment (C)	
	text	Display text or set new text for current experiment	
		(C)	
	write	Write formatted text to a device (C)	

attval Calculate pulse width (M)

Syntax	attval (pw,tpwr)
Description	Calculates the pulse width and B_1 field at every transmitter power. A low transmitter power should be used where the amplifier is not in compression. Calculation is not valid where amplifier is in compression.
Arguments	pw is the pulse width. tpwr is the transmitter power.
Examples	attval(7.0,59)

atune ProTune Present (P)

Description Hardware configuration parameter specifying if ProTune is or is not present. Parameter is set in the System Configuration window.

131

Arguments	'y' Pro' 'n' Pro'	Tune is present Tune not is present
See also	VnmrJ 1	nstallation and Administration
Related	wtune	Specify when to tune (P)
	tupwr	Transmitter power used in tuning (P)

Submit experiment to acquisition and process data (M)

Syntax au<(<'nocheck'><,'next'><,'wait'>)>

Description Performs the experiment described by the current acquisition parameters, checking the parameters loc, spin, gain, wshim, load, and method to determine the necessity to perform various actions in addition to simple data acquisition. This may involve a single FID or multiple FIDs, as in the case of arrays or 2D experiments. au causes the data to automatically be processed according to the following parameters:

- wbs specifies what happens after each block.
- wnt specifies what happens after each FID is collected.
- wexp specifies what happens when the entire acquisition is complete (which may involve several complete FIDs in the case of 1D arrays or 2D experiments).

Before starting the experiment, au executes the two user-created macros if they exist. The first is usergo, a macro that allows the user to set up general conditions for the experiment. The second is a macro whose name is formed by go_ followed by the name of the pulse sequence (from seqfil) to be used (e.g., go_s2pul, go_dept). This macro allows a user to set up experiment conditions suited to a particular sequence.

Arguments 'nocheck' is a keyword to override checking if there is insufficient free disk space for the complete 1D or 2D FID data set to be acquired.

'next' is a keyword to put the experiment started with au('next') at the head of the queue of experiments to be submitted to acquisition.

'wait' is a keyword to stop submission of experiments to acquisition until wexp processing of the experiment, started with au('wait'), is finished.

Examples au

au('wait')

See also NMR Spectroscopy User Guide

Related	auto_au	Controlling macro for automation (M)
	change	Submit a change sample experiment to acquisition (M)
	ga	Submit experiment to acquisition and FT the result (M)
	gain	Receiver gain (P)
	go	Submit experiment to acquisition (M)
	go_	Pulse sequence setup macro called by go, ga, and au (M)
	load	Load status of displayed shims (P)

au

loc	Location of sample in tray (P)
lock	Submit an Autolock experiment to acquisition (C)
method	Autoshim method (P)
sample	Submit change sample, Autoshim experiment to acquisition (M)
seqfil	Pulse sequence name (P)
shim	Submit an Autoshim experiment to acquisition (C)
spin	Submit a spin setup experiment to acquisition (C)
spin	Sample spin rate (P)
su	Submit a setup experiment to acquisition (M)
usergo	Experiment setup macro called by go, ga, and au $\left(M\right)$
wbs	Specify action when bs transients accumulate (C)
wexp	Specify action when experiment completes (C)
wnt	Specify action when nt transients accumulate (C)
wshim	Conditions when shimming is performed (P)

AuCALch3i Set up autocalibration with CH3I sample (M)

SyntaxAuCALch3iDescriptionRetrieves standard proton parameter set and setup for automatic
calibration of proton (observe and decouple), carbon (observe and
decouple), gcal, and C/H gradient ratio. The AuCALch3i macro is the
same as the AuCALch3i1 macro.

AuCALch3i1 Get autocalibration with CH₃I sample (M)

Syntax	AuCALch3i1
Description	Retrieves standard proton parameter set and setup for automatic
	calibration of proton (observe and decouple), carbon (observe and
	decouple), gcal, and C/H gradient ratio. The AuCALch3i1 macro is
	the same as the AuCALch3i macro.

AuCALch3oh Set up autocalibration with Autotest sample (M)

Syntax	AuCALch3oh		
Description	Retrieves standard proton parameter set and setup for automat calibration of proton (observe), carbon (decouple), gcal and C		
	gradient ratio. The AuCALch30h macro is the same as the		
	AuCALch30h1 macro.		

AuCALch3oh1 Get autocalibration with Autotest sample (M)

Syntax AuCALch3oh1

Α

Description Retrieves standard proton parameter set and setup for automatic calibration of proton (observe), carbon (decouple), gcal and C/H gradient ratio. The AuCALch30h1 macro is the same as the AuCALch30h macro.

Aucalibz0 Automatic Hz to DAC calibration for ZO (M)

Applicability	Autocalibration routine		
Syntax	Called by Augmapz0 calibration routine.		
Description	Called by Augmapz0 calibration routine. Automatically calibrates lock frequency change per Z0 DAC unit change. The calibrated value is written out in the probe file as lkhzdac parameter		
See also	System Administration.		
Related	Augmapz0 Automatic lock gradient map generation and Z0 calibration (M)		
	Aufindz0 Automatic adjustment of Z0 (M)		

AuCdec Carbon decoupler calibration macro (M)

Syntax	AuCdec	
Description	Used by AuCALch3i and AuCALch3oh autocalibration routines to do carbon decoupler calibrations. Calibrates high-power pulse widths and dmf.	
See also	System Administration	
Related	AuCALch3i AuCALch3oh dmf	Get autocalibration with CH ₃ I sample (M) Get autocalibration with Autotest sample (M) Decoupler modulation frequency for first decoupler (P)

AuCgrad Carbon/proton gradient ratio calibration macro (M)

Syntax	AuCgrad		
Description	Used by AuCALch3i1 and AuCALch3oh1 autocalibration routines for C/H gradient ratio calibrations.		
See also	System Administration		
Related	AuCALch3i1	Get autocalibration with CH_3I sample (M) Get autocalibration with Autotest sample (M)	
	AUCALCIISOIII	det autocambration with Autotest sample (m)	

AuCobs Carbon observe calibration macro (M)

Syntax	AuCobs	
Description	Used by AuCALch3i1 autocalibration routines for carbon observe calibrations.	
See also	System Administration	
Related	AuCALch3i1 Get autocalibration with CH ₃ I sample (M)	

audiofilter Audio filter board type (P)

Description	Sets the type of audio filter board used where the spectral width (sw) is less than 100 kHz. The filter type is set in the Spectrometer Configuration window (opened from config) using the label Audio Filter Type.			
Values	'b' indicates the system has a 100-kHz Butterworth filter board (100 kHz Butterworth choice in the Spectrometer Configuration window.).			
	'e' indicates the system has a 100-kHz elliptical filter board (100 Elliptical choice in the Spectrometer Configuration window).			
	'2' indicates the system has a 200-kHz Butterworth filter board (200 kHz Butterworth choice in the Spectrometer Configuration window).			
	'5' indicates the system has a 500-kHz elliptical filter board (500 kHz Elliptical choice in the Spectrometer Configuration window).			
See also	System Administration			
Related	configDisplay current configuration and possibly change it (M)swSpectral width in directly detected dimension (P)			

Aufindz0 Automatic adjustment of ZO (M)

Syntax	Aufindz0	
Description	Finds z0 by doing lock 1D spectrum. The frequency is then used along with the lkhzdac value in the probe file to calculate the z0 value for a given solvent and autolocking is done. This requires previous calibration of the hzdac value done using the Aucalibz0 macro.	
See also	System Administration	
Related	Aucalibz0 Automatic Hz to DAC calibration for Z0 (M)	

AugcalProbe gcal calibration macro (M)

Syntax Augcal

Description	Used by AuCAL	ch3i1 and AuCALch3oh1 autocalibration routines for	
	probe gcal cal	ibrations.	
See also	System Administration		
Related	AuCALch3i1	Get autocalibration with CH ₃ I sample (M)	
	AuCALch3oh1	Get autocalibration with Autotest sample (M)	
	gcal	Gradient calibration constant (P)	

Augmap Automated gradient map generation (M)

Syntax	Augmap	
Description	Automatically adjusts gradient level, offset, window, and pulse width to generate a $z1-z4$ gradient map using a 2-Hz D_2O sample. This macro is used by the Aumakegmap auto gradient map generation macro and is applicable only for a lock gradient map.	
See also	System Administration	
Related	AumakegmapAuto lock gradient map generation (M)gzsizeNumber of z-axis shims used by gradient shimming (P)	

Augmapz0Automatic lock gradient map generation and z0 calibration
(M)

Syntax	Augmapz0		
Description	Using the 2-Hz D_2O sample, the augmapz0 macro automatically creates a lock gradient map, followed by Hz to DAC calibration of Z0 for the autolocking procedure.		
See also	System Administration		
Related	Aucalibz0	Automatic Hz to DAC calibration for Z0 (M)	
	Aufindz0	Automatic adjustment of Z0 (M)	

AuHdec Proton decoupler calibration (M)

Syntax	AuHdec	
Description	Used by AuCALch3i autocalibration routine to do proton decoupler calibrations. Calibrates high-power pulse widths and dmf.	
See also	System Administration	
Related	AuCALch3i	Get autocalibration with CH3I sample (M)
	dmf	Decoupler modulation frequency for first decoupler (P)

Proton observe calibration macro (M) AuHobs

Syntax AuHobs

Description Used by AuCALch3i and AuCALch3oh autocalibration routines for proton observe calibrations.

Auto lock gradient map generation (M) Aumakegmap

Aumakegmap(<1k or hs or H1>) Syntax

Description Generates z1–z4 lock gradient ('1k' argument), lock homospoil ('hs' argument), or ¹H gradient map ('H1' argument). If no argument is given, the defaults is 'lk', if gradtype='nnh' to 'hs'. The doped 2-Hz D_2O should be used for hs and 1k maps. H1 map is typically done on the sample. Automatically adjusts gradient level, offset, window, and pulse width. The map name is automatically stored in the probe file.

Get parameters for a given nucleus (M) AuNuc

Syntax AuNuc (nucleus, solvent) Description Retrieves standard parameter set for a given nucleus and adds all required parameters for Tcl/dg driven parameters. If no parameter set exists in stdpar, then carbon parameters are retrieved and tn changed.

Prepare for an automation run (C) auto

Applicability Systems with an automatic sample changer. Syntax

auto<(automation_directory)>

Description Prepares the automation directory for an automation run. auto aborts if the spectrometer is already in automation mode.

automation_directory is the name of the automation directory, Arguments either an absolute UNIX path (i.e.the first character is a "/") or a relative path (the first character is not a "/"). The default is the value of the parameter autodir. If for some reason autodir is not defined, you are prompted to provide the location of the automation directory. If not given as an argument, you are prompted for the path. If the automation directory is not present, it is created with full access for all users. auto aborts if it fails to create this directory.

Examples auto auto('/home/vnmr1/autorun_620')

See also	NMR Spectroscopy User Guide, VnmrJ User Programming, Vnn Automation User Guide		
Related	auto_au	Controlling macro for automation (M)	
	autodir	Automation directory absolute pathname (P)	
	autogo	Start an automation run (C)	
	autoname	Prefix for automation data file (P)	

auto Automation mode active (P)

Applicability Systems with an automatic sample changer.

Description A global variable that shows whether or not an automation run is in progress. Macros typically test this parameter because actions can differ between the automation and non-automation modes. The value of auto is not enterable by the user. An automation experiment is initiated with the autogo command. The auto parameter is only set to 'y' for those macros and commands that are run as part of an automation experiment.

- Values 'y' indicates automation mode is active.
 - 'n' indicates automation mode is inactive
- See also NMR Spectroscopy User Guide, VnmrJ User Programming, VnmrJ Automation User Guide.
 - Relatedauto_auControlling macro for automation (M)autogoStart an automation run (C)autoraResume suspended automation run (C)autosaSuspend current automation run (C)

autoaa Abort an automation run with no error

Syntax	autoaa	
Applicability	VnmrJ 3.1	
Description	This command is used to abort an experiment that has been submitted to automation. The currently running experiment will not be interrupted, but when it is over, the automation run will be terminated.	
Arguments	The macro co	nsists of autosa and aa, run sequentially.
See also	For further information on autosa or aa, see the manual.	
Related	halt	halt acquisition with no error
	autora	resume the interrupted automation run

auto_au Controlling macro for automation (M)

Applicability Systems with an automatic sample changer.

Syntax auto_au

- Description Reads sampleinfo file (defines an automation experiment) using the lookup facility, sets the solvent and loc parameters based on the SOLVENT and SAMPLE# fields of sampleinfo, runs exec on the entry in the MACRO field, and writes the experiment text based on the TEXT field. After that, auto_au examines the value of the wexp parameter:
 - If wexp is set to 'procplot', then auto_au calls au.
 - If wexp is set to 'autolist', then auto_au inserts 'auto' as the first argument to autolist and calls au('wait').
 - If wexp is set to anything else, auto_au does not call au.

If no data is generated from the requested MACRO field, due to an error or some other reason, auto_au sets the STATUS field to "No Data Requested."

auto_au is used only during automation and should not be called directly. It provides a starting point for all automation experiments. As such, it is a convenient point for user customization of automation.

See also NMR Spectroscopy User Guide, VnmrJ User Programming, VnmrJ Automation User Guide

Related au Submit experiment to acquisition and process data (M) Prepare for an automation run (C) auto autolist Set up and start chained acquisition (M) exec Execute a VnmrJ command (C) Location of sample in tray (P) loc Look up words and lines from a text file (C) lookup Lock solvent (P) solvent wexp When experiment completes (P)

autog Utility commands for the automation queue

Syntax	autoq
Applicability	VnmrJ 3.1
Arguments	This command can contain the following arguments:
	• autoq('add',pathname): adds the sampleinfo file at pathname to the automation queue (enterQ). The pathname may contain multiple sampleinfo entries. An implicit lock is placed on the queue. An autosa / autora pair is not needed.
	• autoq('add',pathname,'priority'): adds the sampleinfo file at pathname to the automation queue (enterQ) with queue name 'priority'. The pathname may contain multiple sampleinfo entries. For the enterQ, 'priority' is interpreted as adding it to the top of the file. An implicit

lock is placed on the queue. An autosa / autora pair is not needed.

- autoq('lock',seconds): locks the automation queue (enterQ) so other processes will not access it. By default, all locks expire after 5 seconds. A second argument can set the expiration time between 1 and 15 seconds.
- autoq('unlock'): removes the lock.
- autoq('get',pathname): gets the next sampleinfo file from the automation queue (enterQ) and places it at pathname. An implicit lock is placed on the queue. An autosa / autora pair is not needed. This option will generally not be needed by user macros. This function is currently performed by Autoproc.
- autoq('sendmsg',message): Send "message" to whatever Vnmr session is listening. This is often used by background automation if it wants to send a message to a foreground Vnmr.
- autoq('recvmsg','on'): Turn on receiving messages from an autoq('sendmsg',message) command.
- autoq('recvmsg','off'): Turn off receiving messages from an autoq('sendmsg',message) command.

Autobackup Back up current probe file (M)

Syntax	Autobackup

Description Makes a copy of the probe file before starting the calibrations and prints the current calibration file. Autobackup is called by the autocalibration routines AuCALch3i1 and AuCALch3oh1

autodept Automated complete analysis of DEPT data (M)

Syntax	autodept			
Description	Processes DEPT spectra, plots the unedited spectra, edits the spectra, plots the edited spectra, and prints outs editing information.			
See also	NMR Spectroscopy User Guide, VnmrJ Automation User Guide			
Related	adept Automatic DEPT analysis and spectrum editing (C)			
	Dept Set up parameters for DEPT experiment			
deptproc		Process DEPT data (M)		
	padept	Perform adept analysis and plot resulting spectra		
		(C)		
	pldept Plot DEPT data, edited or unedited (M)			

autodir Automation directory absolute path (P)

Applicability Systems with an automatic sample changer or LC-NMR accessory. Description When using a sample changer, autodir is a global variable that holds the absolute path of the currently active automation directory. When VnmrJ is started, autodir is set to the absolute path of the last automation run. When using the LC-NMR accessory, autodir specifies a directory in which experiments using a stored queue are saved. See also NMR Spectroscopy User Guide Related auto Set up an automation directory (C) autoname Prefix for automation data file (P) globalauto Automation directory name (P) walkup Walkup automation (M)

autogo Start automation run (C)

Applicability	Systems with an automatic sample changer.		
Syntax	<pre>autogo<(file<,automation_directory>)></pre>		
Description	Starts an automation run. The autogo parameter cannot be entered while the spectrometer is in automation mode. You must have an enter queue prepared to start an automation run. The queue is checked to verify that it was prepared using the enter command (autogo aborts if an error in the format is found.) Your automation directory is also checked for the presence of a non-empty enter queue (autogo aborts if the current queue in the automation directory is present and not empty). Finally, autogo checks the automation directory and runs the auto command if this directory is not present or another problem is found. When autogo completes, the system is in automation mode and your automation run starts.		
Arguments	file is the file name of your <u>enter</u> queue. The default is that the system prompts you for the location of the <u>enter</u> queue.		
	automation The default i	_directory is the pathname of the automation directory. s the current value of the parameter <u>autodir</u> .	
Examples	autogo autogo('My autogo('My	Samples') Samples','/home/vnmr1/AutoRun_621')	
See also	NMR Spectroscopy User Guide		
Related	auto autodir autoname	Set up an automation directory (C) Automation directory absolute path (P) Prefix for automation data file (P)	
	enter	Enter sample information for automation run (C)	

Α

autolist Set up and start chained acquisition (M)

autolist(<options,>experiment1<,experiment2<,>) Sets up parameters for chained experiments by executing the experiments given as arguments and then starting a chained acquisition. Note that the macro au is executed as part of autolist</options,>		
Ind should not be included in the arguments to autolist. Options is one or more of the following keywords: Fauto' is a keyword to add 'wait' to the au call (e.g,		
au('wait', 'next')). 'start' is a keyword to make the first experiment in the list as one that needs to be acquired rather than processed.		
experiment1, experiment2, are experiments written as strings e.g., 'dept' or 'c13'). experiment1 is the current experiment and, when it finishes, the macro procplot is called to process the data. If experiment2 is listed, that experiment is executed and then the nacro au('next') is performed. For subsequent experiments, the ext, solvent and temp are used from the preceding experiment. Also, the wexp parameter is reset to 'autolist' with the first experiment removed.		
autolist('h1','c13','dept') autolist('h1','hcosy')		
NMR Spectroscopy User Guide		
Auto_auControlling macro for automation (M)Automated proton and carbon acquisition (M)Automated proton, carbon, and APT acquisition (M)Automated proton, carbon, and HETCOR acquisitionAutomated proton and COSY acquisition (M)Automated proton and COSY acquisition (M)Automatically process FIDs (M)		
eaac'a'te(verttraalaakkk k		

automerge Merges overniteQ with daytimeQ

Description This option is useful for sorting the short runs from longruns and merging the longruns at the back of short runs before doing <u>autogo</u> on an existing enter file. Alterntively, this macro can be used in a cron job to merge overniteQ with daytimeQ in a current automation run at a specified time. An optional 2nd argument will suppress <u>autosa/autora</u>. Optional argument allows one to merge overniteQ with daytimeQ for an enter file - i.e., non walkup mode.

Arguments add Arguments

Automkdir Creates Data Directory from Template

See also This macro is executed by automation at runtime to create the directory path specified in the Preferences/Templates panel.

autoname Create path for data storage (C)

Applicability Syntax	Automation autoname:\$path autoname(name_template):\$path autoname(name_template,sample_info_file):\$path autoname (name_template,sample_info_file, <'keepspaces' 'replacespaces'>):\$path autoname(name_template,sample_info_file, <,'excluded_suffixes'<,'keepspaces' 'replace spaces'>):\$path
	<pre>Svfname:\$path Svfname(name_template):\$path Svfname(name_template, suffix):\$path Svfname(name_template, suffix, 'excluded_suffixes'):\$path Svfname(name_template, suffix, 'excluded_suffixes', 'keepspaces' 'replacespaces'):\$path chkname(name_template, 'characters', 'par or tmpl or str','replacechar'):\$s1,\$s2,\$par,\$req chkname('fileChars', 'characters')</pre>
Description	The autoname command determines the path for data storage during an automation run and uses the value of a naming template (the autoname parameter by default) and the contents of a sample info file (default is sampleinfo in the current experiment) to determine this path. The path name is stored in the return argument or displayed on line 3 if no return argument is present.
	The name is prefaced with using the value of the parameter autodir or userdir+'/data/' if autodir is equal to ".
	The default excluded_suffix is.fid.
Arguments	No argument provided. The command uses the default autoname parameter and sampleinfo in the current experiment directory for the path to the sample info file. If the autoname parameter does not exist or is set to ", the default template is%SAMPLE#:%%PEAK#:%.
	<pre>name_template (no quotes) is string that contains keywords separated by substitution specifiers to represent the data storage path. Substitution specifiers in this template are either a percent sign (%) or a dollar sign (\$). The keywords are obtained from the sample_info_file file, if it exists, using % substitution specifiers or VNMR parameters using \$ substitution specifiers.</pre>

A template is passed directly using: autoname('\$owner\$/\$sample\$'):\$path.

Percent sign (%) substitution specifier is used with the autoname command to scan the sample_info_file for the text specific by keyword between the first percent sign in the template string and the next percent sign. The text specified by the keyword between the % substitution specifiers is passed to \$path.

The following percent substitutions (% keywords) for time and date are obtained from the system clock, not from the sample info file:

Keyword	Format	Description
%DATE% %TIME%	YYYYMMDD HHMMSS	4 digit year 2 digit month 2 digit day 2 digit each for hours minutes and
0111110		seconds
%YR%	YYYY	4-digit year
%YR2%	YY	2-digit year
%MO%	MM	2-digit month
%DAY%	DD	2-digit day
%HR%	HH	2-digit hour
%MIN%	MM	2-digit month
%SEC%	SS	2-digit second

The following are some of the percent substitutions (% keywords) are obtained from the second argument, sample_info_file.

Keyword	Description
%USER%	user name
%MACRO%	macro name
%SAMPLE%	sample name
SOLVENT%	solvent name

Version number is specified by Rn where n is an integer from 0 to 9 (default 2), as follows:

n=	Description
0	no revision digits are appended (all names must be
1 to 9	uniquely constructed without these revision digits). revision number is padded with leading zeroes to form
	an n-digit number. If more places are needed than
>9	specified, more zeroes are used. Rnn is still used as a search string in the sampleinfo
(more than one digit)	file. %Rn% must be specified at the end of the
	<pre>name_template string. The revision digits are</pre>
no %Rn%	always appended except if %R0% is used. default of %R2% is used

Specify the starting number to be used when constructing the version number by appending a colon : and start number after Rn.

The default starting value is 1. A zero is not allowed.
Dollar sign (\$) substitution specifiers works in manner analogous to the percent substitution specifier, except that the text between the dollar signs is interpreted as the name of a VNMR parameter. The value of this parameter is substituted for the substitution specifier.

Numeric parameters are represented as a string and truncated to an integer value. The template, pw=\$pw\$usec, with vnmr parameter pw having a value of 12.3 produces pw=12usec01 which is appended to.fid and passed to \$path. The 01 following usec is added by the %R2% default setting.

sample_info_file (no quotes) is the name of a text file to read for the % substitutions passed to autoname. The file must exist.

Using the keyword 'replacespaces' uses underscores (_) in place of spaces ' ' in the resulting path name or the keyword 'keepspaces' retains spaces in the resulting path name.

The keyword, 'keepspaces' or 'replacespaces' is an optional argument (includes quotes). The argument is accepted as the third or fourth argument.

Solaris and Linux operating systems default to replacespaces.

A comma separated list of excluded suffixes the new path name will not use or match is specified if the third keyword is not 'keepspaces' or 'replacespaces'.

Examples Using a \$ substitution specifier:

autoname(pw=\$pw\$usec):\$path

A substitution specifier, pw=\$pw\$usec, is the name_template and a relative path. The vnmr parameter, pw, has a value of 12.3 and the resulting filename is: pw=12usec01.fid. The path name is prefaced with the value of the parameter autodir if the name template generates a relative pathname.

Examples Using \$ substitution specifiers and a comma separated list of suffixes:

autoname('\$seqfil\$_\$tn\$_','/vnmr/conpar','.img'):\$path

The \$ substitution specifier is; \$seqfil\$_\$tn\$_ the dummy info filename is; '/vnmr/conpar', and the comma separated list of excluded suffixes is .img. The path name is prefaced with seqfil_tn_index. Each time a file is written to the directory the command changes the index by one (see %Rn% above). The suffix is both .fid and .img. The file is named gems_H1_03.img if target directory contains gems_H1_01.fid and gems_H1_02.img.

See also NMR Spectroscopy User Guide, VnmrJ User Programming, VnmrJ Automation User Guide

Related autoname Temple determining the path where is data stored (P) Svfname Determines the name used to store data (C) svfname Specifies the filename template (P) Applicability Automation

- Description The autoname temple determines the resulting path where the data is stored for an entry in the automation run and uses the contents of a sample info file (the name by default is "sampleinfo" in the current experiment) to determine this path. The path name is stored in the return argument and displayed on line 3 if no return argument is present.
 - See also NMR Spectroscopy User Guide, VnmrJ User Programming, VnmrJ Automation User Guide
 - Related autoname Determines path for data storage during an automation run (C).

autora Resume suspended automation run (C)

Applicability	Systems with an automatic sample changer.
Syntax	autora
Description	Resumes a previously suspended automation run. No matter what caused the interruption (including autosa, power failure, or system boot-up), the system examines the condition of the automation file and resumes acquisition for all experiments that have not finished. If autora is executed while an automation run is in progress, it has no effect.
See also	NMR Spectroscopy User Guide

Related autosa Suspend current automation run (C)

autosa Suspend current automation run (C)

Applicability	Systems with an automatic sample changer.	
Syntax	autosa	
Description	Suspends the automation mode at the conclusion of the current experiment and changes the system to the manual mode. The currently running experiment is not interrupted.	
See also	NMR Spectroscopy User Guide	
Related	autora Resume suspended automation run (C)	

autoscale Resume autoscaling after limits set by scalelimits macro (M)

Syntax	autoscale	
Description	Returns to autoscaling in which the scale limits are determined by the expl command such that all the data in the expl input file is displayed.	
See also	NMR Spectroscopy User Guide	
Related	expl	Display exponential or polynomial curves (C)
	scalelimits	Set limits for scales in regression (M)

autostack Automatic stacking for processing and plotting arrays (M)

Syntax	autostack	
Description	When processing and plotting arrayed 1D spectra, VnmrJ automatically determines whether the stacking mode is horizontal, vertical or diagonal from the number of traces and the number of lines in the spectrum. If this automatic function is not desirable (or makes an undesirable decision), it can be overridden by placing the stack macro in the experiment startup macro or by calling stack before processing (or reprocessing) a spectrum. autostack switches back to automatic determination of the stack mode by destroying the stackmode parameter.	
See also	NMR Spectro	scopy User Guide
Related	procarray	Process arrayed 1D spectra (M)
	plarray	Plot arrayed 1D spectra (M)
	stack	Fix stacking mode for processing / plotting arrayed spectra (M)
	stackmode	Stacking control for processing (P)

autotest Open Auto Test Window (C)

Syntax	autotest
Description	Opens the Auto Test window.
See also	AutoTest Software manual.

autotime Displays approximate time for automation (M)

Syntax autotime(<automation directory>)

Set abs. value mode in 1st indirectly detected dimension (C)

- Α
- Description Displays approximate time for each experiment and for each location in an automation run. If no argument is given, time is calculated for the current automation run (enterQ). See also NMR Spectroscopy User Guide

Related explist Display approximate time for current experiment chain (M)

av

Set abs. value mode in directly detected dimension (C)

Syntax av

Description Selects the absolute-value spectra display mode by setting the parameter dmg to the string value 'av'. In the absolute-value display mode, each real point in the displayed spectrum is calculated as the square root of the sum of the squares of the real and imaginary points comprising each respective complex data point. All information, including noise, is always positive, and the relationship between signal and noise is linear.

> For multidimensional data, av has no effect on data prior to the second Fourier transform. If pmode='full', av acts in concert with commands ph1, av1, or pwr1 to yield the resultant contour display for the 2D data.

See also NMR Spectroscopy User Guide

Related av1 Set abs. value mode in 1st indirectly detected dimension (C) av2 Set abs. value mode in 2nd indirectly detected dimension (C) Display mode in directly detected dimension (C) Absolute-value display of FID data or spectrum in acqi (P) dmqf ft Fourier transform 1D data (C) Fourier transform along f_2 dimension (C) ft1d ft2d Fourier transform 2D data (C) Set phase angle mode in directly detected dimension (C) pa pa1 Set phase angle mode in 1st indirectly detected dimension (C) Set phased mode in directly detected dimension (C) ph Set phased mode in 1st indirectly detected dimension (C) pmode Processing mode for 2D data (P) pwr1 Set power mode in 1st indirectly detected dimension (C) wft Weigh and Fourier transform 1D data (C) wft1d Weigh and Fourier transform of 2D data (C) wft2d Weigh and Fourier transform 2D data (C)

av1

Syntax av1

148

In the *absolute-value display mode*, each real point in the displayed trace is calculated as the square root of the sum of the squares of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the real-real and imaginary-real points from each respective hypercomplex data point are used in the summation. In this mode, all information, including noise, is always positive; and the relationship between signal and noise is linear.

The avl command is only needed if mixed-mode display is desired. If the parameter dmgl does not exist or is set to the null string, the display mode along the first indirectly detected dimension defaults to the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of avl is the same as for traces provided that pmode='partial' or pmode='' (two single quotes with no space between).

See also NMR Spectroscopy User Guide

Related av Set abs. value mode in directly detected dimension (C) dmg1 Data display mode in 1st indirectly detected dimension (P)

av2

Set abs. value mode in 2nd indirectly detected dimension (C)

Syntax av2

Description

Selects absolute-value spectra display mode for the second indirectly detected dimension by setting the parameter dmg2 to the value 'av2'. If dmg2 does not exist or is set to the null string, av2 creates dmg2 and set it equal to 'av2'.

In the *absolute-value display mode*, all information, including noise, is positive; and the relationship between signal and noise is linear. Each real point in the displayed trace is calculated as the square root of the sum of the squares of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the real-real and imaginary-real points from each respective hypercomplex data point are used in the summation.

The av2 command is only needed if mixed-mode display is desired. If the parameter dmg2 does not exist or is set to the null string, the display mode along the second indirectly detected dimension defaults to the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of av2 is the same as for traces provided that

ed av	Set abs. value mode in directly detected dimension
	(C)
dmg2	Data display mode in 2nd indirectly detected
	dimension (P)

averag Calculate average and standard deviation of input (C)

Syntax	<pre>averag(number1,number2,):average,sd, number_arguments,sum_numbers,sum_squares</pre>	
Description	Finds average, standard deviation, and other characteristics of a set of numbers.	
Arguments	number1, number2, is a finite set of numbers.	
	average is the average of the numbers.	
	sd is the standard deviation of the numbers.	
	number_arguments is the number of number1,number2, arguments.	
	sum_numbers is the sum of the numbers	
	sum_squares is the sum of squares of the numbers.	
Examples	averag(3.4,4.3,3.5,5.4):r1,r2	
See also	VnmrJ User Programming	

awc

Additive weighting const. in directly detected dimension (P)

Description Adds the current value of awc to each value of the weighting function along the directly detected dimension. This dimension is often referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, and so forth. awc is applied *after* the sinebell and exponential function, but *before* the Gaussian function. This allows using gf as a Gaussian apodization even when awc is non-zero. Typical value of awc is 'n'.

See also NMR Spectroscopy User Guide

Related awcl Additive weighting const. in 1st indirectly detected dimension (P)

awc2 Additive weighting const. in 2nd indirectly detected dim. (P) gf Gaussian function in directly detected dimension (P)

awc1 Additive weighting const. in 1st indirectly detected dimension (P)

Description Adds the current value of awc1 to each value of the weighting function along the first indirectly detected dimension This dimension is often referred to as the f_1 dimension of a multidimensional data set. awc1is analogous to the parameter awc. The "conventional" parameters (1b, gf, etc.) operate on the detected FIDs, while this "2D" parameter is used during processing of the interferograms.

See also NMR Spectroscopy User Guide

Related awc Additive weighting const. in directly detected dimension (P)

awc2 Additive weighting const. in 2nd indirectly detected dimension (P)

- Description Adds the current value of awc2 to each value of the weighting function along the second indirectly detected dimension This dimension is often referred to as the f_2 dimension of a multidimensional data set. awc2is analogous to the parameter awc. The value of awc2 can be set with wti on the 2D interferogram data.
 - See also NMR Spectroscopy User Guide
 - Related awc Additive weighting const. in directly detected dimension (P) wti Interactive weighting (C)

axis Provide axis labels and scaling factors (C)

Syntax	axis('fn' 'fn1' 'fn2') <:\$axis_label,\$freq_scaling,\$scaling_factor>
Description	Displays or returns values of the axis labels and scaling factors to the calling macro. See the macro rl for an example of using this command.
Arguments	'fn' 'fn1' 'fn2' is the Fourier number parameter for the axis of interest.
	<pre>\$axis_label is the axis label (e.g., ppm, kHz, cm, or ppm(sc)).</pre>
	<pre>\$freq_scaling is the divisor needed to convert from units of Hz to the units defined by the axis parameter with any scaling. axis uses the current value of the axis parameter for that dimension and also checks for axis scaling using the corresponding scalesw, scalesw1, or scalesw2 parameter.</pre>
	\$scaling_factor is a second scaling factor, determined solely by the scalesw type of parameter. This last scaling factor is independent of the value of the axis parameter.

Examples	axis('fn') axis('fn1'):\$lab,\$fr,\$scl	
See also	VnmrJ User Programming	
Related	axis rl scalesw scalesw1 scalesw2	Axis label for displays and plots (P) Set reference line (M) Scale spectral width in directly detected dimension (P) Scale spectral width in 1st indirectly detected dimension (P) Scale spectral width in 2nd indirectly detected dimension (P)

axis Axis label for displays and plots (P)

Applicability	Certain arguments work only if system has the proper hardware.
Description	Specifies the units for the axis display and plot.
	For 1D experiments, axis uses a single letter that includes 'h' for Hz, 'p' for ppm, and 'k' for kHz (e.g., $axis='h'$).
	For 2D experiments, axis uses two letters, with the first letter describing the detected spectral axis (f_2), and the second letter describing the indirectly detected axis (f_1). Thus axis='ph' is appropriate for a homonuclear 2D-J experiment, with a referenced ppm scale along the spectral axis and an axis in Hz ('h') along the J-axis. axis='pp' is appropriate for COSY or NOESY experiments.
	For 3D experiments, axis uses three letters with the first letter describing the detected spectral axis (f_3) , the second letter describing the first indirectly detected axis (f_1) , and the third letter specifying the second indirectly detected axis (f_2) .
	The special letter d is used to reference the indirectly detected axis to the parts per million of the decoupler channel, as appropriate for heteronuclear chemical shift correlation experiments, which would typically have axis='pd'. The letter n is used to suppress the axis display on one or both axes (e.g., axis='nn', axis='pn').
	For systems with multiple decouplers, the characters '1', '2', and '3' can be used to reference an axis relative to the frequency of that decoupler. Setting axis='p1' is effectively the same as axis='pd'.
Values	'1' sets the axis label for units of ppm relative to the first decoupler. '2' sets the axis label for units of ppm relative to the second decoupler.
	'3' sets the axis label for units of ppm relative to the third decoupler.
	 'c' sets the axis label for units of centimeters. 'd' sets the axis label for units of ppm relative to the first decoupler. 'h' sets the axis label for units of hertz.
	'k' sets the axis label for units of kilohertz. 'm' sets the axis label for units of millimeters. 'n' sets no axis label display.

A

'p' sets the axis label for units of ppm relative to the observe transmitter.
 'u' sets the axis label for units of micrometers.
 See also NMR Spectroscopy User Guide
 Related axis Provide axis labels and scaling factors (C) axisf Axis label for FID displays and plots (P) dscale
 Display scale below spectrum or FID (C)

Plot scale below spectrum or FID (C)

axisf Axis label for FID displays and plots (P)

pscale

Description	Specifies the units for the FID axis display and plot. To create the FID
	display parameters axisf, dotflag, vpf, vpfi, crf, and deltaf (if
	the parameter set is older and lacks these parameters), enter addpar('fid').
Values	's' sets the axis label for units of seconds.
	'm' sets the axis label for units of ms

- 'u' sets the axis label for units of μ s.
- 'n' sets no axis label display.
- See also NMR Spectroscopy User Guide
- Related addpar Add selected parameters to the current experiment (M) axis Axis label for displays and plots (P) dscale Display scale below spectrum or FID (C)
 - pscale Plot scale below spectrum or FID (C)



Agilent VnmrJ 4 Command and Parameter Reference Guide

B

bandinfo	Shaped pulse information for calibration (M)
banner	Display message with large characters (C)
bc	1D and 2D baseline correction (C)
beepoff	Turn beeper off (C)
beepon	Turn beeper on (C)
bigendian	Determine system byte order (C)
binom	Set up parameters for BINOM pulse sequence (M)
bioref	Bio-NMR Referencing (P)
bootup	Macro executed automatically (M)
box	Draw a box on a plotter or graphics display (C)
boxes	Draw boxes selected by the mark command (M)
bpa	Plot boxed parameters (M)
bph	Individually phase each trace of arrayed 1D data (C)
br24	Set up parameters for BR24 pulse sequence (M)
bs	Block size (P)

bandinfo Shaped pulse information for calibration (M)

Applicability Syntax	<pre>Information only useful on systems capable of shaped pulse generation. bandinfo<(shape,width<,ref_power>)>:duration,power</pre>
Description	Displays a table containing the duration and the predicted 90° pulse power setting for the pulse shape and bandwidth given by the arguments. No parameter settings are changed. The necessary data is contained in the shapeinfo file in the shapelib subdirectory.
Arguments	If bandinfo is run without arguments, prompts operator for input
	shape is the name of the shape. The default is system prompts for a name.
	width is the bandwidth, in Hz, desired for the pulse.
	<code>ref_power</code> is value of <code>tpwr</code> to which <code>pw90</code> is set. The default is 55 dB.



	duration is the duration, in μs , of the pulse.		
	power is th	e predicted 90° pulse power setting.	
Examples	bandinfo bandinfo('sinc' 10) nw towr	
See also	User Programming		
Delated	1	Shanad nulse information for colibration (M)	
Related	pulseinio pw90	90° pulse width (P)	
	tpwr	Observe transmitter power level with linear amplifiers	
		(P)	

banner Display message with large characters (C)

Syntax	<pre>banner(message<,color>)</pre>	
Description	Displays text as large-size characters on the graphics windows.	
Arguments	s message is the text to be displayed. If the text includes a single quotation mark ('), it must be preceded by a backslash (\'). Multilin displays are available by inserting two backslashes (\\) between line Any undefined characters are displayed as a "bug" shape.	
	color is the color of text on a color display: 'red', 'yellow', 'green', 'cyan', 'blue', 'magenta', and 'white'. The default is 'yellow'.	
Examples	banner('banner sample') banner('Don\'t Touch','blue')	
See also	User Programming	

bc

1D and 2D baseline correction (C)

Description Makes 1D or 2D baseline correction using a spline or a second to twentieth order polynomial fitting of predefined baseline regions. bc defines every odd-numbered integral (those integrals that disappear when intmod='partial') as baseline and attempts to correct these points to zero.

Arguments

<> indicates that an argument is optional; the absence therefore indicates that an argument is mandatory.

The optional argument 'ifnotddr' will skip the baseline correction if the data were acquired with a system with a direct-digital receiver. On these systems, the baseline is usually flat enough not to warrant additional corrections. If the data were not acquired with a system with a direct-digital receiver, the argument is ignored. The 'ifnotddr' argument must follow the 'f1' or 'f2' arguments if they are used. It must precede all other arguments. The bc command will return to the calling macro its status. It returns a 0 if it failed. It returns a 1 if it succeeds. If the 'ifnotddr' argument is given and the bc operation is skipped, because the data were acquired with a direct-digital receiver system, it returns a 2. If the bc command fails and a return value is not requested, it will abort the calling macro.

1D

bc(<arg1>):

arg1 - (A) order of the polynomial used in the fitting procedure; the default value is 1, which obtains a spline fit; ranges from 1 to 20. (B) 'unbc'; does not require that the order is specified; BC reads in the order and the coefficients used in the previous BC operation and reverses the BC operation; only functional for 1D baseline correction operations on either 1D spectra or individual 2D traces.

bc('alt'<scale<, smooth>>) - An alternative automatic baseline correction method developed by Carlos Cobas, etc... This method automatically finds signal-free regions using CWT (Continuous Wavelet Transform derivatives), and models the baseline using uses Whittaker smoother algorithm. This method should be used with proper selection of "scale" and "smooth" parameters, in order to avoid "over correction" (resulting in an unrealistically flat baseline).

scale - A scaling factor for calculating derivatives for CWT. Increasing "scale" reduces noise, but broadens the signals. "scale" may be calculated as a proportional function of the broadest signal. If scale is too small, the broad peaks will be treated as baseline instead of peaks. The value of "scale" may range from 1 to a few hundreds. The default is 200 (a relatively large value to avoid losing broad peaks).

smooth - A smooth factor for baseline modeling. It increases smoothness at the expense of spectral fidelity (worse fitting of the data). The value of "smooth" may range from 100 to tens of thousands. The default value is 10000.

bc('dis') -calculate baseline of displayed 1D trace according to parameter bcmode, and display the baseline with spectrum (without correction).

bc('all') - calculate and apply baseline correction for each trace of arrayed 1D spectra, and save baseline(s) in curexp+'/datdir/bc.fdf'.

bc('apply',name) - apply baseline correction according to baseline file *.fdf. The baseline file needs to be loaded prior to calling this command. The command to load baseline file isaipLoadSpec(fdfFile,name).

bc('cancel') - undo baseline correction.

2D

bc(arg1,<arg2>,<arg3>,<arg4>):

arg1 - 'f1' or 'f2'; specifies the direction along which the 2D BC is to take place.

arg2 - order of the polynomial used in the fitting procedure; the default value is 1,which obtains a spline fit; ranges from 1to 20.

arg3 - trace number for the spectrum on which the 2D BC is to commence; must lie within the appropriate range or an error will result.

 $\arg 4$ - trace number for the spectrum on which the 2D BC is to terminate; must lie within the appropriate range or an error will result.

See also NMR Spectroscopy User Guide

Related	dc	Calculate spectral drift correction (C)
	fn	Fourier number in directly detected dimension (P)
	intmod	Integral display mode (P)
	trace	Mode for 2D data display (P)
	wft1da	Weight and Fourier transform phase-sensitive data (M)
	wft2da	Weight and Fourier transform phase-sensitive data (M)

beepoff Turn beeper off (C)

Description	Turns off the beeper sound so that the system does not use sound to
	warn the user when errors occur. The default is the beeper is turned
	on.
See also	User Programming
Related	beepon Turn beeper on (C)

beepon Turn beeper on (C)

Syntax	beepon	
Description	Turns on the beeper sound so that the user hears a sound when errors occur. The default is the beeper is turned on.	
See also	User Programming	
Related	beepoff Turn beeper off (C)	

bigendian Determine system byte order (C)

Syntax	bigendian:\$type
Description	The bigendian
Description	command determines the system byte order for storing numbers. One architecture is Big Endian, used by Sun computers with the "Sparc" CPU'S. The other architecture is Little Endian, used by most PCs.

Return values to argument \$type:

1 if it is a "Big Endian" system.

0 if it is a "Little Endian" system.

This command should rarely be used. Its only current use is when imaging .fdf files are created. The .fdf file headers can specify whether the data is stored as big or little endian.

binom Set up parameters for BINOM pulse sequence (M)

Description Sets up a binomial water suppression pulse sequence. See also NMR Spectroscopy User Guide

bioref Bio-NMR Referencing (P)

Applicability	All		
Syntax	bioref=' <y n="" or="">'</y>		
Description	Flag, global or local, for Bio-NMR Referencing. Setting the flag (bioref='y') sets the system to bio-NMR referencing (based on nuctables/nuctabrefBio) rather than standard IUPAC / organic chemistry referencing (based on nuctables/nuctabref). Bio-NMR referencing uses DSS for nuclei such as ¹³ C and liquid NH ₃ for ¹⁵ N.		
	Creating bioref as a local parameter (create('bioref','flag') creates a local flag) permits its use for a specific case. The parameter can be created as a local parameter and saved with a standard parameter set (stdpar/N15) to enable bio-NMR referencing for a specific nucleus. The local value of the parameter takes precedence over the global parameter.		
	<pre>create('bioref','flag','global') - creates a global flag. setenumeral('bioref',2,'y','n','global') - sets the possible values of a string parameter in a parameter tree.</pre>		
Examples	bioref='y' sets referencing to use nuctables/nuctabrefBio		
Related	Create new parameter in a parameter tree (C)		

bootup Macro executed automatically (M)

Syntaxbootup<(foreground)>DescriptionExecuted automatically when VnmrJ is started up. The bootup macro
displays a message, looks for a macro login in the user's local maclib
directory and executes it (if found), starts Acqstat and acqi (acqi

is not run if system is configured as a workstation), and then starts the menu system. This set of actions can be modified on a per user basis by constructing custom bootup or login macros in the user's maclib directory. A custom login macro is preferred because all custom bootup macros are overridden whenever a new VnmrJ release is installed.

- Arguments foreground is 0 if VnmrJ is being run in the foreground or nonzero if being run in the background. This argument is passed to the login macro.
 - See also User Programming

Related	acqi	Interactive acquisition display process (C)
	Acqstat	Bring up the acquisition status display (U)

box Draw a box on a plotter or graphics display (C)

Syntax	<pre>box(<'keywords',>x1mm,x2mm,y1mm,y2mm <,'nolimit'>)<:r1,r2></pre>
Description	Draws a box on a plotter or a graphics display.
Arguments	<pre>'keywords' identifies the output device ('graphics' 'plotter'), drawing mode ('xor' 'normal'), and drawing capability ('newovly' 'ovly' 'ovlyC').</pre>
	• 'graphics' 'plotter' is a keyword for the output device. The default is 'plotter'. The output selected is passed to subsequent pen, move, or draw commands and remains active until a different output is specified.
	• 'xor', 'normal' is a keyword for the drawing mode when using the 'graphics' output device. The default is 'normal'. In the 'xor' mode, if a line is drawn such that one or more points of the line are in common with a previous 'xor' line, the common points are erased. In the normal mode, the common points remain. The mode selected is passed to subsequent pen, move, and draw commands and remains active until a different mode is specified.
	• 'newovly', 'ovly' and 'ovlyC' are keywords that specify an interactive drawing capability that is slightly slower than the 'xor' mode but more consistent in color. 'newovly' clears any previous draws, boxes, and writes made with the 'ovly' modes and draws the figure. 'ovly' draws without clearing so that multi-segment figures can be created. 'ovlyC' clears without drawing.
	x1mm is the left edge of the box, x2mm is the right edge, y1mm is the bottom, and y2mm is the top. The location of the edges are given in plotter units (mm on most plots) and are scaled in mm for the graphics display. (If units are in Hz or ppm, you can use the hztomm command to convert units.)
	'nolimit' allows the box to extend outside the limits determined by the parameters sc , wc , $sc2$, and $wc2$.
	r1, r2 return the location of the upper left corner of the box.

Examples	box('plotter',20,100,40,150) box(25,105,45,155,'nolimit'):r1,r2	
See also	NMR Spectroscopy User Guide	
Related	gin	Return current mouse position and button values
		(C)
	hztomm	Convert positions from Hz or ppm to plotter units
		(C)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	WC	Width of chart (P)
	wc2	Width of chart in second direction (P)
	wcmax	Maximum width of chart (P)

boxes Draw boxes selected by the mark command (M)

Syntax	<pre>boxes<('graphics' 'plotter')></pre>
Description	Draws boxes on a plotter or a graphics display with the location of the edges given in Hz. The data to make the boxes is stored in the mark2d.out file produced by the mark command. If there is no data in mark2d.out, a box is drawn from the current cursor positions. The boxes command also numbers the boxes above the upper left corner.
Arguments	'graphics' 'plotter' is a keyword to send output to the graphics display or to the plotter, respectively. The default is 'graphics'.
Examples	boxes boxes('plotter')
See also	NMR Spectroscopy User Guide
Related	mark Determine intensity of spectrum at a point (C)

bpa Plot boxed parameters (M)

Syntax	bpa:\$sc2_minimum	
Description	Plots a box around the entire chart (assuming blank paper) and then plots "chemist-style" parameters in boxes along the lower edge of the chart. bpa is the same as ppa, but with a different layout. Both ppa and bpa behave somewhat naively if the pulse sequence is more complex, but they were designed primarily for chemists, not for spectroscopists.	
Arguments	$sc2_minimum$ returns the minimum value for $sc2$ to plot a scale properly. To use the command pir, vp has to be set to a non-zero value.	
See also	NMR Spectroscopy User Guide	
Related	apaPlot parameters automatically (M)papPlot out "all" parameters (C)	

pir	Plot integral amplitudes below spectrum (C)
ppa	Plot a parameter list in "English" (M)
sc2	Start of chart in second direction (P)
vp	Vertical position of spectrum (P)

bph Individually phase each trace of arrayed 1D data

Syntax bph<(trace)> - auto phase a given trace, or each trace of an arrayed 1D data.

bph0<(trace)> - zero-order auto phase a given trace, or each trace of an arrayed 1D data.

bph('write'<,path>) - save phase parameters in a text file. Default
path is curexp/datdir/bph.txt

bph('read'<,path>) - read phase parameters from a text file.

bph('on') - turn on bph mode.

bph('off') - turn off bph mode.

Description This command is implemented to individually phase each trace of arrayed 1D data.

Phase parameters are stored in block headers (each block contains a single trace).

All bph commands, except bph('off'), automatically turn on bph mode. In bph mode, phase parameters in block headers will be used. User may interactively phase a selected trace.

When bph mode is off, rp, lp parameters will be used for all blocks (traces).

bph<(trace)> or bph0<(trace)> automatically write out curexp/datdir/bph.txt.

phase file format is (for example):

comment # trace-index bph-flag rp lp 0 1 -138.523163 0.000000 1 1 -138.406555 0.000000 2 1 -138.799194 0.000000 3 1 -138.848038 0.000000 4 1 39.744347 0.000000 5 1 40.512058 0.000000 6 1 40.141430 0.000000

•••••

bph-flag is 1 if aph algorithm returns successfully, 0 if failed. This flag only serve informative purpose.

If phase file is written with bph('write'<,path>) command, bph-flag is zero for all traces.

br24 Set up parameters for BR24 pulse sequence (M)

Applicability	Systems with solids hardware.	
Description	Converts a FLIPFLOP, MREV8, or S2PUL parameter set into a BR24 solids line-narrowing multiple-pulse sequence.	
See also	User Guide:	Solid-State NMR
Related	cylbr24	Set up parameters for cycled BR24 pulse sequence (M)
	flipflop	Set up parameters for FLIPFLOP pulse sequence (M)
	mrev8	Set up parameters for MREV8 pulse sequence (M)
	s2pul	Set up standard two-pulse sequence (M)

bs Block size (P)

Description Directs the acquisition computer, as data are acquired, to periodically store a block of data on the disk, from where it can be read by the host computer.

CAUTION

If bs='n', block size storage is disabled and data are stored on disk only at the end of the experiment. If the experiment is aborted prior to termination, data will be lost.

Values	1 to 32767 tr	ansients, 'n'
See also	NMR Spectro	scopy User Guide
Related	wbs wbs	Specify action when <i>bs</i> transients accumulate (C) When block size (P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

c13	Automated carbon acquisition (M)
c13p	Process 1D carbon spectra (M)
calcECC	Calculate ECC corrections (C)
calcdim	Calculate dimension of experiment (C)
calfa	Recalculate alfa so that first-order phase is zero (M)
calibflag	Correct systematic errors in DOSY experiments (P)
calibrate	Start a dialog for autocalibration routines (M)
callacq	Utility macro to call Acq command (M)
capt	Automated carbon and APT acquisition (M)
Carbon	Set up parameters for 13C experiment (M)
cat	Display one or more text files in text window (C)
cattn	Coarse attenuator type (P)
cd	Change working directory (C)
cdc	Cancel drift correction (C)
cdept	Automated carbon and DEPT acquisition (M)
cdump	Prints the current graphics screen (M)
celem	Completed FID elements (P)
center	Set display limits for center of screen (C)
centerprobe	Calculates probe position relative to the ISO-Center.
centersw	Move cursor to center of spectrum (M)
centersw1	Move cursor to center of spectrum in 1st indirect dimension (M)
centersw2	Move cursor to center of spectrum in 2nd indirect dimension (M)
cexp	Create an experiment (M)
cf	Current FID (P)



chkname	Parse the template and return substituted strings and lists of parameters defined by the template
cfpmult	Calculate first-point multiplier for 2D experiments (M)
change	Submit a change sample experiment to acquisition (M)
checkstring	Find and replace unwanted characters (C)
chiliConf	Control flag set by ecc_on and ecc_off (P)
Cigar2j3j	Convert the parameter to a CIGAR2j3j experiment (M)
ckresloc	Macro to Reserve Specific Locations
ckstring	Utility to Check String Variables for Illeagal Characters
cla	Clear all line assignments (M)
cla	Calculated transition number (P)
clamp	Calculated transition amplitude (P)
cleanexp	Remove old files and directories from an experiment (M)
clear	Clear a window (C)
cleardosy	Delete temporarily saved data in current sub experiment (M)
clfreq	Calculated transition frequency (P)
clindex	Index of experimental frequency of a transition (P)
clradd	Clear add/subtract experiment (C)
color	Select plotting colors from a graphical interface (M)
cmdlineOK	Determine if an operator has a command line
coldprobe	Tells system a coldprobe is present.
combiplate	View a color map for visual analysis of VAST microtiter plate (U)
combishow	Display regions (red, green, and blue) in CombiPlate window (M)
compressfid	Compress double-precision FID data (M,U)
config	Display current configuration and possibly change it (M)
confirm	Confirm message using the mouse (C)
Console	System console type (P)
contact_time	MAS cross-polarization spin-lock contact time (M)

continflag	The command ddif creates a CONTIN display if continflag='y'.
continprepare	Called by the macro dosy to prepare the input file for the CONTIN programme.
continread	Called by the macro dosy to take the output of the CONTIN programme and create an input file for ddif.
continueMovie	Continue movie in either forward or backward direction (C)
convert	Convert data set from a VXR-style system (M,U)
convertbru	Convert Bruker data (M,U)
сору	Copy a file (C)
cos	Find cosine value of an angle (C)
Cosy	Convert the parameter to a COSY experiment (M)
cosyps	Set up parameters for phase-sensitive COSY pulse sequence (M)
ср	Copy a file (C)
ср	Cycle phase (P)
cpdone	Macro called upon study completion (M)
cpgo	Macro called upon study completion (M)
cpmgt2	Set up parameters for CPMGT2 pulse sequence (M)
cpos_cvt	Convert data set from a VXR-style system (M,U)
cptmp	Copy experiment data into experiment subfile (M)
cptmpltdefaults	Defaults for Save Data Template
срх	Create pbox shape file (M)
cqexp	Load experiment from protocol (M)
cqfindz0	Run an experiment to find the value of z0 (M)
cqgmap	Perform gradient shimming utility functions (M)
cqinit	Initialize liquids study queue (M)
cqpars	Create study queue parameters for liquids (M)
cqplot	Macro to perform generic 2D plot (M)
cqprotocol	Macro to create protocols (M)
cqreset	Reset study queue parameters (M)
cqsavestudy	Macro to save study queue parameters (M
cqwtmenu	Macro to set weighting functions from a panel (M)
cqreset cqsavestudy cqwtmenu	Reset study queue parameters (M) Macro to save study queue parameters (M Macro to set weighting functions from a panel (M)

cr	Cursor position in directly detected dimension (P)
crl	Cursor position in 1st indirectly detected dimension (P)
cr2	Cursor position in 2nd indirectly detected dimension (P)
crcom	Create user macro without using text editor (M)
create	Create new parameter in a parameter tree (C)
create(P)	Parameter used for RF transmitter board temperature compensation
createqcomp	Create qcomp parameter (M)
crf	Current time-domain cursor position (P)
crl	Clear reference line in directly detected dimension (M)
crl1	Clear reference line in 1st indirectly detected dimension (M)
crl2	Clear reference line in 2nd indirectly detected dimension (M)
crmode	Current state of the cursors in df, ds, or dconi programs (P)
crof2	Recalculate rof2 so that lp = 0 (M)
cryo_noisetest	Run Cold Probe conditioning experiments (M)
cryoclient	Start the CryoBay Monitor program (M, U)
CSschedule	Generates a NUS schedule
csv2cpQ	Imports CSV Data (M)
ct	Completed transients (P)
ctext	Clear the text of the current experiment (C)
curexp	Current experiment directory (P)
curscan	Scan currently in progress (P)
curwin	Current window (P)
cutoff	Data truncation limit (P)
cyclenoe	Set up parameters for CYCLENOE pulse sequence (M)
cylbr24	Set up parameters for cycled BR24 pulse sequence (M)
cylmrev	Set up parameters for cycled MREV8 pulse sequence (M)
cz	Clear integral reset points (C)

C

c13 Automated carbon acquisition (M)

Syntax c13<(solvent)>

Description	Prepares p	arameters for automatically acquiring a standard $^{13}\mathrm{C}$	
	spectrum.	The parameter wexp is set to 'procplot' for standard	
	processing.	If c13 is used as the command for automation via the	
	enter con	mand, the au is supplied automatically and should not be	
	entered on	the MACRO line of the enter program. However, it is	
	possible to customize the standard c13 macro on the MACRO line by		
	following it with additional commands and parameters. For example,		
	c13 nt=1	uses the standard c13 setup but with only one transient.	
Arguments	solvent i	solvent is the name of the solvent. In automation mode the solvent	
	is supplied	by the enter program. The default is 'CDC13'.	
Examples	c13		
•	c13('DMS0))	
See also	NMR Spectroscopy User Guide		
Related	au	Submit experiment to acquisition and process data (M)	
	c13p	Process of 1D carbon spectra (M)	
	enter	Enter sample information for automation run (C)	
	proc1d	Processing macro for simple (non-arrayed) 1D spectra (M)	
	procplot	Automatically process FIDs (M)	
	wexp	When experiment completes (P)	

c13p Process 1D carbon spectra (M)

Syntax	c13p	
Description	Processes non-arrayed 1D carbon spectra using a set of standard macros. c13p is called by the proc1d macro, but can also be used directly. Fully automatic processing (up to a point where a spectrum could be plotted) is provided: Fourier transformation (using pre-set weighting functions), automatic phasing (aphx macro), automatic integration (integrate macro if required only), vertical scale adjustment (vsadjc macro), avoiding excessive noise (nois1m macro), threshold adjustment (thadj macro), and referencing to the TMS signal if present (setref macro then tmsref macro).	
See also	NMR Spectros	scopy User Guide
Related	aphx I c13 I integrate I proc1d I	Perform optimized automatic phasing (M) Automated carbon acquisition (M) Automatically integrate 1D spectrum (M) Limit noise in spectrum (M) Processing macro for simple (non-arrayed) 1D spectra (M)

setref	Set frequency referencing for proton spectra (M)
thadj	Adjust threshold (M)
tmsref	Reference spectrum to TMS line (M)
vsadjc	Adjust vertical scale for carbon spectra (M)

calcdim Calculate dimension of experiment (C)

Syntax calcdim

Description Calculates the dimension of an experiment and puts the result into the parameter arraydim. If an experiment is arrayed, arraydim is the product of the size of the arrays.
 See also NMR Spectroscopy User Guide

Related arraydim Dimension of experiment (P)

calcECC Calculate ECC corrections (C)

Syntax calcECC(infile, outfile)

The calcECC command requires two arguments. The first is a pathname to a reference data set to be used to do the ECC corrections. The first argument should be the name of a ".fid" directory, containing a data set saved by VnmrJ. The second argument is a filename where to place the results. A typical value would be curexp+'/eccref', as in

calcECC(userdir+'/data/waterref.fid',curexp+'/ecc
ref')

In this case, the parameter fidecc=curexp+'/eccref' accesses this information for the ft('ftargs') and wft('ftargs') commands. The calcECC commands calculated the phase angle for each data point in the FID and writes it to the output file.

calfa Recalculate alfa so that first-order phase is zero (M)

Syntax	calfa
Description	Based upon the current alfa and lp values, calfa calculates a new value for alfa so that the first-order phase parameter lp is rendered approximately 0. When digital filtering is active $(dsp='r' \text{ or } dsp='i')$, calfa also adjusts rof2 as well as alfa. For calfa to work properly, a trial spectrum must be obtained and phased to pure absorption. This spectrum provides calfa with the current alfa and lp values. calfa pertains to processing 2D data. Unless lp is approximately 0, fpmult will affect both the dc offset and the
	curvature of the spectrum.

See also	so NMR Spectroscopy User Guide		
Related	alfa	Set alfa delay before acquisition (P)	
	cfpmult	Calculate first-point multiplier for 2D experiments (M)	
	crof2	Recalculate rof2 so that $lp = 0$ (M)	
	dc	Calculate spectral drift correction (C)	
	dsp	Type of DSP for data acquisition (P)	
	fpmult	First-point multiplier for np FID data (P)	
	hoult	Set parameters alfa and rof2 according to Hoult (M)	
	lp	First-order phase in directly detected dimension (P)	
	rof2	Receiver gating time following pulse (P)	

calibflag Correct systematic errors in DOSY experiments (P)

Syntax	calibflag		
Description	Corrects systematic errors in DOSY experiments.		
Values	'y' corrects systematic deviations in DOSY analysis.		
	'n' omits gradient correction in DOSY analysis.		
See also	NMR Spectroscopy User Guide		
Related	dosy Process DOSY experiments (M)		

calibrate Start a dialog for autocalibration routines (M)

Syntax	calibrate
Description	Starts a dialog for autocalibration routines

callacq Utility macro to call Acq command (M)

Syntax	callacq(arg_string)		
Description	Utility macro to construct a string to pass to psg via the $Acq()$ command. This macro should be used only by users with advanced knowledge. A well-constructed argument string is required. The motivation for this macro is to make the 'go' macro re-entrant, while still synchronizing with VnmrJ.		
Arguments	arg_string is a character string constructed from a macro.		
Examples	callacq(\$callback)		
Related	goSubmit experiment to acquisition (M)reqparcheckFlag which enables/disables required parameters (P)		

reqparclear	Clears the parameters in required parameter list (M)
reqparlist	List of required parameters (P)
reqpartest	Tests whether required parameters are set (M)

capt Automated carbon and APT acquisition (M)

Syntax capt<(solvent)>

Description Prepares parameters for automatically acquiring a standard ¹³C spectrum, followed by an APT experiment. In non-automation mode, the carbon and APT spectra are acquired in the experiment in which capt is entered. Following acquisition completes, the commands rttmp('C13') and rttmp('apt') can be used for further processing of the carbon and APT spectra, respectively. solvent is name of the solvent used. In automation mode, the enter Arguments program supplies name. In non-automation mode, the default is 'cdc13'. Syntax capt au capt('dmso') See also NMR Spectroscopy User Guide Related Apt Prepare parameters for APT experiment (M) Automated carbon acquisition (M) Enter sample information for automation run (C) enter Retrieve experiment subfile (M) rttmp

Carbon Set up parameters for 13C experiment (M)

Description Set up parameters for ¹³C experiment

cat Display one or more text files in text window (C)

Syntax	<pre>cat(file1<, file2>)</pre>
Description	Displays the contents of one or more text files on the text window. It pauses after the window has filled and waits for the user to indicate whether it should display more or should terminate.
Arguments	file1, file2, are the names of the files to be displayed.
Examples	cat('/vnmr/manual/cat') cat('/vnmr/manual/cat','/vnmr/manual/cattn')
See also	NMR Spectroscopy User Guide

cattn Coarse attenuator type (P)

Applicability Systems with a coarse attenuator.

- Description Identifies the type of coarse attenuator if this attenuator is present on the current rf channel. The value of cattn is set in the Spectrometer Configuration window (opened by entering config) using the label Coarse Attenuator.
 - Values 0 for no coarse attenuator, as in the case with class C amplifiers (Not Present choice in Spectrometer Configuration window).

79 for standard systems (79 dB choice in Spectrometer Configuration window).

127 for imaging attenuator (63.5 dB SIS choice in Spectrometer Configuration window).

63 for deuterium decoupler channel.

- See also VnmrJ Installation and Administration
 - Related config Display current configuration and possibly change it (M) fattn Fine attenuator (P) tpwr Observe transmitter power level with linear amplifiers (P)

cđ

Change working directory (C)

Syntax	cd<(directory)>		
Description	Changes current working directory to another directory.		
Arguments	directory is the name of the directory that becomes the new current working directory. The change is made only if the directory name already exists and the user has permission to be in the directory. If no argument is included, cd changes the current working directory to the user's home directory.		
Examples	cd cd(userdir+'/exp1') cd('/home/george/vnmrsys')		
See also	NMR Spectroscopy User Guide		
Related	pwd Display current working directory (C)		

cdc Cancel drift correction (C)

SyntaxcdcDescriptionTurns off the drift correction started by the dc command and resets
the spectral drift correction parameters lvl (level) and tlt (tilt) to
zero.

See also	NMR S	pectroscopy User Guide
Related	dc	Calculate spectral drift correction (C)
	dcg	Drift correction group (P)

acg	Drift correction group (P)
lvl	Zero-order baseline correction (P)
tlt	First-order baseline correction (P)

cdept Automated carbon and DEPT acquisition (M)

Syntax cdept<(solvent)>

Prepares parameters for automatically acquiring a standard ¹³C Description spectrum, followed by a DEPT experiment. In non-automation mode, the carbon and DEPT spectra are acquired in the experiment in which cdept was entered. Following the completion of the acquisition, the rttmp('C13') and rttmp('dept') commands can be used for further processing of the carbon and DEPT spectra, respectively. Arguments solvent is name of the solvent used. In automation mode, the enter program supplies name. In non-automation mode, the default is 'cdc13'. Examples cdept au cdept('DMSO') See also NMR Spectroscopy User Guide Automatic DEPT analysis and spectrum editing (C) Related adept Automated carbon acquisition (M) Prepare parameters for DEPT experiment (M) dept enter Enter sample information for automation run (C) Retrieve experiment subfile (M) rttmp

cdump Prints the current graphics screen (M)

Syntax cdump(`filename')

Description cdump takes the current display and sends it to the current printer. If an optional filename is passed as an argument, the current display will be saved in the print subdirectory of the user's vnmrsys directory. This directory will be created if is does not already exist. If the filename passed to the cdump macro is an absolute pathname, i.e., it starts with a '/' character, that pathname will be used.

> If the current display is saved as a file, the format of the file is specified by the printformat parameter. It can be set to the following values. as for PostScript formatted output.

japed for Joint Photographic Experts Group JFIF formatted output. nag for Portable Network Graphics formatted output.

celem Completed FID elements (P)

Description Indicates the current number of completed FIDs in an experiment. When go or au is entered, celem is set to 0. As each FID acquisition is completed, celem is updated to reflect this. This parameter is most useful in conjunction with wbs, wnt, wexp, and werr processing commands.

See also NMR Spectroscopy User Guide

Related	arraydim	Dimension of experiment (P)	
	au	Submit experiment to acquisition and process data (C)	
	go	Submit experiment to acquisition (C)	
	ni	Number of increments in 1st indirectly detected dimension	
		(P)	
	wbs	Specify action when bs transients accumulate (C)	
	werr	Specify action when error occurs (C)	
	wexp	Specify action when experiment completes (C)	
	wnt	Specify action when nt transients accumulate (C)	

center Set display limits for center of screen (C)

Description Sets parameters sc and wc (horizontal control) and parameters sc2 and wc2 (vertical control) to produce a display (and subsequent plot) in the center portion of the screen (and page). For 2D data, space is left for the scales.

NMR Spectroscopy User Guide See also Related full Set display limits for a full screen (C) Set display limits for full screen with room for traces (C) fullt left Set display limits for left half of screen (C) Set display limits for right half of screen (C) right Start of chart (P) sc Start of chart in second direction (P) sc2 Width of chart (P) WC Width of chart in second direction (P) wc2

centerprobe Calculates probe position relative to the ISO-Center

Syntax	centerprobe
Applicability	VnmrJ 3.1
Description	A macro that calculates the rf probe position relative to the ISO-center. Using the Z2 plot from the current gradient map this macro calculates how far out of center the probe is.
Arguments	centerprobe will display the results on the graphics screen.

centerprobe('plot') will plot the results on the selected plotter.

centersw Move cursor to center of spectrum (M)

Description	Sets cursor position parameter cr in the directly detected dimension for the center of the spectrum.	
See also	NMR Spectroscopy User Guide	
Related	centersw1	Move cursor to center of spectrum in 1st indirect dimension (M)
	centersw2	Move cursor to center of spectrum in 2nd indirect dimension (M)
	cr	Cursor position in directly detected dimension (P)

centersw1 Move cursor to center of spectrum in 1st indirect dimension (M)

Description	Sets cursor position parameter cr1 in the first indirectly detected				
	dimension t	o the center of the spectrum.			
See also	NMR Spectroscopy User Guide				
Related	centersw	Move cursor to center of spectrum (M)			
	cr1	Cursor position in 1st indirectly detected dimension (P)			

centersw2 Move cursor to center of spectrum in 2nd indirect dimension (M)

Description	Sets cursor position parameter cr2 in the second indirectly detected		
	dimension to the center of the spectrum.		
See also	NMR Spectroscopy User Guide		
Related	centersw	Move cursor to center of spectrum (M)	
	cr2	Cursor position in 2nd indirectly detected dimension (P)	

cexp Create aVnmr experiment (M)

Syntax cexp(<exp_dir,>exp_no):\$stat,\$message Description cexp creates a VNMR experiment which is used as a temporary work space and can hold a complete 1D, 2D, or 3D data set. The cexp macro will copy the "current" and "processed" parameter trees to the newly created experiment's curpar and procpar files. If the current

experiment is exp0, or experiments 1-4 are being created as part of the "auto" command, the curpar and procpar files for the newly created experiment are copied from the procpar file in /vnmr/fidlib/fidld.fid. The cexp macro calls the CEXP command to create the experiment and copy the parameters. The cexp macro then does the additional step of creating an appropriate jexp<N> macro in the user's maclib, where the <N> is the exp_no and if exp_no is greater than 9. The macros jexp1 to jexp9 do not need to be created since they exist as standard VnmrJ macros. Both the CEXP command and cexp macro will return two optional values to the calling macro. The first (\$stat) is set to 0 if the command / macro fails and it is set to 1 if the command / macro succeeds. The second return value is a text message that can be used. exp_no specifies the VNMR experiment number and must be between 1 and 9999. exp_dir is an optional argument which specifies the path of the directory in which the particular experiment is to be created. If exp_dir is not entered, the default directory is the VNMR user directory (userdir). cexp(3)Examples cexp('/data',2) cexp(32):\$stat,\$msg // silently create exp32 and the //jexp32 macro. \$stat reports //success or failure. The \$msg //message could be displayed. CEXP(33):\$stat,\$msg // silently create exp33, but do not //make the jexp33 macro. \$stat //reports success or failure. The //\$msg message could be displayed. See also NMR Spectroscopy User Guide Related delexp Delete an experiment (C) Join existing experiment (C) jexp userdir User directory (P)

cf Current FID (P)

Description Specifies which FID to operate on when working with multi-FID data. All subsequent operations such as Fourier transformation are applied to the selected data block.
When an experiment acquires nf number of data segments through explicit acquisition, cf indicates the cfth FID to use. For example, in the COSY-NOESY experiment with nf=2, cf=1 would select the COSY part of the experiment, and cf=2 would select the NOESY part.
Values 1 through the value of parameter nf.

С

cfpmult Calculate first-point multiplier for 2D experiments (M)

Description Calculates an fpmult value for the dataset, which is then used by wft2da. For 2D experiments, such as NOESY, run cfpmult on the transformed first increment, prior to entering wft2da, to minimize "f₂ ridges" in the final 2D spectrum. To do this manually for a 2D dataset, enter fpmult=1.0 wft(1) cdc in the command line and note whether the spectrum (essentially the baseline) moves up or down when dc is typed. Vary the value of fpmult until the dc correction (jump in the baseline) is as small as possible. With care, fpmult can be set to two decimal places. Typical values for fpmult range from 1.00 to 2.00. The default value is 1.0.

This calculation only needs to be performed for cosine-type experiments, such as NOESY, where both the t_2 FID and the t_1 interferogram decay. cfpmult might give incorrect values for first increments of experiments having baseline distortions (e.g., water suppression with 11-echo or 1331); in such cases, manual optimization of fpmult is more suitable.

When processing 2D data, unless the parameter lp is approximately 0, fpmult affects both the dc offset and the curvature of the spectrum. See the entries for alfa and calfa for more information.

See also NMR Spectroscopy User Guide

Related	alfa	Set alfa delay before acquisition (P)	
	calfa	Recalculate alfa so that first-order phase is zero (M)	
	crof2	Recalculate rof2 so that $lp = 0$ (M)	
	dc	Calculate spectral drift correction (C)	
	fpmult	First point multiplier for np FID data (P)	
	lp	First-order phase in directly detected dimension (P)	
	wft2da	Weight and Fourier transform phase-sensitive data (M)	

change Submit a change sample experiment to acquisition (M)

Systems with automatic sample changer.		
Removes the sample currently in the probe and loads the sample currently in sample location loc. change runs in the acquisition computer and is inoperative if loc is 0 and/or traymax is 'n' or 0. change also sets all hardware according to the current parameters.		
NMR Spectroscopy User Guide		
auSubmit experiment to acquisition and process data (C)gaSubmit experiment to acquisition and FT the result (C)		
	Systems with automatic sample changer. Removes the sample currently in the probe and loads the sample currently in sample location loc. change runs in the acquisition computer and is inoperative if loc is 0 and/or traymax is 'n' or change also sets all hardware according to the current parameters <i>NMR Spectroscopy User Guide</i> au Submit experiment to acquisition and process data (C) ga Submit experiment to acquisition and FT the result (C)	

go	Submit experiment to acquisition (C)
loc	Location of sample in tray (P)
lock	Submit an autolock experiment to acquisition (C)
sample	Submit change sample, Autoshim experiment to acquisition
	(M)
shim	Submit an Autoshim experiment to acquisition (C)
spin	Submit a spin setup experiment to acquisition (C)
su	Submit a setup experiment to acquisition (M)
traymax	Sample changer tray size (P)

checkstring Find and replace unwanted characters (C)

Syntax	checkstring('\$VALUE',variable):variable			
Description	checkstring is used panel to check and replace user-entered strings like samplename, notebook, or page for Unix-unfriendly characters:			
	" " (blank space) , ; : * ! ? (" ") [" "] {" "} <" "> ~ # \$ & /			
	Data may be saved to unexpected directories (or not at all) with Save Data Setup (used for automatic saving of NMR data) if operating system special characters are used within a filename.			
	An error/warning message is issued and the respective character(s) is/are replaced with an underscore, Multiple consecutive characters are replaced by one single underscore. Example: samplename = 'special			
	type of (new) sample' becomes 'special_type_of_new_sample'.			

chiliConf Control flag set by ecc_on and ecc_off (P)

Applicability	Systems with Varian, Inc. Cold Probes		
Description	Control flag set by ecc_on and ecc_off macros		
Values	E — enable PSG control of ECC n — disable PSG control of ECC		
Related	ecc_on ecc_off	Turns on eddy current compensation for Cold Probes (M) Turns off eddy current compensation for Cold Probes (M)	

chkname Parse the template and return substituted strings and lists of parameters defined by the template

Syntax chkname(name_template, 'characters', 'par or tmpl or str', 'replacechar'):\$s1,\$s2,\$par,\$req chkname('fileChars', 'characters')

Applicability VnmrJ 3.1

- Description The chkname command takes an argument that is a template type of the form used by the <u>Svfname</u> command. It parses the template and returns substituted strings and lists of parameters defined by the template. The chkname command will substitute parameters enclosed in the "\$" substitution parameters. If the string of characters between the "\$ pair" does not correspond to an existing parameter, the first \$ character will be treated as a simple character. The chkname command will also substitute the time and date "% pairs" described above. It does this for all "% pairs" except the %Rn% or %Rn:number% specifiers. Like the <u>Svfname</u> command, the chkname command does not read a sample info file.
- Arguments Following any substitutions, the chkname command will substitute characters in the resulting string, based on specified rules. The second argument to chkname specifies the characters that are allowed in the resulting string, or those that are disallowed in the resulting string. This is done because certain characters in filenames are either not allowed or they may make tools that use filenames difficult to use. Characters that often cause problems are:

' ', '!', '"', '\$', '&', '\'', '(', ')', '*', ';', '<', '>', '?', '\\', '[', ']', '^', '`', '{', '}', '|', ',', '\0'

This second optional argument is either a keyword, a keyword plus modifiers, or a list of disallowed characters. Below are the keywords and the subset of characters they allow. The default is the keyword 'dir'.

'file' allows all alphanumeric characters [a-z, A-Z, and 0-9] and _ and . characters. These are typically used for file names.

'dir' allows all characters allowed by the 'file' keyword plus the directory specifier '/'.

'alnum' allows all alphanumeric characters [a-z, A-Z, and 0-9] The alnum keyword can be followed by a list of other allowable characters.

'alnum_.' is identical to 'file'.

'alnum_./' is identical to 'dir'

'alnum_./:@%?=-&' might be used for email or web addresses. 'none' disables the character substitutions.

Supplying a list of characters is interpreted as disallowed characters. An example may be ' . , ; : * ! ? () [] { } <>~# %&/ '

A special incantation of the chkname command will set the allowed characters selected by the 'file' and 'dir' keywords. The command chkname('fileChars','_') specifies the _ character in addition to alphanumerics as allowed characters for the 'file' keyword. The 'dir' keyword adds the '/' character to the 'file' set.

The third optional argument is the keyword 'par' or 'tmpl' or 'str' Actually, only the initial 'p', 't', or 's' is required, the longer name suggests the usage. The 'tmpl' (or 't', or 'template') keyword will do the "% pair" and "\$ pair" substitutions before replacing disallowed

characters. The 'str' (or 's', or 'string') keyword does not give any special meaning to the '%' or '\$' characters. If they are found and they are in the disallowed list, they will be replaced with the replacement
character. The 'par' (or 'p' or 'parameter') keyword is identical to the 'tmpl' keyword with respect to the first returned string. However, for the optional second returned string, it does not do any "\$ pair" or "% pair" substitutions. It also does not replace the '\$' characters with '#' characters (see below) as is the case with the 'tmpl' keyword. The 'par' keyword is the default.

A fourth and final optional argument specifies the replacement character for any disallowed characters . The default is an underscore ('_'). A null string will remove any disallowed characters from the string.

The chkname command returns up to four values to the calling macro.

The first value returned is the expanded template. All places where a parameter is defined are substituted with the value of that parameter. If a used string parameter is an empty string, an empty string will be substituted for the template parameter. All "% pairs", except the %Rn% pairs will be substituted.

The second returned value depends on the value of the fourth optional argument to chkname. In the case of the 'tmpl' fourth argument, the second returned value is the same as the first returned value, except for the way in which empty string parameters are handled. In this case, instead of replacing the \$parname\$ with an empty string, it is replaced with the parameter name enclosed in # symbols. In the case of the 'str' fourth argument, the second returned value is identical to the first returned value. In the case of the default 'par' fourth argument, the second returned value is identical to the second returned value has no "\$ pair" nor "% pair" substitutions.

The third returned value is a list of parameters defined by the template.

The fourth returned value is a subset all the defined parameters that are set to an empty string. This can be used to identify parameters that must be set before a template can be fully expanded.

In summary, the chkname command with a single argument will do the "\$ pair" and "\$ pair" substitutions (except for the %Rn% pairs) and replace any characters other than alphanumerics [a-zA-ZO-9] and '_', '.', and '/'. The chkname command should always be called before the Svfname command, which will do the final %Rn% pair substitution. The default values for the second, third, and fourth arguments will often be correct.

Examples operator='vnmr1'

```
comment='A special compound'
samplename='C17H21NO4'
ident=''
$val='local'
chkname('$samplename$ $$val$
$ident$/$operator$_$comment$_%R2%'):$s1,$s2,$p,$r
```

```
sets $s1 =
'C17H21N04_local_/vnmr1_A_special_compound_%R2%'
    $s2 =
'C17H21N04_local_#ident#/vnmr1_A_special_compound_%R2%'
    $p = 'samplename $val ident operator comment'
    $r = 'ident'
    chkname('$samplename$
$ident$/$operator$_$comment$_%R2%','file','par','.'):$s
1,$s2,$p,$r
    sets $s1 = 'C17H21N04.vnmr1_A.special.compound_%R2%'
    $s2 =
'$samplename$.$ident$.$operator$_$comment$_%R2%'
    $p = 'samplename ident operator comment'
    $r = 'ident'
```

Cigar2j3j Convert the parameter to a CIGAR2j3j experiment (M)

Syntax Convert the parameter to a CIGAR2j3j experiment.

ckresloc Macro to Reserve Specific Locations

Description This macro checks the automation.conf file for any reserved locations. \$ Operator specific reservations are in the following format: cppref_reserveloc_operator: 1 2 4. Similarly, dayQ and nightQ specific reservations are in the format: cppref_DAYQ_ONLY: 1 2 3 4 and cppref_NIGHTQ_ONLY: 41 42 43 neitherQ (i.e., blocked), respectively. Specific reservations are in the format: cppref_NEITHERQ: 23 38. Any location not specifically assigned are always allowed. If cppref_SMSLOCATION: is set to next, all locations except NEITHERQ are allowed.

Syntax ckresloc

ckstring Utility to Check String Variables for Illegal Characters

Description This macro tests string variables for illegal characters. Syntax ckstring('\$VALUE',<argument2>):\$return. Examples ckstring('samplename',2):\$samplename

cla Clear all line assignments (M)

Syntax cla

Description	Clears the line assignment parameters clindex and slfreq for spin simulation iteration, which matches simulated spectra to actual data.	
See also	NMR Spectroscopy User Guide	
Related	assign	Assign transitions to experimental lines (M)
	dla	Display line assignments (M)
	clindex	Index of experimental frequency of a transition (P)
	slfreq	Measured line frequencies (P)

cla Calculated transition number (P)

Description	A global arrayed parameter that stores the transition number of
	calculated transitions of the spin simulation program when they are
	above a threshold set by sth. In the iterative mode, the cla value of
	an assigned transition is associated with an experimental frequency
	whose index is the clindex value.
See also	NMR Spectroscopy User Guide

Related	clamp	Calculated transition amplitude (P)
	clfreq	Calculated transition frequency (P)
	clindex	Index of experimental frequency of a transition (P)
	sth	Minimum intensity threshold (P)

clamp Calculated transition amplitude (P)

- Description A global arrayed parameter that stores the transition amplitude of calculated transitions of the spin simulation program when they are above a threshold set by the parameter sth. Enter dla('long') to display clamp.
 - See also NMR Spectroscopy User Guide

Related	cla	Calculated transition number (P)
	clfreq	Calculated transition frequency (P)
	clindex	Index of experimental frequency of a transition (P)
	dla	Display line assignments (C)
	sth	Minimum intensity threshold (P)

C

Syntax	<pre>cleanexp<(file1<, file2<,>>)></pre>		
Description	Removes experiment subfiles from chained experiments that exist in an experiment directory. cleanexp only cleans the currently active experiment.		
Arguments	file1, file2, are specific experiment subfiles to be removed. If no argument is given, all files in curexp/subexp are removed.		
Examples	cleanexp cleanexp('H1','relayh')		
See also	NMR Spectroscopy User Guide		
Related	curexpCurrent experiment directory (P)hccorrAutomated proton, carbon, and HETCOR acquisition (M)hcosyAutomated proton and COSY acquisition (M)		

clear Clear a window (C)

Syntax	<pre>clear<(window_number)></pre>
ription	Clears one of the four windows on the G

Description	Clears one of the four windows on the GraphOn terminal (status, input, graphics, text) or one of the two windows on the Sun (text and graphics).
Arguments	window_number is the number $(1 \text{ to } 4)$ of the window to be cleared:
	• 1 clears the status window (GraphOn only)

- 2 clears the graphics window
- 3 clears the input window (GraphOn only)
- 4 clears the text window (the default value).

Examples clear clear(2)

See also User Programming

cleardosy Delete temporarily saved data in current sub experiment (M)

Syntax	cleardosy
Description	Deletes any copies of DOSY data temporarily saved in the current sub experiment.
See also	NMR Spectroscopy User Guide
Related	dosy Process DOSY experiments (M)

Calculated transition frequency (P) clfreq

sth

- Description A global arrayed parameter that stores the transition frequency of calculated transitions of the spin simulation program when they are above a threshold set by the parameter sth. Enter dla to display clfreq.
 - See also NMR Spectroscopy User Guide Related cla Calculated transition number (P) Calculated transition amplitude (P) clindex Index of experimental frequency of a transition (P) dla Display line assignments (M) Minimum intensity threshold (P)

Index of experimental frequency of a transition (P) clindex

Description	A global arrayed parameter where each value contains the index of an experimental frequency assigned to the associated calculated transition for use in iterative spin simulation. Use assign to make the assignments. A value of zero indicates no assignment.	
See also	NMR Spectroscopy User Guide	
Related	assign	Assign transitions to experimental lines (M)
	cla	Clear line assignments (M)
	cla	Calculated transition number (P)
	dla	Display line assignments (M)

clradd Clear add/subtract experiment (C)

Description Deletes the add/subtract experiment (exp5). The add-subtract experiment number is defined by the global addsubexp parameter. The clradd program uses the delexp command to delete the add-subtract experiment. It takes the same return values as the delexp command. These can be used to suppress messages. See "delexp Delete an experiment (M)" on page 241 for a description of the return values. See also NMR Spectroscopy User Guide Related add Add current FID to add/subtract experiment (C) Subtract current FID from add/subtract experiment (C) sub

cmdlineOK Determine if an operator has a command line

Applicability VnmrJ 3.1

The VnmrJ administrator interface is used to grant access to the command line.

If the operator has access to the command line, the cmdlineOK program will return a 1 to the calling macro.

In the example below, sok will be set to 1. It will be set to 0 if the command line is not available.

cmdlineOK:\$ok

An optional argument can be provided. This will be the return value if command line access is not granted. For example,

cmdlineOK(-1): \$ok will set \$ok to -1 if command line access is not granted. This can be used by the interface designed so that a button may be either "grayed out" or removed if command line access is not granted.

coldprobe Tells system a coldprobe is present

Applicability VnmrJ 3.1

Description The coldprobe macro tells the system that a coldprobe is present so that the rof2 rule is enforced.

Arguments If a C13 observe coldprobe is being used, the value of rof2 should not be less than 350 usec.

color Select plotting colors from a graphical interface (M)

- DescriptionDisplays a window with color palettes for selecting colors for plotting
the background of the display screen, spectrum, integral, FID, etc.See alsoNMR Spectroscopy User GuideRelatedplPlot spectra (C)
 - setcolor Set colors for graphics window and for plotters (C)

combiplate View a color map for visual analysis of VAST microtiter plate (U)

Syntax(From UNIX) combiplateDescriptionOpens the CombiPlate window, which provides a map of microtiter
plate, allowing data to be viewed from individual sample wells. The
window enables viewing integral region intensities by colors and color
densities.

See also	NMR Spectroscopy User Guide	
Related	combishow	Display regions as red, green, and blue in CombiPlate window (M)
	dlivast	Produce text file and process last wells $\left(M\right)$

combishow Display regions (red, green, and blue) in CombiPlate window (M)

Syntax	combishow()	r,g,b)
Description	Displays integral regions shown on the spectrum as red (r) , green (g) , and blue (b) in the CombiPlate window. CombiPlate reads the regions automatically. 1, 2, or 3 integral regions can be designated. At least one integral region must be specified. Combishow displays spectra associated with individual wells.	
See also	NMR Spectroscopy User Guide	
Related	combiplate	View a color map for visual analysis of VAST microtiter plate (U)
	dlivast	Produce text file and process last wells (M)

compressfid Compress double-precision FID data (M,U)

Syntax	compressfid(<infiddir,>outFIDdir) (From UNIX) compressfid -i inFIDdir -o outFIDdir -f (From UNIX) compressfid -e exp_number -o outFIDdir -f</infiddir,>
Description	Compresses double-precision FID data to single-precision and updates the parameter dp in the file propcar. compressfid can be run through a macro interface in VnmrJ or directly at the UNIX level. In entering FID directory names, leave off the .fid directory extension.
Arguments	inFIDdir is the double-precision FID directory to be compressed. If inFIDdir is not entered, the default FID directory is curexp/acqfil.
	outFIDdir is the FID directory to receive the output.
	<code>exp_number</code> is the number of the experiment that contains the FID data.
	-i specifies that the next argument is the input FID directory.
	$- \circ $ specifies that the next argument is the output FID directory.
	$-{\rm e}$ specifies that the next argument is the number of the experiment that contains the FID data. The $-{\rm e}$ and the $-{\rm i}$ options are mutually exclusive.
	-f specifies that any existing directory with the name outFIDdir.fid is to be overwritten. Note that the macro interface

always overwrites any preexisting directory with the name specified by outFIDdir.fid. Examples compressfid('/vnmr/fidlib/fidld', 'testfidld')compressfid('testfidld') (From UNIX) compressfid -e 5 -o testfidld -f (From UNIX) compressfid -i /vnmr/fidlib/fidld -o testfidld -f See also NMR Spectroscopy User Guide Related dp Double precision (P)

config Display current configuration and possibly change it (M)

Syntax	config <('display')>				
Applicability	VnmrJ 3.2				
Description	The "config" command displays the current configuration. The configuration can be changed if the console is in use and the user has write access to the following:				
	system global parameter file				
	the "stdpar" link in the VNMR system directory				
	the file that the "stdpark" link points to				
	the "tests" link" in the VNMR system directory,				
	the file that the "tests" link points to				
	• the VNMR system directory				
	In this situation, a window will appear on top of the VNMR windows. The single argument 'display' will PREVENT the interactive mode from operating.				
	Usually, the VNMR system manager will configure the system once, and the set the protection on the parameter file to permit read access only by other users.				
	In interactive mode, a separate panel appears. In non-interactive mode, the current choices are displayed in the text window. See installation manual for details on the choices.	e the			
Arguments	'display' is a keyword that the system administrator can use a make config run in the display mode rather than the interactive mode.	to e			
Examples	config config('display')				
See also	VnmrJ Installation and Administration				
Related	amptypeAmplifier type (P)audiofilterAudio filter type (P)cattnCoarse attenuator (P)				
	Console System console type (P)				
	rine attenuator (r)				

fifolpsize	FIFO loop size (P)
gradtype	Gradients for X, Y, and Z axes (P)
hlfreq	Proton frequency of spectrometer (P)
latch	Frequency synthesizer latching (P)
lockfreq	Lock frequency (P)
numrfch	Number of rf channels (P)
overrange	Frequency synthesizer overrange (P)
parmax	Parameter maximum values (P)
parmin	Parameter minimum values (P)
parstep	Parameter step size values (P)
ptsval	PTS frequency synthesizer value (P)
rfchtype	Type of rf channel (P)
rftype	Type of rf generation (P)
rfwg	RF waveform generator (P)
rotorsync	Rotor synchronization (P)
shimset	Type of shim set (P)
sysgcoil	System gradient coil (P)
system	System type (P)
traymax	Sample changer tray slots (P)
vttype	Variable temperature controller present (P)

confirm Confirm message using the mouse (C)

Syntax	confirm(message):response		
Description	Displays a dialog box with the specified message and two buttons: Confirm and Cancel. Clicking on the buttons with the mouse produces a return value.		
Arguments	message is a single-line muticharacter string to be shown in the dialog box.		
	response is 1 if the user clicks the left button of the mouse on the Confirm button or presses the Return key; response is 0 if the user clicks the mouse on the Cancel button.		
Examples	confirm('Are you sure you want pw>100?'):\$response		
See also	User Programming		

Console System console type (P)

Description A global parameter that sets the type of system console The value is usually set using the Console label in the Spectrometer Configuration window (opened from config).When go, au, or ga is entered, the value of the Console parameter is copied from the systemglobal parameter tree to the current

experiment and named as the console parameter (lowercase c). If

console does not exist in an old parameter set, rt via fixpar creates it and sets it to ''. Both console and Console are type acquisition. Macros can use Console and console to take conditional action based on spectrometer type.

See also VnmrJ Installation and Administration Related au Submit experiment to acquisition and process data (M) config Display current configuration and possibly change it (M) fixpar Correct parameter characteristics in experiment (M) ga Submit experiment to acquisition and FT the results (M) rt Retrieve FIDs (M) go Submit experiment to acquisition (M)

system System type (P)

contact_timeMAS cross-polarization spin-lock contact time (M)

Applicability	Systems with solids module.				
Description	Processes data obtained using an array of values for a pulse-length parameter. It runs the UNIX program expfit, which does an exponential curve fitting that determines the value of <i>Tch</i> and <i>T1rho</i> . The output is matched to the equation				
	$I = [S0 - (S0 - S_inf)^*exp(-T/Tch))^*exp(-T/T1rho)) + S_inf$				
	where Tch is the time constant of a spin-locked cross-polarization process, and $T1rho$ is relaxation time of ¹³ C polarization in the proton rotating field.				
	The required input is file fp.out from the program fp and the values of the arrayed parameter. The output table is file analyze.list in the current experiment. The file analyze.out is used by the expl to display the results.				
See also	User Guide: Solid-State NMR				
Related	expfitLeast-squares fit to polynomial or exponential curve (U)explDisplay polynomial/exponential curves (C)fpFind peak heights (C)				

continflag The command ddif creates a CONTIN display if continflag='y'.

Syntax	continflag
Applicability	VnmrJ 3.1
Description	Tells the command ddif to create a 2D display using data produced by the CONTIN program. Set by the dosy macros, does not normally need to be set manually.
Arguments	continflag = 'y'

continflag = 'n'

See also dosyproc

continprepareCalled by the macro dosy to prepare the input file for the CONTIN programme.

Syntax	continprepare
Applicability	VnmrJ 3.1
Description	continprepare takes the dosy_in file created in dosy and creates the file dosy_contin.in the format required by the CONTIN programme (http://s-provencher.com/index.shtml).
See also	continread dosy
	splmodprepare

continread Called by the macro dosy to take the output of the CONTIN programme and create an input file for ddif.

Syntax	continread
Applicability	VnmrJ 3.1
Description	continread takes a file dosy_contin.out as created by CONTIN programme, run by the continrun shell script from the dosy macro, and creates the files diffusion_display. contin and diffusion_spectrum used by ddif and sdp to display DOSY results.
Arguments	continread takes no arguments
See also	ddif
	dosy

Syntax	continueMovie(rate)
Description	Like startMovie, but can continueMovie can play a movie forward or back ward, and, instead of always starting from the beginning, it starts from the beginning if movie has not started yet, or continues from where it was stopped (by stopMovie). Movie direction is controlled by parameter aipMovieSetting[3]=1 or -1 .
Arguments	aipMovieRate, or a number for the rate
See also	startMovie, stopMovie, resetMovie.

convert Convert data set from a VXR-style system (M,U)

Syntax	convert(VXR_file) (From UNIX) cpos_cvt VXR_file				
Description	Converts data stored on a VXR-style system (VXR, XL, or Gemini) to the format used in software. The macro convert loads the data from VXR_file into the current experiment and converts it to the new format. The UNIX command cpos_cvt writes the converted data in a subdirectory of the current working directory, using the original name of the data set.				
Arguments	VXR_file is the name of a VXR-style file to be converted to VnmrJ style				
See also	NMR Spectroscopy User Guide				
Related	cpos_cvt decomp	Convert data set from a VXR-style system (C,U) Decompose a VXR-style directory (C)			

convertbru Convert Bruker data (M,U)

Description A C-language program for converting 32-bit Bruker AMX data and 24and 32-bit Bruker AM data into a 32-bit format compatible with the Varian sread program. After converting the Bruker data into the new format, the converted data can be read into VnmrJ using sread and can then be processed normally. The parameters proc and proc1 are set appropriately by sread, so that wft or wft2da correctly processes the data.

> Bruker AM parameters are converted to Varian parameters as shown in the table "AM Parameter Conversion." Bruker parameter names that do not conflict with a Varian parameter name are converted under the original name: td, fw, ds, o1, o2, ns, te, id, sfo1, sfo2, and ro. Parameters proc and proc1 are set to 'rft' for all spectra (assuming TPPI data in both dimensions).

AM Parameter Conversion

Bruker	Varian	Bruker	Varian
sweeps completed	ct	sp	satdly
td	np	dp	dpwr
dw	dw	te	temp=te-273
fw	fb=1.1*sw/2	id	sw1=1/id
ds	SS	sfo1	<pre>sfrq=sfo1+o1</pre>
SW	SW	sfo2	dfrq=sfo2+o2

Table 1.

Bruker	Varian	Bruker	Varian	
experiments done	ni	p#	p#	
01	tof	d#	d#	
02	dof	s#	s#	
rd (or d1 if rd=0)	rd	ro	spin	
pw (or p0 if pw=0)	pw	rg	gain	
p1	pw90	date	date	
de	de	time	time	
ns	nt			

Table 1.

Bruker AMX parameters are converted to Varian parameters as shown in the table "AMX Parameter Conversion." All Bruker parameters are converted under their original names if the name doesn't conflict with the name of a Varian parameter. Arrayed Bruker parameters like P and D are converted to the names P# and D#, where # is the index into the array.

Because sread is limited to 8-character parameter names, the parameters routwd1# and routwd2# are converted to rtwd1# and rtwd2#.

The parameter proc is set to 'ft' when the Bruker parameter aq_mod is 1, and proc is set to 'rft' when aq_mod is 2. proc1 is always set to rft, assuming TPPI in t1.

If there is a file named info in the directory with the Bruker data, it is read in and put into the text file for the converted data set.

AMX Parameter Conversion

Bruker	Varian	Bruker	Varian
ns (from acqu)	nt	te	temp=te-273
ns (from acqus)	ct	sfol	sfrq=sfo1
td (from acqus)	np	sfo2	dfrq=sfo2
td (from acqu2s)	ni	01	tof
sw_h	SW	02	dof
sw_h	dw=1.0e6/sw	ro	spin
sw_h (from acqu2s)	swl	rg	gain
fw	fb=1.1*sw/2	date	date
ds	SS	date	time
rd (or d1 if rd=0)	rd	nucleus	tn
de	de	decnuc	dn

Table 2.

Table 2.

Bruker	Varian	Bruker	Varian
pw (or p0 if pw=0)	pw	pulprog	pslabel
p1	pw90	pulprog	seqfil

Arguments file is the input file name. For AMX data, file should be the name of the directory that contains the acqus, acqu2s, and fid or ser files. For AM data, file should be the name of the file containing the AM data. The file argument is not required to have a .bru extension, but if it does, the .bru extension is removed before creating the output file. Unless the -cfile option is present, the output file will have the same name as the input file, but with a .cv extension, and will be written into the current working directory.

> options for AMX and AM data are the following, which can be entered in any order as long as file comes first (options are usually not necessary, but can be used to override the default actions of convertbru):

- -bam or -bamx specifies whether input is AM or AMX data. The default is determined from name of the input file given.
- •-cfile specifies that the output file is given the name specified by file and is written with .cv appended to the name
- \bullet -dxxx, where xxx is the decoupler frequency (it must be a value between 10.0 and 640.0 MHz). The default is to read from data set.
- -f specifies that old output file is to be overwritten. The default is to not overwrite old files.
- •-olsb or -omsb specifies whether the data has the least- or mostsignificant byte first. For AM data, the default is determined from data set. For AMX data, the default is -olsb.
- -pxxx, where xxx is the number of 24- or 32-bit words to skip before converting data. This option is for use with -t option to skip the header in AM data without converting it. Typical header sizes are 216 or 256 words. The default is 0.
- -s3 or -s4 specifies if AM data is 24-bit (3-byte) or 32-bit (4-byte). All AMX data is 32-bit. The default is determined from the data set.
- •-tall, -thdr, or -tdata specifies whether convertbru should convert the header and the data, just the header, or just the data. The default is -tall.
- Examples Convert AM data from a UNIX shell (in all these examples, the file name is arbitrarily named br_data):
 - convertbru br_data determines the file format and converts the header and data in the file br_data.
 - convertbru br_data -d250.0 -cout determines the file format, converts the header and data in the br_data, sets the decoupler frequency to 250.0 MHz, and writes to an output file named out.cv in the current working directory.

- convertbru br_data -thdr determines file format and converts only the header in the file br_data.
- convertbru br_data -tdata -p256 -s3 -omsb converts only the data in br_data after skipping the 256-word header. The data is converted assuming it is 24-bit AM data words with the most-significant byte first.

Convert AM data from VnmrJ:

• convertbru('br_data', '-tdata', '-p256', '-s3', '-omsb') converts only the data in br_data after skipping the 256-word header. The data is converted assuming it is 24-bit AM data words with the most-significant byte first.

Convert AMX data from a UNIX shell:

• convertbru br_data -f converts acqus and acqu2s files to ASCII, if needed, and then converts data and overwrites the existing br_data.cv file.

Convert AMX data from VnmrJ:

- convertbru('br_data','-f') converts acqus and acqu2s files to ASCII, if needed, and then converts data and overwrites the existing br_data.cv file.
- convertbru('br_data','-c/home/vnmr1/bdata/data1') converts acqus and acqu2s files to ASCII, if needed, and then converts the data and writes it to /home/vnmr1/bdata/data1.cv.

See also NMR Spectroscopy User Guide

Related readbrutape Read Bruker data files from 9-track tape (U) sread Read converted data into VnmrJ (C) wft2da Weight and Fourier transform phase-sensitive data (M)

сору

Copy a file (C)

Syntax	<pre>copy(<'-r',>from_file,to_file)<:\$res></pre>			
Description	Makes a copy of a file and is identical to the cp command. All arguments are passed. Command will abort with no return value if an illegal file name is used.			
Arguments	'-r' – keyword requesting a recursive copy (i.e., copy a directory).			
	<pre>from_file - name of the file (or directory if '-r' used) to be copied.</pre>			
	to_file - name of the copy of the file (or directory). If the from_file argument has an extension (e.g., .fid), be sure the to_file argument has the same extension.			
	: $sres$ - variable to hold the result of the copy process.			
	1 is returned if the copy is successful.			
	0 is returned if the copy failed.			
Examples	<pre>copy('-r','/home/vnmr1/vnmrsys/seqlib','/vnmr/seqlib')</pre>			

cos Find cosine value of an angle (C)

Syntax	<pre>cos(angle)<:n></pre>		
Description	Finds the cosine of an angle.		
Arguments	angle is the angle, given in radians.		
	n is the return value with the cosine of angle. The default is to display the cosine value in the status window.		
Examples	cos(.5) cos(val):cos_val		
See also	User Programming		
Related	sin Find sine value of an angle (C)		

Cosy Convert the parameter to a COSY experiment (M)

Description	Convert the parameter to a COSY experiment.		
See also	NMR Spectroscopy User Guide		
Related	cosyps Set up parameters for phase-sensitive COSY pulse sequence (M)		
	Dqcosy	Set up parameters for double-quantum filtered COSY (M)	
	relayh	Set up parameters for RELAYH pulse sequence (M)	

cosyps Set up parameters for phase-sensitive COSY pulse sequence (M)

Description	Sets up a phase-sensitive COSY (homonuclear correlation) experiment		
See also	NMR Spectroscopy User Guide		
Related	Cosy	Set up parameters for COSY pulse sequence (M)	
	Dqcosy	Set up parameters for double-quantum filtered COSY (M)	
	relayh	Set up parameters for RELAYH pulse sequence (M)	

Copy a file (C)

Syntax	cp(<'-r',>from_file,to_file)<:\$res>			
Description	Makes a copy of a file and is identical to the copy command. All arguments are passed. Command will abort with no return value if an illegal file name is used.			
Arguments	'-r' is a keyword requesting a recursive copy (i.e., copy a directory).			
	from_file is the name of the file (or directory if '-r' used) to be copied.			
	to_file is the name of the copy of the file (or directory). If the from_file argument has an extension (e.g., .fid), be sure the to_file argument has the same extension.			
	:\$res variable to hold the result of the copy process.			
	1 is returned if the copy is successfully			
	0 is returned if the copy failed			
Examples	<pre>cp('/home/vnmr1/vnmrsys/seqlib/d2pul', \</pre>			
See also	NMR Spectroscopy User Guide			
Related	copy Copy a file (C)			

ср

ср

Cycle phase (P)

Description	Sets the values that real-time variable oph is calculated as, either			
	0,1,2,3 (cp='y') or 0 (cp='n'). The only circumstance where setting			
	cp='n' may be useful is when displaying an FID with acqi. If there			
	is an imbalance between the two receiver channels, the FID displayed			
	for acqi may show alternating dc levels. The standard gf macro that			
	prepares parameters for the FID display in acqi automatically handles			
	this issue.			
Values	'y' makes oph calculate as $0,1,2,3$; this is the typical value. 'n' makes oph calculate as 0.			
See also	User Programming			
Related	acqi Interactive acquisition display process (C)			
	go Submit experiment to acquisition (C)			
	gf Prepare parameters for FID/spectrum display in acqi (M)			

cpdone Macro called upon study completion (M)

Syntax Applicability VnmrJ 3.1

Description	This macro is called when a study is completed or paused. This system macro should never be edited. Changes should be implemented in the usercpdone macro.
See also	User Guide: Automation-User Space Customization
Related	usercpdone (m)

cpgo Macro called when acquisition is started (M)

Syntax	
Applicability	VnmrJ 3.1
Description	This macro is called when an acquisition is started.
	This system macro should never be edited.
	Changes should be implemented in the usergo macro.
Examples	User Guide: Automation-User Space Customization
See also	
Related	usercpgo (m)

cpmgt2 Set up parameters for CPMGT2 pulse sequence (M)

Description	Macro to set up a CPMGT2 (Carr-Purcell Meiboom-Gill T_2) experiment.
See also	NMR Spectroscopy User Guide
Related	t2 T_2 exponential analysis (M)

cpos_cvt Convert data set from a VXR-style system (M,U)

Syntax	(From UNIX) cpos_cvt VXR_file convert(VXR_file)		
Description	Converts data stored on a VXR-style system (Gemini, VXR, or XL) to the format used in VnmrJ software. cpos_cvt writes the converted data in a subdirectory of the current working directory, using the original name of the data set. The command convert loads the data from VXR_file into the current experiment and converts it to the new format.		
Arguments	$\ensuremath{\texttt{VXR_file}}$ is the file name in the VXR-style format to be converted to the VnmrJ style.		
Related	convert decomp rt	Convert data set from a VXR-style system (C,U) Decompose a VXR-style directory (C) Retrieve FIDs (C)	

cptmp Copy experiment data into experiment subfile (M)

Syntax	cptmp<(file)>		
Description	Copies the data (parameters, FID, and transformed spectrum) from the current experiment into a subdirectory inside curexp+'/subexp'.		
Arguments	file is the name of the subfile to receive the data. The default is to take the name from the transmitter nucleus (if seqfil='s2pul') or to use the pulse sequence name.		
Examples	cptmp cptmp('cosy')		
Related	curexp rttmp seqfil svtmp	Current experiment directory (P) Retrieve experiment data from experiment subfile (M) Pulse sequence name (P) Move experiment data into experiment subfile (M)	

cptmpltdefaultsDefaults for Save Data Template

Examples This macro sets the default values used for creating the save-data template in the Preferences/Templates popup. It is called when the "Restore to Defaults" button on the Preferences/Templates popup is clicked.

cpx Create pbox shape file (M)

Syntax	cpx<(ref_pw90,ref_pwr)> or cpx<('g')>		
Description	Calls UNIX command Pbox, which generates the specified pulse shap		
	or decoupling/spin locking pattern, as defined by the shapelib/Pbox.inp file.		
Arguments	ref_pw90 is the reference 90° pulse width		
	ref_pwr is the reference power level.		
	'g' is a keyword that is required only when generating gradient shapes and if the file type is not specified otherwise.		
Examples	cpx cpx('g') cpx(pw90*compH,tpwr)		
See also	NMR Spectroscopy User Guide		
Related	Pbox Pulse shaping software (U)		

cqexp Load experiment from protocol (M)

Applicability	Liquids		
Description	Macro to load an experiment from a protocol.		
Syntax	cqexp(experiment <, apptype>)		
	The first argument is the experiment name, and the second argument is the apptype. If the apptype is not specified, the previous apptype is used.		
Examples	cqexp('Proton', 'std1d')		
Related	apptypeApplication type (P)execparsSet up the exec parameters (M)		

cqfindz0 Run an experiment to find the value of z0 (M)

Applicability	Liquids	
Description	A macro to run a z0 for a given so number of deuter calibration for lk macro may be acc panels.	a deuterium experiment to find the correct value of lvent. It requires an entry in the probe file for the ium Hz per DAC. Run the appropriate probe Hz per DAC to set the value in the probe file. The essed through the Find z0 button available on several
Related	solvent z0	Lock solvent (P) Z0 field position (P)

cqgmap Perform gradient shimming utility functions (M)

Applicability	Liquids	
Description	Macro runs	gradient shimming utility functions.
Related	gmapshim	Run gradient autoshimming, set parameters,
		map shims (M)

cqinit Initialize liquids study queue (M)

Applicability	Liquids	
Description	Initializes the liqu	uids study queue.
Related	cqreset	Reset study queue parameters (M)
	sqfilemenu	Study queue file menu commands (M)

cqpars Create study queue parameters for liquids (M)

Applicability	Liquids	
Description	A macro t	o create study queue parameters for the Walkup interface.
See also	VnmrJ Wa	lkup
Related	fixpar	Correct parameter characteristics in experiment (M)

caplot Macro to perform generic 2D plot (M)

Applicability	Liquids	
Description	A macro to perfo traces. Usually ca line.	orm generic 2D plotting, including 1D experiment lled by other macros, and not used from the command
Related	plot	Automatically plot spectra (M)
	plot2D	Plot results of 2D peak picking (C)
	plt2Darg	Plot 2D arguments (P)

cqprotocol Macro to create protocols (M)

Applicability	Liquids	
Description	A macro to create protocols for liquids applications.	Called by the
	Make protocols dialogs in the Utilities menu.	

cgreset Reset study queue parameters (M)

Applicability	Liquids	
Description	Reset liquids stud when starting a r	y queue parameters. Usually called by other macros new study.
Related	cqinit sqfilemenu	Initialize liquids study queue (M) Study queue file menu commands (M)

cqsavestudy Macro to save study queue parameters (M)

Applicability Liquids

ulaicu	scuaypar	Study parameters (1)
	xmsubmit	Submit sample(s) to the study queue (M)
	xmendq	End a chained study queue (M)

cqwtmenu Macro to set weighting functions from a panel (M)

ApplicabilityLiquids, ImagingDescriptionA macro to set weighting functions from a panel. It is used for both
1D and 2D weighting parameters. Called by processing parameter
panels.

cr Cursor position in directly detected dimension (P)

Description	Contains the current cursor position. The rl macro uses cr to set the reference line.		
See also	NMR Spectroscopy User Guide		
Related	centersw	Move cursor to center of spectrum (M)	
	crf	Current time-domain cursor position (P)	
crl Clear ref. line in directly detected dim		Clear ref. line in directly detected dimension (M)	
	delta	Difference of two frequency cursors (P)	
	rl	Set reference line in directly detected dimension (M)	

cr1 Cursor position in 1st indirectly detected dimension (P)

Description	Contains the current cursor position along the first indirectly detected dimension. Analogous to the cr parameter except that cr1 applies to the first indirectly detected dimension of a multidimensional data set. The rl1 macro uses cr1 to set the reference line along this dimension.		
See also	NMR Spectroscopy User Guide		
Related centersw1 Move cursor to dimension (M)		Move cursor to center of spectrum in 1st indirect dimension (M)	
	cr	Cursor position in directly detected dimension (P)	
	cr2	Cursor position in 2nd indirectly detected dimension (P)	
	rl1	Set ref. line in 1st indirectly detected dimension (M)	

Cursor position in 2nd indirectly detected dimension (P)

Description Contains the current cursor position along the second indirectly detected dimension. Analogous to the cr parameter except that cr2 applies to the second indirectly detected dimension of a multidimensional data set. The rl2 macro uses cr2 to set the reference line along this dimension.

See also NMR Spectroscopy User Guide

Related	centersw2	Move cursor to center of spectrum in 2nd indirect
		dimension (M)
	cr	Cursor position in directly detected dimension (P)

- cr1 Cursor position in 1st indirectly detected dimension (P)
- r12 Set ref. line in 2nd indirectly detected dimension (M)

crcom Create user macro without using text editor (M)

Syntax	<pre>crcom(file,actions)</pre>	

Description Creates a macro file in the user's macro library (maclib) with the contents given in the actions argument.

Arguments file is the file name of the user macro to be created. If a macro of the same name already exists, the user is asked whether or not to overwrite it.

actions is a string containing the actions making up the user macro. The string cannot include a carriage return. If a single quote is needed within the string, it must be preceded by a backslash (see second example below).

- See also User Programming

create Create new parameter in a parameter tree (C)

Syntax	create	(parameter<,	type<,	tree>>)
--------	--------	--------------	--------	---------

- Description Creates a parameter in one of the parameter trees. A *parameter tree* is a UNIX file containing the attributes of parameters as formatted text. Refer to the command paramvi for a description of the file contents.
- Arguments parameter is the name of the parameter to be created.
 - type is the type of values in the parameter to be created and can be one of the following values (default is 'real'):
 - 'real' is a value with no limits on range and can be positive or negative.

- 'string' is a value composed of characters. Entry of strings can be limited to selected words by enumerating the possible values with the command setenumeral. For example, the enumerated values of intmod are 'off', 'partial', and 'full'. Therefore, intmod can be set only to one of these three string values, such as intmod='full'.
- 'delay' is a value from 0 to 8190, in unit of seconds.
- 'frequency' is a positive real number value.
- 'flag', like 'string', is a value composed of characters. Entry of flags can be limited to selected characters by enumerating the possible values with the command setenumeral. For example, the enumerated values of dmm are 'c', 'f', 'g', 'm', 'p', 'r', 'u', 'w', and 'x'. Therefore, dmm can only be set to a combinations of these nine characters, such as dmm='ccw'. If enumerated values are not set, the 'string' and 'flag' types are identical.
- 'pulse' is a value from 0 to 8190, in units of μ s.
- 'integer' is a value composed of integers (0,1,2,3,...).

- 'current' contains parameters that are adjusted to set up an experiment. The parameters are from the file curpar in the current experiment.
- 'global' contains user-specific parameters from the file global in the vnmrsys directory of the present UNIX user.
- 'processed' contains parameters with which the data was obtained. These parameters are from the file procpar in the current experiment.
- 'systemglobal' contains instrument-specific parameters from the text file /vnmr/conpar. Most of these parameters are defined using the config program. All users have the same systemglobal tree. Note that conpar is not written out when you exit; the only time conpar is ever modified is by the config program. Thus, any changes you make to conpar using create (or destroy, setvalue, etc.) are not permanent. To permanently create a parameter in conpar, you must use a text editor to change /vnmr/conpar.

Examples create('a')

```
create('b','string')
create('c','real','global')
```

See also User Programming

Related	destroy	Destroy a parameter (C)
	display	Display parameters and their attributes (C)
	fread	Read parameters from file and load them into a tree
		(C)
	fsave	Save parameters from a tree to a file (C)
	paramvi	Edit a parameter and its attributes using vi text
		editor (M)
	prune	Prune extra parameters from current tree (C)

tree is one of the following types of parameter trees (default is
'current'):

setenumeralSet values of a string variable in a tree (C)setgroupSet group of a parameter in a tree (C)setprotectSet protection mode of a parameter (C)

create (P) Parameter used for RF transmitter board temperature compensation

Syntaxcreate('rftempcomp', 'string', 'global')ApplicabilityVnmrJ 3.1ArgumentsIf rftempcomp='n' temperature compensation on the RF transmitter
board is turned off.If rftempcomp='y' temperature compensation on the RF transmitter
board is turned on and will make a single compensation.If rftempcomp='c' temperature compensation on the RF transmitter
board is turned on continuously and will continuously update until it

createqcomp Create qcomp parameter (M)

is turned off.

Applicability	Systems with Varian, Inc. Cold Probes
Description	Macro to create the gcomp parameter with the appropriate attributes.
	qcomp is created as a flag parameter in the global tree.

crf Current time-domain cursor position (P)

- Description Contains current time-domain cursor position. To create crf and the other FID display parameters axisf, dotflag, vpf, vpfi, and deltaf (if the parameter set is older and lacks these parameters), enter addpar('fid').
 - Values Number, in seconds.
 - See also NMR Spectroscopy User Guide
 - Related addpar Add selected parameters to the current experiment (M) crl1 Clear ref. line in 1st indirectly detected dimension (C) deltaf Difference of two time cursors (P)
 - fidpar Add parameters for FID display in current experiment (M)

cr1 Clear reference line in directly detected dimension (M)

Description	Clears frequency referencing along the directly detected dimension by setting the reference parameters rfl and rfp to zero. crl also resets the referencing parameters refpos and reffrq.	
See also	NMR Spe	ctroscopy User Guide
Related	crl1	Clear ref. line in 1st indirectly detected dimension (C)
	crl2	Clear ref. line in 2nd indirectly detected dimension (C)
	rl	Set ref. line in directly detected dimension (M)
	reffrq	Reference frequency of reference line (P)
	refpos	Position of reference frequency (P)
	rfl	Ref. peak position in directly detected dimension (P)
	rfp	Ref. peak frequency in directly detected dimension (P)

Clear reference line in 1st indirectly detected dimension (M)

Description	Clears frequency referencing along the first indirectly detected dimension by setting the reference parameters rfl1 and rfp1 to zero crl1 also resets the referencing parameters refpos1 and reffrq1	
See also	NMR Spec	troscopy User Guide
Related	crl rl1 reffrq1 refpos1 rfl1 rfp1	Clear ref. line in directly detected dimension (C) Set ref. line in 1st indirectly detected dimension (M) Ref. frequency of reference line in 1st indirect dimension (P) Position of reference frequency in 1st indirect dimension (P) Ref. peak position in 1st indirectly detected dimension (P) Ref. peak frequency in 1st indirectly detected dimension (P)

cr12Clear reference line in 2nd indirectly detected dimension
(M)

DescriptionClears frequency referencing along the second indirectly detected
dimension by setting the reference parameters rfl2 and rfp2 to zero.
crl2 also resets the referencing parameters refpos2 and reffrq2.See alsoNMR Spectroscopy User GuideRelatedcrlcrlClear ref. line in directly detected dimension (C)
rl2reffrq2Ref. frequency of reference line in 2nd indirect dimension
(P)

refpos2	Position of reference frequency in 2nd indirect dimension
	(P)
rfl2	Ref. peak position in 2nd indirectly detected dimension
	(P)
rfp2	Ref. peak frequency in 2nd indirectly detected dimension
	(P)

crmode Current state of the cursors in df, ds, or dconi programs (P)

Description	Stores the current state (box mode or cursor mode) of cursors in the df, ds, or dconi interactive display programs. crmode is mostly used by programmable menus to determine the status of the cursors. It i		
	stored in t	he file vnmrsys/global.	
Values	'b' signifies the box mode, 'c' signifies the cursor mode.		
See also	User Progr	ramming	
Related	dconi	Interactive 2D data display (C)	
	df	Display a single FID (C)	
	ds	Display a spectrum (C)	

crof2 Recalculate rof2 so that lp = 0 (M)

Syntax	crof2<(alfa)>
бущал	CIUI2 (alla

Description Recalculates a new value for rof2 (receiver gating time following a pulse) based upon the current rof2 and lp (first-order phase) values, so that lp is rendered approximately 0. For crof2 to work properly, a trial spectrum must be obtained and phased to pure absorption. This spectrum provides the current rof2 and lp values for crof2. The value of the alfa delay is left constant, provided rof2 does not become less than 1 μs.

crof2 pertains to processing 2D data. Unless lp is approximately 0, fpmult affects both the dc offset and the curvature of the spectrum.

Arguments alfa specifies a value for the alfa delay before acquisition.

Related	alfa	Set alfa delay before acquisition (P)
	cfpmult	Calculate first point multiplier for 2D experiments (P)
	fpmult	First point multiplier for np FID data (P)
	lp	First-order phase along directly detected dimension (P)
	rof2	Receiver gating time following a pulse (P)

cryo_noisetestRun Cold Probe conditioning experiments (M)

Applicability Systems with Varian, Inc. Cold Probes

cryoclient Start the CryoBay Monitor program (M, U)

Systems with Cold Flobes and Cryobay Monitor Software.
Starts the CryoBay Monitor software in a separate window. This program is a CORBA client that requires an active CORBA server running on the CryoBay PC
Cruogenic Systems Installation and Operation

CSschedule Generates a NUS schedule

Description The CSschedule command will generate a schedule for non-uniform sampling.

With no arguments, it uses the following parameters.

ni, ni2, and ni3 are used to define the total size of the data matrix. If they exist, are greater than 1 and not set to "Not used", they will beused to define the matrix.

CSdensity is the percentage of points in the data matrix that are acquired. All other points will be treated as zeros. Default is 100.0, that is, useuniform sampling.

The CSpars parameter will override the default selection of the indirect dimensions. For example, if the parameter set is a 3D parameter set with both ni and ni2 set, the CSparsparameter can be set to make just the first or second or both indirect dimensions sparse by setting CSpars='d2' or CSpars='d3' or

CSpars='d2', 'd3', respectively. If the CSpars parameterdoes not exist or is set to the empty string (CSpars="), the default behavior with ni, ni2, etcwill be used.

CSseed parameter (integer) containing the seed for the random numbergenerator used to create the sampling schedules. A positive number indicates that the sampling schedule is executed sequentially. A negative number indicates that the sampling schedule is applied randomly. Default value is 169.

It writes the schedule as a text file in the current experiment directory as sampling.sch.

Arguments can be supplied to the CSsampling command to override the defaults.

Values NOBURN – waits the operator input period of time between tests. No arguments – macro will prompt for a time in minutes.

If an argument starts with 'seed ', then the number in the 'seed ' string will override the CSseed parameter. This will also set the value of the CSseed parameter.

Any string parameter that does not start with 'seed' will be used as the filename of the schedule as it is stored in the current experiment.

The first real number overrides the CSdensity parameter. Its range is $0 < \text{density} \leq 100.0$.

If it falls outside this range, it will be set to 100.0. This will also set the value of the CSdensity parameter.

Subsequent real numbers define the sizes of the first, second, etc indirect dimensions. They override the values of ni, ni2, etc. respectively.

Acquisition related parameters.

The sampling parameter is used to select sparse sampling. sampling='sparse' selects sparse data sampling. sampling='e' selects ellipitical data sampling, used by imaging applications.

Any other value of sampling selects standard uniform data sampling.

The CStype parameter specifies the type of sampling schedule that will be used by acquisition.

Sampling schedules are of two types. One contains a list of indexes of the increments in the indirect dimensions. For the first indirect dimension, these indexes would fall between 0 and ni-1. Each index is multiplied by the dwell time in the indirect dimension (1/sw1).

The second type of schedule is a list of the actual delay times for the indirect dimension.

CStype='a' to automatically generate an index schedule.

CStype='i' to use a pre-existing index schedule.

CStype='d' to use a pre-existing delay schedule.

The default is CStype='a'. If CStype='a', the above parameters (CSdensity, CDpars, CSseed) will be used to generate the sampling schedule.

Examples

CSschedule

CSschedule('seed 33') CSschedule('seed 33',10.0, 512, 256) CSschedule('seed -78')

csv2cpQ Imports CSV data (M)

Syntax Applicability VnmrJ 3.1 The CSV file used by the csv2cpQ macro is an ASCII text file containing, as the name implies, text values separated by commas. The first line of this file defines how the comma separated values in subsequent lines are to be interpreted. The first line can contain VnmrJ parameter names or keywords. There are no required fields. Any field that does not correspond to a VnmrJ parameter name is considered a keyword. Keywords are looked up in a synonym table to see if they should be re-interpreted as a VnmrJ parameter or value. This synonym translation file is in an appdir directory with the name <appdir>/adm/walkupadm/csv2cpQ_synonym.

These CSV files may be generated manually, or they may be exported from a spread-sheet. Often, the values available to the spread-sheet do not correspond directly to a VnmrJ parameter. The synonym feature allows VnmrJ to translate the spread-sheet value to something VnmrJ can use. For example, the spread-sheet might define a solvent as MeOH. The synonym table allows csv2cpQ to translate that into 'cd3od'. Any field that is not a vnmr parameter or keyword will be ignored.

Arguments The actions can be submitted to an automation run, to a file for use in a future automation run, or directly to the foreground VnmrJ.

This selection is controlled by the optional argument 'auto', 'enter', or 'acq', respectively. The default is 'auto'. Another optional argument is 'print' or 'noprint'. This controls whether submission information is printed or not. The default is 'noprint'.

Examples "SAMPLE", "DAY", "NIGHT", "solvent", "operator", "samplename", "notebook", "page", "Comments"

1,"PROTON",,"CDCl3","John","johnstuff","Johns book","p32","csv2cpQ test location 1"

2,"PROTON gCOSY","gHMBCAD","DMSO","Paul","paulstuff","Pauls book","p42","csv2cpQ test location 2"

3,"PROTON-HSQCAD",,"D2O","George","Georges book","Georges book","p23","csv2cpQ test sample 3"

4,"PROTON gHSQCAD","CARBON","DMSO","Ringo","ringostuff", "Ringos book","p38","Ringos Sample 4"

ct Completed transients (P)

Description Stores a nonuser-enterable informational parameter that changes during the course of an experiment to reflect the number of completed transients. During most experiments, an accurate transient counter is displayed in the acquisition status window, updated every five seconds. The value of ct is displayed in the acquisition parameter group by the dg command and is only updated when data processing occurs on the FID. In an experiment that is accumulating and not processed until the acquisition is complete, ct always indicates 0 until the end of the acquisition.

See also NMR Spectroscopy User Guide

Related dg Display parameters of acquisition/processing group (C)

ctext Clear the text of the current experiment (C)

Description	Clears the text from the current experiment text file (a block of text that may be used to describe the sample and experiment).	
See also	NMR Spectroscopy User Guide	
Related	atext text	Append string to the current experiment text (M) Display text or set new text for current experiment (C)

curexp Current experiment directory (P)

Description	Contains the	full UNIX path to the currently active experiment. This
	parameter is	useful when accessing text files generated by various
	commands (e.g., cat(curexp+'/fp.out')).	
See also	NMR Spectroscopy User Guide	
Related	systemdir	VnmrJ system directory (P)
	userdir	VnmrJ user directory (P)

curscan Scan currently in progress (P)

Applicability	Systems with LC-NMR accessory.	
Description	Keeps track of which "scan" is currently in progress. If curscan does not exist, the parlc macro can create it.	
See also	NMR Spectroscopy User Guide	
Related	parlc Create LC-NMR parameters (M)	

curwin Current window (P)

Description An arrayed global parameter. The first value is the index of the selected window pane in the graphics window. The second value is the

number of window pane rows. The third value is the number of columns.

See also	NMR Spectroscopy User Guide	
Related	fontselect	Open FontSelect window (C)
	jwin	Activate current window (M)
	mapwin	List of experiment numbers (P)
	setgrid	Activate selected window (M)
	setwin	Activate selected window (C)

cutoff Data truncation limit (P)

Description	Defines the distance above and below the current vertical position vp at which spectra and integrals are truncated. By arraying cutoff to have two different values, the truncation limits above and below the current vertical position can be controlled independently (e.g., cutoff=50 truncates data at $vp+50$ mm and $vp-50$ mm, and cutoff=50,10 truncates data at $vp+50$ mm and $vp-10$ mm). cutoff='n' disables the action of cutoff.		
	cutoff is not active during interactive spectral displays (i.e., for the ds command), but is active during non-interactive spectral displays and plots (for the dss and pl commands).		
Values	'n', number in mm.		
See also	NMR Spectroscopy User Guide		
Related	ds	Display a spectrum (C)	
	dss	Display stacked spectra (C)	
	pl	Plot spectra (C)	
	vp	Vertical position of spectrum (P)	

cyclence Set up parameters for CYCLENOE pulse sequence (M)

Applicability	Systems in which the observe channel is equipped with direct synthesis		
	ri and a linear amplifier.		
Description	Sets up a difference NOE experiment.		

cylbr24 Set up parameters for cycled BR24 pulse sequence (M)

Applicability	Systems with solids module.
Description	Sets up a BR24 sequence with quadrature detection and prepulse for
	solids multiple-pulse line narrowing.

See alsoUser Guide: Solid-State NMRRelatedbr24Set up parameters for BR24 pulse sequence (M)

cylmrev Set up parameters for cycled MREV8 pulse sequence (M)

Applicability	Systems with	a solids module.
Description	Sets up a MREV8 sequence with quadrature detection and prepulse for solids multiple-pulse line narrowing.	
See also	User Guide: Solid-State NMR	
Related	mrev8	Set up parameters for MREV8 pulse sequence (M)

Clear integral reset points (C)

cz<(freque	ncy1,frequency2,)>
Removes currently defined integral reset points.	
frequency1, frequency2, are reset points corresponding to specified frequencies to be removed. The default is remove all reset points.	
cz cz(800,600,250,60)	
NMR Spectroscopy User Guide	
dli	Display listed integral values (C)
dlni	Display listed normalized integral values (C)
nli	Find normalized integral values (C)
Z	Add integral reset point at the cursor position (C)
	cz<(frequer Removes curr frequency1 specified freq points. cz cz(800,600 <i>NMR Spectro</i> dli dlni nli z

CZ



Agilent VnmrJ 4 Command and Parameter Reference Guide

0b	Overhead delay between FIDs (P)
d1	First delay (P)
d2	Incremented delay in 1st indirectly detected dimension (P)
d2pul	Set up parameters for D2PUL pulse sequence (M)
d3	Incremented delay for 2nd indirectly detected dimension (P)
d4	Incremented delay for 3rd indirectly detected dimension (P)
DAC_to_G	Store gradient calibration value in DOSY sequences (P)
da	Display acquisition parameter arrays (C)
daslp	Increment for t1 dependent first-order phase correction (P)
date	Date (P)
daxis	Display horizontal LC axis (M)
Dbppste	Set up parameters for Dbppste pulse sequence (M)
Dbppsteinept	Set up parameters for Dbppsteinept pulse sequence (M)
dbsetup	Set up VnmrJ database (U)
dbupdate	Update the VnmrJ database (U)
dc	Calculate spectral drift correction (C)
dc2d	Apply drift correction to 2D spectra (C)
dcg	Drift correction group (P)
dcon	Display non interactive color intensity map (C)
dconi	Interactive 2D data display (C)
dconi	Control display selection for the dconi program (P)
dconn	Display color intensity map without screen erase (C)
dcrmv	Remove dc offsets from FIDs in special cases (P)



ddf	Display data file in current experiment (C)
ddff	Display FID file in current experiment (C)
ddfp	Display phase file in current experiment (C)
ddif	Synthesize and show DOSY plot (C)
ddrcr	Direct digital receiver coefficient ratio (P)
ddrpm	Set ddr precession mode (P)
ddrtc	Set ddr time constant (P)
dds	Default display (M)
dds_seqfil	Sequence-specific default display (M)
debug	Trace order of macro and command execution (C)
decasynctype	Select the type of decoupler asynchronous mode (P)
decay_gen	Calculates the form of diffusional attenuation expected for the measured gradient and signal maps in non-uniform gradient calibration.
deccwarnings	Control reporting of DECC warnings from PSG (P)
decomp	Decompose a VXR-style directory (M)
def_osfilt	Default value of osfilt parameter (P)
defaultdir	Default directory for Files menu system (P)
delcom	Delete a user macro (M)
delete	Delete a file, parameter directory, or FID directory (C)
delexp	Delete an experiment (M)
delexpdata	Delete data from the current experiment
deletenucleus	Removes nucleus entry to probe file (M)
dels	Delete spectra from T_1 or T_2 analysis (C)
delta	Cursor difference in directly detected dimension (P)
delta1	Cursor difference in 1st indirectly detected dimension (P)
delta2	Cursor difference in 2nd indirectly detected dimension (P)
deltaf	Difference of two time-domain cursors (P)
Dept	Set up parameters for DEPT experiment (M)
deptgl	Set up parameters for DEPTGL pulse sequence (M)
deptproc	Process array of DEPT spectra (M)
destroy	Destroy a parameter (C)
destroygroup	Destroy parameters of a group in a tree (C)
df	Display a single FID (C)
------------	---
df2d	Display FIDs of 2D experiment (C)
dfid	Display a single FID (C)
dfmode	Current state of display of imaginary part of a FID (P)
dfrq2	Transmitter frequency of second decoupler (P)
dfrq3	Transmitter frequency of third decoupler (P)
dfrq4	Transmitter frequency of fourth decoupler (P)
dfs	Display stacked FIDs (C)
dfsa	Display stacked FIDs automatically (C)
dfsan	Display stacked FIDs automatically without screen erase (C)
dfsh	Display stacked FIDs horizontally (C)
dfshn	Display stacked FIDs horizontally without screen erase (C)
dfsn	Display stacked FIDs without screen erase (C)
dfww	Display FIDs in whitewash mode (C)
dg	Display group of acquisition/processing parameters (C)
dg	Control dg parameter group display (P)
dg1	Display group of display parameters (M)
dg1	Control dg1 parameter group display (P)
dg2	Display group of 3rd and 4th rf channel/3D parameters (M)
dg2	Control dg2 parameter group display (P)
dga	Display group of spin simulation parameters (M)
DgcsteSL	Set up parameters for DgcsteSL pulse sequence (M)
Dgcstecosy	Set up parameters for Dgcstecosy pulse sequence (M)
Dgcstehmqc	Set up parameters for Dgcstehmqc pulse sequence (M)
dglc	Display group of LC-NMR parameters (M)
dglc	Control $dglc$ parameter group display (P)
dglp	Control \mathtt{dglp} parameter group of linear prediction parameters (P)
dgs	Display group of shims and automation parameters (M)
dgs	Control dgs parameter group display (P)
dhp	Decoupler high-power control with class C amplifier (P)

diagth2d	Exclude diagonal peaks when peak picking
dialog	Display a dialog box from a macro (C)
diffparam	Report differences between parameter sets (U)
diffparams	Report differences between two parameter sets (U)
diffshims	Compare two sets of shims (M,U)
digfilt	Write digitally filtered FIDs to another experiment (M)
dir	List files in directory (C)
display	Display parameters and their attributes (C)
dla	Display spin simulation parameter arrays (M)
dlalong	Long display of spin simulation parameter arrays (C)
dlC	Display LC detector trace(s) in a horizontal format.
dlCNMR	Display all forms of LC-NMR data
dli	Display list of integrals (C)
dlivast	Produce text file and process wells (M)
dll	Display listed line frequencies and intensities (C)
dlni	Display list of normalized integrals (M)
dlp	Decoupler low-power control with class C amplifier (P)
dm	Decoupler mode for first decoupler (P)
dm2	Decoupler mode for second decoupler (P)
dm3	Decoupler mode for third decoupler (P)
dm4	Decoupler mode for fourth decoupler (P)
dmf	Decoupler modulation frequency for first decoupler (P)
dmf2	Decoupler modulation frequency for second decoupler (P)
dmf3	Decoupler modulation frequency for third decoupler (P)
dmf4	Decoupler modulation frequency for fourth decoupler (P)
dmfadj	Adjust tip-angle resolution time for first decoupler (M)
dmf2adj	Adjust tip-angle resolution time for second decoupler (M)
dmf3adj	Adjust tip-angle resolution time for third decoupler (M)
dmf4adj	Adjust tip-angle resolution time for fourth decoupler (M)
dmg	Data display mode in directly detected dimension (P)
dmg1	Data display mode in 1st indirectly detected dimension (P)

218

dmg2	Data display mode in 2nd indirectly detected dimension (P)
dmgf	Absolute-value display of FID data or spectrum in acqi (P)
dmm	Decoupler modulation mode for first decoupler (P)
dmm2	Decoupler modulation mode for second decoupler (P)
dmm3	Decoupler modulation mode for third decoupler (P)
dmm4	Decoupler modulation mode for fourth decoupler (P)
dn	Nucleus for first decoupler (P)
dn2	Nucleus for second decoupler (P)
dn3	Nucleus for third decoupler (P)
dn4	Nucleus for fourth decoupler (P)
dndfid	Retrieve and process fid data from the locator (M)
dndjoin	Join a work space from the locator (M)
dndpar	Retrieve a parameter set from the locator (M)
dndshims	Retrieve a shimset set from the locator (M)
doautodialog	Start a dialog window using ${\tt def}$ file (M)
dodialog	Start a dialog window with dialoglib file (M)
dof	Frequency offset for first decoupler (P)
dof2	Frequency offset for second decoupler (P)
dof3	Frequency offset for third decoupler (P)
dof4	Frequency offset for fourth decoupler (P)
Doneshot	Set up parameters for Doneshot pulse sequence (M)
dopardialog	Start a dialog with dialoglib/experiment def file (M)
do_pcss	Calculate proton chemical shifts spectrum (C)
dosy	Process DOSY experiments (M)
dosy2d	Apptype macro for dosy 2D experiments (M)
dosy3Dflag	Used by the dosy macro to determine whether to use 2D or 3D DOSY processing
dosy3Dproc	Used by the dosy macro to determine whether to use 2D or 3D processing
dosybypoints	Determines whether peak picking is used by the dosy macro
dosyfit	Fits 2D or 3D DOSY data to obtain diffusion coefficients, amplitudes and statistics

D

dosyfrq	Larmor frequency of phase encoded nucleus in DOSY (P)
dosygamma	Gyromagnetic constant of phase encoded nucleus in DOSY (P)
dosyproc	Determines the type of processing performed by the dosy macro
dosytimecubed	Gyromagnetic constant of phase encoded nucleus in DOSY (P)
dot1	Set up a T ₁ experiment (M)
dotflag	Display FID as connected dots (P)
downsamp	Downsampling factor applied after digital filtering (P)
dp	Double precision (P)
dpcon	Display plotted contours (C)
dpconn	Display plotted contours without screen erase (C)
dpf	Display peak frequencies over spectrum (C)
dpir	Display integral amplitudes below spectrum (C)
dpirn	Display normalized integral amplitudes below spectrum (M)
dpiv	Display integral amplitudes below spectrum (M)
dpirn	Display normalized integral amplitudes below spectrum (C)
dpl	Default plot (M)
dpl_seqfil	Sequence-specific default plot (M)
dplane	Display a 3D plane (M)
dpr	Default process (M)
dpr_seqfil	Sequence-specific default process (M)
dprofile	Display pulse excitation profile (M)
dproj	Display a 3D plane projection (M)
dps	Display pulse sequence (C)
dpwr	Power level for first decoupler with linear amplifier (P)
dpwr2	Power level for second decoupler with linear amplifier (P)
dpwr3	Power level for third decoupler with linear amplifier (P)
dpwr4	Power level for fourth decoupler amplifier (P)
dpwrf	First decoupler fine power (P)
dpwrf2	Second decoupler fine power (P)

dpwrf3	Third decoupler fine power (P)
dpwrm	First decoupler linear modulator power (P)
dpwrm2	Second decoupler linear modulator power (P)
dpwrm3	Third decoupler linear modulator power (P)
Dqcosy	Convert the parameter to a DQCOSY experiment (M)
draw	Draw line from current location to another location (C)
dres	Measure linewidth and digital resolution (C)
dres	Tip-angle resolution for first decoupler (P)
dres2	Tip-angle resolution for second decoupler (P)
dres3	Tip-angle resolution for third decoupler (P)
dres4	Tip-angle resolution for fourth decoupler (P)
ds	Display a spectrum (C)
ds2d	Display 2D spectra in whitewash mode (C)
ds2dn	Display 2D spectra in whitewash mode without screen erase (C)
dscale	Display scale below spectrum or FID (C)
dsnarray	Report statistical signal-to-noise for Cold Probes (M)
dscoef	Digital filter coefficients for downsampling (P)
dseq	Decoupler sequence for first decoupler (P)
dseq2	Decoupler sequence for second decoupler (P)
dseq3	Decoupler sequence for third decoupler (P)
dseq4	Decoupler sequence for fourth decoupler (P)
dsfb	Digital filter bandwidth for downsampling (P)
dshape	Display pulse shape or modulation pattern (M)
dshapef	Display last generated pulse shape (M)
dshapei	Display pulse shape or modulation pattern interactively (M)
dshim	Display a shim "method" string (M)
dslsfrq	Bandpass filter offset for downsampling (P)
dsn	Measure signal-to-noise (C)
dsnmax	Calculate maximum signal-to-noise (M)
dsplanes	Display a series of 3D planes (M)
dsptype	Type of DSP (P)

dss	Display stacked spectra (C)
dssa	Display stacked spectra automatically (C)
dssan	Display stacked spectra automatically without erasing (C)
dssh	Display stacked spectra horizontally (C)
dsshn	Display stacked spectra horizontally without erasing (C)
dssl	Label a display of stacked spectra (M)
dssn	Display stacked spectra without screen erase (C)
dsvast	Display VAST Data in a stacked 1D-NMR matrix format
dsvast2d	Display VAST data in a pseudo-2D format
dsww	Display spectra in whitewash mode (C)
dtext	Display a text file in graphics window (M)
dtrig	Delay to wait for another trigger or acquire a spectrum (P)
dutyc	Duty cycle for homodecoupling (optional) (P)

d0 Overhead delay between FIDs (P)

Description Defines the extra overhead delay at the start of each FID or array element. Overhead times between increments and transients are deterministic, i.e., both known and constant. However, the time between increments (typically x) is longer than the time between transients (y, not including times that are actually part of the pulse sequence, such as d1). Some experiments may benefit if it is ensured that these two times are not only constant but equal. To ensure that the times are constant and equal, insert the time d0 at the start of each transient (before the pulse sequence actually starts); the actual delay is then y+d0. However, the overhead time may differ with different system configurations. To keep the d0 delay consistent across systems, set d0 greater than the overhead delay. The inter-FID delay x is then padded so that y+d0=x+(d0-(x-y)).

> Currently, d0 only takes into account the extra delay at the start of each array element. It does not take into account the overhead delays at the start and end of each scan. It also does not take into account delays when arraying status statements, shims, or spinner speeds.

> The d0 parameter does not exist in any parameter set and must be created by the user. To create d0, enter create('d0','delay'). If d0 is nonexistent, do not insert a delay between transients.

Values 'n', 'y', or 0 to the maximum delay time (in seconds).

If d0='n', the software calculates the overhead time for an array element and then delays that length of time at the beginning of subsequent transients for every array element. The calculated value of d0 can be viewed by entering d0='y' in the input window.

If d0 is set to a value, that value is the length of delay time at the beginning of subsequent transients for every array element. If the value is greater than the array overhead time, the array overhead time is padded to d0.

See also User Programming

Related create Create new parameter in parameter tree (C)

d1 First delay (P)

Description	Length of the first delay in the standard two-pulse sequence and most other pulse sequences. This delay is used to allow recovery of magnetization back to equilibrium, if such a delay is desired.	
Values	0.1 μs to 8190 sec, smallest value possible is 0.1 $\mu s,$ finest increment possible is 12.5 ns.	
See also	NMR Spectroscopy User Guide	
Related	alfa	Set alfa delay before acquisition (P)
	d2	Incremented delay in 1st indirectly detected dimension (P)
	d3	Incremented delay in 2nd indirectly detected dimension (P)
	d4	Incremented delay in 3rd indirectly detected dimension (P)
	pad	Preacquisition delay (P)

d2 Incremented delay in 1st indirectly detected dimension (P)

Length of the second delay in the standard two-pulse sequence. The delay is controlled by the parameters ni and $sw1$ in a 2D experiment	
0.1 μs to 8190 sec, smallest value possible is 0.1 $\mu s,$ finest increment possible is 12.5 ns.	
NMR Spectroscopy User Guide	
d1 ni sw1	First delay (P) Number of increments in 1st indirectly detected dimension (P) Spectral width in 1st indirectly detected dimension (P)
	Length o delay is o 0.1 µs to possible NMR Spe d1 ni swl

d2pul Set up parameters for D2PUL pulse sequence (M)

Description Sets up a standard two-pulse sequence using the decoupler as transmitter.

See also	NMR Sp	ectroscopy User Guide
Related	dhp	Decoupler high power with class C amplifier (P)
	dn	Nucleus for the first decoupler (P)
	dof	Frequency offset for first decoupler (P)
	dpwr	Power level for first decoupler with linear amplifiers (P)
	s2pul	Set up parameters for standard two-pulse sequence (M)
	tn	Nucleus for the observe transmitter (P)
	tof	Frequency offset for observe transmitter (P)
	tpwr	Power level of observe transmitter with linear amplifiers (P)

d3 Incremented delay for 2nd indirectly detected dimension (P)

Description	Length of a delay controlled by the parameters ni2 and sw2 in a 3D experiment. The d2 delay, which is controlled by ni and sw1, is incremented through its entire implicit array first before d3 is incremented. To create parameters d3, ni2, phase2, and sw2 to acquire a 3D data set in the current experiment, enter addpar ('3d')	
Values	0.1 μs to 8190 sec, smallest value possible is 0.1 $\mu s,$ finest increment possible is 12.5 ns.	
See also	NMR Spectroscopy User Guide	
Related	addpar d1 ni2	Add selected parameters to the current experiment (M) First delay (P) Number of increments in 2nd indirectly detected dimension (P)
	par3d phase2 sw2	Create 3D acquisition, processing, display parameters (C) Phase selection for 3D acquisition (P) Spectral width in 2nd indirectly detected dimension (P)

d4

Incremented delay for 3rd indirectly detected dimension (P)

- Description Length of a delay controlled by the parameters ni3 and sw3 in a 4D experiment. The d3 delay, which is controlled by ni2 and sw2, is incremented through its entire implicit array first before d4 is incremented. To create parameters d4, ni3, phase3, and sw3 to acquire a 4D data set in the current experiment, enter addpar('4d').
 Values 0.1 us to \$100 sec smallest value possible is 0.1 us fixed increment.
 - Values $0.1 \ \mu s$ to 8190 sec, smallest value possible is $0.1 \ \mu s$, finest increment possible is 12.5 ns.
 - See also NMR Spectroscopy User Guide
 - Related addpar Add selected parameters to the current experiment (M) d1 First delay (P) ni3 Number of increments in 3rd indirectly detected dimension (P)

par4d	Create 4D acquisition parameters (C)
phase3	Phase selection for 4D acquisition (P)
sw3	Spectral width in 3rd indirectly detected dimension (P)

DAC_to_G Store gradient calibration value in DOSY sequences (P)

Description DAG_to_G is automatically set by the setup_dosy macro by retrieving the gradient strength from the probe calibration file if probe<>'' and storing it in DAC_to_G. If probe=' ' (i.e., the probe is not defined), then DAC_to_G is set to the current value of the global parameter gcal See also NMR Spectroscopy User Guide.

RelateddosyProcess DOSY experiments (M)setup_dosySet up gradient levels for DOSY experiments (M)setgcalSet the gradient calibration constant (M)

da Display acquisition parameter arrays (C)

Syntax	da<(par1<,par2><,par3>)>
Description	Displays arrayed acquisition parameters.
Arguments	par1,par2,par3, are names of parameters to be displayed. The default is to display all such parameters.
Examples	da da('d2')
See also	NMR Spectroscopy User Guide
Related	dg Display parameters of acquisition/processing group (C)

daslp Increment for t1 dependent first-order phase correction (P)

Description Causes "shearing" of f_1 traces of a 2D dataset and is used to rotate the narrow projection of some solids correlations into the f_1 dimension. Several solids experiments for Dynamic Angle Spinning (DAS) and a triple-quantum filtered 2D MAS experiment require the use of daslp. (Note that the command rotate shears two traces and is inapplicable for these experiments.)

When created, the value of lp for each increment of a 2D experiment is incremented by the value of daslp after the first Fourier transformation. The incremented phase correction is applied to the interferogram created from the coefficient table by ftld, ft2d, wftld and wft2d, when coefficients are present. daslp is also used with ftlda, ft2da, wftlda and wft2da.

Values Real values, typically similar in size to the value of parameter 1p. See also NMR Spectroscopy User Guide Related ftld Fourier transform along f_2 dimension (C) ft1da Fourier transform phase-sensitive data (M) ft2d Fourier transform 2D data (C) ft2da Fourier transform phase-sensitive data (M) First-order phase in directly detected dimension (P) rotate Rotate 2D data (C) Weight and Fourier transform f2 for 2D data (C) wft1d wftlda Weight and Fourier transform phase-sensitive data (M)

- wft2d Weight and Fourier transform 2D data (C)
- wft2da Weight and Fourier transform phase-sensitive data (M)

date Date (P)

Description An informational parameter taken from the UNIX-level calendar (which is set by the UNIX system operator only and cannot be entered by the user). Whenever data are acquired, the date is copied from UNIX and written into the acquisition parameters, thus maintaining a record of the date of acquisition.

See also NMR Spectroscopy User Guide

daxis Display horizontal LC axis (M)

Applicability	Systems with LC-NMR accessory.		
Syntax	<pre>daxis(time,major_tic,minor_tic)</pre>		
Description	Displays a horizontal LC axis. Horizontal axes are assumed to be use with "LC plots" of an entire LC run and are labeled accordingly.		
Arguments time is the time scale, in minutes (decimal values are fine)			
	<pre>major_tic is spacing, in minutes (decimal values are fine), of major tics.</pre>		
	<pre>minor_tic is spacing, in minutes (decimal values are fine), of minor tics.</pre>		
See also	NMR Spectroscopy User Guide		
Related	paxis Display horizontal LC axis (M)		

Dbppste Set up parameters for Dbppste pulse sequence (M)

Description Converts a parameter set to Dbppste experiment; replaces the macro bppste.

See also	NMR Spectroscopy User Guide	
Related	dosy	Process DOSY experiments (M)
	fiddle	Perform reference deconvolution (M)
	setup_dosy	Set up gradient levels for DOSY experiments (M)

DbppsteineptSet up parameters for Dbppsteinept pulse sequence (M)

Description	Converts a parameter set to Dbppsteinept experiment.		
See also	NMR Spectroscopy User Guide		
Related	dosy	Process DOSY experiments (M)	
	fiddle	Perform reference deconvolution (M)	
	setup_dosy	Set up gradient levels for DOSY experiments (M)	

dbsetup Set up VnmrJ database (U)

Syntax	dbsetup <vnmr_adm remove standard imaging> dbsetup vnmr_adm <remove standard imaging> As Root:</remove standard imaging></vnmr_adm remove standard imaging>		
Arguments	dbsetup vnmr_adm VnmrJ_Home_dir <standard imaging> vnmr_adm is the login ID of the VnmrJ system administrator. remove only removes the data-database; does not recreate a database standard creates the database for standard use. imaging creates the database for imaging spectroscopy.</standard imaging>		
Description	 The UNIX script dbsetup is used during the installation of VnmrJ software and can only be run by the VnmrJ administrator (vnmr_adm) or the UNIX administrator (root). Normally it is never used again. dbsetup creates and deletes the data-database in /vnmr/pgsql/data and the user information in /vnmr/adm/users. When run as root at least two arguments must be supplied, the login ID of the VnmrJ administrator and the VnmrJ home directory. When run as root dbsetup will delete and recreate the data-database in /vnmr/pgsql/data for all users in /vnmr/adm/users. If no user list exists yet, the list is created with the VnmrJ administrator as the only user. The mode can be specified with the third argument as 'standard' or 'imaging'; if neither is specified the mode is taken from the global file of the VnmrJ administrator. It defaults to standard. The VnmrJ administrator does not need to supply any of the arguments. 		
	Note that additional users are created using vnmrj adm.		
Examples	dbsetup dbsetup vnmr1		

See also NMR Spectroscopy User Guide VnmrJ Imaging NMR VnmrJ Installation and Administration

dbupdate Update the VnmrJ database (U)

Applicability	Systems with the VnmrJ software.			
Syntax	dbsupdate stop once [slow_ms] forever [slow_ms]			
Arguments	<pre>slow_ms is an optional argument used to slow down the database update so as not to use all of the available CPU time. slow_ms=0 is full speed. slow_ms=1000 uses about 2-5% of the CPU. The dbupdate command is runs under nice so that any other process will be able to take the CPU away from this update anyway. The default slow_ms for forever is 1000. The default slow_ms for once is 0.</pre>			
Description	A UNIX command to start and stop a program to update the VnmrJ database used by the Locator. This command might be needed at a data station to view newly acquired data. The database at the spectrometer will automatically be updated.			

Calculate spectral drift correction (C)

Description Turns on a linear baseline correction. The beginning and end of the straight line to be used for baseline correction are determined from the display parameters sp and wp. dc applies this correction to the spectrum and stores the definition of the straight line in the parameters lvl (level) and tlt (tilt). The correction is turned off by the command cdc.

Care must be taken to ensure that a resonance does not appear too close to either end of the spectrum, or dc can produce the opposite effect from that intended; namely, it induces a sloping baseline where none was present!

See also NMR Spectroscopy User Guide

Related	bc	1D and 2D baseline correction (C)
	cdc	Cancel drift correction (C)
	dc	Drift correction group (P)
	lvl	Zero-order baseline correction (P)
	sp	Start of plot (P)
	tlt	First-order baseline correction (P)
	qw	Width of plot (P)

dc

dc2d Apply drift correction to 2D spectra (C)

Syntax	dc2d('f1' 'f2')		
Description	Computes a drift correction and applies it to each individual trace.		
Arguments	'fl' is a keyword to apply drift correction in the \mathbf{f}_1 axis direction		
	'f2' is a keyword to apply drift correction in the \mathbf{f}_2 axis direction.		
Examples	dc2d('f1') dc2d('f2')		
See also	NMR Spectroscopy User Guide		
Related	Axis label for displays and plots (P)		
	bc 1D and 2D baseline correction (C)		

dcg Drift correction group (P)

Description	Contains the results of the dc or cdc command. This parameter cannot be set in the usual way but it can be queried by entering dcg? to determine whether drift correction is active.	
Values	'dc' indicates drift correction is active. 'cdc' indicates drift correction is inactive.	
See also	NMR Spectroscopy User Guide	
Related	cdcCancel drift correction (C)dcCalculate spectral drift correction (C)	

dcon Display noninteractive color intensity map (C)

Syntax dcon<(options)>

Description Produces a "contour plot," actually a color intensity map, in the graphics window. The parameters sp and wp, sp1 and wp1, and sp2 and wp2 control which portion of the spectrum is displayed. The parameters sf and wf, sf1 and wf1, and sf2 and wf2 control which portion of time-domain data (FIDs and interferograms) is displayed. The parameter trace selects which dimension is displayed along the horizontal axis. The parameters sc, wc, sc2, and wc2 control where on the screen the display occurs. The parameter th is active as a threshold to black out all contours whose intensity is below th. That is, if th=7, the colors 1 to 6 are not used for the display. The parameter vs controls the vertical scale of the spectrum.

dcon displays either absolute-value mode or phase-sensitive 2D data. In av mode, data are shown in 15 different colors (starting with black), with each color representing a factor of two in intensity (a single color is used on monochrome screens). In the ph mode, the normal display of colors ranges from -6 to +6, each representing a factor of two in intensity, with the color black representing intensity 0 in the center.

Arguments options can be any of the following:

- 'linear' is a keyword to use linear instead of logarithmic increments.
- 'phcolor' is a keyword to use a phased color set with positive and negative peaks.
- 'avcolor' is a keyword to use an absolute-value color set with positive peaks. Negative contours only *cannot* be displayed, but if the data can be rephased, 180° added to rpl, and dcon('avcolor') entered again, the same thing is accomplished by inverting the phase of all peaks. Alternatively, dpcon can display negative peaks only.
- 'gray' is a keyword to use a gray scale color set.
- 'noaxis' is a keyword to omit the display outline and any horizontal or vertical axis.
- 'plot' causes the dcon display to be sent to the plotter instead of being drawn on the graphics window.

```
Examples dcon
```

```
dcon('gray')
dcon('linear','phcolor','plot')
```

```
See also NMR Spectroscopy User Guide
```

Related	dconi	Interactive 2D data display (C)
	dconi	Control display selection for the dconi program (P)
	dconn	Display color intensity map without screen erase
		(C)
	dpcon	Display plotted contours (C)
	imageprint	Plot noninteractive gray scale image (M)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	sf	Start of FID (P)
	sp	Start of plot (P)
	sp1	Start of plot in 1st indirectly detected dimension
		(P)
	sp2	Start of plot in 2nd indirectly detected dimension
		(P)
	th	Threshold (P)
	trace	Mode for n -dimensional data display (P)
	WC	Width of chart (P)
	wc2	Width of chart in second direction (P)
	wf	Width of FID (P)
	wp	Width of plot (P)
	wp1	Width of plot in 1st indirectly detected dimension
		(P)
	wp2	Width of plot in 2nd indirectly detected dimension
		(P)

dconi Interactive 2D data display (C)

Syntax dconi<(options)>

Description Opens a 2D data display that can be interactively adjusted. The dconi program can accommodate any data set that can be displayed by dcon, dpcon, and ds2d, including 2D FIDs, interferograms, 2D spectra, planes from 3D data sets, and images. These data sets are generated by the commands df2d, ft1d, ft2d, and ft3d.

Arguments options can be any of the following (note that the dconi parameter is also available to control the dconi program display):

- 'dcon' is a keyword to display a color intensity map; this is the default mode, but 'dcon' is provided for compatibility with certain macros. If 'dcon' is the first argument, it can be followed by any of the keywords 'linear', 'phcolor', 'avcolor', 'gray', and 'noaxis'; all of these keywords have the same meaning as when used with dcon.
- 'dpcon' is a keyword to display a true contour plot. If 'dpcon' is the first argument, it can be followed by any of the keywords 'pos', 'neg', and 'noaxis', and then followed by values for levels and spacing. All of these options have the same meaning as when used with dpcon.
- 'ds2d' is a keyword to display a stacked plot in whitewash mode (after the first spectra, each spectra is blanked out in regions in which it is behind an earlier spectra). If 'ds2d' is the first argument, it can be followed by any of the keywords 'nobase', 'fill', 'fillnb', and 'noaxis'. All of these keywords have the same meaning as used with ds2d.
- 'again' is a keyword to make dconi identify which display mode is currently being used and redraw the screen in that mode.
- 'restart' is a keyword to activate dconi without redrawing the 2D data set. This action causes dconi to make sure that 2D data is already displayed.
- 'toggle' is a keyword to toggle between the cursor and box modes.
- 'trace' is a keyword to draw a trace above the spectrum.
- 'expand' is a keyword to toggle between the expand and full views of the spectrum.
- 'plot' is a keyword to plot a projection or a trace.
- 'hproj_max' is a keyword to do a horizontal projection of the maximum trace.
- 'hproj_sum' is a keyword to do a horizontal projection of the sum of all traces.
- 'vproj_max' is a keyword to do a vertical projection of the maximum trace.
- 'vproj_sum' is a keyword to do a vertical projection of the sum of all traces.

Examples	dconi dconi('d dconi('d	con','gray','linear') pcon')		
See also	NMR Spec	NMR Spectroscopy User Guide		
Related	boxes	Draw boxes selected by the mark command (C)		
	crmode	Current state of cursors in dfid, ds, or dconi (P)		
	dcon	Display noninteractive color intensity map (C)		
	dconi	Control display selection for the dconi program (P)		
	dconn	Display color intensity map without screen erase (C)		
	delta1	Cursor difference in 1st indirectly detected dimension (P)		
	df2d	Display FIDs of 2D experiment (C)		
	dpcon	Display plotted contours (C)		
	ds2d	Display 2D spectra in whitewash mode (C)		
	ft1d	Fourier transform along f ₂ dimension (C)		
	ft2d	Fourier transform 2D data (C)		
ft3d	ft3d	Perform a 3D Fourier transform on a 3D FID data set		
		(M,U)		
	imconi	Display 2D data in interactive gray-scale mode (M)		
	is	Integral scale (P)		
	112d	Automatic and interactive 2D peak picking (C)		
	proj	Project 2D data (C)		
	sf	Start of FID (P)		
	sp	Start of plot (P)		
	sp1	Start of plot in 1st indirectly detected dimension (P)		
	th	Threshold (P)		
	vs2d	Vertical scale for 2D displays (P)		
	vsadj	Automatic vertical scale adjustment (M)		
	wf	Width of FID (P)		
	WD	Width of plot (P)		
	wp1	Width of plot in 1st indirectly detected dimension (P)		

dconi Control display selection for the dconi program (P)

Description Controls the selection of the 2D display that follows entering the dconi command. Because dconi is implicitly executed by ft2d, the dconi parameter also controls the display that follows the ft2d or wft2d command.

dconi can be a string parameter in the "current" parameter set. Its syntax is similar to an argument string passed to the dconi program. For example, if dconi = 'dpcon, pos, 12, 1.2', the dconi command displays twelve positive contours with dpcon, using a spacing of 1.2. The first component of the dconi string must be the name of the display program, such as dcon, dconn, dpcon, dpconn, ds2d, or ds2dn. Subsequent components of the string are arguments appropriate for that display program. Because the entire dconi parameter is a string, single quotes around words are not necessary and mixing words and numbers is not a problem, as the example above shows.

If the dconi parameter does not exist or is set to the null string (''), the dconi program uses its normal default. If the dconi parameter is set to a string (e.g., dconi='dcon, gray, linear' for image display), and arguments are supplied to the dconi program, (e.g.,

dconi ('dpcon')), the supplied arguments to the command take precedence. In the case of the examples above, a contour map, not an image, is displayed.

If the dconi parameter does not exist in the current experiment, it can be created by the commands create('dconi','string') setgroup('dconi','display')

Values '' (two single quotes) indicates that this parameter is ignored. String 'display_program' selects the named program for 2D displays. String 'display_program, option1, option2' selects the named

program for 2D displays with options appropriate to the program.

- Examples dconi='dpcon' selects contour drawing rather than default color map dconi='dcon,gray,linear' selects image display mode.
- See also NMR Spectroscopy User Guide; VnmrJ Imaging NMR

Related	dcon	Display noninteractive color intensity map (C)
	dconi	Interactive 2D data display (C)
dconnDisplay color intensity map without screen eradpconDisplay plotted contours (C)		Display color intensity map without screen erase (C)
		Display plotted contours (C)
	dpconn	Display plotted contours without screen erase (C)
	ds2d	Display 2D spectra in whitewash mode (C)
	ds2dn	Display 2D spectra in whitewash mode without screen erase
		(C)
	ft2d	Fourier transform 2D data (C)
	imconi	Display 2D data in interactive gray-scale mode (M)
	wft2d	Weight and Fourier transform 2D data (C)

dconn Display color intensity map without screen erase (C)

Suntar	dapprox(optiona)		
Syntax	aconn<(options)>		
Description	Produces a "contour plot," actually a color intensity map, on the screen the same as the dcon command, but without erasing the screen before starting the plot. The options available are the same as the dcon command.		
See also	NMR Spectroscopy User Guide		
Related	dcon	Display noninteractive color intensity map (C)	
	dconi	Control display selection for the dconi program (P)	

dcrmv Remove dc offsets from FIDs in special cases (P)

Description	If dcrmv ex remove the systems with particular ec create('d setgroup(dcrmv exists and is set to 'y', hardware information is used to move the dc offset from the FID providing ct=1. This only works on stems with sw less than 100 kHz. If this feature is desired for a rticular experiment, create dcrmv in that experiment by entering ceate('dcrmv', 'string') etgroup('dcrmv', 'processing') dcrmv='y'		
See also	To create image parameters dcrmv, grayctr and graysl in the current experiment, enter addpar('image'). NMR Spectroscopy User Guide; VnmrJ Imaging NMR			
Related	addpar create ct dc setgroup	Add selected parameters to the current experiment (M) Create new parameter in a parameter tree (C) Completed transients (P) Calculate spectral drift correction (C) Set group of a variable in a tree (C)		

ddf Display data file in current experiment (C)

Syntax	ddf<(block_number,trace_number,first_number)>		
Description	Displays the file header of the data file in the current experiment. If entered with arguments, it also displays a block header and part of the data file of that block.		
Arguments	block_number is the block number. Default is 1.		
	trace_number is the trace number within the block. Default is 1.		
	first_number is the first data element number within the trace. Default is 1.		
See also	User Programming		
Related	ddff Display FID file in current experiment (C)		
	ddfp Display phase file in current experiment (C)		

ddff Display FID file in current experiment (C)

Syntax	ddff<(block_number,trace_number,first_number)>
Description	Displays the file header of the FID file in the current experiment. If entered with arguments, it also displays a block header and part of the FID data of the block.
Arguments	block_number is the block number. Default is 1.
	trace_number is the trace number within the block. Default is 1.
	first_number is the first data element number within the trace. Default is 1.

See also	User Program	nming
Related	ddf	Display data file in current experiment (C)
	ddfp	Display phase file in current experiment (C)

ddfp Display phase file in current experiment (C)

Description Displays the file header of the phase file in the current experiment. With arguments, it also display a block header and part of the phase file data of that block.

Arguments	block_number is the block number. Default is 1.
	$\verb"trace_number"$ is the trace number within the block. Default is 1.
	first_number is the first data element number within the trace. Default is 1.
See also	User Programming
Related	ddf Display data file in current experiment (C)

ddif Synthesize and show DOSY plot (C)

ddff

Syntax	ddif(<pre><option></option></pre>	lowerlimit	,upperlimit)
--------	-------	------------------------------	------------	--------------

Description Synthesizes a 2D spectrum from 1D spectra using the information produced by the dosy macro. ddif takes the 1D spectrum and a table of diffusion data stored in the file diffusion_display.inp in the current experiment and synthesizes a 2D DOSY spectrum. It is normally run by dosy, but can be directly run, for example, to recalculate a 2D DOSY spectrum with different digitization.

Arguments option is either 'i' or 'c'.

'i' is for a display in which the 2D peak volume is proportional to 1D peak height.

Display FID file in current experiment (C)

'c' is for a display in which the 2D peak height equals the 1D.

lowerlimit is the lower diffusion limit (in units of 10^{-10} m²/s).

upperlimit is the upper diffusion limit (in units of 10^{-10} m²/s).

If arguments are not supplied, ddif defaults to showing the full range of diffusion coefficients in the file diffusion_display.inp in the current experiment. Make sure that the first increment of the DOSY data set has been transformed with the desired fn2D before using ddif. Digitization of the resultant spectrum is determined by fn2D in the spectral (F2) domain and fn1 in the diffusion (F1) domain. Make sure that the product fn2D*fn1 is not too large, or memory and processing time problems might result. Typical values are fn2D=16384 (max: 64k) and fn1=512. After dosy or ddif, 1D data is overwritten by the 2D (the dosy macro keeps a copy of the 1D data, which can be retrieved with the command undosy). Similarly, after a DOSY spectrum has been calculated, it can be retrieved with the command redosy.
 See also NMR Spectroscopy User Guide
 Related dosy Process DOSY experiments (M) fn2D Fourier number to build up 2D DOSY display in frequency domain (P)

- redosy Restore the previous 2D DOSY display from the subexperiment (M)
- undosy Restore original 1D NMR data from the subexperiment (M)

ddrcr Direct digital receiver coefficient ratio (P)

Applicability	VNMRS systems and 400 - MR systems
Syntax	ddrcr= <value></value>
Description	Sets the filter sharpness or filter coefficient ratio. The default value of 75 is used if the parameter does not exist.
Examples	create('ddrce','integer') setlimit('ddrcr',1000,2,1) ddrcr=300
Values	Integer values between 2 and 1000
See also	NMR Spectroscopy User Guide and VnmrJ User Programming.
Related	sw Spectral width in directly detected dimension (P)

ddrpm Set ddr precession mode (P)

Applicability	VNMRS sys	tems	
Syntax	ddrpm=<'mode'>		
Values	mode can b	e either of following:	
Mod	e Description		
р	Pulse — de	fault if no argument is supplied.	
	The value is	calculated as follows if ddrpm does not exist or	
	ddrpm='p	·':	
	ddrtc =	alfa + rof2 + 2 * pw[1] / π	
e	ECNO — The	e value is calculated as follows: ddrtc = alfa.	
See also	VnmrJ Use	r Programming	
Related	setrc	Set frequency referencing based upon lock signal shift	
		(M)	
	ddrtc	Set ddr precession mode (P)	

ddrtc Set ddr time constant (P)

Applicability	VNMRS systems		
Syntax	ddrtc=<'value'>		
Description	The value of ddrtc is set in the setrc macro and is determined by the ddrpm parameter.		
	A value of a not exist.	ddrtc = alfa is used by psg if the ddrtc parameter does	
Values	value 0 to 1000 $\mu sec.$		
See also	VnmrJ User Programming		
Related	setrc	Set frequency referencing based upon lock signal shift (M)	
	setlp0 ddrpm	Set parameters for zero linear phase (M) Set ddr precession mode (P)	

dds Default display (M)

- Description Looks for sequence-specific default display macro (dds_seqfil) and executes if one is found. If not, the dds macro displays 1D, 2D, or array spectrum as the case may be.
 - Related dds_seqfil Sequence-specific default display (M) dpl Default plot (M) dpr Default process (M)

dds_seqfil Sequence-specific default display (M)

Description	Sequence-spe	cific default display. These macros are called by the dds
	macro.	
Examples	dds_NOESY1D	
	dds_TOCSY1	
Related	dds	Default display (M)
	dpl	Default plot (M)
	dpr	Default process (M)

debug Trace order of macro and command execution (C)

Syntaxdebug('c'|'C')DescriptionControls VnmrJ command and macro tracing. When turned on, debug
displays a list of each command and macro in the shell tool from which
VnmrJ was started. If VnmrJ is started when the user logs in, or if it

was started from a drop-down menu or the CDE tool, the output goes to a Console window. If no Console window is present, the output goes into a file in the /var/tmp directory. This last option is not recommended. Nesting of the calls is indicated by indentation of the output. This feature is primarily a debugging tool for MAGICAL programming.
To associate the debut('c') output with a particular terminal, enter tty. The system responds with /dev/pts/yyy, where yyy is a numerical value. On the VnmrJ command line, enter jFunc(55, '/dev/pts/yyy'), substituting the numerical value for the yyy.
'c' is a keyword to turn on command and macro tracing.
'C' is a keyword to turn off command and macro tracing.
debug('c') debug('C')

See also User Programming

decasynctype Decoupler asynchronous scheme (P)

Applicability	VnmrJ	3.1
---------------	-------	-----

Description Specifies the decoupler asynchronous scheme. This flag parameter is optional, and can be used to select between different schemes to implement asynchronous decoupling. This parameter will be applicable to decoupling on all of the RF channels. If the decoupling mode (dm), 's' is selected, the decoupling is synchronous and this parameter has no effect.

Values 'p' selects the "progressive offset" scheme, which is the default. This simulates a free running decoupler modulation with respect to the acquisition window.

'b' selects the "bit reversal" scheme. This scheme uses the bit reversal algorithm to implement asynchronous decoupling. It attempts to efficiently sample various phases of the decoupling cycle and hence may be more appropriate when number of transients (nt) is a small number that is a power of two.

'r' selects a random scheme for implementing asynchronous decoupling.

decay_gen Calculates the form of diffusional attenuation expected for the measured gradient and signal maps in non-uniform gradient calibration.

Syntax decay_gen(D,ngrads) Applicability VnmrJ 3.1

Description	decay_gen takes the measured signal profile and gradient map as a function of position and calculates the predicted signal attenuation as a function of gradient strength.
Arguments	decay_gen takes two arguments: the diffusion coefficient (D) of the calibrant, and the number of gradient levels (ngrads) for which the attenuation is to be calculated. decay_gen is normally run only by the nugcalib macro.
See also	nugcalib gradfit powerfit

deccwarningsControl reporting of DECC warnings from PSG (P)

Applicability	Systems with DECC (Digital Eddy Current Compensation) boards for gradient compensation.
Description	A global parameter that controls whether PSG will warn the user when the ECC corrections are large enough that they could exceed the capabilities of the DECC board. By default, this parameter does not exist, and a warning is printed whenever an experiment is started if the ECC amplitudes are possibly too large. The warning does indicate a definite be a problem, only that not enough ECC drive capability is available to compensate for an instantaneous gradient swing from minus the maximum gradient strength to the maximum positive gradient.
	To disable the warnings, create this global string parameter and set it to 'n'.

Values 'n' or 'N' to suppress warnings. If the value starts with any other character, the normal warnings are printed.

decomp Decompose a VXR-style directory (M)

Syntax	<pre>decomp<(VXR_file)></pre>	
Description	Takes a library, as loaded from a VXR-style system (VXR, XL, or Gemini), and extracts each entry into a separate UNIX file. The file can be obtained from a magnetic tape or over limNET. decomp creates a UNIX subdirectory in the current working directory and uses that to write each entry as a UNIX file. The name of the UNIX subdirectory is derived from the library name.	
Arguments	VXR_file is the name of the original file. It must have an extension in the form .NNN, where NNN is the number of entries in the original library. A limit of 432 entries is imposed.	
See also	NMR Spectroscopy User Guide	
Related	convert Convert data set from a VXR-style system (C,U)	

def_osfilt Default value of osfilt parameter (P)

- Description A global parameter that establishes the default type of digital filter, AnalogPlus[™] or brickwall, when DSP is configured. The *actual* filter used in any experiment is set by the local parameter osfilt. Usually, def_osfilt is set to the value for normal use, and then osfilt is changed within a given experiment if different filter characteristics are desired.
 - Values 'a' or 'A' for the Analog*Plus* digital filter. This filter is flatter in the passband and drops off somewhat more sharply than analog filters.

'b' or 'B' for the brickwall digital filter. This filter is extremely flat across the passband and drops off sharply on the edge; however, the enhanced filtering comes at the expense of somewhat reduced baseline performance.

- See also NMR Spectroscopy User Guide
 - Related dsp Type of DSP for data acquisition (P) osfilt Oversampling filter for real-time DSP (P)

defaultdir Default directory for Files menu system (P)

- Description Stores the name to the default directory for use with the Directory Menu in the Files menu system. Initial value for defaultdir is the home or login directory of the user. Selecting the Default button in the Directory Menu sets the current directory to the value of defaultdir. The opposite action, setting the value of defaultdir to the current directory, occurs when the Set Default button in the Directory Menu is selected. If the entry for a directory is marked and the Set Default button is selected, the directory marked becomes the new value of defaultdir.
 - See also NMR Spectroscopy User Guide

delcom Delete a user macro (M)

Syntax	delcom(file	e)
Description	Deletes a ma delcom will	cro file in a user's macro library (maclib). Note that not delete a macro in the VnmrJ system macro library.
Arguments	file is the file name of the user's macro to be deleted.	
Examples	delcom('lds')	
See also	User Programming	
Related	crcom	Create user macro without using a text editor (C)
	macrorm	Remove a user macro (C)

Syntax delete(file1<, file2, ...>)

Description Delete files and directories in a somewhat safer manner than the rm command. Using rm is not recommended in VnmrJ because rm allows wildcard characters (* and ?) in the file description and recursive file deletion with the -r option. The delete command does not allow wildcard characters or the -r option, but you can still use the delete command to delete a file as well as remove .fid and .par directories, normally the only directories that need to be removed (experiment directories are deleted with the delexp macro).

- Arguments file1, file2, ... are the names of one or more files or directories to be deleted. When the delete command is entered, it first searches for file1. If it finds that file and it is not a directory, file1 is deleted. If file1 is not found, .fid is appended to the file name and delete searches for the file in that .fid directory. If the file is found, it is removed; otherwise, .par is appended to the file name and delete searches for the file in that .par directory. If the file is found, it is removed; otherwise, the command takes no action and continues to the next file name. The process is repeated for each file name given as an argument.
- Examples delete('/home/vnmr1/memo') delete('/vnmr/fidlib/fidld')

See also NMR Spectroscopy User Guide

RelateddelexpDelete an experiment (M)rmDelete file (C)rmdirRemove directory (C)

delexp Delete an experiment (M)

Syntax delexp(exp_no):\$stat,\$message

Description Deletes an experiment. Only experiments 2-9999 can be deleted; experiment 1 cannot be deleted by delexp.

The delexp macro calls the DELEXP command to delete the experiment The delexp macro then does the additional step of deleting an appropriate jexp<N> macro from the user's maclib, where the <N> is the exp_no and if exp_no is greater than 9.

Both the DELEXP command and delexp macro will return two optional values to the calling macro. The first (\$stat) is set to 0 if the command / macro fails and it is set to 1 if the command / macro succeeds. The second return value is text message that can be used.

The DELEXP command has one special syntax, DELEXP('auto'). This schedules the current experiment to be deleted when the current background Vnmrbg finishes executing its command. This is only available from a background Vnmrbg. A typical usage would be for a

foreground macro to assign a task to a background Vnmrbg in a separate experiment. For example,

	<pre>nextexp:\$e // find number for a new experiment</pre>	
	CEXP(\$e):\$stat // silently create the experiment	
	<pre>write('line3',`Vnmrbg -mback -n%d "DELEXP('auto')</pre>	
Arguments	$\exp(n0)$ is the number (from 2 through 9999) of the experiment to be deleted (experiment 1 cannot be deleted). delexp also deletes the corresponding $jexpXXX$ macro if necessary.	
Examples	delexp(2)	
	<pre>delexp(75):\$stat,\$msg // Silently delete exp75. If the</pre>	
	<pre>DELEXP(77):\$stat,\$msg // Silently delete exp77, but do</pre>	
See also	NMR Spectroscopy User Guide	
Related	cexpCreate an experiment (M)jexpJoin existing experiment (C)	

delexpdata Delete data from the current experiment

Syntax	delexpdata
Applicability	VnmrJ 3.1
Description	The delexpdata command will remove data from the current
	experiment. It will delete 3D data, if present. This command will not
	execute if an acquisition is active or queued in the current experiment.

deletenucleus Removes nucleus entry from current probe file (M)

Applicability	ALL	
Description	All lines for the specified nucleus are removed from the current probe file. The argument should correspond to an entry in the probe file.	
Syntax	<pre>deletenucleus('nucleus')</pre>	
Arguments	nucleus - name followed by atomic number, e.g. c13 not 13C.	
Examples	deletenucleus('Si29')	
Related	addnucleusAdds nucleus entry to probe file (M)addprobeCreate new probe directory and probe file (M)	

dels Delete spectra from T_1 or T_2 analysis (C)

Syntax	<pre>dels(index1<, index2,>)</pre>	
Description	Deletes the spectra selected from the file fp.out (the output file of fp) used by the t1 or t2 analysis. Spectra may be restored by rerunning fp.	
Arguments	index1, index2, are the indexes of the spectra to be deleted.	
Examples	dels(7) dels(2,5)	
See also	NMR Spectroscopy User Guide	
Related	d11	Display listed line frequencies and intensities (C)
	fp	Find peak heights or phases (C)
	getll	Get frequency and intensity of a line (C)
	t1	T_1 exponential analysis (M)
	t2	T_2 exponential analysis (M)

delta Cursor difference in directly detected dimension (P)

Description	Difference between two frequency cursors along the directly detected dimension. The value is changed by moving the right cursor, relative to the left, in the ds or dconi display.	
Values	Positive number, in Hz.	
See also	NMR Spectroscopy User Guide	
Related	dconi	Interactive 2D data display (C)
	delta1	Cursor difference in 1st indirectly detected dimension (P)
	delta2	Cursor difference in 2nd indirectly detected dimension
		(P)
	ds	Display a spectrum (C)
	split	Split difference between two cursors (M)

delta1 Cursor difference in 1st indirectly detected dimension (P)

Description	Difference of two frequency cursors along the first indirectly detected dimension. Analogous to the delta parameter except that delta1 applies to the first indirectly detected dimension of a multidimensional data set.	
Values	Positive number, in Hz.	
See also	NMR Spectroscopy User Guide	
Related	delta Cursor difference in directly detected dimension (P)	

delta2 Cursor difference in 2nd indirectly detected dimension (P)

Description	Difference of two frequency cursors along the second indirectly detected dimension. Analogous to the delta parameter except that delta2 applies to the second indirectly detected dimension of a multidimensional data set.	
Values	Positive number, in Hz.	
See also	NMR Spectroscopy User Guide	
Related	delta Cursor difference in directly detected dimension (P)	

deltaf Difference of two time-domain cursors (P)

Description	Difference between the two time-domain cursors of the df (or dfid) display. To create this parameter and the other FID display parameters axisf, dotflag, vpf, vpfi, and crf (if the parameter set is older and lacks these parameters), enter addpar('fid').		
Values	Number, in seconds.		
See also	NMR Spectroscopy User Guide		
Related	addpar crf df dfid	Add selected parameters to the current experiment (M) Current time-domain cursor position (P) Display a single FID (C) Display a single FID (C)	

dept Set up parameters for DEPT experiment (M)

Description	Set up parameters for DEPT experiment	
See also	NMR Spectroscopy User Guide	
Related	adept	Automatic DEPT analysis and spectrum editing (C)
	autodept	Automated complete analysis of DEPT data (M)
	deptgl	Set up parameters for DEPTGL pulse sequence (M)
	deptproc	Process array of DEPT spectra (M)
	padept	Plot automatic DEPT analysis (C)
	ppcal	Proton decoupler pulse calibration (M)

deptg1 Set up parameters for DEPTGL pulse sequence (M)

Description Macro for the DEPTGL pulse sequence for spectral editing and polarization transfer experiments.

See also NMR Spectroscopy User Guide

Related Dept Set up parameters for DEPT pulse sequence (M)

deptproc Process array of DEPT spectra (M)

Description Automatically processes arrays of DEPT-type spectra. The FIDs are transformed (using lb=2.5), phased, and scaled. In foreground operation, a stacked display is produced. By default, an automatic DEPT analysis (adept) is performed.

See also NMR Spe	ctroscopy User	Guide
------------------	----------------	-------

Related	adept	Automatically edit DEPT spectra (C)
	Dept	Set up parameters for DEPT experiment
	lb	Line broadening along the directly detected dimension
		(P)
	pldept	Plot DEPT type spectra (M)
	procplot	Automatically process FIDs (M)

destroy Destroy a parameter (C)

Syntax	<pre>destroy(parameter<,tree>)</pre>		
Description	Removes a parameter from one of the parameter trees. If the destroyed parameter was an array, the array parameter is automatically updated.		
	If destroy abort with indicate st	is called for a non-existent parameter, the command will a message. If an optional return value is given, it will uccess (1) or failure (0) and the command will not abort.	
Arguments	paramete	r is the name of the parameter to be destroyed.	
	tree is a 'current 'current types of t	keyword for the type of parameter tree: 'global', ', 'processed', or 'systemglobal'. The default is '. Refer to the create command for more information on rees.	
Examples	destroy('a')		
	destroy('c','global')	
See also	User Prog	ramming	
Related	array	Parameter order and precedence (P)	
	create	Create new parameter in a parameter tree (C)	
	display	Display parameters and their attributes (C)	
	paramvi	Edit a variable and its attributes using vi text editor (C)	
	prune	Prune extra parameters from current tree (C)	

destroygroupDestroy parameters of a group in a tree (C)

Syntax	destroygroup(group<,tree>)		
Description	Removes parameters of a group from one of the parameters trees.		
Arguments	group is a keyword for the type of parameter group: 'all', 'sample', 'acquisition', 'processing', 'display', or 'spin'.		
	tree is a ke 'current', create com	yword for the type of parameter tree: 'global', or 'processed'. The default is 'current'. Refer to the mand for more information on trees.	
Examples	destroygroup('sample') destroygroup('all','global')		
See also	User Programming		
Related	create	Create new parameter in a parameter tree (C)	
	destroy	Destroy a parameter (C)	
	display	Display parameters and their attributes (C)	
	groupcopy	Copy parameters of group from one tree to another (C)	
	setgroup	Set group of a variable in a tree (C)	

Display a single FID (C)

- Syntax (1) df<(index)> (2) df(options)
- Description Displays a single FID. Parameter entry after an FID has been displayed causes the display to be updated. The FID is left-shifted by the number of complex data points specified by the parameter lsfid. The FID is also phase-rotated (zero-order only) by the number of degrees specified by the parameter phfid. Left shifting and phasing can be avoided by setting lsfid and phfid to 'n'. df is identical in function to the dfid command.

Arguments index (used with syntax 1) is the number of a particular FID for arrayed 1D experiments or for 2D experiments. Default is 1.

options (used with syntax 2) is any of the following:

- 'toggle' is a keyword to switch between box and cursor modes.
- 'restart' is a keyword to redraw the cursor if it has been turned off.
- 'expand' is a keyword to switch between expanded and full views of the FID.
- 'imaginary' is a keyword to switch on and off the display of the imaginary FID.
- 'sfwf' is a keyword to interactively adjust the start and width of the FID display.
- 'phase' is a keyword to enter an interactive phasing mode.

đf

	•'dscale'	is a keyword to toggle the scale below the FID on and off.	
Examples	df		
	df(4)		
	df('rest	art')	
See also	NMR Spectroscopy User Guide		
Related	crmode	Current state of cursors in dfid, ds, or dconi (P)	
	dfid	Display a single FID (C)	
	df2d	Display FIDs of 2D experiment (C)	
	dfmode Current state of display of imaginary part of a FID		
		(P)	
	lsfid	Number of complex points to left-shift the np FID	
		(P)	
	phfid	Zero-order phasing constant for the np FID (P)	

df2d Display FIDs of 2D experiment (C)

Syntax	df2d<(<'nf',> <array_index>)></array_index>	
Description	Produces a color intensity map of the raw 2D FIDs as a function of t_1 and t_2 . The display can be modified by subsequent display commands, for example, df2d dconn will display the 2D FIDs without clearing the graphics screen.	
Arguments	'nf' is a keyword specifying that the data has been collected in the compressed form using nf. In other words, each array element is collected as one 2D FID or image comprised of nf FIDs or traces.	
Examples	df2d(1)	
See also	NMR Spectroscopy User Guide	
Related	dconiInteractive 2D data display (C)dfDisplay a single FID (C)	

dfid Display a single FID (C)

Syntax	<pre>(1) dfid<(index)> (2) dfid<(options)></pre>
Description	Functions the same as the df command. See df for information.
See also	NMR Spectroscopy User Guide
Related	df Display a single FID (C)

Description	Holds a string variable that reflects the state of display of the imaginary part of a FID. dfmode is primarily used by the programmable menu dfid to determine the status of the display of the imaginary part of a FID.	
Values	'r' indicates the current display is real only.	
	'i' indicates the current display is imaginary.	
	'z' indicates the display is zero imaginary.	
See also	User Programming	

dfrq Transmitter frequency of first decoupler (P)

Description	Contains the automatically be necessary	transmitter frequency for the first decoupler. $dfrq$ is y set when the parameter dn is changed and should not f for the user to manually set.
Values	Frequency, in channel.	n MHz. The value is limited by synthesizer used with the
See also	NMR Spectroscopy User Guide	
Related	dfrq2	Transmitter frequency of second decoupler (P)
	dfrq3	Transmitter frequency of third decoupler (P)
	dfrq4	Transmitter frequency of fourth decoupler (P)
	dn	Nucleus for first decoupler (P)
	dof	Frequency offset for first decoupler (P)
	sfrq	Transmitter frequency of observe nucleus (P)
	spcfrq	Display frequencies of rf channels (M)

dfrq2 Transmitter frequency of second decoupler (P)

Applicability Systems with a second decoupler.

- Description Contains the transmitter frequency for the second decoupler. dfrq2 is automatically set when parameter dn2 is changed and should not be necessary for the user to manually set.
 - Values Frequency, in MHz. Value is limited by synthesizer used with the channel. If dn2=' (two single quotes with no space in between) and a second decoupler is present in the console, dfrq2 is internally set to 1 MHz.
 - See also NMR Spectroscopy User Guide

Related	dn2	Nucleus for second decoupler (P)
	dof2	Frequency offset for second decoupler (P)

dfrq3 Transmitter frequency of third decoupler (P)

Applicability	Systems with a third decoupler.
Description	Contains the transmitter frequency for the third decoupler. dfrq3 is automatically set when the parameter dn3 is changed and should not be necessary for the user to manually set.
Values	Frequency, in MHz. Value is limited by synthesizer used with the channel. If $dn3=''$ (two single quotes with no space in between) and a third decoupler is present in the console, $dfrq3$ is internally set to 1 MHz.
See also	NMR Spectroscopy User Guide
Related	dn3Nucleus for third decoupler (P)dof3Frequency offset for third decoupler (P)

dfrq4 Transmitter frequency of fourth decoupler (P)

- Applicability Systems with a deuterium decoupler channel as the fourth decoupler.
 Description Contains the transmitter frequency for the fourth decoupler. dfrq4 is automatically set when the parameter dn4 is changed and should not be necessary for the user to manually set.
 Values Frequency, in MHz. Value is limited by a synthesizer used with the
 - channel. If dn4='' (two single quotes with no space in between) and a fourth decoupler is present in the console, dfrq4 is internally set to 1 MHz.
 - See also NMR Spectroscopy User Guide

Related	dn4	Nucleus for fourth decoupler (P)
	dof4	Frequency offset for fourth decoupler (P)
	spcfrq	Display frequencies of rf channels (M)
	rftype	type of rf generation

dfs Display stacked FIDs (C)

Syntax	dfs<(<start><,finish><,step><,'all' 'imag'><,color>)></start>
Description	Displays one or more FIDs. The position of the first FIDs is governed by the parameters wc, sc, and vpf. A subsequent FID is positioned relative to the preceding FID by the parameters vo and ho.
Arguments	start is the index number of the first FID for multiple FIDs. It can also be the index number of a particular FID for arrayed 1D or 2D data sets.
	finish is the index number of the last FID for multiple FIDs. To include all FIDs, set start to 1 and finish to arraydim (see example below).

	step is th	e increment for the FID index. The default is 1.
	'all' is a	keyword to display all of the FIDs. This is the default.
	'imag' is	a keyword to display only the imaginary FID channel.
	color is t 'magenta	<pre>he color of the display: 'red', 'green', 'blue', 'cyan', ', 'yellow', 'black', or 'white'.</pre>
Examples	dfs(1,arı dfs('imag	caydim,3) g')
See also	NMR Spect	roscopy User Guide
Related	arraydim	Dimension of experiment (P)
	dfsa	Display stacked FIDs automatically (C)
	dfsan	Display stacked FIDs automatically without screen erase
		(C)
	dfsh	Display stacked FIDs horizontally (C)
	dfshn	Display stacked FIDs horizontally without screen erase
		(C)
	dfsn	Display stacked FIDs without screen erase (C)
	dfww	Display FIDs in whitewash mode (C)
	ho	Horizontal offset (P)
	plfid	Plot FID (C)
	pfww	Plot FIDs in whitewash mode (C)
	SC	Start of chart (P)
	vo	Vertical offset (P)
	vpf	Current vertical position of FID (P)
	WC	Width of chart (P)

dfsa Display stacked FIDs automatically (C)

Syntax	dfsa<(<start><,finish><,step><,'all' 'imag'><,color>)></start>
Description	Displays one or more FIDs automatically by adjusting the parameters vo and ho to fill the screen in a lower left to upper right presentation (wc must be set to less than full screen width for this to work). The position of the first FID is governed by parameters wc, sc, and vpf.
Arguments	start is the index number of the first FID for multiple FIDs. It can also be the index number of a particular FID for arrayed 1D or 2D data sets.
	finish is the index number of the last FID for multiple FIDs.
	step is the increment for the FID index. The default is 1.
	'all' is a keyword to display all of the FIDs. This is the default.
	'imag' is a keyword to display only the imaginary FID channel.
	color is the color of the display: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', or 'white'.

dfsan Display stacked FIDs automatically without screen erase (C)

Syntax	dfsan<(<start><,finish><,step><,'all' 'imag'><,color>)></start>
Description	Functions the same as the command dfsa except the graphics window is not erased before starting the display. This allows composite displays of many FIDs to be created. The arguments are the same as dfsa.
See also	NMR Spectroscopy User Guide
Related	dfsa Display stacked FIDs automatically (C)

dfsh Display stacked FIDs horizontally (C)

Syntax	dfsh<(<start><,finish><,step><,'all' 'imag'><,color>)></start>
Description	Displays one or more FIDs horizontally by setting vo to zero and adjusting ho, sc, and wc to fill the screen from left to right with the entire array. The position of the first FID is governed by parameters wc, sc, and vpf.
Arguments	start is the index number of the first FID for multiple FIDs. It can also be the index number of a particular FID for arrayed 1D or 2D data sets.
	finish is the index number of the last FID for multiple FIDs. To display all FIDs, set finish to the parameter arraydim.
	step is the increment for the FID index. The default is 1.
	'all' is a keyword to display all of the FIDs. This is the default.
	'imag' is a keyword to display only the imaginary FID channel.
	color is the color of the display: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', or 'white'.
See also	NMR Spectroscopy User Guide
Related	dfsDisplay stacked FIDs (C)dfshnDisplay stacked FIDs horizontally without screen erase (C)

dfshn Display stacked FIDs horizontally without screen erase (C)

Syntax dfshn<(<start><,finish><,step><,'all'|'imag'><,color>)>

dfsn Display stacked FIDs without screen erase (C)

Syntax	dfsn<(<start><,finish><,step><,'all' 'imag'><,color>)></start>
Description	Functions the same as the command dfs except the graphics window is not erased before starting the display. This allows composite displays of many FIDs to be created. The arguments are the same as dfs .
See also	NMR Spectroscopy User Guide
Related	dfs Display stacked FIDs (C)

dfww Display FIDs in whitewash mode (C)

Syntax	dfww<(<start><,finish><,step><,'all' 'imag'><,color>)></start>
Description	Displays FIDs in whitewash mode (after the first FID, each FID is blanked out in regions in which it is behind an earlier FID). The position of the first FIDs is governed by parameters wc, sc, and vpf.
Arguments	start is the index number of the first FID for multiple FIDs. It can also be the index number of a particular FID for arrayed 1D or 2D data sets.
	finish is the index number of the last FID for multiple FIDs.
	step is the increment for the FID index. The default is 1.
	'all' is a keyword to display all of the FIDs. This is the default.
	'imag' is a keyword to display only the imaginary FID channel.
	color is the color of the display: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', or 'white'.
See also	NMR Spectroscopy User Guide
Related	dfsDisplay stacked FIDs (C)pfwwPlot FIDs in whitewash mode (C)

dg

Display group of acquisition/processing parameters (C)

Syntax dg('template',<'file_name'>)

Description Displays the group of acquisition and 1D/2D processing parameters. To display an individual parameter, enter the name of the parameter
	followed by a question mark (e.g., sw?). Parameters do not have to be displayed in order to be entered or changed. The dg display is controlled by the string parameter dg.	
Arguments	template is the name of the template parameter. The default is 'dg'. See the manual <i>User Programming</i> for rules on constructing a template. The macros dg dg1, dg2, dg1p, and dgs activate dg with a template argument such as 'dg','dg1', 'dg2', 'dg1p', 'dgs', etc. or a user defined template.	
	file_name is the name of the file to which the dg command will write the parameters specified by template.	
Examples	dg dg('dgexp') dg('dg','dgout')	
See also	NMR Spectroscopy User Guide; User Programming	
Related	 Pisplay the value of an individual parameter (C) Display acquisition parameter arrays (C) Display group of linear prediction parameters (C) Display acquisition parameter arrays (P) Control dg parameter group display (P) Control dglp parameter group of linear prediction parameters (P) 	
	dg1 Display group of display parameters (M)	
	(M)	

dgs Display group of special/automation parameters (M)

dg Control dg parameter group display (P)

- Description Controls the display of the dg command for the group of acquisition and 1D/2D processing parameters. dg, a string parameter, can be modified with the command paramvi('dg').
 - See also NMR Spectroscopy User Guide
 - Related dg Display group of acquisition/processing parameters (C) paramvi Edit a parameter and its attributes with vi text editor (C)

dg1 Display group of display parameters (M)

Description Displays the group of display parameters. To display an individual parameter, enter the name of the parameter followed by a question mark (e.g., sp?). Parameters do not have to be displayed in order to be entered or changed. The dg1 display is controlled by the string parameter dg1.

See also	NMR Spectroscopy User Guide
Related	? Display individual parameter value (C)
	dg1 Control dg1 parameter group display (P)
	dg Display group of acquisition/processing parameters (C)

dg1Control dg1 parameter group display (P)

Description	Controls the display of the dgl command for the group of display
	parameters. dg1, a string parameter, can be modified with
	paramvi('dg1').
See also	NMR Spectroscopy User Guide

Related dg1 Display group of display parameters (M) paramvi Edit a parameter and its attributes with vi text editor (C)

dg2 Display group of 3rd and 4th rf channel/3D parameters (M)

- Description Displays the group of acquisition parameters associated with a second decoupler channel on a system with a third rf channel. It also displays the group of parameters associated with selective 2D processing of 3D data sets. To display an individual parameter, enter the name of the parameter followed by a question mark (e.g., sw?). Parameters do not have to be displayed in order to be entered or changed. The dg2 display is controlled by the string parameter dg2.
 See also NMR Spectroscopy User Guide
 - RelateddgDisplay group of acquisition/processing parameters (C)dg2Control dg2 parameter group display (P)

dg2 Control dg2 parameter group display (P)

DescriptionControls the display of the dg2 command for the group of 3rd and 4th
rf channel/3D parameters. dg2, a string parameter, can be modified
with the command paramvi ('dg2'). To retrieve the dg2 and ap
display templates for the current experiment, enter addpar('3rf').See alsoNMR Spectroscopy User GuideRelatedaddparAdd selected parameters to the current experiment (M)
dg2Display group of 3rd and 4th rf channel/3D parameters
(M)paramviEdit a parameter and its attributes with vi text editor
(M)

dga Display group of spin simulation parameters (M)

- Description Displays the file of spin simulation parameters (Group A). There is one such group of parameters in the data system, not one per experiment as with normal NMR parameters.
 - See also NMR Spectroscopy User Guide
 - Related dgDisplay group of acquisition/processing parameters (C)dlaDisplay spin simulation parameter arrays (C)

dgcsteSL Set up parameters for DgcsteSL pulse sequence (M)

Description	Converts a parameter set to DgcsteSL experiment.	
See also	NMR Spectroscopy User Guide	
Related	dosy	Process DOSY experiments (M)
	fiddle	Perform reference deconvolution (M)
	setup_dosy	Set up gradient levels for DOSY experiments (M)

dgcstecosy Set up parameters for Dgcstecosy pulse sequence (M)

Description	Converts a parameter set to Dgcstecosy experiment	
See also	NMR Spectroscopy User Guide	
Related	dosy	Process DOSY experiments (M)
	makeslice	Synthesize 2D projection of a 3D DOSY spectrum
		(C)
	setup_dosy	Set up gradient levels for DOSY experiments (M)
	showoriginal	Restore first 2D spectrum in 3D DOSY spectrum
		(M)

dgcstehmqc Set up parameters for Dgcstehmqc pulse sequence (M)

Description	Converts a parameter set to Dgcstehmqc experiment	
See also	NMR Spectroscopy User Guide	
Related	dosy makeslice setup_dosy showoriginal	Process DOSY experiments (M) Synthesize 2D projection of 3D DOSY spectrum (C) Set up gradient levels for DOSY experiments (M) Restore first 2D spectrum in 3D DOSY spectrum (M)

Applicability	Systems with LC-NMR accessory.	
Description	Displays parameters related to LC-NMR on a separate screen. This macro is equivalent to the command dg('dglc').	
See also	NMR Spectroscopy User Guide	
Related	dglc Control LC-NMR parameter display (P)	

dglc Control dglc parameter group display (P)

Applicability	Systems with LC-NMR accessory.	
Description	Controls the display of the LC-NMR parameters by the macro dglc and the equivalent command dg('dglc'). If this parameter does not exist, the parlc macro can create it.	
See also	NMR Spectroscopy User Guide	
Related	dglc parlc	Display LC-NMR parameters (M) Create LC-NMR parameters (M)

dglp Display group of linear prediction parameters (C)

Syntax	dglp	
Description	Displays the linear prediction parameters group. Parameters do not have to be displayed in order to be entered or changed. The dglp display is controlled by the string parameter dglp.	
Examples	dglp	
See also	NMR Spectroscopy User Guide; User Programming	
Related	dg Control dg parameter group display (P)	

dgs Display group of shims and automation parameters (M)

Description Displays the group of shims and automation parameters. To display an individual parameter, enter name of the parameter followed by a question mark (e.g., sw?). Parameters do not have to be displayed in order to be entered or changed. The dgs display is controlled by the parameter dgs.
See also NMR Spectroscopy User Guide

dgs Control dgs parameter group display (P)

Description	Controls display of the dgs command for the group of shims and
	automation parameters. dgs, a string parameter, can be modified by
	paramvi('dgs').
See also	NMR Spectroscopy User Guide

Related dgs Display group of special/automation parameters (M) paramvi Edit a parameter and its attributes with vi text editor (C)

dhp Decoupler high-power control with class C amplifier (P)

Applicability	System with a class C amplifier.		
Description	dhp selects a decoupler high-power level for systems with class C amplifiers on the decoupler channel. Specific values of dhp should be calibrated periodically for any particular instrument and probe combination. As a rough guide, dhp=75 corresponds to approximate 2 watts at 200 MHz.		
CAUTION	Decoupler power greater than 2 watts in a switchable probe will damage the probe. Always carefully calibrate high-power decoupling to avoid exceeding 2 watts of power.		
	For systems equipped with a linear amplifier on the decoupler channel, dhp is nonfunctional and is replaced by the parameter $dpwr$.		
	Note that dhp runs in the opposite direction from dlp (i.e., for dhp a higher number means more power, for dlp a higher number means less power).		
Values	0 to 255 (where 255 is maximum power) in uncalibrated, non-linear units.		
	'n' selects low-power decoupling under the control of the parameter dlp.		
See also	NMR Spectroscopy User Guide		
Related	dlpDecoupler low power with class C amplifier (P)dpwrPower level for first decoupler with linear amplifier (P)tnNucleus for observe transmitter (P)		

diagth2d Exclude diagonal peaks when peak picking

Applicability VnmrJ 3.1

Description This parameter is used by 112d to exclude diagonal peaks when peak picking. Peaks within diagth2d Hertz of the diagonal will not be picked by 112d. Setting diagth2d to 0.0 will cause 112d to pick all peaks including diagonal peaks.

Related 112d

dialog Display a dialog box from a macro (C)

Syntax	<pre>dialog(definition_file,output_file<,'nowait'>)</pre>	
Description	Opens a dialog box from a macro. The output is written to a file that can be read by the macro using the lookup command.	
Arguments	definition_file is the name of the file (specified by an absolute path) that defines the layout of the dialog box.	
	output_file is the name of the file (specified by an absolute path) where the results of the dialog box are written.	
	'nowait' is a keyword to return immediately, without waiting for input into the dialog box.	
Examples	dialog(userdir+'/dialoglib/array,'/tmp/array')	
See also	User Programming	
Related	Look up words and lines from a text file (C)	

diffparam Report differences between parameter sets (UNIX)

Syntax	diffparam file1 file2 <parametergroup></parametergroup>		
Applicability	VnmrJ 3.1		
Description	Reports differences between VNMR parameter sets, based on the output of the listparam command.		
Arguments	<pre>file1 and file2 are VNMR parameter files, like \$HOME/vnmrsys/exp1/procpar \$HOME/vnmrsys/exp1/curpar \$HOME/vnmrsys/global /vnmr/conpar xyz.fid/procpar</pre>		
	file1 and file2 can also be directories (xyz.fid or xyz.par, or a local experiment like ~/vnmrsys/exp1); in this case diffpar will look for a subfile procpar in these directories. parametergroup is an optional argument that permits specifying the parameter type. By default, only acquisition parameters are compared. The following options exist (only the first two characters are relevant):		
	• acquisition - compare acquisition parameters (default)		
	• processing - compare processing parameters only		
	• display - compare display parameters only		

	• spsim - compare spin simulation parameters only
	• sample - compare sample parameters only
	\bullet all - compare ALL parameters (output indicates group for) for each parameter
	• JCAMP - compare acquisition and processing parameters in JCAMP-DX compatible format. Inactive parameters are suppressed.
Examples	diffparam abc.fid xyz.fid diffparam ~/vnmrsys/exp[13] processing diffparam ~/vnmrsys/exp[12]/curpar
Related	list parameters in simple format (UNIX) vnmr2jcamp create JCAMP parameters from VNMR parameters

diffparams Report differences between two parameter sets (U)

(UNIX)

Syntax diffparams <-list> file1 file2 <macroname>

- Description Reports differences between parameter sets. A macro can optionally be created that will convert file1 into file2.
- Arguments file1 and file2 are parameter files, like \$HOME/vnmrsys/exp1/procpar \$HOME/vnmrsys/exp1/curpar \$HOME/vnmrsys/global /vnmr/conpar xyz.fid/procpar file1 and file2 can also be directories (xyz.fid or xyz.par, or a local experiment like ~/vnmrsys/exp1); in this case diffparams will look for a subfile procpar in these directories. The optional -list argument will cause a list of the parameters which are different to be printed. If the *-list* option is used, the macro feature is turned off. If a parameter exists in file1 but not file2, it is not listed. If a parameter exists in file2 but not file1, it is listed. If the parameter exists in both files, it is listed if the values are different. It is not listed if other information associated with the parameter is different. This other information is things like protection bits, maximum values, group, type, etc. An optional third argument specifies the pathname of a macro to output. This macro will contain the MAGICAL commands necessary to

convert file1 into file2. Examples diffparams abc.fid xyz.fid diffparams -list abc.fid xyz.fid diffparams ~/vnmrsys/exp1 ~/vnmrsys/exp3 diffparams ~/vnmrsys/exp1 ~/vnmrsys/exp3

~/vnmrsys/maclib/change1to3

diffshims Compare two sets of shims (M,U)

Syntax	diffshims(shimfile1,shimfile2)	
	(From UNIX) diffshims shimfile1 shimfile2	
Description	Compares values for room-temperature shims stored in two separate files.	
Arguments	shimfile1 and shimfile2 are names of separate files containing shim values. Both files must have been written using the svs command.	
See also	NMR Spectroscopy User Guide	
Related	svs Save shim coil settings (C)	

digfilt Write digitally filtered FIDs to another experiment (M)

Syntax	digfilt(exp_number<,option>)		
Description	Saves digitally filtered FIDs to another experiment.		
Arguments	<code>exp_number</code> specifies the number of the experiment, from 1 to 9, for saving the FIDs.		
	option is one of the keywords 'nodc', 'zero', 'lfs', 'zfs', or 't2dc'. Use a keyword for an option if the same option was used when processing the data with ft, wft, ft2d, or wft2d.		
See also	NMR Spectroscopy User Guide		
Related	downsamp	Sampling factor applied after digital filtering (P)	
	ft	Fourier transform 1D data (C)	
	ft2d Fourier transform 2D data (C)		
	wft	Weight and Fourier transform 1D data (C)	
	wft2d	Weight and Fourier transform 2D data (C)	

dir List files in directory (C)

Syntax	dir<(string)>
Description	Displays files in a directory on the text window. The dir command is identical to the ls and lf commands.
Arguments	<pre>string is a string argument containing the options and/or directory names used if this were the UNIX 1s command (e.g., dir('-1 *.fid') requests a long listing (-1) of all files ending with .fid (*.fid)). If no argument is entered, dir lists all files in the current working directory.</pre>
Examples	dir dir('data') dir('-l *.fid')

See also	NMR Spectr	roscopy User Guide
Related	lf	List files in directory (C)
	ls	List files in directory (C)

display Display parameters and their attributes (C)

Syntax	display(parameter '*' '**'<,tree>)			
Description	Displays or tree.	Displays one or more parameters and their attributes from a parameter ree.		
Arguments	Three level	s of display are available: parameter, '*', and '**'.		
	• paramete: its attribut parameter	c is the name of a single parameter and the display is of es (e.g., display('a') displays the attributes of a in the (default) current tree).		
	• ' * ' is a ke a tree (e.g. and values	eyword to display the name and values of all parameters in , display('*','global') displays all parameter names in the global tree).		
	• ' * * ' is a tree (e.g., o all parame	keyword to display the attributes of all parameters in a display('**','processed') displays the attributes of ters in the processed tree).		
	tree is th 'processe to the cre	e type of parameter tree and can be 'global', 'current', ed', or 'systemglobal'. The default is 'current'. Refer ate command for more information on types of trees.		
Examples	display('a') display('*','global') display('**','processed')			
See also	User Programming			
Related	create destroy paramvi	Create new parameter in a parameter tree (C) Destroy a parameter (C) Edit a parameter and its attributes with the vi text		
	prune	Prune extra parameters from current tree (C)		

dla Display spin simulation parameter arrays (M)

Syntax	dla<('long')>
Description	Displays the parameters containing the line assignments for spin simulation iteration (matching simulated spectra to actual data). A clindex value of a calculated transition gives the index of the assigned measured line. The value is zero for unassigned transitions.
Arguments	'long' is a keyword to display the parameters containing the line assignments for spin simulation iteration (matching simulated spectra to actual data) and put the line assignments into the file spini.la. This option is most useful when the dla display is too large to display

	all the calculated transitions in the text window. The dlalong command operates the same as the dla('long') command.	
Examples	dla	
	dla('long'	
See also	NMR Spectroscopy User Guide	
Related	assign	Assign transitions to experimental lines (M)
	clindex	Index of experimental frequency of a transition (P)
	dga	Display parameters of spin simulation group (C)
	dlalong	Long display of spin simulation parameter arrays
		(C)

dlalong Long display of spin simulation parameter arrays (C)

Syntax	dlalong	
Description	Puts line assignments into the file spini.la in a more complete form, then displays this file in the text window. It is most useful when the dla display is too large to display all the calculated transitions in the text window. The dla('long') command operates the same as dlalong.	
See also	NMR Spectroscopy User Guide	
Related	dla Display spin simulation parameter arrays (M)	

dLC Display LC detector trace(s) in a horizontal format

Applicability	VnmrJ 3.1
See also	pLC
	dlcnmr
	pLCNMR

dlCNMR Displays all forms of LC-NMR data

Applicability	VnmrJ 3.1
Applicability Description	VnmrJ 3.1 This macro is executed with a button on the LC-NMR display pane (labeled spare). Displays on-flow and stopped-flow 1D LC-NMR data. With on-flow data, dconi is used to display the NMR data with the time-aligned LC detector trace(s) along the left side. In the stopped-flow mode, dLC displays the 1D NMR data for each stop code at a position that it is time-aligned with the relevant LC peak. If arguments are supplied, dLCNMR passes the supplied arguments to
	dconi and forces a contour plot display. With no arguments, or when activated by the "Display LC & NMR" button, the dconi display uses

the dconi parameter to determine the default display mode. The "Contour" check-box can be used to select the contour map (dpcon) display mode instead of the default color intensity map (dconi) display. Examples dLCNMR(<number of contours>,<contour spacing>) See also dLC pLC dLCNMR pLCNMR

dli Display list of integrals (C)

Description Displays a list of integrals at the integral reset points. The frequency units of the displayed list of integrals is controlled by the parameter axis. The reset points may be defined with the z command and these frequencies are stored in lifrq. The calculated amplitudes of the integral region are stored in liamp. The reset points are stored as hertz and are not referenced to rfl and rfp. The amplitudes are stored as the actual value; they are not scaled by ins or by insref. When the integral blanking mode is used (i.e., intmod='partial'), only the integrals corresponding to the displayed integral regions are listed.

The displayed integral value can be scaled with the setint macro. The integral is scaled by the parameters ins and insref.

See also NMR Spectroscopy User Guide

axis	Axis label for displays and plots (P)
CZ	Clear integral reset points (C)
dlni	Display list of normalized integrals (M)
ins	Integral normalization scale (P)
insref	Fourier number scaled value of an integral (P)
liamp	Amplitudes of integral reset points (P)
lifrq	Frequencies of integral reset points (P)
nli	Find integral values (C)
rfl	Reference peak position in directly detected dimension (P)
rfp	Reference peak frequency in directly detected dimension
	(P)
setint	Set value of an integral (M)
Z	Add integral reset point at cursor position (C)
	axis cz dlni ins insref liamp lifrq nli rfl rfp setint z

dlivast Produce text file and process wells (M)

Applicability VAST accessory. Syntax dlivast<(last)>

Description	Produces a text file containing the integral of the partial regions and processes the wells.		
Arguments	last is the number of the last well. The default is 96.		
See also	NMR Spectroscopy User Guide		
Related	combiplate View a color map for visual analysis of VAST microtiter plate (U)		
	combishow Display regions as red, green, and blue in CombiPlate window (M)		

dll Display listed line frequencies and intensities (C)

Syntax	dll<('pos'<,noise_mult>)><:number_lines,scale>		
Description	Displays a list of line frequencies and amplitudes that are above a threshold defined by th. Frequency units are defined by the parameter axis. The results of this calculation are stored in llfrq and llamp. The frequencies are stored as Hz and are not referenced to rfl and rfp. Amplitudes are stored as the actual data point value; they are not scaled by vs.		
Arguments	'pos' is a keyword to list only positive lines.		
	noise_mult is a numerical value that determines the number of noise peaks listed for broad, noisy peaks. The default value is 3. A smalle value results in more peaks, a larger value results in fewer peaks, and a value of 0.0 results in a line listing containing all peaks above the threshold th. Negative values of noise_mult are changed to 3.		
	number_lines is a return argument with the number of lines above the threshold.		
	scale is a return argument with a scaling factor for line amplitudes. This scaling factor accounts for vs and whether the lines are listed in absolute intensity mode or normalized mode.		
Examples	dll dll('pos') dll(2.5) dll:r1.sc		
See also	NMR Spectroscopy User Guide		
Related	axisAxis label for displays and plots (P)delsDelete spectra from T_1 or T_2 analysis (C)fpFind peak heights (C)get11Get frequency and intensity of a line (C)llampList of line amplitudes (P)llfrqList of line frequencies (P)nlPosition the cursor at the nearest line (C)nl1Find line frequencies and intensities (C)rf1Reference peak position in directly detected dimension (P)		
	rip Reference peak frequency in directly detected dimension (P)		

th	Threshold (P)
VS	Vertical scale (P)

dlni Display list of normalized integrals (M)

Description Displays integrals in a normalized format. The parameter ins represents the value of the sum of all the integrals. When the integral blanking mode is used (i.e., intmod='partial'), only the integrals corresponding to the displayed integral regions are listed and are used in the summation.

See also	NMR	Spectroscopy	User	Guide
----------	-----	--------------	------	-------

CZ	Clear integral reset points (C)
dli	Display list of integrals (C)
ins	Integral normalization scale (P)
nli	Find integral values (C)
Z	Add integral reset point at cursor position (C)

dlp Decoupler low-power control with class C amplifier (P)

Systems with a class C amplifier.			
dlp controls the decoupler power level for systems with a class C decoupler amplifier in the low-power mode, generally used for homonuclear decoupling. dlp specifies dB of attenuation of the decoupler, below a nominal 1 watt value. dlp is active only if $dhp='n'$.			
On systems with a decoupler linear amplifier, dlp is nonfunctional and dpwr controls decoupler power.			
0 to 39 (in dB of attenuation, 0 is maximum power).			
NMR Spectroscopy User Guide			
dhpDecoupler high-power control with class C amplifier (PdmDecoupler mode for first decoupler (P)dmfDecoupler modulation frequency for first decoupler (P)dpwrPower level for first decoupler with linear amplifier (P)			

dm Decoupler mode for first decoupler (P)

ApplicabilityVNMRS systemsDescriptionDetermines the state of first decoupler during different status periods
within a pulse sequence (refer to the manual User Programming for
a discussion of status periods). Pulse sequences may require one, two,
three, or more different decoupler states. The number of letters that
make up the dm parameter vary appropriately, with each letter

representing a status period (e.g., dm='yny' or dm='ns'). If the decoupler status is constant for the entire pulse sequence, it can be entered as a single letter (e.g., dm='n').

Values 'n', 'y', 'a', or 's' (or a combination of these values), where:

'n' specifies no decoupler rf.

'y' specifies the asynchronous mode. In this mode, the decoupler rf is gated on and modulation is started at a random places in the modulation sequence.

On the VNMRS system, the default asynchronous decoupling uses a "progressive offset" scheme. Other asynchronous schemes are also implemented on the VNMRS. They can be selected using an optional flag parameter "decasynctype". Create "decasynctype" as a flag parameter in the current tree and set the following:

decasynctype = 'p' selects the "progressive offset" scheme (default)

= 'b' selects the "bit reversed" scheme, and

= 'r' selects the random scheme.

'a' specifies the asynchronous mode, the same as 'y'.

's' specifies the synchronous mode in which the decoupler rf is gated on and modulation is started at the beginning of the modulation sequence.

See also NMR Spectroscopy User Guide

Related	dm2	Decoupler mode for second decoupler (P)
	dm3	Decoupler mode for third decoupler (P)
	dm4	Decoupler mode for fourth decoupler (P)
	dmf	Decoupler modulation frequency for first decoupler
		(P)
	dmm	Decoupler modulation mode for first decoupler (P)
	dn	Nucleus for first decoupler (P)
	decasynctype	Decoupler asynchronous mode (P)

dm2 Decoupler mode for second decoupler (P)

Applicability	Systems with a second decoupler.			
Description	Determines the state of second decoupler during different status periods within a pulse sequence. It functions analogously to dm.			
Values	Same as dm , except that if $dn2='$ (two single quotes with no space in between) and a second decoupler is present in the console, $dm2$ assumes a default value of 'n' when go is executed.			
See also	NMR Spectroscopy User Guide			
Related	dm Decoupler mode of first decoupler (P)			
	dmf2 Decoupler modulation frequency for second decoupler (P)			
	dmm2 Decoupler modulation mode for second decoupler (P)			
	dn2 Nucleus for second decoupler (P)			

dm3 Decoupler mode for third decoupler (P)

Applicability	Systems with a third decoupler.		
Description	Determines the state of third decoupler during different status periods within a pulse sequence. It functions analogously to dm .		
Values	Same as dm , except that if $dn3=$ '' (two single quotes with no space in between) and a third decoupler is present in the console, $dm3$ assumes a default value of 'n' when go is executed.		
See also	NMR Spectroscopy User Guide		
Related	d dm Decoupler mode of first decoupler (P) dmf3 Decoupler modulation frequency for third deco (P)		
	dmm3 dn3 decasynctype	Decoupler modulation mode for third decoupler (P) Nucleus for third decoupler (P) Select the type of decoupler asynchronous mode (P)	

dm4 Decoupler mode for fourth decoupler (P)

Applicability	Systems with a deuterium decoupler channel as the fourth decoupler		
Description	Determines the state of fourth decoupler during different status periods within a pulse sequence. It functions analogously to dm.		
Values	Same as dm , except that if $dn4=$ ' (two single quotes with no space in between) and a fourth decoupler is present in the console, $dm4$ assumes a default value of 'n' when go is executed.		
See also	NMR Spectroscopy User Guide		
Related	dm Decoupler mode of first decoupler (P)		
	dmf4	Decoupler modulation frequency for fourth decoupler (P)	
	dmm4	Decoupler modulation mode for fourth decoupler (P)	
	dn4	Nucleus for fourth decoupler (P)	
	decasynctype	Select the type of decoupler asynchronous mode (P)	

dmf Decoupler modulation frequency for first decoupler (P)

Description Controls modulation frequency of the first decoupler. It specifies 1/pw90 at the particular power level used. After calibrating the decoupler field strength γH_2 (expressed in units of Hz), dmf should be set equal to $4*\gamma H_2$ for WALTZ, MLEV16, GARP, and XY32 (when available). dmf is inactive for CW mode decoupling (dmm='c'). dmf is also active for square wave mode decoupling (dmm='r') and fm-fm mode (dmm='f') decoupling. For dmm='f', the modulation frequency is swept back and forth between about 0.5% and 5% of the dmf frequency (e.g., if dmf is 100 kHz, the modulation is swept between approximately 500 Hz and 5 kHz). A reasonable optimum value for dmf when dmm='f' is the decoupler frequency divided by 4000.

Values 5 Hz to 2 MHz in steps of 5 Hz (steps are actually approximately 4.768 Hz).

For GARP modulation, the dmf value is internally multiplied by 45, making the limit of possible dmf values to 5 Hz to 44.4 kHz when dmm='g'.

- See also NMR Spectroscopy User Guide
 - Relateddmf2Decoupler modulation frequency for second decoupler (P)dmf3Decoupler modulation frequency for third decoupler (P)dmf4Decoupler modulation frequency for fourth decoupler (P)dmmDecoupler modulation mode for first decoupler (P)pw9090° pulse width (P)

dmf2 Decoupler modulation frequency for second decoupler (P)

Applicability	Systems with a second decoupler.		
Description	Controls the modulation frequency of the second decoupler. It functions analogously to the parameter dmf .		
Values	Same as dmf except that if $dn2='$ ' (two single quotes with no space in between) and a second decoupler is present in the console (numrfch greater than 2), $dmf2$ assumes a default value of 1000 Hz when go is executed.		
See also	NMR Spectroscopy User Guide		
Related	dm2	Decoupler mode for second channel (P)	
	dmf	Decoupler modulation frequency for first decoupler (P)	
	dmm2	Decoupler modulation mode for second decoupler (P)	
	dn2	Nucleus for second decoupler (P)	
	numrfch	Number of rf channels (P)	

dmf3 Decoupler modulation frequency for third decoupler (P)

Applicability	Systems with a third decoupler.
Description	Controls the modulation frequency of the third decoupler. It functions analogously to the parameter dmf.
Values	Same as dmf except that if $dn3='$ (two single quotes with no space in between) and a third decoupler is present in the console (numrfch equals 4), $dmf3$ assumes a default value of 1000 Hz when go is executed.

See also	NMR Spect	roscopy User Guide
Related	dm3	Decoupler mode for third channel (P)
	dmf	Decoupler modulation frequency for first decoupler (P)
	dmm3	Decoupler modulation mode for third decoupler (P)
	dn3	Nucleus for third decoupler (P)
	numrfch	Number of rf channels (P)

dmf4 Decoupler modulation frequency for fourth decoupler (P)

Systems with a deuterium decoupler channel as the fourth decoupler.		
Controls the modulation frequency of the fourth decoupler. It functions analogously to the parameter dmf .		
Same as dmf except that if $dn4=$ ' ' (two single quotes with no space in between) and a fourth decoupler is present in the console (numrfch equals 5), $dmf4$ assumes a default value of 1000 Hz when go is executed.		
NMR Spectroscopy User Guide		
dm4	Decoupler mode for fourth channel (P)	
dmf	Decoupler modulation frequency for first decoupler (P)	
dmm4	Decoupler modulation mode for fourth decoupler (P)	
dn4	Nucleus for fourth decoupler (P)	
numrfch	Number of rf channels (P)	
	Systems with Controls the r analogously t Same as dmf in between) a equals 5), dm executed. <i>NMR Spectroo</i> dm4 dmf dm4 dm4 numrfch	

dmfadj Adjusts the parameter 'dmf'

```
Syntax dmfadj(<tipangle_resoln>)
```

Applicability VnmrJ 3.1

Description `dmfadj` adjusts the parameter 'dmf' so that the time associated with the tip-angle resolution is an integral multiple of 100 ns. This insures that there is no truncation error in time in the execution of the programmable decoupling or spin-locking sequence by the waveform generator. The optional argument 'tipangle_resoln' specifies the necessary tip-angle resolution for the programmable decoupling or spin-locking sequence which is to be executed. For example, the tip-angle resolution for an MLEV-16 decoupling sequence should be 90.0 degrees since every pulse in that sequence can be represented as an integral multiple of 90.0 degrees; the tip-angle resolution for a GARP decoupling sequence, however, should be 1.0 degrees. Arguments If the argument 'tipangle_resoln' is not specified when the macro `dmfadj` is called, the default value therefore is taken from the parameter 'dres'.

Related	dmf2adj	adjusts the parameter 'dmf2
	pwsadj	adjusts 'pulse_parameter'

Applicability	Systems with a second decoupler.		
Syntax	dmf2adj<(tipangle_resolution)>		
Description	Adjusts the parameter dmf2 to make time associated with the second decoupler tip-angle resolution an integral multiple of 50 ns. dmf2adj functions analogously to the macro dmfadj.		
Arguments	tipangle_resolution specifies the necessary tip-angle resolution for the programmable decoupling or spin-locking sequence to be executed. The default value is the current value of the parameter dres2.		
Examples	dmf2adj dmf2adj(90.0)		
See also	NMR Spectroscopy User Guide		
Related	dmf2	Decoupler modulation frequency for second decoupler (P)	
	dmfadj dres2	Adjust decoupler tip-angle resolution time (M) Tip angle resolution for second decoupler (P)	

dmf3adj Adjust tip-angle resolution time for third decoupler (M)

Applicability	Systems with a third decoupler.		
Syntax	dmf3adj<(tipangle_resolution)>		
Description	Adjusts the parameter $dmf3$ to make time associated with the third decoupler tip-angle resolution an integral multiple of 50 ns. $dmf3ad3$ functions analogously to the macro $dmfad3$.		
Arguments	tipangle_resolution specifies the necessary tip-angle resolution for the programmable decoupling or spin-locking sequence to be executed. The default value is the current value of the parameter dres3.		
Examples	dmf3adj dmf3adj(90.0)		
See also	NMR Spectroscopy User Guide		
Related	dmf3Decoupler modulation frequency for third decoupler (P)dres3Tip-angle resolution for third decoupler (P)		

dmf4adj Adjust tip-angle resolution time for fourth decoupler (M)

Applicability Systems with a deuterium decoupler as the fourth decoupler. Syntax dmf4adj<(tipangle_resolution)>

Description	Adjusts the parameter $dmf4$ to make time associated with the four decoupler tip-angle resolution an integral multiple of 50 ns. $dmf4a$ functions analogously to the macro $dmfadj$.	
Arguments	tipangle_resolution specifies the necessary tip-angle resolution for the programmable decoupling or spin-locking sequence to be executed. The default value is the current value of the parameter dres4.	
Examples	dmf4adj	
See also	NMR Spectroscopy User Guide	
Related	dmf4Decoupler modulation frequency for fourth decoupler (P)dres4Tip-angle resolution for fourth decoupler (P)	

dmg Data display mode in directly detected dimension (P)

Description Controls the mode of data display along the directly detected dimension. dmg is in the display group and can be set manually or by executing the commands ph, av, pwr, or pa for the values 'ph', 'av', 'pwr', or 'pa', respectively.

Values 'ph' sets the *phased mode* in which each real point in the displayed spectrum is calculated from a linear combination of real and imaginary points comprising each respective complex data point.

'av' sets the *absolute-value mode* in which each real point in the displayed spectrum is calculated as the square root of the sum of squares of the real and imaginary points comprising each respective complex data point.

'pwr' sets the *power mode* in which each real point in the displayed spectrum is calculated as the sum of squares of the real and imaginary points comprising each respective complex data point.

'pa' sets the *phase angle* mode in which each real point in the displayed spectrum is calculated as the phase angle from the arc tangent of the real and imaginary points comprising each respective complex data point.

See also NMR Spectroscopy User Guide

Related	aig	Absolute intensity group (P)
	av	Set absolute-value mode in directly detected dimension (C)
	dcg	Drift correction group (P)
	dmg1	Data display mode in 1st indirectly detected dimension (P)
	dmg2	Data display mode in 2nd indirectly detected dimension
		(P)
	ft	Fourier transform 1D data (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ft2d	Fourier transform 2D data (C)
	pa	Set phase angle mode in directly detected dimension (C)
	ph	Set phased mode in directly detected dimension (C)
	pmode	Processing mode for 2D data (P)

pwrSet power mode in directly detected dimension (C)wftWeigh and Fourier transform 1D data (C)wft1dWeigh and Fourier transform of 2D data (C)wft2dWeigh and Fourier transform 2D data (C)

dmg1 Data display mode in 1st indirectly detected dimension (P)

- Description Controls the mode of data display along the first indirectly detected dimension of a multidimensional data set. dmg1 is in the display group and can be set manually or by executing the commands ph1, av1, pwr1, or pa1 for the values 'ph1', 'av1', 'pwr1', or 'pa1', respectively. If dmg1 does not exist or if it is set to the empty string (dmg1=''), VnmrJ uses the value of dmg to decide the display mode along the first indirectly detected dimension.
 - Values 'ph1' sets phased mode.
 - 'av1' sets absolute-value mode.
 - 'pwr1' sets power mode.
 - 'pa1' sets phase angle mode.

See also NMR Spectroscopy User Guide

Relatedav1Set absolute-value mode in 1st indirectly det. dim. (C)dmgData display mode in directly detected dimension (P)pa1Set phase angle mode in 1st indirectly detected dimension (C)ph1Set phased mode in 1st indirectly detected dimension (C)pwr1Set power mode in 1st indirectly detected dimension (C)

dmg2 Data display mode in 2nd indirectly detected dimension (P)

Description Controls the mode of data display along the second indirectly detected dimension of a multidimensional data set. dmg2 is in the display group and can be set manually or by executing the commands ph2, av2, or pwr2 for the values 'ph2', 'av2', or 'pwr2', respectively. If dmg2 does not exist or if it is set to the empty string (dmg2=''), VnmrJ uses the value of the parameter dmg instead of dmg2 to decide the display mode along the second indirectly detected dimension.

Values 'ph2' sets phased mode.

'av2' sets absolute-value mode.

'pwr2' sets power mode.

See also	NMR Spectro	scopy User Guide
Related	av2	Set absolute-value mode in 2nd indirectly det. dim. (C)
	dmg	Data display mode in directly detected dimension (P)
	ph2	Set phased mode in 2nd indirectly det. dim. (C)
	pwr2	Set power mode in 2nd indirectly det. dim. (C)

dmgf Absolute-value display of FID data or spectrum in acqi (P)

Description	If the parameter dmgf exists and is set to 'av', the FID display in				
	the acqi program is set to the absolute-value mode, which displays				
	the square root of the sum of the squares of the real and imaginary				
	channels. dmgf has no function outside of the acqi program. This				
	display mode may cause the displayed FID to exceed the displayed ADC				
	limits in acqi by as much as a factor of the square root of 2.				
See also	NMR Spectroscopy User Guide				
Related;	acqi Interactive acquisition display process (C)				
	av Set absolute-value mode in directly detected dimension (C)				
	gf Prepare parameters for FID/spectrum display in acqi (M)				

dmm Decoupler modulation mode for first decoupler (P)

Description Sets the modulation modes for the first decoupler. In the standard two-pulse sequence, dmm typically has a single state because the decoupler modulation is normally not changed during the pulse sequence, but this is not fixed. For example, dmm='ccw' gives single-frequency CW decoupling during the first part of the sequence and WALTZ-16 decoupling during acquisition.

In pulse sequences using the decoupler for pulsing (INEPT, DEPT, HETCOR, etc.), decoupler modulation must be set to 'c' during periods of the pulse sequence when the decoupler is to be pulsed.

- Values 'c', 'f', 'g', 'm', 'p', 'r', 'u', 'w', and 'x' are available;:
 - 'c' sets continuous wave (CW) modulation.
 - 'f' sets fm-fm modulation (swept-square wave).
 - 'g' sets GARP modulation.
 - 'm' sets MLEV-16 modulation.
 - 'n' sets noise modulation.
 - 'p' sets programmable pulse modulation using the dseq parameter to specify the decoupling sequence.
 - 'r' sets square-wave modulation.
 - 'u' sets user-supplied modulation using external hardware.

- 'w' sets WALTZ-16 modulation.
- 'x' sets XY32 modulation.

See also NMR Spectroscopy User Guide

Related	dm	Decoupler mode for first decoupler (P)	
	dmf	Decoupler modulation frequency for first decoupler (P)	
	dmm2	Decoupler modulation mode for second decoupler (P)	
	dmm3	Decoupler modulation mode for third decoupler (P)	
	dmm4	Decoupler modulation mode for fourth decoupler (P)	
	dseq	Decoupler sequence for the first decoupler (P)	

dmm2 Decoupler modulation mode for second decoupler (P)

Applicability	Systems with a second decoupler.		
Description	Sets the type of decoupler modulation for the second decoupler during different status periods within a pulse sequence. It functions analogously to dmm.		
Values	'c', 'f', 'g', 'm', 'p', 'r', 'u', 'w', and 'x' are available. Refer to dmm for the definition of these values (note that if the mode 'p' is selected, dseq2 specifies the decoupling sequence). If dn2='' (two single quotes) and a second decoupler is present in the console (numrfch greater than 2), dmm2 is internally set to 'c' when go is executed.		
See also	NMR Spect	roscopy User Guide	
Related	dm2 dmf2	Decoupler modulation for the second decoupler (P) Decoupler modulation frequency for the second decoupler (P)	
	dmm	Decoupler modulation mode for first decoupler (P)	
	dn2	Nucleus for the second decoupler (P)	
	dseq2	Decoupler sequence for the second decoupler (P)	
	numrfch	Number of rf channels (P)	

dmm3 Decoupler modulation mode for third decoupler (P)

Applicability	Systems with a third decoupler.
Description	Sets type of decoupler modulation for the third decoupler during different status periods within a pulse sequence. It functions analogously to dmm.
Values	'c', 'f', 'g', 'm', 'p', 'r', 'u', 'w', and 'x' are available. Refer to dmm for the definition of these values (note that if the mode 'p' is selected, dseq3 specifies the decoupling sequence). If dn3='' (two single quotes) and a third decoupler is present in the console (numrfch equal to 4), dmm3 is internally set to 'c' when go is executed.

See also	NMR Spectro	pscopy User Guide
Related	dm3	Decoupler modulation for third decoupler (P)
	dmf3	Decoupler modulation frequency for third decoupler (P)
	dmm	Decoupler modulation mode for first decoupler (P)
	dn3	Nucleus for the third decoupler (P)
	dseq3	Decoupler sequence for the third decoupler (P)
	numrfch	Number of rf channels (P)

S

Decoupler modulation mode for fourth decoupler (P) dmm4

Applicability	Systems with a deuterium decoupler channel as the fourth decoupler.		
Description	Sets type of decoupler modulation for the fourth decoupler during different status periods within a pulse sequence. It functions analogously to dmm.		
Values	'c', 'f', 'g', 'm', 'r', 'u', 'w', and 'x' are available. Refer to dmm for the definition of these values. If $dn4=$ '' (two single quotes) and a fourth decoupler is present in the console (numrfch greater than 4), dmm4 is internally set to 'c' when go is executed.		
See also	NMR Spectroscopy User Guide		
Related	dm4Decoupler modulation for the fourth decoupler (P)dmf4Decoupler modulation frequency for the fourth decoupler (P)dmmDecoupler modulation mode for first decoupler (P)dm4Nucleus for the fourth decoupler (P)dseq4Decoupler sequence for the fourth decoupler (P)numrfchNumber of rf channels (P)		

Nucleus for first decoupler (P) dn

Description	Changing the value of dn causes a macro (named _dn) to be executed that extracts values for dfrq and dof from lookup tables. The tables, stored in the directory /vnmr/nuctables, are coded by atomic weights.		
Values	In the lookup tables, typically 'H1', 'c13', 'P31', etc.		
See also	NMR Spectroscopy User Guide		
Related	dfrq Transmitter frequency of first decoupler (P)		
	dn2	Nucleus for second decoupler (P)	
	dn3Nucleus for third decoupler (P)dn4Nucleus for fourth decoupler (P)dofFrequency offset for first decoupler (C)tnNucleus for observe transmitter (P)		

dn2 Nucleus for second decoupler (P)

Applicability Systems with a second decoupler.

- Description Changing the value of dn2 causes a macro (named _dn2) to be executed that extracts values for dfrq2 and dof2 from lookup tables. Otherwise, dn2 functions analogously to the parameters tn and dn. If an experiment does not use the second decoupler channel, the channel can be disabled by setting dn2='' (two single quotes with no space in between). This sets dm2='n', dmm2='c', dmf2=1000 (in Hz), dfrq2=1 (in MHz), dof2=0, dpwr2=0, dseq2='', and dres2=1.
 - See also NMR Spectroscopy User Guide

Related	dfrq2	Transmitter frequency of second decoupler (P)
	dn	Nucleus for first decoupler (P)
	dof2	Frequency offset for second decoupler (C)
	numrfch	Number of rf channels (P)
	tn	Nucleus for observe transmitter (P)
	tn	Nucleus for observe transmitter (P)

dn3 Nucleus for third decoupler (P)

Applicability	Systems with a third decoupler.		
Description	Changing the value of dn3 causes a macro (named _dn3) to be executed that extracts values for dfrq3 and dof3 from lookup tables. Otherwise, dn3 functions analogously to the parameters tn and dn. If an experiment does not use the third decoupler channel, the channel can be disabled by setting $dn3=''$ (two single quotes with no space in between). This sets $dm3='n'$, $dmm3='c'$, $dmf3=1000$ (in Hz), dfrq3=1 (in MHz), $dof3=0$, $dpwr3=0$, $dseq3=''$, and $dres3=1$.		
See also	NMR Spectroscopy User Guide		
Related	dn Nucleus for first decoupler (P)		
	dfrq3	Transmitter frequency of third decoupler (P)	
	dof3	Frequency offset for third decoupler (C)	
	numrfch	Number of rf channels (P)	
	tn Nucleus for observe transmitter (P)		

dn4 Nucleus for fourth decoupler (P)

Applicability Systems with a deuterium decoupler channel as the fourth decoupler. Description Changing the value of dn4 causes a macro (named _dn4) to be executed that extracts values for dfrq4 and dof4 from lookup tables. Otherwise, dn4 functions analogously to the parameters tn and dn except that the only valid value for dn4 is 'H2'. If an experiment does not use the fourth decoupler channel, the channel can be disabled by setting dn4='' (two single quotes with no space in between). This sets

	dm4='n', dmm4='c', dmf4=1000 (in Hz), dfrq4=1 (in MHz), dof4=0.			
	dpwr4=0, dseq4='', and dres4=1.			
See also	NMR Spectro	scopy User Guide		
Related	dfrq4 Transmitter frequency of fourth decoupler (P)			
	dn	Nucleus for first decoupler (P)		
	dof4	Frequency offset for fourth decoupler (C)		
	numrfch	Number of rf channels (P)		
	tn	Nucleus for observe transmitter (P)		

dndfid Retrieve and process fid data from the locator (M)

Applicability	Liquids, Imaging,	Solids
Description	Retrieve fid data processed if Proc the System settir	from an item selected in the locator. Data is also ess data on drag-and-drop from locator is selected in ags dialog in the Utilities menu.
Related	dndjoin	Join a work space from the locator (M)
	dndpar	Retrieve a parameter set from the locator (M)
	dndshims	Retrieve a shimset set from the locator (M)
	locaction	Locator action (M)
	locprotoexec	Execute a protocol from the locator (M)
	xmmakenode	Make a new study queue node (M)

dndjoin Join a work space from the locator (M)

Description	Join the work sp	ace selected by the locator.
Related	dndfid	Retrieve and process fid data from the locator (M)
	dndpar	Retrieve a parameter set from the locator (M)
	dndshims	Retrieve a shimset set from the locator (M)
	locaction	Locator action (M)
	locprotoexec	Execute a protocol from the locator (M)
	xmmakenode	Make a new study queue node (M)

dndpar Retrieve a parameter set from the locator (M)

Retrieve a param	eter set selected by the locator.
dndfid	Retrieve and process fid data from the locator (M)
dndjoin	Join a work space from the locator (M)
dndshims	Retrieve a shimset set from the locator (M)
locaction	Locator action (M)
	Retrieve a param dndfid dndjoin dndshims locaction

locprotoexec Execute a protocol from the locator (M) xmmakenode Make a new study queue node (M)

dndshims Retrieve a shimset set from the locator (M)

Description	Retrieve a shimse	et set selected by the locator.
Related	dndfid	Retrieve and process fid data from the locator (M)
	dndjoin	Join a work space from the locator (M)
	dndpar	Retrieve a parameter set from the locator (M)
	locaction	Locator action (M)
	locprotoexec	Execute a protocol from the locator (M)
	xmmakenode	Make a new study queue node (M)

doautodialogStart a dialog window using def file (M)

Applicability	Systems with automation.
Syntax	doautodialog
Description	Internal macro used by <u>enter</u> to start a dialog window using the def file for an experiment in the dialoglib directory.
Related	enter Enter sample information for automation run (M,U)

dodialog Start a dialog window with dialoglib file (M)

Syntax	dodialog
Description	Internal macro that starts a dialog window using a dialog file in the dialoglib directory.

dof Frequency offset for first decoupler (P)

Description Controls the frequency offset of the first decoupler. Higher numbers move the decoupler to higher frequency (toward the left side of the spectrum). The frequency accuracy of the decoupler offset is generally0.1 Hz. The value is specified in the config program.
Values -100000 to 100000 Hz (approximate, depends on frequency), in steps of 0.1 Hz.

See also	NMR Spectroscopy User Guide	
Related	config	Display current configuration and possible change it (M)
	dof2	Frequency offset for second decoupler (P)
	dof3	Frequency offset for third decoupler (P)
	dof4	Frequency offset for fourth decoupler (P)
	tof	Frequency offset for observe transmitter (P)

dof2 Frequency offset for second decoupler (P)

Applicability	Systems with a second decoupler.	
Description	Controls the fanalogously to	requency offset for the second decoupler. dof2 functions to the parameters tof and dof.
Values	-100000 to 100000 Hz (approximate, depends on frequency), in steps of 0.1 Hz. If $dn2=''$ (two single quotes with no space in between) and a second decoupler channel is present in the console, dof2 assumes a default value of 0 when go is executed.	
See also	NMR Spectroscopy User Guide	
Related	dn2	Nucleus for second decoupler (P)
	dof	Frequency offset for first decoupler (P)
	tof	Frequency offset for observe transmitter (P)

dof3 Frequency offset for third decoupler (P)

Applicability	Systems with a third decoupler.	
Description	Controls the frequency offset for the third decoupler. dof3 functions analogously to the parameters tof and dof.	
Values	-100000 to 100000 Hz (approximate, depends on frequency), in steps of 0.1 Hz. If $dn3=''$ (two single quotes with no space in between) and a third decoupler channel is present in the console, dof3 assumes a default value of 0 when go is executed.	
See also	NMR Spectroscopy User Guide	
Related	dn3 Nucleus for third decoupler (P)	
	dof Frequency offset for first decoupler (P)	
	tof Frequency offset for observe transmitter (P)	

doff Frequency offset for fourth decoupler (P)

Applicability Systems with a deuterium decoupler channel as the fourth decoupler.

Description	Controls the frequency offset for the fourth decoupler. $dof4$ function analogously to the parameters tof and dof.	
Values	-100000 to 100000 Hz (approximate, depends on frequency), in steps of 2.384 Hz. If $dn4=$ '' (two single quotes with no space in between) and a fourth decoupler channel is present in the console, dof4 assumes a default value of 0 when go is executed.	
See also	NMR Spectroscopy User Guide	
Related	dn4 Nucleus for fourth decoupler (P)	
	dof Frequency offset for first decoupler (P)	
	tof Frequency offset for observe transmitter (P)	

doneshot Set up parameters for Doneshot pulse sequence (M)

Converts a parameter set to Doneshot experiment.	
NMR Spectroscopy User Guide	
dosy	Process DOSY experiments (M)
fiddle	Perform reference deconvolution (M)
setup_dosy	Set up gradient levels for DOSY experiments (M)
	Converts a pa NMR Spectro dosy fiddle setup_dosy

dopardialog Start a dialog with dialoglib/experiment def file (M)

Description Internal macro that starts a dialog window using a def file in the directory dialoglib/experiment.

do_pcss Calculate proton chemical shifts spectrum (C)

Syntax	do_pcss<(<threshold><,max_cc><,max_width)></threshold>
Description	Strips a high-resolution proton spectrum down to a list of chemical shifts. The list is saved in the file pcss.outpar. If no argument is given, do_pcss automatically calculates the threshold and uses default values for the maximum allowable coupling constant and the maximum width of a spin multiplet.
Arguments	threshold sets the level whether a point belongs to a peak or is noise.
	${\tt max_cc}$ is the maximum allowable coupling constant in the spectrum. Default is 20 Hz.
	${\tt max_width}$ is the maximum width of a spin multiplet in the spectrum. Default is 60 Hz.
Examples	do_pcss do_pcss(10) do_pcss(9,20,80)

See also NMR Spectroscopy User Guide

Related pcss Calculate and show proton chemical shifts spectrum (M)

dosy Process DOSY experiments (M)

Syntax	<pre>dosy(<'prune'>,<lowerlimit,upperlimit>)</lowerlimit,upperlimit></pre>	
Description	Performs a DOSY (diffusion ordered spectroscopy) analysis of the data in an array of spectra.	
	dosy uses the commands dll and fp to determine the heights of all signals above the threshold defined by the parameter th and then fits the decay curve for each signal to a Gaussian using the program dosyfit. It stores a summary of all diffusion coefficients and their estimated standard errors and various other results as follows:	
	• In the directory \$HOME/vnmrsys/Dosy: diffusion_display.inp, general_dosy_stats, calibrated_gradients, fit_errors, and diffusion_spectrum	
	• In the current experiment: a second copy of diffusion_display.inp.	
	The command showdosy has been incorporated into dosy.	
Arguments	prune starts a dialog to allow one or more spectra to be omitted from	
	the analysis. lowerlimit is the lower diffusion limit (in units of 10^{-10} m ² /s) to be displayed	
	upperlimit is the upper diffusion limit (in units of 10^{-10} m ² /s) to be displayed.	
	Without arguments, $dosy$ uses all the experimental spectra and covers the whole diffusion range seen in the experimental peaks.	
See also	NMR Spectroscopy User Guide	
Related	ddifSynthesize and display DOSY plot (C)fiddlePerform reference deconvolution (M)setup_dosySet up gradient levels for DOSY experiments (M)	

dosy2d Apptype macro for dosy 2D experiments (M)

Applicability	Liquids	
Description	Performs the action experiments. It is	ons for 2D dosy protocols to set up, process, and plot a only available if the Dosy software is installed.
Related	apptype execpars	Application type (PM) Set up the exec parameters (M)

dosy3Dflag Used by the dosy macro to determine whether to use 2D or 3D DOSY processing

Syntax	dosy3Dflag
Applicability	VnmrJ 3.1
Description	dosy3Dflag is a parameter used by the dosy macro to determine whether to use 2D or 3D processing. It is normally set automatically, but can also be set manually, e.g. to force 2D processing of one increment of a 3D dataset.
Arguments	dosy3Dflag='y'
See also	dosy3D11ag='n'
Dec aibu	

dosy3Dproc Used by the dosy macro to determine whether to use 2D or 3D processing

Syntax	dosy3Dproc	
Applicability	VnmrJ 3.1	
Description	dosy3Dproc is a parameter used by the dosy macro to determine whether to use 2D or 3D processing, and what type of the latter. It is normally set automatically, but can also be set manually, e.g. to force 2D processing of an increment extracted from a 3D dataset.	
Arguments	dosy3Dproc='n'	
	dosy3Dproc='ntype'	
	dosy3Dproc='ptype'	
	dosy3Dproc='y'	
See also	dosy	

dosybypointsDetermines whether peak picking is used by the dosy macro

Syntax	dosybypoints
Applicability	VnmrJ 3.1
Description	Determines whether dosy produces a 2D display based on whole peaks (the default) or point by point (much slower) in the spectral dimension.
Arguments	'n' divides the spectrum into individual peaks, creating one cross-peak for each individual peak found in the 1D spectrum. 'y' performs a diffusion fit for every point in the displayed region of the spectrum that lies above the threshold th.
See also	ddif
	dosy

dosyfit fits 2D or 3D DOSY data to obtain diffusion coefficients, amplitudes and statistics

Syntax	dosyfit
	dosyfit('version')
	dosyfit('3D')
	dosyfit('3D', avgnoise)
Applicability	VnmrJ 3.1
Description	dosyfit performs monoexponential least squares fitting on signal intensities from 2D and 3D datasets, summarising the results in various files.
Arguments	dosyfit takes 0, 1, or 2 arguments: 'version' returns the version number of the software, '3D' invokes processing of cross-peak volumes stored in the files peaks.bin. <n> rather than peak heights stored in the file dosy_in. In the case of 3D processing, the parameter avgnoise allows correction for the average baseplane noise in absolute value data</n>
See also	ddif
	dosy

dosyfrg Larmor frequency of phase encoded nucleus in DOSY (P)

Description	Stores the NMR frequency of the phase encoded nucleus in DOSY experiments. It is directly set by the DOSY sequences.	
See also	NMR Spectroscopy User Guide	
Related	dosy	Process DOSY experiments (M)

dosygamma Gyromagnetic constant of phase encoded nucleus in DOSY (P)

Description	Stores the gyromagnetic constant of the phase encoded nucleus in	
	DOSY experiments. It is automatically set by the DOSY sequences and	
	used by the dosy macro.	
See also	NMR Spectroscopy User Guide	
Related	dosy Process DOSY experiments (M)	

dosypeaks Determines whether peak picking is used by the dosy macro

Syntax dosypeaks

Applicability	VnmrJ 3.1
Description	Determines whether dosy produces a 2D display based on whole peaks (the default) or point by point (much slower) in the spectral dimension.
Arguments	'y' divides the spectrum into individual peaks, creating one cross-peak for each individual peak found in the 1D spectrum. 'n' performs a diffusion fit for every point in the displayed region of the spectrum that lies above the threshold th.
See also	ddif dosy

dosyproc Determines the type of processing performed by the dosy macro

Syntax	dosyproc
Applicability	VnmrJ 3.1
Description	Determines whether $dosy$ produces a discrete or a continuous diffusion spectrum.
Arguments	'discrete' invokes monoexponential fitting with dosyfit if ncomp=1, and multiexponential fitting with the external programme SPLMOD if ncomp>1. 'continuous' invokes processing with the external programme CONTIN and gives a continuous distribution in the diffusion domain.
See also	dosy
	For information about the programmes SPLMOD and CONTIN please visit http://s-provencher.com/index.shtml.

dosytimecubedGyromagnetic constant of phase encoded nucleus in DOSY (P)

Description	n Time cubed factor in the expression for diffusional attenua		
	automatically	r set by the DOSY sequences and used by the dosy macro.	
See also	NMR Spectroscopy User Guide		
Related	dosy	Process DOSY experiments (M)	

dot1 Set up a T₁ experiment (M)

Syntaxdot1<(min_T1_estimate, max_T1_estimate, time)>DescriptionSets up all parameters to perform a T_1 experiment, including d1, pw,
p1, nt, and an array of d2 values, based on information entered you
enter. Make sure that the parameter pw90 is set properly and contains
the correctly calibrated 90° pulse width because dot1 uses this

information. If you have not done a pulse width calibration recently, you may wish to do so now.

Minimum and maximum T_1 for the peaks of interest are estimates. Do the best you can. Your estimates are used to select optimum values of d2. If the T_1 does not fall between your two guesses, your experiment may not be optimum, but it should still be usable unless your estimates are extremely far off. When you are satisfied with the parameters, enter ga or au to acquire the data.

Arguments min_T1_estimate is the estimated minimum expected T_1 . The default is the system prompts the user for the value.

max_T1_estimate is the estimated maximum expected T_1 . The default is the system prompts the user for the value.

time is the total time in hours that the experiment should take. The default is the system prompts the user for the value.

Examples dot1

dot1(1,2,.5)

See also NMR Spectroscopy User Guide

on (P)
(C)

dotflag Display FID as connected dots (P)

Description When sparse FID data points are displayed, they are displayed as unconnected dots. If dotflag exists and is set to 'n', the FID dots will be connected. To create dotflag, enter create ('dotflag', 'flag'). To create dotflag and the FID display parameters axisf, vpf, vpfi, crf, and deltaf (if the parameter set is older and lacks these parameters), enter addpar('fid'). Values 'n' sets connecting the dots. 'y' sets not connecting the dots. See also NMR Spectroscopy User Guide Related addpar Add selected parameters to the current experiment (M) create Create new parameter in a parameter tree (C) df Display a single FID (C)

dousermacro Mechanism to provide customization to VnmrJ operations

Syntax dousermacro('rootName' <,args>)

Applicability VnmrJ 3.1

Description Certain VnmrJ operations have software hooks to allow for easy user customization. For example, the svf operation will call a macro named usersvf, if it exists. That usersvf macro could copy additional files into the .fid directory, write a log file, or email a message. It is up to the user to decide how they may want to customize the operation.

The mechanism we use to provide this customization is dousermacro. This macro is often called with the syntax dousermacro(\$0) where \$0 is the name of the macro being executed (svf in the example above.) The dousermacro prepends the string 'user' to the first passed argument and then checks if that macro exists. If it does, it is executed. If any additional arguments are passed to dousermacro, these are passed along to the 'user'+rootName macro.

Some of the operations that have these dousermacro hooks include:

- bootup
- calibrate
- operatorlogin
- operatorlogout
- plot
- process
- rt
- rtp
- savefid
- svf
- updateprobe

Creating a local macro named, for example, userplot will allow customization any time the plot macro is called. There are several other macros that call dousermacro. They generally require a fairly good understanding of how these other macros are used in order to effectively use the dousermacro tool. You can find all the macros that call dousermacro by executing: grep dousermacro /vnmr/maclib/* from a shell tool.

downsamp Downsampling factor applied after digital filtering (P)

Description Specifies the downsampling factor applied after digital filtering. The spectral width of the data set after digital filtering and downsampling is sw divided by downsamp, where sw is the acquired spectral width. If downsamp does not exist in the current experiment, enter addpar('downsamp') to add it. addpar('downsamp') creates the digital filtering and downsampling parameters downsamp, dscoef, dsfb, dslsfrg, and filtfile.

Values Number for the downsampling factor. 1 sets digital filtering with a filter bandwidth specified by dsfb without downsampling.

	'n' sets normal data processing without digital filtering.	
See also	NMR Spectroscopy User Guide	
Related	addpar	Add selected parameters to current experiment (M)
	digfilt	Write digitally filtered FID to another experiment (M)
	dscoef	Digital filter coefficients for downsampling (P)
	dsfb	Digital filter bandwidth for downsampling (P)
	dslsfrq	Bandpass filter offset for downsampling (P)
	filtfile	File of FIR digital filter coefficients (P)
	pards	Create additional parameters used by downsampling
		(M)
	SW	Spectral width in directly detected dimension (P)

Double precision (P)

đp

Description	Sets whether data are acquired in a 16-bit or 32-bit integer format.		
Values	'n' sets 16-bit format, 'y' sets 32-bit format. If the 200-kHz receiver option is installed (Max. Narrowband Width set to 200 kHz in the Spectrometer Configuration window), dp is forced to 'n' if $120000 < sw < = 200000$. If $sw > 200000$, dp is forced to 'y'. On wideline systems dp='y' is required when $sw > 100000$		
See also	NMR Spectroscopy User Guide		
Related	sw Spectral width in directly detected dimension (P)		

dpcon Display plotted contours (C)

Syntax	<pre>dpcon(<options,><levels,spacing>)</levels,spacing></options,></pre>		
Description	Produces a true contour plot display.		
Arguments	options must precede levels and spacing in the argument list and can be one or more of the following:		
	• 'pos' is a keyword to limit the display to positive peaks only in phased spectra. The default is both positive and negative peaks.		
	• 'neg' is a keyword to limit the display to negative peaks only in phased spectra.		
	• 'noaxis' is a keyword to omit outlining the display and drawing the horizontal or vertical axis.		
	levels is the maximum number of contours to be shown. The default is 4.		
	spacing is the spacing by relative intensity of successive contour levels. The default is 2.		
Examples	dpcon dpcon('pos',6) dpcon(15,1.4)		

See also	NMR Spectroscopy User Guide	
Related	dcon	Display noninteractive color intensity map (C)
	dconi	Control display selection for the dconi program (P)
	dpconn	Display plotted contours without screen erase (C)
	pcon	Plot contours on plotter (C)

dpconn Display plotted contours without screen erase (C)

Syntax dpconn(<options, ><levels, spacing>)

- Description Produces a true contour plot display exactly the same as the dpcon command, but without erasing the screen before drawing. The arguments are entered the same as dpcon.
 See also NMR Spectroscopy User Guide
 - Related dpcon Display plotted contours (C)

dpf Display peak frequencies over spectrum (C)

- Description Displays peak frequencies in the graphics window, with units specified by the axis parameter. Only those peaks greater than th high are selected. If the interactive command ds is active, dpf deactivates it.

Two basic modes of label positioning are available: labels placed at the top, with *long leaders* extending down to the tops of the lines (syntax 1 using 'top' keyword) or labels positioned just above each peak, with *short leaders* (syntax 2 using 'leader' keyword). The default is short leaders.

Arguments 'noll' is a keyword to display frequencies using last previous line listing.

'pos' (or 'noneg') is a keyword to display positive peaks only.

noise_mult is a numerical value that determines the number of noise peaks displayed for broad, noisy peaks. The default is 3. A smaller value results in more peaks, a larger value results in fewer peaks, and a value of 0.0 results in a line listing containing all peaks above the threshold th. Negative values of noise_mult are changed to a value of 3. The noise_mult argument is inactive when the 'noll' keyword is specified.

'top' is a keyword to display peak labels at the top with long leaders. In this mode, the height of labels is varied by changing the parameter wc2.

'leader' is a keyword to display labels positioned just above each peak.
length specifies the leader length, in mm, if labels are positioned just above each peak. The default is 20.

Examples	dpf('pos')					
	dpf('leader',30)					
	dpf('top	p','noll')				
	dpf('pos	s',0.0,'leader',30)				
See also	NMR Spee	ctroscopy User Guide				
Related	Axis label for displays and plots (P)					
	dpir	Display integral amplitudes below spectrum (C)				
	dpirn	irn Display normalized integral amplitudes below spectrum (M)				
	pir	Plot integral amplitudes below spectrum (C)				
	pirn	Plot normalized integral amplitudes below spectrum (M)				
	ppf Plot peak frequencies over spectrum (M)					
	th	Threshold (P)				
	vp	Vertical position of spectrum (P)				
	wc2	Width of chart in second direction (P)				

dpir Display integral amplitudes below spectrum (C)

Description	Displays integral amplitudes below the appropriate spectral regions.		
See also	NMR Spectroscopy User Guide		
Related	dpf	Display peak frequencies over spectrum (C)	
	dpirn	Display normalized integral amplitudes below spectrum	
		(M)	
	pir	Plot integral amplitudes below spectrum (C)	
	pirn	Plot normalized integral amplitudes below spectrum (M)	
	ppf	Plot peak frequencies over spectrum (M)	

dpirnDisplay normalized integral amplitudes below spectrum(M)

Description	Equivalent to is normalized	the command dpir except that the sum of the integrals to the value of the parameter ins.			
See also	NMR Spectro	NMR Spectroscopy User Guide			
Related	dpir Display integral amplitudes below spectrum (C)				
	ins	Integral normalization scale (P)			
	pirn	Plot normalized integral amplitudes below spectrum			
		(M)			

dpiv Display integral values below spectrum (M)

Syntax	dpiv<(ve	rtical_position)>
Description	Labels int number in	egrals with a bracket below the spectrum and a vertical dicating the integral value.
	• vertical la	bels for narrower regions
	• avoids lab	el overlap by label shifting
	• more flexi	ble vertical positioning
	The vertic labels, ass position of that the in message is aborts. No The minim and in the	al position defaults to a location just underneath the scale uming there is enough room below the scale. If the vertical is too low, the vertical position is allowed to approach the f the spectrum up to 1 mm. If the spectral position is so low ntegral labels would overlap with the spectrum, an error is produced (indicating the minimum vp), and the command error message is produced in case of overlap with the scale. num for vp depends on the plotter and the character size, e case of dpiv also on the size of the graphics window.
	Use an op no checkin overlap. p 2 mm belo	tional argument to force the vertical position to any value; ng is done, and no error message is produced in case of $iv(vp-2)$ produces integral labels with the brackets ending by the position of the spectrum.
	dpiv follo insref an (local or g	ws this convention: the output is controlled by ins and nd not by is. Restore the is integration mode by creating a global) parameter oldint and set oldint= 'y':
	create(' oldint='	oldint','flag','global') y'
	oldint=' integratior	$\ensuremath{\texttt{n}}\xspace$ (or destroy the parameter) switches back to the default $\ensuremath{\texttt{n}}\xspace$ mode.
Examples	vp=25 dp	iv
	vp=50 pl	pscale piv(0)
Related	dpir dpirn	Display integral amplitudes below spectrum (C) Display normalized integral amplitudes below spectrum (C)
	dpivn	Display normalized integral amplitudes below spectrum (M)
	pirn	Plot normalized integral amplitudes below spectrum (C)
	pir	Plot integral amplitudes below spectrum (C)
	piv	Plot integral amplitudes below spectrum (M)
	pivn	Plot normalized integral amplitudes below spectrum (M)

dpivn

Display normalized integral values below spectrum (M)

Syntax dpivn<(vertical_position)>

Description	Labels integrals with a bracket below the spectrum and a vertical number indicating the integral value.			
	See dpiv	7 for description and use.		
Related	dpir	Display integral amplitudes below spectrum (C)		
	dpirn	Display normalized integral amplitudes below spectrum (C)		
	dpiv	Display integral amplitudes below spectrum (M)		
	pirn	Plot normalized integral amplitudes below spectrum (C)		
	pir	Plot integral amplitudes below spectrum (C)		
	piv	Plot integral amplitudes below spectrum (M)		
	pivn	Plot normalized integral amplitudes below spectrum (M)		

dpl Default plot (M)

Description	Looks for sequence-specific default plot macro (dpl_seqfil) and executes if one is found.		
Related	dpl_seqfil	Sequence-specific default plot (M)	
	dpr	Default process (M)	
	dds	Default display (M)	

dpl_seqfil Sequence-specific default plot (M)

Description	Sequence-spe macro.	ecific default plot.	These	macros	are	called	by	the	dpl
Examples	dpl_NOESY1 dpl_TOCSY1	D D							
Related	dpl dpr dds	Default plot (M) Default process Default display ((M) M)						

dplane Display a 3D plane (M)

Syntax	dplane(<plane_type,>plane_number)</plane_type,>
Description	Displays the 2D color map of a particular data plane from a 3D spectral data set. The 3D parameters are loaded into VnmrJ each time dplane is executed. The parameter path3d specifies the absolute path to the directory (without the .extr file extension) where the 2D planes extracted from the 3D spectral data set reside.
Arguments	plane_type is one of the keywords 'f1f3', 'f2f3', and 'f1f2' for the f_1f_3 , f_2, f_3 , and f_1f_2 planes, respectively. If plane_type is specified, the parameter plane is updated with that new value. plane is then used to determine the type of 3D plane to be displayed.

	plane_numb displayed:	er specifies which plane of a particular type is to be				
	• For plane f ₁ f	For plane f_1f_3 , the range of plane_number is 1 to $fn2/2$				
	• For plane f ₂ f	S_3 , the range of plane_number is 1 to fn1/2				
	• For plane f ₁ f	f_2 , the range of plane_number is 1 to fn/2				
Examples	dplane(3) dplane('f1	f2',2)				
See also	NMR Spectro	oscopy User Guide				
Related	dsplanes	Display a series of 3D planes (M)				
	dproj	Display a 3D plane projection (M)				
	getplane	Extract planes from a 3D spectral data set (M)				
	nextpl	Display the next 3D plane (M)				
	path3d	Path to currently displayed 2D planes from a 3D data set (P)				
	plane	Currently displayed 3D plane type (P)				
	prevpl	Display the previous 3D plane (M)				
	plplanes	Plot a series of 3D planes (M)				

dpr Default process (M)

Description	Looks for sequence-specific default plot macro (dpr_seqfil) and executes if one is found.		
Related	dpr_seqfil	Sequence-specific default process (M)	
	dpl	Default plot (M)	
	dds	Default display (M)	

dpr_seqfil Sequence-specific default process (M)

Description	Sequence-spe macro.	cific default plot. These macros are called by the dpr
Examples	dpr_NOESY11 dpr_TOCSY11	
Related	dpr	Default process (M)
	dpl	Default plot (M)
	dds	Default display (M)

dprofile Display pulse excitation profile (M)

Syntax dprofile<(axisflag<,profile<,shapefile>>)>

Description	Displays the X, Y and Z excitation (inversion) profile for a pulse shape generated by the Pbox software. If shapefile is not provided, the last simulation data stored in the shapelib/pbox.sim file are displayed.
Arguments	The axisflag and profile arguments can be given in any order.
	axisflag is 'y' to display the full spectrum and a frequency scale, or 'n' to suppress the scale and spectrum. The default is 'n'.
	profile is a character string identifying the desired profile. 'xyz' selects X, Y, and Z (inversion) profiles; 'xy' selects only the excitation (transverse) profiles; 'x' selects only the X transverse excitation profile; and 'z' selects only the inversion profile. The default is 'xyz'.
	shapefile is the name of a *.RF or *.DEC file, including the extension.
Examples	dprofile dprofile('y','xy') dprofile('xy','n','softpls.RF')
See also	NMR Spectroscopy User Guide
Related	pprofilePlot pulse excitation profile (M)PboxPulse shaping software (U)

dproj Display a 3D plane projection (M)

Syntax	dproj<(pla	ne_type)>	
Description	Displays 2D color map of the 2D projection plane from a 3D spectral data set. The projection is a skyline projection. The 3D parameters are loaded into VnmrJ each time dproj is executed. For this macro, the parameter path3d specifies the directory (without the .extr extension) where the 2D projection resides that has been created from the 3D spectral data set.		
Arguments	plane_type the f_1f_3 , f_2 , f_3 ; the parameter to determine	is one of the keywords 'flf3', 'f2f3', and 'flf2' for and f_1f_2 planes, respectively. If plane_type is specified, er plane is updated with that value. plane is then used the type of 2D projection to be displayed.	
Examples	dproj dproj('f1f	2 ')	
See also	NMR Spectre	oscopy User Guide	
Related	dplane dsplanes getplane nextpl path3d plane plplanes	Display a 3D plane (M) Display a series of 3D planes (M) Extract planes from a 3D spectral data set (M) Display the next 3D plane (M) Path to currently displayed 2D planes from a 3D data set (P) Currently displayed 3D plane type (P) Plot a series of 3D planes (M) Display the previous 3D plane (M)	
	prevpi	Display the previous 5D plane (M)	

293

Syntax dps<(file),x,y,width,height> Description Displays a picture of pulse sequences consisting of three to five parts. The top part is the transmitter pulse sequence (Tx). The second part is the decoupler pulse sequence (Dec). The third part might be the second or third decoupler (Dec2 or Dec3) pulse sequence or gradients (X, Y, or Z), depending on the program. The lowest part is the status. The pulse parameters are displayed if there is enough space an if the length of the parameter name is less than thirty letters. The value of each pulse is also displayed. If the value delay or width is less than zero, a question mark (?) is displayed. The time units are displayed in color (on a color monitor). The height of pulses is scaled according to their power level. dps also displays spin lock, transmitter gating, observe transmitter power, and other information. Arguments file specifies the name of the file containing the pulse sequences. The default is the file seqfil. x, y specifies the start of the position with respect to the lower-left corner of the window. width, height are in proportion to wcmax and wc2max. See also NMR Spectroscopy User Guide Related pps Plot pulse sequence (C) seqfil Pulse sequence name (P) Width of chart (P) WC Maximum width of chart (P) wcmax wc2max Maximum width of chart in second direction (P)

dpwr Power level for first decoupler with linear amplifier (P)

Applicability Systems with a linear amplifier.

Description On systems equipped with a linear amplifier, a 63-dB or 79-dB attenuator between the decoupler transmitter and the amplifier controls the power level.

The system value for the attenuator upper safety limit is set fin the Spectrometer Configuration window (opened by config). The Upper Limit entry sets this value. For broadband decoupling of ¹H nuclei, typical values range from 36 to 49 dB. For homonuclear decoupling, typical values range from 5 to 15 dB.

Values 79 dB, -16 to +63, in steps of 1 dB.

Decoupler power greater than 2 watts in a switchable probe will damage the probe. Always carefully calibrate decoupling to avoid exceeding 2 watts. The maximum value for dpwr on a 200-, 300-, or 400-MHz system with a linear amplifier on the decoupler channel has

been set to 49, corresponding to about 2 watts of power. Before using dpwr=49 for continuous decoupling, ensure safe operation by measuring the output power. This should be done during system installation and checked periodically by the user.

See also VnmrJ Installation and Administration

Related	cattn	Coarse attenuator (P)
	config	Display current configuration and possible change it (M)
	dpwrf	First decoupler fine power (P)
	dpwr2	Power level for second decoupler (P)
	dpwr3	Power level for third decoupler (P)
	dpwr4	Power level for fourth decoupler (P)
	fattn	Fine attenuator (P)
	tpwr	Power level of observe transmitter with linear amplifiers
		(P)
	tpwrf	Observe transmitter fine power (P)

dpwr2 Power level for second decoupler with linear amplifier (P)

ApplicabilitySystems with a linear amplifier as the second decoupler.DescriptionControls the coarse attenuator (63 dB or 79 dB) that resides between
the transmitter board and the linear amplifier associated with the
second decoupler. The system value for the attenuator upper safety
limit is set in the Spectrometer Configuration window (opened by
config).

Values 79 dB, -16 to +63, in steps of 1 dB.

If dn2='' (two single quotes) and a second decoupler channel is present in the console, dpwr2 assumes a default value of 0 when go is executed.

CAUTION Decoupler power greater than 2 watts in a switchable probe will damage the probe. Always carefully calibrate decoupling to avoid exceeding 2 watts. The maximum value for dpwr2 on a 200-, 300-, or 400-MHz system with a linear amplifier on the decoupler channel has been set to 49, corresponding to about 2 watts of power. Before using dpwr2=49 for continuous decoupling, ensure safe operation by measuring the output power. This should be done during system installation and checked periodically by the user.

See also	NMR Spectroscopy User Guide	
Related	cattn	Coarse attenuator type (P)
	config	Display current configuration and possible change it
		(M)
	dn2	Nucleus for second decoupler (P)

dpwr3 Power level for third decoupler with linear amplifier (P)

ApplicabilitySystems with a linear amplifier as the third decoupler.DescriptionControls the coarse attenuator (63 dB or 79 dB) that resides between
the transmitter board and the linear amplifier associated with the third
decoupler. The system value for the attenuator upper safety limit is set
in the Spectrometer Configuration window (opened by config).

Values If 63-dB attenuator installed: 0 to 63 (63 is max. power), in units of dB. If 79-dB attenuator installed: -16 to 63 (63 is max. power), in units of dB. If dn3='' (two single quotes) and a third decoupler channel is present in the console, dpwr3 assumes a default value of 0 when go is executed.

CAUTION Decoupler power greater than 2 watts in a switchable probe will damage the probe. Always carefully calibrate decoupling to avoid exceeding 2 watts. The maximum value for dpwr3 on a 200-, 300-, or 400-MHz system with a linear amplifier on the decoupler channel has been set to 49, corresponding to about 2 watts of power. Before using dpwr3=49 for continuous decoupling, ensure safe operation by measuring the output power. This should be done during system installation and checked periodically by the user.

See also	NMR Spectroscopy User Guide	
Related	cattn	Coarse attenuator type (P)
	config	Display current configuration and possible change it
		(M)
	dn3	Nucleus for third decoupler (P)

dpwr4 Power level for fourth decoupler amplifier (P)

Applicability Systems with deuterium decoupler channel as the fourth decoupler.
Description Controls the coarse attenuator (45 dB range) that resides on the Lock Transceiver board and the amplifier associated with the fourth decoupler. The system value for the attenuator upper safety limit is set in the Spectrometer Configuration window (opened by config).
Values 48-dB attenuator: 15 to 63 (63 is max. power), in units of dB. If dn4='' (two single quotes) and a third decoupler channel is present in the console, dpwr4 assumes a default value of 0 when go is executed.

CAUTION

Decoupling power greater than 5 watts applied to a triple-resonance probe will damage the probe. The maximum value for dpwr4 is 63, corresponding to about 35 watts to the probe. A value of dpwr4 equal to 52 corresponds to about 5 watts and will produce approximately a 1 kHz decoupling field. Always carefully calibrate decoupling power to avoid exceeding 5 watts. Before using dpwr4=52 continuous decoupling, ensure safe operation by measuring the output power. Measurement should be taken during system installation and checked periodically by the user.

See also	NMR Spectroscopy User Guide	
Related	cattn	Coarse attenuator type (P)
	config	Display current configuration and possible change it
		(M)
	dn3	Nucleus for third decoupler (P)

dpwrf First decoupler fine power (P)

Applicability	Systems with an optional fine attenuator on the decoupler channel.	
Description	Controls the first decouple fine attenuator. Systems with this attenuator are designated within the Spectrometer Configuration window (opened by config) by the status of the Fine Attenuator entry. The fine attenuator is linear and spans 6 dB.	
Values	0 to 4095 (where 4095 is maximum power). If dpwrf does not exist in the parameter table, a value of 4095 is assumed.	
See also	User Programming, User Guide: Solids; CP/MAS Installation,	
Related	config	Display current configuration and possibly change it (M)
	dpwr	Power level for first decoupler with linear amplifiers (P)
	dpwrf2	Second decoupler fine power (P)
	dpwrf3	Third decoupler fine power (P)
	dpwrm	First decoupler linear modulator power (P)
	fattn	Fine attenuator (P)
	tpwr	Power level of observe transmitter with linear amplifiers (P)
	tpwrf	Transmitter fine power (P)

dpwrf2 Second decoupler fine power (P)

Applicability	Systems with an optional fine attenuator on the second decoupler channel.
Description	Controls the second decoupler fine attenuator, functioning analogously to dpwrf.

Values	0 to 4095 (w	here 4095 is maximum power). If dpwrf2 does not exist
	in the param	eter table, a value of 4095 is assumed.
See also	User Program	nming
Related	dpwrf	First decoupler fine power (P)

dpwrf3 Third decoupler fine power (P)

Applicability	Systems with an optional fine attenuator on the third decoupler channel.
Description	Controls the third decoupler fine attenuator, functioning analogously to $dpwrf$.
Values	0 to 4095 (where 4095 is maximum power). If dpwrf3 does not exist in the parameter table, a value of 4095 is assumed.
See also	User Programming
Related	dpwrf First decoupler fine power (P)

dpwrm First decoupler linear modulator power (P)

Applicability	Systems with a first decoupler linear modulator. The fine power control is linear and spans 0 to dpwr.	
Values	0 to 4095 (w) in the parame	here 4095 is maximum power). If dpwrm does not exist eter table, a value of 4095 is assumed.
See also	User Programming; User Guide: Solids; CP/MAS Installation	
Related	dpwrm2 dpwrm3 tpwrm	Second decoupler linear modulator power (P) Third decoupler linear modulator power (P) Observe transmitter linear modulator power (P)

dpwrm2 Second decoupler linear modulator power (P)

Applicability	Systems with a second decoupler linear modulator.		
Description	Controls the second decoupler linear modulator systems.		
Values	0 to 4095 (where 4095 is maximum power). If dpwrm2 does not exist in the parameter table, a value of 4095 is assumed.		
See also	User Programming		
Related	dpwrm First decoupler linear modulator power (P)		

dpwrm3 Third decoupler linear modulator power (P)

Applicability	Systems with a third decoupler linear modulator.
Description	Controls the third decoupler linear modulator systems.
Values	0 to 4095 (where 4095 is maximum power). If dpwrm3 does not exist in the parameter table, a value of 4095 is assumed.
See also	User Programming
Related	dpwrm First decoupler linear modulator power (P)

Dgcosy Convert the parameter to a DQCOSY experiment (M)

Description	Convert the parameter to a double-quantum filtered (DQCOSY) experiment	
See also	NMR Spectroscopy User Guide	
Related	cosyps	Set up parameters for phase-sensitive COSY (M)
	Cosy	Set up parameters for COSY pulse sequence (M)
	relayh	Set up parameters for COSY pulse sequence (M)

draw Draw line from current location to another location (C)

Syntax	draw(<'keywords'>x,y)
Description	Draws a line from the current location to the absolute location with coordinates given by the arguments.
Arguments	<pre>'keywords' identifies the output device ('graphics' 'plotter'), drawing mode ('xor' 'normal'), and drawing capability ('newovly' 'ovly' 'ovlyC').</pre>
	• 'graphics' 'plotter' is a keyword for the output device. The default is 'plotter'. The output selected is passed to subsequent pen, move, or draw commands and remains active until a different output is specified.
	• 'xor', 'normal' is a keyword for the drawing mode when using the 'graphics' output device. The default is 'normal'. In the 'xor' mode, if a line is drawn such that one or more points of the line are in common with a previous 'xor' line, the common points are erased. In the normal mode, the common points remain. The mode selected is passed to subsequent draw, pen, and move commands and remains active until a different mode is specified.

	'newovly',	'ovly', and 'ovlyC' are keywords that specify an
	interactive d	rawing capability that is slightly slower than the 'xor'
	mode but mo	ore consistent in color. 'newovly' clears any previous
	draws, boxes	, and writes made with the 'ovly' modes and draws
	the figure. '	ovly' draws without clearing so that multisegment
	figures can b	e created. 'ovlyC' clears without drawing.
	x,y are the to be drawn. wcmax at the	absolute coordinates, in mm, of the endpoint of the line The range of x is 0 at the left edge of the chart and e right edge. The range of y is -20 at the bottom of the
	chart and wo	22max at the top.
Examples	draw('grap	hics','xor'.wcmax-sc,vp+th)
	draw(wcmax	-sc-wc*(cr-delta-sp)/wp,wc2max)
See also	NMR Spectroscopy User Guide	
Related	gin	Return current mouse position and button values
		(C)
	move	Move to an absolute location (C)

move	Move to an absolute location (C)
pen	Select a pen or color for drawing (C)
wcmax	Maximum width of chart (P)
wc2max	Maximum width of chart in second direction (P))

dres Measure linewidth and digital resolution (C)

Syntax	<pre>dres<(<freq<,fractional_height>>)> :linewidth,digital_resolution</freq<,fractional_height></pre>		
Description	Analyzes the line defined by the current cursor position for its linewidth (width at half-height) and digital resolution.		
Arguments	freq is the frequency of the line. The default is the parameter cr. This overrides using the current cursor position as the frequency.		
	fractional_height is the linewidth is measured at this height.		
	linewidth is the value returned for the linewidth of the line.		
	digital_resolution is the value returned for the digital resolution of the line.		
Examples	dres:\$width,\$res dres(cr,0.55)		
See also	NMR Spectroscopy User Guide; User Programming		
Related	crCurrent cursor position (P)dsnMeasure signal-to-noise (C)		

dres Tip-angle resolution for first decoupler (P)

Applicability Systems with waveform generators.

- Description Controls the tip-angle resolution to be used within a waveform generator decoupling sequence on the first decoupler. The optimum value is a function of the decoupling sequence to be used: for WALTZ-16, dres=90.0; for MLEV16-240, dres=30.0; and for GARP1, dres=1.0.
 Values 1.0 to 90.0, in units of degrees. In reality, dres can assume values as
 - values 1.0 to 90.0, in units of degrees. In reality, dress can assume values as small of 0.7 (but no smaller) and can be specified in units of 0.1°. To use this capability, change the limits of dres by using destroy('dres') create('dres', 'real') setlimit('dres', 360, 0.7, 0.1). Making corresponding changes within the fixpar macro ensures that dres is created in the desired way with each new parameter set.
 - See also NMR Spectroscopy User Guide

dmfadj	Adjust decoupler tip-angle resolution time (M)
dres2	Tip angle resolution for second decoupler (P)
dres3	Tip angle resolution for third decoupler (P)
fixpar	Correct parameter characteristics in experiment (M)
	dmfadj dres2 dres3 fixpar

dres2 Tip-angle resolution for second decoupler (P)

Applicability	Systems with waveform generators.	
Description	Controls the tip-angle resolution to be used within a waveform generator decoupling sequence on the second decoupler. The optimum value is a function of the decoupling sequence to be used: for WALTZ-16, dres2=90.0; for MLEV16-240, dres2=30.0; and for GARP1, dres2=1.0.	
Values	1.0 to 90.0, in units of degrees.	
See also	NMR Spectroscopy User Guide	
Related	dmf2adj Adjust second decoupler tip-angle resolution time (M)	
	dres Tip-angle resolution for first decoupler (P)	

dres3 Tip-angle resolution for third decoupler (P)

Applicability Systems with waveform generators.

- Description Controls the tip-angle resolution to be used within a waveform generator decoupling sequence on the third decoupler. The optimum value is a function of the decoupling sequence to be used: for WALTZ-16, dres3=90.0; for MLEV16-240, dres3=30.0; and for GARP1, dres3=1.0.
 - Values 1.0 to 90.0, in units of degrees.

See also	NMR Spectroscopy User Guide	
Related	dmf3adj	Adjust third decoupler tip-angle resolution time (M)
	dres	Tip-angle resolution for first decoupler (P)

dres4 Tip-angle resolution for fourth decoupler (P)

Applicability	Systems with	a deuterium decoupler channel as the fourth decoupler.
Description	Controls the tip-angle resolution to be used for the decoupling sequence on the fourth decoupler. The optimum value is a function of the decoupling sequence to be used: for WALTZ-16, dres4=90.0; for MLEV16-240, dres4=30.0; and for GARP1, dres4=1.0.	
Values	1.0 to 90.0, in units of degrees.	
See also	NMR Spectroscopy User Guide	
Related	dmf4adj	Adjust fourth decoupler tip-angle resolution time (M)
	dres	Tip-angle resolution for first decoupler (P)

ds

Display a spectrum (C)

Syntax	<pre>(1) ds<(index)> (2) ds<(options)></pre>
Description	Displays a single spectrum. Parameter intmod controls integral display:
	• intmod='off' turns off the integral display
	• intmod='full' displays the entire integral
	• intmod='partial' displays every other integral region
	Parameter entry after a spectrum has been displayed with the ds command causes the spectrum to be updated.
	Two additional parameters control the behavior of the $\ensuremath{\texttt{ds}}$ command:
	• The parameter phasing (in the "global" parameter set) controls the percentage of the spectrum updated during interactive phasing. This parameter can be set in the range of 10 to 100. A value of 100 causes the entire spectrum to be updated. A value of 20 causes the area

between the two horizontal cursors to be updated.

• The parameter lvltlt (in the "current" parameter set) controls the sensitivity of the interactive lvl and tlt adjustments. lvltlt can be set to any positive real number. It is basically a multiplier for the sensitivity. The default value is 1.0. Larger values make the adjustments larger. Smaller values make the adjustments smaller.

For arrayed 1D spectra or for 2D spectra, a particular trace can be viewed by supplying the index number as an argument. For 2D data sets, spectra can be displayed from either the f_1 or f_2 domain by setting

the parameter trace equal to 'f1' or 'f2', respectively. After entering ft1d, interferograms can be viewed by setting trace='f1' and then typing ds.

Spectra are scaled according to the number of completed transients ct. If nt is arrayed (nt=1,2,4,8), each spectrum is scaled by its own ct.

Arguments index (used with syntax 1) is the index number of a particular trace to be displayed in arrayed 1D spectra or in 2D spectra (syntax 1).

options (used with syntax 2) is any of the following keywords:

- 'toggle' switches between the box and the cursor modes.
- 'restart' redraws the cursor if it has been turned off.
- 'expand' toggles between expanded and full view of the spectrum.
- 'spwp' interactively adjusts start and width of the spectrum display.
- 'phase' enters an interactive phasing mode.
- 'thresh' interactively adjusts the threshold.
- 'z' interactively sets integral resets.
- 'dscale' toggles the scale below the spectrum on and off.
- 'lvltlt' interactively adjusts the lvl and tlt parameters.
- 'scwc' interactively adjusts the start and width of chart.
- 'noclear' start or restart the ds display without clearing the graphics screen
- 'exists' exit the ds display, leaving a non-interactive dss display.

Examples ds

ds(7) ds('restart')

See also NMR Spectroscopy User Guide

Related	crmode	Current state of cursors in dfid, ds, or dconi (P)
	ct	Completed transients (P)
	exists	
	ft1d	Fourier transform along f_2 dimension (C)
	intmod	Integral display mode (P)
	lp	First-order phase in directly detected dimension (P)
	lvl	Zero-order baseline correction (P)
	lvltlt	Control sensitivity of lvl and tlt adjustments (P)
	nt	Number of transients (P)
	phasing	Control update region during ds phasing (P)
	rp	Zero-order phase in directly detected dimension (P)
	select	Select a spectrum without displaying It (C)
	tlt	First-order baseline correction (P)
	trace	Mode for n-dimensional data display (P)
	wft1d	Weight and Fourier transform f ₂ for 2D data (C)

ds2d Display 2D spectra in whitewash mode (C)

Syntax ds2d<(options)>

Description Displays a stacked plot of 2D spectra in whitewash mode (after the first spectra, each spectra is blanked out in regions in which it is behind an earlier spectra). Color does not represent intensity (unlike dcon), because intensity can be seen visually, but instead successive traces are displayed in different colors so that color represents frequency.

Arguments options can be any of the following keywords:

- 'nobase' is a keyword to activate the th parameter to suppress all intensity below the th level.
- 'fill' is a keyword to fill in the peaks. When using 'fill', th operates linearly and not logarithmically (factors of 2) as it does in the contour or color intensity displays.
- 'fillnb' is a keyword to combine base suppression and peak filling. When using 'fillnb', th operates linearly and not logarithmically (factors of 2) as it does in the contour or color intensity displays.
- 'noaxis' is a keyword to omit outlining the display and drawing the horizontal and vertical axis.

Examples ds2d

ds2d('fillnb')

See also NMR Spectroscopy User Guide

- RelateddconDisplay noninteractive color intensity map (C)dconiControl display selection for the dconi program (P)ds2dnDisplay 2D spectra in whitewash mode without screen erase (C)pl2dPlot 2D spectra in whitewash mode (C)
 - th Threshold (P)

ds2dn Display 2D spectra in whitewash mode without screen erase (C)

Syntax	ds2dn<(options)>	
Description	Displays a stacked plot of 2D spectra in whitewash mode (after the first spectra, each spectra is blanked out in regions in which it is behind an earlier spectra) the same as ds_2d but without erasing the screen before drawing. The arguments are the same as ds_2d .	
Examples	ds2dn ds2dn('fillnb')	
See also	NMR Spectroscopy User Guide	
Related	ds2d Display 2D spectra in whitewash mode (C)	

dscale Display scale below spectrum or FID (C)

Syntax dscale<(<rev><,axis><,label><,vp0><,sp0><,color><,pen>)> Description Displays a scale under a spectrum or FID. rev – reverses the direction of the scale. That is, the smaller numbers Arguments will be at the left side of the scale. If used, 'rev' must be the first argument. axis – If the letter p, h, k, etc. is supplied, it will be used instead of the current value of the parameter axis. For an FID scale, if the letter s, m, or u is supplied, it will be used instead of the current value of the parameter axisf. label – If a string of 2 or more characters is supplied, it will be used as the axis label. vp0 – This is supplied as the first real number. It defines the vertical position where the scale is drawn. The default is 5 mm below the current value of the parameter vp. sp0 - This is supplied as the second real number. It is a modified start of plot. If, for example, the display is from 347 to 447 hz, but the scale is desired to read 0 to 100 hz., sp0 would be input as 0. wp0 – This is supplied as the third real number. It is a modified width of plot. If, for example, the display is from 347 to 447 hz, but the scale is desired to read 0 to 550 Units. sp0 would be input as 0, wp0 would be 550, and the label would be 'Units'. An optional color or pen number can be supplied to dscale or pscale. The available colors and pens are: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', 'white' 'pen1', 'pen2', 'pen3',..., 'pen8' Examples dscale dscale('rev') dscale('h',0,'green') dscale('h',vp-10,0) See also NMR Spectroscopy User Guide Related axis Axis label for displays and plots (P) axisf Axis label for FID displays and plots (P) pscale Plot scale below spectrum or FID (C) Vertical position of spectrum (P)

dsnarray Report statistical signal-to-noise for Cold Probes (M)

ApplicabilitySystems with Varian, Inc. Cold ProbesDescriptionReport the statistical S/N of a series of repeated gNhsqc data sets
acquired with a labeled protein sample.

Digital filter coefficients for downsampling (P) dscoef

Description	Specifies the number of coefficients used in the digital filter. This			
	parameter does not need to be changed as the parameter downsamp			
	is changed, t	because dscoef is automatically adjusted by vnmrJ to give		
	filter cutoffs	that are the same, regardless of the value of downsamp.		
	This is done	by using dscoef*downsamp/2 coefficients in the digital		
	filter. VnmrJ	always rounds dscoef*downsamp/2 to an odd number.		
	If dscoef d	oes not exist in the current experiment, enter		
	addpar('do	ownsamp') to add it. Entering addpar('downsamp')		
	creates the o	digital filtering and downsampling parameters downsamp,		
	dscoef, dsfb, dslsfrq, and filtfile.			
Values	Number of digital filter coefficients. The default is 61. A larger number			
	of coefficients gives a filter with sharper cutoffs; a smaller			
	gives a filter	ilter with more gradual cutoffs.		
See also	NMR Spectroscopy User Guide			
Related	addpar	Add selected parameters to current experiment (M)		
	downsamp	Downsampling factor applied after digital filtering (P)		
	dsfb	Digital filter bandwidth for downsampling (P)		
	dslsfrq	Bandpass filter offset for downsampling (P)		
	filtfile	File of FIR digital filter coefficients (P)		
	pards	Create additional parameters used for downsampling		
		(M)		

Decoupler sequence for first decoupler (P) dseq

Applicability	Systems with waveform generators.		
Description	Specifies the decoupling sequence (without the .DEC file extension) to be used during any period of programmable decoupling on the first decoupler under status control (i.e., dmm='p'). The decoupling sequence must be located in the user's shapelib directory or in the VnmrJ system's shapelib directory.		
See also	NMR Spectroscopy User Guide		
Related	dmm Decoupler modulation mode for first decoupler (P)		
	dseq2 Decoupler sequence for second decoupler (P)		
	dseq3 Decoupler sequence for third decoupler (P)		

dseq2 **Decoupler sequence for second decoupler (P)**

Applicability Systems with waveform generators.

Description Specifies the decoupling sequence (without the .DEC file extension) to be used during any period of programmable decoupling on the second decoupler under status control (i.e., dmm2='p'). The decoupling

to

sequence must be located in the user's shapelib directory or in the VnmrJ system shapelib directory. See also NMR Spectroscopy User Guide Related dmm2 Decoupler modulation mode for second decoupler (P)

dseq Decoupler sequence for first decoupler (P)

dseq3 Decoupler sequence for third decoupler (P)

Applicability	Systems with waveform generators.		
Description	Specifies the decoupling sequence (without the .DEC file extension) to be used during any period of programmable decoupling on the third decoupler under status control (i.e., dmm3='p'). The decoupling sequence must be located in the user's shapelib directory or in the shapelib directory.		
See also	NMR Spectroscopy User Guide		
Related	dmm3 Decoupler modulation mode for third decoupler (P)		
	dseq Decoupler sequence for first decoupler (P)		

dseq4 Decoupler sequence for fourth decoupler (P)

Applicability	Systems with waveform generators.		
Description	Specifies the decoupling sequence (without the .DEC file extension) to be used during any period of programmable decoupling on the third decoupler under status control (i.e., dmm4='p'). The decoupling sequence must be located in the user's shapelib directory or in the system's shapelib directory.		
See also	NMR Spectroscopy User Guide		
Related	dmm4Decoupler modulation mode for third decoupler (P)dseqDecoupler sequence for first decoupler (P)		

dsfb Digital filter bandwidth for downsampling (P)

of $\pm \frac{sw}{2}$.

Description	Specifies the bandwidth of the digital filter used for downsampling. If
	dsfb does not exist in the current experiment, enter
	<pre>addpar('downsamp') to add it. addpar('downsamp') creates the</pre>
	digital filtering and downsampling parameters downsamp, dscoef,
	dsfb, dslsfrq, and filtfile.
Values	Number, in Hz. A smaller value rejects frequencies at the spectrum
	edges; a larger value aliases noise and signals at frequencies outside

'n'	makes	dsfb	default	to	the	final	<mark>sw</mark> /2
-----	-------	------	---------	----	-----	-------	--------------------

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to current experiment (M)
	downsamp	Downsampling factor applied after digital filtering (P)
	dscoef	Digital filter coefficients for downsampling (P)
	dslsfrq	Bandpass filter offset for downsampling (P)
	filtfile	File of FIR digital filter coefficients (P)
	pards	Create additional parameters used for downsampling
		(M)
	SW	Spectral width in directly detected dimension (P)

dshape Display pulse shape or modulation pattern (M)

Syntax	dshape<(pattern.ext)>	
Description	Displays the real (X) and imaginary (Y) components of a shaped pulse. Any type of waveform (.RF, .DEC or .GRD) can be displayed.	
Arguments	pattern is the name of a shape or pattern file specified by an absolute file name, relative file name, or a simple pattern file name. ext is a file name extension that specifies the file type. In the case of a simple file name, dshape searches for the file in the local directory, then in the user's shapelib, and finally in the directory /vnmr/shapelib. If pattern.ext is not given, dshape displays the last created waveform stored in the pbox.fid file.	
Examples	dshape dshape('Pbox.RF')	
See also	NMR Spectroscopy User Guide	
Related	PboxPulse shaping software (U)pshapePlot pulse shape or modulation pattern (M)	

dshapef Display last generated pulse shape (M)

Description	Displays the real (X) and imaginary (Y) components of last generated shaped pulse, stored in pbox.fid file.		
See also	NMR Spectroscopy User Guide		
Related	PboxPulse shaping software (U)pshapefPlot last generated pulse shape (M)		

dshapei Display pulse shape or modulation pattern interactively (M)

Syntax dshapei<(pattern.ext)>

- Description Displays the real (X) and imaginary (Y) components of a pulse shape, modulation pattern or gradient shape interactively. dshapei overwrites the existing data (FID) after the permission is granted by the user. It also asks for the duration of the waveform and displays the timescale.
- Arguments pattern is the name of a shape or pattern file specified by an absolute file name, relative file name, or a simple pattern file name. ext is a file name extension that specifies the file type. In the case of a simple file name, dshapei searches for the file in the local directory, then in the user's shapelib, and finally in the directory /vnmr/shapelib. If no file name is given, dshapei displays the last created waveform stored in the pbox.fid file.

Examples dshapei dshapei('myfile.DEC') See also NMR Spectroscopy User Guide Related Pbox Pulse shaping software (U)

dshim Display a shim "method" string (M)

Syntax	<pre>(1) dshim<(file)></pre>		
	(2) dshim('1	method' 'help')	
Description	Looks in the user's shimmethods directory and then in the system shimmethods directory for a file and displays the file (syntax 1) or displays information about method strings (syntax 2).		
Arguments	file is the name of a file to be searched for in the shimmethods directories. The default is to display the contents of the shimmethods directories.		
	'method' is a keyword to explain the structure of method strings.		
	'help' is a keyword to describe the method strings in the system's shimmethods directory.		
Examples	es dshim dshim('method')		
1			
	dshim('hel	p')	
See also	NMR Spectroscopy User Guide		
Related	method	Autoshim method (P)	
	newshm	Interactively create a shim "method" with options	
	-1-1	(M) Submit on Autochim ormaniment to convisition (C)	
	SHIM	Submit an Autosnim experiment to acquisition (C)	
	stdshm	Interactively create a shim "method" (M)	

- Description For downsampling, selects a bandpass filter that is not centered about the transmitter frequency. In this way, dslsfrq works much like lsfrq. If dslsfrq does not exist in the current experiment, add it by entering addpar('downsamp'). The command addpar('downsamp') creates the digital filtering and downsampling parameters downsamp, dscoef, dsfb, dslsfrq, and filtfile.
 - Values A number, in Hz. A positive value selects a region upfield from the transmitter frequency; a negative value selects a downfield region.
 - See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to current experiment (M)
	downsamp	Downsampling factor applied after digital filtering (P)
	dscoef	Digital filter coefficients for downsampling (P)
	dsfb	Digital filter bandwidth for downsampling (P)
	filtfile	File of FIR digital filter coefficients (P)
	lsfrq	Frequency shift of the fn spectrum in Hz (P)
	movedssw	Set parameters for digital filtering and downsampling
		(M)
	pards	Create additional parameters used by downsampling
		(M)

dsn Measure signal-to-noise (C)

Syntax dsn<(low_field,high_field)>:signal_to_noise,noise

Description Measures the signal-to-noise ratio of the spectrum by first measuring the intensity of the largest peak in the spectral range defined by sp and wp, and then measuring the noise in the spectral region defined by the position of the two cursors. The noise value returned from dsn is not scaled by vs. The interrelations between the signal-to-noise ratio, the noise, and peak intensities can be illustrated by comparing dsn:\$sn,\$noise and peak:\$signal. In this case, \$sn is equal to (\$signal /\$noise)/vs.

> Calculate noise by first doing a drift correction on the noise region. Noise is defined as:

$$noise = 2x \left(\left(\sum_{i=1}^{np} Y_i^2 \right) / np \right)^{\frac{1}{2}}$$

 Y_i^2 values are the square of the drift-corrected amplitude and np is the number of points in the noise region.

Arguments low_field and high_field are the upper and lower frequencies of the noise region to be measured. The default is the position of the two cursors.

signal_to_noise is the calculated value of signal-to-noise ratio.

	noise is the	noise value measured within the defined spectral region.			
Examples	dsn:\$ston				
	dsn(sp+sp,	sp+wp-100)			
	dsn(10000,8000):r1				
See also	User Programming				
Related	dres	Measure linewidth and digital resolution (C)			
	peak	Find tallest peak in specified region (C)			
	sp	Start of plot (P)			
	VS	Vertical scale (P)			
	wp	Width of plot (P)			

dsnmax Calculate maximum signal-to-noise (M)

Syntax	dsnmax<(noi	.se_region)>
Description	Finds the bes	t signal-to-noise in a specified region.
Arguments	noise_region region betwee	on is the size, in Hz, of the region. The default is the n the cursors as defined by the parameter delta.
Examples	dsnmax dsnmax(400)	
See also	User Program	ıming
Related	delta	Cursor difference in directly detected dimension (P)

dsp Display calculated spectrum (C)

Syntax dsp<(file<, 'nods'>)>

Description Using the current table of transitions and intensities, dsp recalculates the simulated spectrum (using the current value for the linewidth slw) and displays the spectrum. dsp can only be used after the spins program has been run. If only the linewidth slw or vertical scale svs have been changed, dsp can be used to redisplay the spectrum. If a chemical shift or coupling constant has been changed, however, dsp will not display a spectrum reflecting the changes in the parameter; spins must be run again to recalculate the new spectrum. The number of points in the calculated spectrum is fn/2. To increase

The number of points in the calculated spectrum is fn/2. To increase the number of points, change fn and rerun dsp without doing a transform.

To display a synthetic spectrum, prepare a file in the following format:

Freq1, Intens1, LineWidth1, GaussFrac1
Freq2, Intens2, LineWidth2, GaussFrac2
...
FreqN, IntensN, LineWidthN, GaussFracN

	which is the per- Lorentzian) shou pure Gaussian). Given numbers i dsp('myshape' often preferred o large peaks.	centage of the line shape that is Gaussian (the rest is ld be between 0 and 1 (i.e., 0 is pure Lorentzian, 1 is Units for intensity are not particularly important. n a file myshape, it is only necessary to enter) to display the synthetic spectrum. This approach is over deconvolution for quantifying small shoulders on
Arguments	file is the name the result of a sy can be used to a spins.outdata information abou Gaussian/Lorent	e of a file containing spectral information that displays pectrum deconvolution. Any file in the proper format generate a display. The default is the file in the experiment directory. This file contains at frequencies, intensities, line widths, and zian fractions.
	'nods' is a key but not to displa the ds or dss c	word for dsp to recalculate the simulated spectrum by the spectrum. The spectrum can be displayed with ommand.
Examples	dsp dsp('fitspec.	outpar')
See also	NMR Spectrosco	py User Guide
Related	ds Di	splay a spectrum (C)
	dss Di	splay stacked spectra (C)
	fn Fo	urier number in directly detected dimension (P)
	slw Sp	in simulation linewidth (P)
	- D	

The units for frequency and line width are Hz. The Gaussian fraction,

spins Perform spin simulation calculation (C)

svs Spin simulation vertical scale (P)

dsp Type of DSP for data acquisition (P)

Description Selects the type of DSP (digital signal processing) for data acquisition:

- Inline DSP performs digital filtering and downsampling on the workstation immediately after each oversampled FID is transferred from the console. sw and at should be set to the values desired for the final spectrum. Only the digital filtered and downsampled data is written to the disk. Selective detection of a region of a spectrum is available using the moveossw macro.
 - *Real-time DSP* uses optional hardware (not available on all systems) to filter the data prior to summing to memory. Real-time DSP is not compatible with pulse sequences that use explicit acquisition to acquire less than the full number of data points (np) in a single acquire statement (e.g., solids sequences such as BR24 and FLIPFLOP).

If either type is active, the filter bandwidth parameter fb is not active. The actual analog filter *is* active and is automatically set by the software to a value that matches (sw/2)*oversamp as closely as possible.

Another type of DSP is available that allows post-processing of data. See the description of the pards macro for details.

Values 'i' selects inline DSP and calls addpar('oversamp') to create the DSP parameters def_osfilt, filtfile, oscoef, osfb, osfilt, oslsfrq, and oversamp. A value of oversamp greater than 1 causes the next experiment run to be oversampled, digitally filtered, and downsampled back to the selected sw prior to saving it to disk.

'r' selects real-time DSP and calls the macro addpar('oversamp') to create the DSP parameters def_osfilt, filtfile, oscoef, osfb, osfilt, oslsfrq, and oversamp (although only oversamp and osfilt are user adjustable for real-time DSP). Use dsp='r' only if the optional DSP hardware is present in the system. Set fsq='y' to use frequency-shifted quadrature detection.

'n' (or parameter dsp is not present) disables both types of DSP. Set dsp='n' if you wish to turn off DSP on a permanent or semi-permanent basis. To turn off DSP within just a single experiment, set oversamp='n'.

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to current experiment (M)
	at	Acquisition time (P)
	def_osfilt	Default value of osfilt (P)
	fb	Filter bandwidth (P)
	filtfile	File of FIR digital filter coefficients (P)
	fsq	Frequency-shifted quadrature detection (P)
	il	Interleave arrayed and 2D experiments (P)
	moveossw	Set oversampling parameters for selected spectral
		region (M)
	np	Number of data points (P)
	oscoef	Digital filter coefficients for oversampling (P)
	osfb	Digital filter bandwidth for oversampling (P)
	osfilt	Oversampling filter for real-time DSP (P)
	oslsfrq	Bandpass filter offset for oversampling (P)
	oversamp	Oversampling factor for acquisition (P)
	pards	Create additional parameters used by downsampling
		(M)
	paros	Create additional parameters used by oversampling (M)
	ra	Resume acquisition stopped with sa command (C)
	sa	Stop acquisition (C)
	SW	Spectral width in the directly detected dimension (P)

dsplanes Display a series of 3D planes (M)

Syntaxdsplanes(start_plane,stop_plane)DescriptionProduces a graphical 2D color or contour map for a subset of 3D
planes. The dconi program is used to display the planes.

D

Arguments start_plane specifies the number of the 3D plane with which display is to begin. It must be greater than 0.

stop_plane specifies the number of the 3D plane with which the display is to end. If start_plane is greater than stop_plane, only the first plane, whose number is start_plane, is plotted. The range of stop_plane depends on the value of the parameter plane as follows:

- If plane='flf3', range of stop_plane is between 0 and fn2/2
- If plane='f2f3', range of stop_plane is between 0 and fn1/2
- If plane='flf2', range of stop_plane is between 0 and fn/2

Examples dsplanes(1,3)

See also NMR Spectroscopy User Guide

Related	dconi	Interactive 2D data display (C)
	dplane	Display a 3D plane (M)
	dproj	Display a 3D plane projection (M)
	getplane	Extract planes from 3D spectral data set (M)
	nextpl	Display the next 3D plane (M)
	plane	Currently displayed 3D plane type (P)
	plplanes	Plot a series of 3D planes (M)
	prevpl	Display the previous 3D plane (M)

dsptype Type of DSP (P)

Description	Indicates the existence of digital signal processing (DSP).
Values	0 indicates no digital signal processing. 1 indicates DSP exists.
Examples	dsptype?=0 dsptype?=1
See also	NMR Spectroscopy User Guide
Related	dsp Type of DSP for data acquisition (P)

dss Display stacked spectra (C)

Syntax	dss<	(<start,< th=""><th>finish<</th><th>,step>><</th><th>,options></th><th>) ></th></start,<>	finish<	,step>><	,options>) >
--------	------	--	---------	----------	-----------	-----

Description Displays one or more spectra on the screen.

The display is not interactive like the command ds. Integral display is controlled by the parameter intmod when a single spectrum is displayed (see 'int' option below). The following values are accepted for intmod:

- intmod= 'off' turns off the integral display.
- intmod='full' displays the entire integral.
- intmod='partial' displays every other integral region.

An individual trace is displayed from and arrayed 1D spectra or 2D spectra by supplying the index number as an argument. Spectra from 2D data set are displayed from either the f_1 or f_2 domain by setting the parameter trace equal to 'f1' or 'f2', respectively. Enter ft1d, trace='f1', and dss to view the interferogram. Multiple spectra are displayed by supplying indexes of the first and last spectra.

The position of the first spectrum is governed by the parameters wc, sc, and vp. For 1D data, subsequent spectra are positioned relative to the preceding spectrum by the parameters vo (vertical offset) and ho (horizontal offset). For 2D data, ho defines the total horizontal offset between the first and last spectrum. Also for 2D data, vo is inactive while the parameter wc2 defines the total vertical offset between the first and last spectrum.

The parameter cutoff, if it exists and is active, defines the distance above and below the current vertical position vp at which peaks are truncated. By arraying cutoff to have two different values, the truncation limits above and below the current vertical position can be controlled independently. For example, cutoff=50 truncates peaks at vp+50 mm and vp-50 mm. cutoff=50,10 truncates peaks at vp+50 mm and vp-10 mm.

Arguments start is the index of the first spectra when displaying multiple spectra. It is also the index number of a particular trace to be viewed when displaying arrayed 1D spectra or 2D spectra.

finish is the index of the last spectra when displaying multiple spectra. Since the parameter arraydim is automatically set to the total number of spectra, it can be used to set finish to include all spectra (e.g., dss(1,arraydim,3)).

step is the increment for the spectral index when displaying multiple spectra. The default is 1.

options can be any of the following:

- 'all' is a keyword to display all of the spectra.
- 'int' is a keyword to display only the integral, independently of the value of the parameter intmod
- 'top' or 'side' are keywords that cause the spectrum to be displayed either above or at the left edge, respectively, of a contour plot. This assumes that the parameters sc, wc, sc2, and wc2 are those used to position the contour plot.
- 'dodc' is a keyword for all spectra to be drift corrected independently.
- 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', and 'white' are keywords that select a color.
- 'pen1', 'pen2', 'pen2' ... are keywords that pens.
- 'nopars' prevents the display commands from drawing the parameters at the bottom of the graphics screen.
- 'custom' uses the parameters shownumx (x position) and shownumy (y position), counting from bottom left of every spectrum.

•	'reve	rse	' – r	otate	the text	by	90°	- use	ful	if the	e arrayed	parameter
	values	are	long	with	respect	to	the	width	of	the	individual	
	sub-sp	ectr	a.									

• 'value' —The values of up to two simultaneous arrays are displayed. Diagonal arrays are allowed. The second parameter is shown in different color). The name of the arrayed parameter(s) is also shown. If used on a one-dimensional array representation of a 2D spectrum, ni and phase (in case of phase sensitive 2Ds) parameters are shown.

Examples dss(1,3)

dss(1,12,3,'green')

Related	cutoff	Data truncation limit (P)
	dssa	Display stacked spectra automatically (C
	dssan	Display stacked spectra automatically without erasing
		(C)
	dssh	Display stacked spectra horizontally (C)
	dsshn	Display stacked spectra horizontally without erasing (C)
	dssn	Display stacked spectra without screen erase (C)
	dsww	Display spectra in whitewash mode (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ho	Horizontal offset (P)
	intmod	Integral display mode (P)
	pl	Plot spectra (C)
	plww	Plot spectra in whitewash mode (C)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	shownumx	x position counting from bottom left of every spectrum
		(P)
	shownumy	y position counting from bottom left of every spectrum
		(P)
	trace	Mode for 2D data display (P)
	vo	Vertical offset (P)
	vp	Vertical position of spectrum (P)
	WC	Width of chart (P)
	wc2	Width of chart in second direction (P)

dssa Display stacked spectra automatically (C)

Syntax dssa<(<start,finish<,step>><,options>)>
Description Displays one or more spectra automatically.
Integral display is controlled by the parameter intmod when a single
spectrum is displayed (see 'int' option below). The following values
are accepted for intmod:

- intmod='off' turns off the integral display.
- intmod='full' displays the entire integral.

See also NMR Spectroscopy User Guide

• intmod='partial' displays every other integral region.

An individual trace is displayed from and arrayed 1D spectra or 2D spectra by supplying the index number as an argument. Spectra from 2D data set are displayed from either the f_1 or f_2 domain by setting the parameter trace equal to 'f1' or 'f2', respectively. Enter ft1d, trace='f1', and dss to view the interferogram. Multiple spectra are displayed by supplying indexes of the first and last spectra.

The position of the first spectrum is governed by the parameters wc, sc, and vp. For 1D data, subsequent spectra are positioned relative to the preceding spectrum by the parameters vo (vertical offset) and ho (horizontal offset). For 2D data, ho defines the total horizontal offset between the first and last spectrum.

Also for 2D data, vo is inactive while the parameter wc2 defines the total vertical offset between the first and last spectrum. To display spectra "automatically," the command dssa adjusts the parameters vo and ho to fill the screen in a lower left to upper right presentation (wc must be set to less than full screen width for this to work)

The parameter cutoff, if it exists and is active, defines the distance above and below the current vertical position vp at which peaks are truncated. By arraying cutoff to have two different values, the truncation limits above and below the current vertical position can be controlled independently. For example, cutoff=50 truncates peaks at vp+50 mm and vp-50 mm. cutoff=50,10 truncates peaks at vp+50 mm and vp-10 mm.

Arguments start is the index of the first spectra when displaying multiple spectra. It is also the index number of a particular trace to be viewed when displaying arrayed 1D spectra or 2D spectra.

finish is the index of the last spectra when displaying multiple spectra.

step is the increment for the spectral index when displaying multiple spectra. The default is 1.

options can be any of the following:

- 'all' is a keyword to display all of the spectra.
- 'int' is a keyword to only display the integral, independently of the value of the parameter intmod
- 'dodc' is a keyword for all spectra to be drift corrected independently.
- 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', and 'white' are keywords that select a color.
- 'pen1', 'pen2', 'pen2' ... are keywords that pens.
- 'nopars' prevents the display commands from drawing the parameters at the bottom of the graphics screen.

Examples dssa(1,3)

See also NMR Spectroscopy User Guide

Related cutoff Data truncation limit (P) dss Display stacked spectra (C)

dssan	Display stacked spectra automatically without erasing
	(C)
dssh	Display stacked spectra horizontally (C)
dsshn	Display stacked spectra horizontally without erasing
	(C)
dssn	Display stacked spectra without screen erase (C)
dsww	Display spectra in whitewash mode (C)
ft1d	Fourier transform along f_2 dimension (C)
ho	Horizontal offset (P)
intmod	Integral display mode (P)
pl	Plot spectra (C)
plww	Plot spectra in whitewash mode (C)
SC	Start of chart (P)
sc2	Start of chart in second direction (P)
shownumx	\boldsymbol{x} position counting from bottom left of every spectrum
	(P)
shownumy	y position counting from bottom left of every spectrum
	(P)
trace	Mode for 2D data display (P)
vo	Vertical offset (P)
vp	Vertical position of spectrum (P)
WC	Width of chart (P)
wc2	Width of chart in second direction (P)

dssan Display stacked spectra automatically without erasing (C)

Syntax	dssan<(<start,finish<,step>><,options>)></start,finish<,step>
Description	Functions the same as the command dssa except the graphics window is not erased before starting the display. This allows composite displays of many spectra to be created. The arguments are the same as dssa.
Examples	dssan(1,3)
See also	NMR Spectroscopy User Guide
Related	dssa Display stacked spectra automatically (C)

dssh Display stacked spectra horizontally (C)

Syntax	dssh<(<start,finish<,step>><,options>)></start,finish<,step>
Description	Displays one or more spectra horizontally.
	Integral display is controlled by the parameter intmod when a single spectrum is displayed (see 'int' option below). The following values are accepted for intmod:
	• intmod='off' turns off the integral display.

• intmod= 'partial' displays every other integral region.

An individual trace is displayed from and arrayed 1D spectra or 2D spectra by supplying the index number as an argument. Spectra from 2D data set are displayed from either the f_1 or f_2 domain by setting the parameter trace equal to 'f1' or 'f2', respectively. Enter ft1d, trace='f1', and dss to view the interferogram. Multiple spectra are displayed by supplying indexes of the first and last spectra.

The position of the first spectrum is governed by the parameters wc, sc, and vp. For 1D data, subsequent spectra are positioned relative to the preceding spectrum by the parameters vo (vertical offset) and ho (horizontal offset). For 2D data, ho defines the total horizontal offset between the first and last spectrum. Also for 2D data, vo is inactive while the parameter wc2 defines the total vertical offset between the first and last spectrum. To display spectra horizontally, the command dssh causes vo to be set to zero and for ho, sc, and wc to be adjusted to fill the screen from left to right with the entire array.

The parameter cutoff, if it exists and is active, defines the distance above and below the current vertical position vp at which peaks are truncated. By arraying cutoff to have two different values, the truncation limits above and below the current vertical position may be controlled independently. For example, cutoff=50 truncates peaks at vp+50 mm and vp-50 mm, and cutoff=50,10 truncates peaks at vp+50 mm and vp-10 mm.

Arguments start is the index of the first spectra when displaying multiple spectra. It is also the index number of a particular trace to be viewed when displaying arrayed 1D spectra or 2D spectra.

finish is the index of the last spectra when displaying multiple spectra.

step is the increment for the spectral index when displaying multiple spectra. The default is 1.

options can be any of the following:

- 'all' is a keyword to display all of the spectra.
- 'int' is a keyword to only display the integral, independently of the value of the parameter intmod
- 'dodc' is a keyword that causes all spectra to be drift corrected independently.
- 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', and 'white' are keywords that select a color.
- 'pen1', 'pen2', 'pen2' ... are keywords that pens.
- 'nopars' prevents the display commands from drawing the parameters at the bottom of the graphics screen.

Examples dssh(1,3)

See also NMR Spectroscopy User Guide

Related	cutoff	Data truncation limit (P)
	dss	Display stacked spectra (C)
	dssa	Display stacked spectra automatically (C)

dssan	Display stacked spectra automatically without erasing
	(C)
dsshn	Display stacked spectra horizontally without erasing
	(C)
dssn	Display stacked spectra without screen erase (C)
dsww	Display spectra in whitewash mode (C)
ft1d	Fourier transform along f_2 dimension (C)
ho	Horizontal offset (P)
intmod	Integral display mode (P)
pl	Plot spectra (C)
plww	Plot spectra in whitewash mode (C)
SC	Start of chart (P)
sc2	Start of chart in second direction (P)
shownumx	x position counting from bottom left of every spectrum
	(P)
shownumy	y position counting from bottom left of every spectrum
	(P)
trace	Mode for 2D data display (P)
vo	Vertical offset (P)
vp	Vertical position of spectrum (P)
WC	Width of chart (P)
wc2	Width of chart in second direction (P)

dsshn Display stacked spectra horizontally without erasing (C)

Syntax	dsshn<(<start,finish<,step>><,options>)></start,finish<,step>
Description	Functions the same as the command dssh except the graphics window is not erased before starting the display. This allows composite displays of many spectra to be created. The arguments are the same as dssh.
Examples	dssh(1,3)
See also	NMR Spectroscopy User Guide
Related	dssh Display stacked spectra horizontally (C)

dss1 Label a display of stacked spectra (M)

Syntax	dssl(<options>)</options>
Description	Displays a label for each element in a set of stacked spectra. The label is an integer value from 1 up to the number of spectra in the display or the values of parameters up to 2 dimensions.
	Labels can appear at incorrect positions if wysiwyg='n'. The positions are empirically determined for a large screen display and are not guaranteed to be correct for all displays.

Arguments	options co as long as th	ontrol the display (more than one option can be entered he options do not conflict with each other):			
	• 'center', 'below' are relative to e	'left', 'right', 'top', 'bottom', 'above', and e keywords setting the position of the displayed index ach spectrum.			
	• 'custom' - shownumy (- uses the parameters shownumx (x position) and y position), counting from bottom left of every spectrum.			
	• 'list=xxx' arrayed para	produces a display of the values contained in the ameter xxx .			
	• 'format=yy label. See th	• 'format=yyy' uses the format yyy to control the display of each label. See the write command for information about formats.			
	• 'reverse' values are lo sub-spectra.	• 'reverse' - rotate the text by 90° - useful if the arrayed parameter values are long with respect to the width of the individual sub-spectra.			
	• 'value' -T Diagonal arr different col- If used on a ni and phas	he values of up to two simultaneous arrays are displayed. rays are allowed. The second parameter is shown in or). The name of the arrayed parameter(s) is also shown. one-dimensional array representation of a 2D spectrum, se (in case of phase sensitive 2Ds) parameters are shown.			
Examples	dssl dssl('top' dssl('valu	,'left') ne','format=%3.1f') pssl			
See also	NMR Spectre	oscopy User Guide			
Related	dss shownumx	Display stacked spectra (C) x position counting from bottom left of every spectrum (P)			
	shownumy	y position counting from bottom left of every spectrum (P)			
	write	Write formatted text to a device (C)			

dssn Display stacked spectra without screen erase (C)

Syntax	dssn<(<start,finish<,step>><,options>)></start,finish<,step>	
Description	Functions the same as the command dss except the graphics window is not erased before starting the display. This allows composite displays of many spectra to be created. The arguments are the same as dss.	
Examples	dssn(1,3)	
See also	NMR Spectroscopy User Guide	
Related	dss Display stacked spectra (C)	

dsvast Display VAST Data in a stacked 1D-NMR matrix format

Applicability VnmrJ 3.1

Description If an array of 1D spectra have been acquired (in particular if a block of 96 spectra have been acquired using VAST automation, especially in a microtiter-plate format), and if these spectra have been glued into a reconstructed 2D dataset (see vastglue), this macro will arrange and display them (on the screen) in a convenient 8 x 12 sample format (as a matrix of 1D spectra).

Uses a file (template) created by plate_glue to display a matrix of data. The number of spectra displayed, and their order, are controlled by the template file. Each "little spectrum" is labeled with its respective alphanumeric coordinates. The modulo number controls how many spectra appear per row.

Examples dsvast(<display order>, <modulo>)
See also dsvast
 dsvast2d
 plvast
 plvast2d
 intvast
 pintvast
 plateglue
 vastglue
 vastget

dsvast2d Display VAST Data in a pseudo-2D format

Applicability	VnmrJ 3.1	
Description	If an array of 1D spectra have been acquired (in particular if a blo of 96 spectra has been acquired using VAST automation, especially a microtiter-plate format), and if these spectra have been glued into reconstructed 2D dataset (see vastglue), this macro will arrange a display them (on the screen) in a convenient pseudo-2D format (almo- like an LC-NMR chromatogram). The default is to plot all the spectra (from 1 through arraydim). A optional argument (plvast(##)) allows one to specify that only spect from 1 through ## should be plotted.	
See also	dsvast dsvast2d plvast plvast2d pintvast	

dsww Display spectra in whitewash mode (C)

Syntax dsww<(<start,finish<,step>><,'int'>)>

Description Displays one or more spectra in whitewash mode (after the first spectra, each spectra is blanked out in regions in which it is behind a prior spectra).

Arguments start is the index of the first spectra when displaying multiple spectra. It is also the index number of a particular trace to be viewed when displaying arrayed 1D spectra or 2D spectra; default is to display all spectra.

finish is the index of the last spectra when displaying multiple spectra.

step is the increment for the spectral index when displaying multiple spectra. The default is 1.

<code>'int'</code> is a keyword to display only the integral, independently of the value of the parameter intmod

Examples dsww(1,3)

Related	dss	Display stacked spectra (C)
	dssa	Display stacked spectra automatically (C)
	dssan	Display stacked spectra automatically without erasing (C)
	dssh	Display stacked spectra horizontally (C)
	dsshn	Display stacked spectra horizontally without erasing (C)
	dssn	Display stacked spectra without screen erase (C)
	pl	Plot spectra (C)
	plww	Plot spectra in whitewash mode (C)

dtext Display a text file in graphics window (M)

Syntax	<pre>dtext<(file,x,y)><:\$x_next,\$y_next,\$increment></pre>	
Description	Displays a text file in the graphics window.	
Arguments	file is the name of a text file. The default is the current experimer text file.	
	${\rm x}$ and ${\rm y}$ are coordinates of the first line of text. This positions the location of the output. The default is the upper left-hand corner of the screen.	
	x_next and y_next are the coordinates where the start of the next line would have been displayed. This is useful for subsequent character display.	
	\$increment is the increment between lines.	
Examples	dtext dtext(userdir+'/exp3/text')	

dtrig Delay to wait for another trigger or acquire a spectrum (P)

```
Applicability Systems with LC-NMR accessory.
```

- Description If ntrig is greater than 0 after a trigger is detected, a pulse sequence waits for dtrig seconds before either waiting for another trigger or acquiring a spectrum. Typically, after the LC has positioned the sample in the NMR probe and stopped the pump, there is a small time (30 seconds) during which conditions (pressure, etc.) in the NMR probe are still settling; better NMR performance is obtained if an appropriate delay is inserted using dtrig. If dtrig does not exist, a value of 0 is assumed. If dtrig does not exist, the parlc macro can create it.
 - Related ntrig Number of trigger signals to wait before acquisition (P) parlc Create LC-NMR parameters (M)

dutyc Duty cycle for homodecoupling (optional) (P)

VNMRS sys dutvc= <va< th=""><th>tems, 400 MR</th></va<>	tems, 400 MR		
dutvc= <va< td=""><td></td></va<>			
	dutyc= <value></value>		
Sets the rf decoupling. does not ex on period. rof1, rof2	Sets the rf duty cycle fraction $(0.0-0.4)$ for rf on part of homonuclear decoupling. The duty cycle default is 0.1 (or 10% rf on) if the dutyc does not exist. Homonuclear decoupling delay before and after the rf on period. homorof1, homorof2, and homorof3, are equivalent to rof1, rof2 and rof3 and all default to 2 µsec.		
0.0 to 0.4 -	– default is 0.1		
dutyc=0.2	e sets a 20% duty cycle		
homo	Homodecoupling control for observe channel (P)		
hdof	Frequency offset for homodecoupling (P)		
hdpwr	Sets the rf attenuator to control the power for		
	homonuclear decoupling (P)		
hdmf	modulation frequency for the band selective homonuclear		
	decoupling (P)		
hdpwrf	Sets the rf linear modulator fine power for homonuclear		
	decoupling (P)		
hdres	Sets the tip angle resolution (P)		
	Sets the rf decoupling. does not ex on period. rof1, rof2 0.0 to 0.4 - dutyc=0.2 homo hdof hdpwr hdmf hdpwrf hdres		
hdseq	Sets the decoupler waveform filename (P)		
----------	---		
homorof1	Delay before turning on homo decoupling rf (P)		
homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)		
homorof3	Delay between setting T/R switch to receive gating on the receiver (P) $% \left(P\right) =\left(T^{2}\right) \left(T^$		
tn	Nucleus for observe transmitter (P)		



Agilent VnmrJ 4 Command and Parameter Reference Guide

e	Eject sample (M)
ecc_on	Turns on eddy current compensation for Cold Probes (M)
ecc_off	Turns off eddy current compensation for Cold Probes (M)
echo	Simple echo command similar to unix echo
edit	Edit a file with user-selectable editor (M)
editht	Create and edit a Hadamard frequency list
editLog	Customize the log details.
editparlib	This macro has been superseded by the Clone utilities. (M)
eject	Eject sample (M)
email	Email address (P)
enter	Enter sample information for automation run (M,U)
enterdialog	Start a dialog window using enterexp file (M)
epage	Emails Output
eplot	Emails PostScript
ernst	Calculate the Ernst angle pulse (C)
errlog	Display recent error messages (C)
errloglen	Number of lines in error message display (P)
exec	Execute a command (C)
execpars	Set up the exec parameters (M)
execplot	Execute plotting macro (P)
execprep	Execute prepare macro (P)
execprescan	Execute prescan macro (P)
execproc	Execute processing macro (P)



execprocess	Execute processing macro (P)
execsetup	Execute setup macro (P)
exists	Checks if parameter, file, or macro exists and file type (C)
exit	Call the vnmrexit command (M)
exp	Find exponential value of a number (C)
expl	Display data on the screen
expladd	Add another diffusion analysis to current display (M)
explib	Display experiment library (M)
explist	Display current experiment chain and approx. time for each (M)

Eject sample (macro)

Syntax	e
Applicability	VnmrJ 3.1
Description	Turns on the eject and slow drop air to eject the sample from the probe.
Arguments	This command is valid on Mercury and GEMINI 2000 only if the optional spin control hardware is installed.

ecc_on Turns on eddy current compensation for Cold Probes (M)

Applicability Systems with Varian, Inc. Cold Prob	es
---	----

Description Turns on eddy current compensation

ecc_off Turns off eddy current compensation for Cold Probes (M)

Applicability	Systems with Varian, Inc. Cold Probes
Description	Turns off eddy current compensation.

Related ecc_on Turns on eddy current compensation for Cold Probes (M)

е

echo Simple echo command similar to unix echo

Syntax	echo[([-n,]arg1, arg2,)]
Applicability	VnmrJ 3.1
Description	This command will display strings and variable values on the output window. The echo command automatically advances to the next line after displaying (it sends a newline character).
Arguments	args can be strings surrounded by single quotes and variables. The -n option prevents the echo command from sending a newline character.
Examples	<pre>echo: Advance to next line (send newline) echo('hello'): Display string. echo('variable a=',a): Display string and variable echo('-n','Please enter a number:'): Display string without a newline.</pre>

edit Edit file or a macro with user-selectable editor

Syntax edit('myfile') - edit a file with user-selectable editor macroedit('mycmd') - edit a macro with user-selectable
editor

Applicability VnmrJ 3.1

Description The edit command will edit a file, letting you select the editor program to be used. Set the environmental parameter "vnmreditor" to be the desired editor program. The default is "vi".

You must provide a vnmr_<editor> script in the bin subdirectory of the VNMR system directory. For example, if "emacs" is to be used, a script named "vnmr_emacs" would need to be present. The major task for this script is determining if a GUI is in use and making required adjustments. The scripts "vnmr_vi" and "vnmr_textedit" provide a mode for non-window and window-based editor interface respectively.

The command macroedit will edit a Magical macro in your personal macro library. System macros cannot be directly edited with this command; they must first be copied to your personal library first.

editht Create and edit a Hadamard frequency list.

Syntax	
Applicability	VnmrJ 3.1
Description	The editht macro opens the Edit HT Freq dialog, for interactively creating and editing a Hadamard frequency line list.
	To set up a Hadamard experiment starting from a 1D experiment, do the following:

1. First run a Proton, Carbon, or other 1D experiment, depending on the type of Hadamard experiment you wish to run (homonuclear or heteronuclear).

2. When the acquisition is finished, process and phase the spectrum.

3. Run the editht macro to open the Edit HT Freq dialog. Create a Hadamard frequency list for the nucleus of interest. Save the frequency list.

4. For a heteronuclear Hadamard experiment, run a Proton experiment, and adjust spectral width and decoupling as desired.

5. Load the desired Hadamard experiment. Check the Hadamard frequency list and other parameters.

6. Start the acquisition of the Hadamard experiment.

7. When acquisition is complete, process with proc1='ht' wft2da.

How to use the Edit HT Freq dialog.

To make a Hadamard frequency list from a 1D spectrum (step 3 above), use the buttons in the Edit HT Freq dialog.

Create Line List: Processes the current spectrum as follows:

- Fourier transform with wft. Multiplet structures can be smoothed out using line broadening.
- Create a line list using nll, greater than the current threshold.
- Keep only frequencies that are the minimum line width apart in the "Min line width" entry box (e.g. 20 Hz).

Nearest line: Place the cursor on the nearest line.

Select: Adds the current cursor position to the line list. (The cursor must be more than the minimum line width from an existing frequency in the line list.)

Remove: Removes the line nearest the cursor position from the line list.

Display: Display the frequency list. If a 1D spectrum is displayed, show the frequencies using dpf in units set by the axis parameter.

CLEAR: Clear all frequencies from the frequency list.

Save HT Frequencies: Saves the current frequency list as a Hadamard line list for the current nucleus (tn). It saves the frequency list, band width, current nucleus, spectral width, and frequency offset in a persistence file. The frequencies and other parameters are loaded from the persistence file when loading a Hadamard experiment (step 5 above).

Line List: The line list is displayed in the text entry window on the right hand side of the page. You may edit the line list directly from this window. Click the 'Set list into parameters' button to set the line list changes into the parameters. The first column of numbers is the Hadamard frequency list, e.g. htfrq1. If there is a second column of numbers, it specifies the bandwidth for each frequency in Hz.

Hz/ppm menu: Select Hz or ppm to display the line list in Hz or ppm. If Hz is selected, the line list is displayed in Hz from the center of the spectrum.

Move HT pars to exp: Move the Hadamard parameters from the current workspace to a new workspace. The workspace number is specified in the entry box.

Set list into parameters: Sets the changes from the line list text entry window into the parameters.

Import list curexp / **htfrq1.ll:** Copies a line list file from curexp into the current line list, and sets the line list into the parameters. The line list file to be copied is named after the frequency parameter, e.g.

/export/home/vnmr1/vnmrsys/exp2/htfrq1.ll

The format of the file is the same as the line list display.

Arguments htfrq1 - Hadamard frequency list in indirect dimension, in Hz from center of spectrum, or ppm.

htbw1 - Hadamard band width in indirect dimension, in Hz. It may be a single value or a list of values for each element in the htfrq1 list.

tn - nucleus used for frequency list.

Examples Example #1:

freq [Hz from center]

1172.37 327.69 - 346.37

-1292.10

In Example #1, the Hadamard frequencies are in Hz from the center of the spectrum.

Example #2:

freq [ppm]		bw	[Hz]
7.930	20		
5.819	16		
4.134	20		
1.770	20		

In Example #2, the Hadamard frequencies are in ppm, referenced to the current spectrum. The bandwidth for each frequency is also specified as 16 Hz for the second frequency, and 20 Hz for the rest. If htbwl is arrayed to two or more values in the parameter set, the values are written to the line list file. If the size of the htbwl array is smaller than the size of the htfrql array, the last value of htbwl is applied to the remaining frequencies.

In a 2D display, the Edit HT Freq dialog may be used to view the Hadamard frequency list in F1. Interactive frequency selection and display from the graphics window may be done. You may also edit frequencies from the Line List window. In a 2D display, frequencies in ppm are referenced to F1.

```
See also ht
HsqcHT
tocsyHT
getht
mht
sethtfrq1
htfrqdisp
dl1
```

editLog Customize the log details

Applicability VnmrJ 3.1, VnmrJ 3.2

Description The sqLog macro records specific events from a study queue. The messages and details of the logging are customizable with the editLog utility.

The sqLog macro is very generic. It gets all of its details from a file written by the editLog utility. This file has the same name as the macro and is in the <appdir>/templates/vnmrj/loginfo directory. sqLog saves logging information only for automation runs. The log editor can handle menus of choices. Files in templates/vnmrj/loginfo with the same name as the keyword will be used to make menus of choices to select from within the editLog editor.

Files prefixed with the name of the logging macro, for example sqLog will make a File menu specific for editLog('sqLog'). The logging macro, along with the editLog editor are very general and can be used to log other events. As an example, suppose one wants to monitor access to the VnmrJ program.

- Description The sqLog facility will record the following events: SampleStart, SampleEnd, ExpStart, ExpEnd, ExpError. Each event recorded in the logfile may may be preceded by header information. This may include things like the date, time, user, etc. This header information is customizable.
 - Examples The sqLog macro is very generic. It gets all of its details from a file written be the editLog utility. This file has the same name as the macro and is in <appdir>/templates/vnmrj/loginfo directory. For example, the current sqLog file is:

Formatting statements for automation log files.
#
1Header Date: %MOC% %DAY% %YR% at %HR%:%MIN%:%SEC%,
User: \$operator\$, Sample: \$samplename\$,
1SampleStart Start new sample at location \$loc\$.
1SampleEnd Finish sample at location \$loc\$\\####
1ExpStart Experiment \$pslabel\$ started.
1ExpEnd Experiment \$pslabel\$ complete.
1ExpError Experiment error: \$\$2\$

1ExpPrescan Prescan:

1File \$autodir\$/logfile

1Ifcondition (auto='y')

Lines starting with a hash mark (#) are comments. The first character of each non-comment line is a 1 or 0, indicating enabled or disabled. The rest of the first word, following the 1 or 0, is a keyword that is passed to the sqLog macro. The remainder of a line is the template for writing the log file. The template is passed to the chkname command for translation.

The File keyword defines where the log file will be saved. If this keyword is disabled, all of the sqLog event logging will be disabled. Disabling other keywords only disables that specific event or feature.

The Ifcondition keyword allows the logging mechanism to make decisions as to whether to log the event. For example, in the case of sqLog, we only log events during an automation run. Logging will occur only if the Ifcondition is true.

A special keyword of "None" for the Ifcondition specifies no special conditions. That is, events are always logged.

The sqLog macro is called from appropriate places in the software. It is called with the keyword as the first argument. If the template uses passed arguments, they can be passed to the sqLog macro. For example, the ExpError template includes the second argument in its templates, which contains the actual error. This would be called as:

geterror:\$err

sqLog('ExpError',\$err)

During an automation run, messages written to 'line3', which puts them into the "acqlog". If sqLog is called with no arguments but one return value, the pathname of the log file, defined by the File keyword, is returned.

If sqLog is passed an event keyword, with optional additional arguments, and requests a return value, the message will not be written into the log file (nor on line3 for automation runs), but will be returned to the calling macro. An example would be: sqLog('SampleStart'):\$res

As defined above, sqLog saves logging information only for automation runs. By changing the File attribute to your **userdir** directory, and setting the Ifcondition to None, all study queue activities will be logged, both automation and foreground.

The log editor can handle menus of choices. Files in **templates/vnmrj/loginfo** with the same name as the keyword will be used to make menus of choices to select from within the editLog editor. Files prefixed with the name of the logging macro, for example sqLog will make a File menu specific for editLog('sqLog')

The logging macro, along with the editLog editor are very general and can be used to log other events. As an example, suppose one wants to monitor access to the VnmrJ program. A "loginLog" could be made as follows.

Make a copy of the sqLog macro called loginLog.

Add a loginLog file describing the events to logged to the
<appdir>/templates/vnmrj/loginfo. An example of such a file may be:
Formatting statements for login log files.
#
IHeader Date: %MOC% %DAY% %YR% at %HR%:%MIN%:%SEC%,
User: \$operator\$
ILogin Login
ILogout Logout
IFile \$systemdir\$/acqqueue/loginLog
IIfcondition ((auto='n') and (jviewport=1))
The only remaining task is to place calls to the loginLog macro in
various other macros. In this case, one might call
loginLog('Login'):\$res fromthe bootup macro and
loginLog('Logout'):\$res
from the exit macro. If one wanted to monitor "operator" logins, one

could and additional keywords such as Operatorlogin and Operatorlogout to the above file and then call loginLog('Operatorlogin'):\$res from the operatorlogin macro and call loginLog('Operatorlogout'):\$res from the operatorlogout macro.

The following are more examples.

sqLog(event<,args>) - log automation events
sqLog(event<,args>):\$res - return automation events to calling
macro
sqLog:\$path - return log file path
editLog - Customize the log details.
See also sqLog

eject Eject sample (M)

Syntax	eject		
Description	Ejects the sample from the probe by turning on the eject air and the slow drop air. The e macro functions the same as the e macro.		
See also	NMR Spectroscopy User Guide		
Related	e Eject sample (M)		
	i Insert sample (M)		
	insert Insert sample (M)		

email Tool to Send Email

Description Called on a filename, this utility prompts for email addresses and sends the specified file. Syntax email(filename)

See also email('textfile')

enter Enter sample information for automation run (M,U)

Applicability	Systems with an automatic sample changer.			
Syntax	enter<(file<,configuration_file>)>			
Decomintion	(FION UNIX) enter <iiie> <configuration_iiie></configuration_iiie></iiie>			
Description	n Enables entry of sample information for automation runs, including sample location user information solvent used experiment or			
	experiments	to run, and arbitrary text information. enter('abc')		
	creates a dire	ectory named abc. In this directory is a file named abc,		
	which contain	ns experiment information.		
Arguments	file is the i	name of the file to be edited. The default is that enter		
	prompts for t	his information. If the file already exists, new entries are		
	appended to	it.		
	configurat	ion_file is the name of a user-supplied file that		
	customizes e	nter for local use. Several configuration files are		
	provided:			
	•enter.conf is used when defining an experiment when an			
	automation run is not currently active.auto.conf is used when defining an experiment for a current automation run. The walkup macro is provided for this style of entering samples.			
	•gilson.con	f is used with the VAST accessory.		
Examples	ples (From VnmrJ or UNIX) enter			
	(From VnmrJ	rJ)enter('mysamples')		
	(From UNIA) enter MySamples			
	MMD Smoothe	acon las Cuida las Programming		
See also	Vmmr.I Autor	scopy Oser Guide, Oser Programming, nation User Guide		
Dalatad	100010 110001			
Related	auto	Stert on outomation mun (C)		
	autogo	Start an automation run (0) Prefix for automation data file (\mathbf{P})		
	autora	Resume a suspended automation run (C)		
	autosa	Suspend current automation run (C)		
	printer	Printer device (P)		
	status	Display status of all experiments (C)		
	walkup	Walkup automation (M)		

enterdialog Start a dialog window using enterexp file (M)

Applicability	Systems with automation.		
Syntax	enterdialog		
Description	Internal macro used by enter to start a dialog window using the enterexp file in the dialoglib directory.		
See also	NMR Spectroscopy User Guide; User Programming, VnmrJ Automation User Guide		
Related	enter En (M	nter sample information for automation run I,U)	

epage Emails Output

Description Used in place of the page command, this macro directs the output to email. Syntax epage Related page, eplot

eplot Emails PostScript

Description	Used in place of the page command, this macro directs PostScript output to email.
See also	eplot
Related	page, epage

ernst Calculate the ernst angle

Syntax	ernst(t1,<90degree>)
Applicability	VnmrJ 3.1
Description	Calculate the ernst angle pulse with a guess at t1 and the 90-degree pulse calibration and sets pw. If there is a parameter $pw90$ and no second parameter is entered, $pw90$ is taken as the 90-degree pulse. An entered 2nd argument resets $pw90$.

errlog Display recent Vnmr error messages

Syntax errlog

-		، بد
orr	$1 \circ \alpha$	Setr
CTTT	LUY.	JOLT

Applicability	VnmrJ 3.1
Description	The errlog command displays the most recent VNMR error messages in the alphanumeric (dg) window. If supplied with a return value, errlog:\$str will return the last message displayed on line 3. A call to errlog:\$str will clear the last message. That is, if two sequential calls are made to errlog:\$str, the second call will always return a blank string.
	The errlog command displays the most recent VNMR error messages in the alphanumeric (dg) window.
Arguments	Use the global parameter "errloglen" to control the number of lines that are displayed. If not defined, the program uses a value of 10 by default.

errloglen Number of lines in error message display (P)

Description	Sets the number of lines in the display of error messages by errlog.	
Values	Integer, default is 10.	
See also	NMR Spectroscopy User Guide	
Related	errlog Display recent error messages (P)	

exec Execute a VNMR command

Syntax	<pre>exec('command') - execute a VNMR command exec('command'):\$ret - execute a VNMR command and report success or failure</pre>
Applicability	VnmrJ 3.1
Description	The exec command allows an arbitrary VNMR command or macro to be executed. It lets a macro construct a character string which is a VNMR command or macro and then execute that command or macro.
	Some macros and commands abort. This causes the calling macro to also abort. By using exec with a return value, whether or not the called macro aborted or not is returned as a macro variable. The calling macro is not aborted.
	For example, in the simple macro macroB
	write('line3','got to here')
	If macroB aborts, the write command is not executed and the calling macro aborts.
	If a return argument is given to $exec$, it will be set to 0 if the called macro aborts and it will be set to 1 if the called macro does not abort. For example, in the following macro
	exec('macroB'):\$ret

if (\$ret = 0) then

write('line3','macroB aborted')

else

write('line3','macroB did not abort')

endif

one or the other write commands will execute, depending on whether macroB aborts. The calling macro does not abort, but continues executing its instructions.

The aborton and abortoff mechanism can also control whether or not the calling macro aborts if its called macro (macroB in the above examples) aborts. However, continued execution of the called macro is not guaranteed. For example,

abortoff

macroB

aborton

write('line3','got to here')

will often execute the write command, whether or not macroB aborts. However, if macroB calls aborton and subsequently aborts, or if macroB calls another macro that calls aborton, and one of those macros aborts, then the calling macro will abort before the write command is executed. Using the exec command with a return argument, as in exec('macro'):\$ret, guarantees that execution of the calling macro will continue.

Examples exec(\$cmdstr):\$ret - execute the contents of \$cmdstr as a VNMR command

execpars Set up the exec parameters (M)

Description Set up the exec parameters as listed in /vnmr/execpars. See also User Programming

Related	apptype	Application type (P)
	execplot	Execute plotting macro (P)
	execprep	Execute prepare macro (P)
	execprescan	Execute prescan macro (p)
	execproc	Execute processing macro (P)
	execsetup	Execute setup macro (P)

execplot Execute plotting macro (P)

Description Defines which plotting macro to use to plot this experiment.

See also	User Programming	
Related	apptype	Applica
	plot	Autom

Application type (P) Automatically plot spectra (M)

execprep Execute prepare macro (P)

Description	Defines which prepar	re macro to use to prescan this experiment.
See also	User Programming	
Related	apptype	Application type (P)
	acquire	Acquire data (M)
	plot	Automatically plot spectra (M)

execprescan Execute prescan macro (P)

Description	Defines which presca	n macro to use t	to prescan	this	experiment
See also	User Programming				
Related	apptype	Application type	e (P)		
	acquire	Acquire data (M	[)		

execproc Execute processing macro (P)

Description	Defines which proces	sing macro to use to process this experiment.
See also	User Programming	
Related	apptype	Application type (P)
	acquire	Acquire data (M)

execprocess Execute processing macro (P)

DescriptionDefines which processing macro to use to process this experiment.See alsoUser Programming

execsetup Execute setup macro (P)

Description Defines which setup macro to use to prescan this experiment.

See also	User Programming	
Related	apptype	Application type (P)
	cqexp	Load experiment from protocol (M)
	sqexp	Load experiment from protocol (M)

exists Checks if parameter, file, or macro exists and file type (C)

Syntax exists(name, 'parameter' [, tree]):\$x - does a parameter exist? exists(name,'file'<,perm>):\$x - does a file exist? exists(name, 'ascii'):\$x - is a file an ASCII text file exists(name, 'directory'):\$x - is a file a directory exists(name, 'parlib'):\$x,\$path - does a parlib entry exist exists(name, 'psglib'): \$x, \$path - does a psglib entry exist exists(name, 'command'): \$x - does a command or macro exist? exists(name, 'maclib'): \$x - does a macro exist? exists(name,directory<,'errval'>):\$x - does a file or directory exist in one of the "applications directories" Applicability VnmrJ 3.1 Description Allows checking for the existence of a parameter, file, command, parlib entry, or macro from within a macro. Allows checking if a file is an ASCII text file or is a directory. Returns 1, if file or parameter exists, or the query is true; else 0. If the 'parameter' keyword is used, an optional variable tree name can be supplied. The variable trees are 'current', 'global', 'processed', 'usertree', and 'systemglobal'. The default tree is 'current'. If the 'file' keyword is used, an optional permission test can be supplied. Without the permission test, simple existence of the file is checked. Access permission can be checked by passing the character \mathbf{r} for read permission, w for write permission, and x for execute permission. One, two, or three characters can be passed in a single argument. For example, exists('/vnmr/conpar','file','rw') checks not only that the file /vnmr/conpar exists, but also that the current user has read and write access to that file. The ascii option checks if the named file is an ascii file. The directory option checks if the named file is a directory. The parlib name will be searched for. If it is not found, a .par will be appended and the appended name will be searched for. The parlib option will also return the absolute path of the parameter set. The

search path for parlib is defined by the VnmrJ administrator interface, using the "applications directories", or appdirs.

The psglib name will be searched for. If it is not found, a .c will be appended and the appended name will be searched for. The psglib option will also return the absolute path of the parameter set. The search path for psglib is defined by the VnmrJ administrator interface, using the "applications directories", or appdirs.

Macros may reside in various places, as determined by the "applications directories", or appdirs. Typical places include the users vnmrsys/maclib directory and /vnmr/maclib.

When macros are executed, the appdirs are searched in order. Exists will return a 0 if the macro is not found in any of the appdirs. It will return a 1, 2, or larger integer, depending on if it is found in the first, second, third, etc appdir.

The command keyword is very similar to the maclib keyword, except that it firsts checks to see if the name represents a built-in Vnmr command.

If the name is neither a built-in command nor a macro, exists will return a 0. If the name represents a built-in command, exists will return a 1. If name is a macro, exists will return either 2, 3, 4, or 5. The return value identifies in which directory the macro is located. The number is 1 greater than the value returned by the maclib keyword. That is, if the command

exists('macroname','maclib'):r1

sets r1 equal to 1, then the command

exists('macroname','command'):r1

will set r1 equal to 2.

The exists command with the maclib keyword is a specific case of a general mechanism to search for files and directories in the "applications directories", or appdirs. The first argument to exists is a file name and the second argument is any subdirectory in an appdir. For example, the second argument could be the following:

shapelib	-	to search for shapes.
manual	-	to search for manuals
probes	-	to search for probes
shims	-	to search for shims

It can be any directory in an appdir. It need not be a standard directory. For example, it could be bin to search for standalone executable programs. One could execute these standalone executable programs using a construction along the following lines.

exists(\$myprog,'bin'):\$e,\$myprogPath

if (\$e) then

shell(\$myprogPath):\$res

else

write('line3','%s: Program %s has not been installed',\$0,\$myprog) endif The second argument to exists can be set to " to search for files in the top-level of the appdirs. For example,

exists('pulsecal','')

will search for pulsecal in the top-level of all appdirs. The directory name can also be multi-level, as in

exists(probename,'probes/'+probe)

The first argument may also be set to ", in which case exists will check for directories in the appdirs.

This generic form of exists will return one or two values to the calling macro. The first return value is an integer indicating in which appdir the file is found. The exists command will return a 0 if the file is not found. It will return a 1, 2, or larger integer, depending on if it is found in the first, second, third, etc appdir. An optional third argument can be provided. This will be the return value if the file is not found. For example,

exists('nomacro', 'maclib',-1):\$ok

will set \$ok to -1 if the "nomacro" does not exist in any of the appdirs. This can be used by the interface designed so that a button may be either "graved out" of removed if a macro or some other file does not exist.

The second optional return value is the absolute path to the found file. If the file does not exist, the second return value will not be set.

See the which macro for an example on the use of the command See also keyword.

Macro to call vnmrexit exit

Syntax	exit
--------	------

vnmrexit

Applicability

Description The command vnmrexit exits from the vnmr system in a graceful manner. It writes parameters and data to the disk, removes lock files and restores the terminals(if on a GraphOn). The macro exit calls the command vnmrexit to exit from vnmr. As a macro, exit provides a user some flexibility in defining other things to do when exiting.

Determine if the experiment has an active acquisition expactive

Syntax expactive(n)<:\$ans> - determine if experiment n has an active acquisition expactive('user')<:\$ans> - determine if current user has an active or queued experiment expactive('auto')<:\$ans> - determine if system is in

VnmrJ 3.1

Е

automation mode expactive('current')<:\$ans> - determine current active experiment number and user expactive<:\$ans> - determine if current experiment has an active acquisition

Applicability VnmrJ 3.1

Description expactive will determine whether an acquisition is active or pending in the current experiment. An experiment number n, where n is a number from 1 to 9999, may be supplied to expactive to determine if an acquisition is active or pending in experiment n.

- Arguments Without a return argument, expactive displays the results on line 3. If a return argument is appended to the expactive command, it will be set to the following:
 - •-1 acquisition is not possible (for example, it is a data station)
 - •0 no acquisition is active in the requested experiment
 - •1 an acquisition is active in the requested experiment
 - 2 or larger if an acquisition is queued in the requested experiment. Subtract 1 from the value to determine its position in the acquisition queue.

If the keyword 'user' is supplied as an argument, expactive will determine if the current user has an active or queued experiment. Without a return argument, expactive('user') displays the results on line 3. If a return argument is appended to the

expactive('user') command, it will be set as in the case above.

If the keyword 'auto' is supplied as an argument, expactive will determine if the system is in automation mode. Without a return argument, expactive('auto') displays the results on line 3. If a return argument is appended to the expactive('auto') command, it will be set to 1 if the system is in automation mode, 0 otherwise.

If the keyword 'current' is supplied as an argument, expactive will determine which experiment, if any, has an active acquisition command running. Without a return argument, expactive('current') displays results on line 3. An experiment is still considered active if it holds up additional acquisitions during its wexp processing by means of the 'wait' flag. If a return argument is appended to the expactive('current'):\$exp command, it will be set to the following:

- •-1 acquisition is not possible (for example, it is a data station)
- •0 no acquisition is active
- n an acquisition is active in experiment "n"

If a second return argument is appended to the

expactive('current'):\$exp,\$user

command, the second argument will be set to the user that started the acquisition. If the system is running in automation mode, this second argument will be set to 'auto'. If no acquisition is running, this second argument will be set to 'nobody'.

expfit Unix program for making a least squares fit to a polynomial or exponential curve.

Syntax expfit option(s) <analyze.inp >analyze.list

Applicability VnmrJ 3.1

Description The program expfit does a least-squares curve fitting to the data supplied in 'analyze.inp'. Macros are available for the specialized uses of analyze such as 't1' and 'kinetics'. They avoid the need for the user to select options and get the correct file format. In the regression mode, the type of curve fitting, ('poly1',...) must be selected. For regression (generalized curve fitting), the regression section in the Operation Manual gives the input file format and describes the menus that permit options choices indirectly through menu buttons. **Files**

The text file analyze.inp which for t1, t2, kinetics, contact_time, and regression, contains:

<optional descriptive text line>

<optional y-axis title - regression only>

number of peaks(data sets) number of (x,y) pairs per peak and,

regression only, x scale type y scale type

<NEXT number of (x,y) pairs for this peak >

peak index

x y (first peak,first pair)

x y (first peak, second pair)

•••••

<NEXT number of (x,y) pairs for this peak > peak index

x y (second peak, first pair)

.....

In the regression mode the line beginning with 'NEXT' is inserted at the start of each data set when the number of pairs per peak is variable. In this case the header contains the maximum number of pairs per peak. For t1, t2, kinetics, and contact_time, information from the file 'fp.out' and from the array 'xarray' are used to construct this file, therefore, it is necessary to run 'fp' prior to 'analyze'. For regression, this file is made by running "expl('regression')". For 'diffusion', 'contact_time', and, if not in regression mode, poly1 and poly2, it is slightly different:

List of <number> x-y data pairs (6 strings)

<Descriptive text line>

<X-values> <Y-values> (2 strings without blanks)

x y (first peak, first pair) (continues as above)

'expfit' also makes a file 'analyze.out', which is used by 'expl' to display the results of the analysis in addition to output to the standard output which is usually directed to 'analyze.list'.

Options

The following options are implemented in 'analyze':

t1, ,Perform T1 analysis (default)

t2Perform T2 analysis

kinetics Perform kinetics analysis decreasing peak height

increment Perform kinetics analysis with increasing peak height listExtended listing for each peak diffusion A special analysis for diffusion experiments contact_time A special analysis for solids cross-polarization spin-lock experiments regression Sets regression mode, signifies generalized curve fitting with choices poly1, poly2, poly3, and exp poly0With regression, calculates mean poly1With regression, a linear fitting poly2With regression, a quadratic fitting poly2With regression, a cubic curve fitting expWith regression, an exponential curve fitting xamples expfit d2 T1 list <analyze.inp >analyze.out

Examples expfit d2 T1 list <analyze.inp >analyze.out expfit regression exp list <analyze.inp >analyze.out

exp1 Display data on the screen

Syntax expl- display the data for all lines on the screen
expl(line#, line#,..)- display selected lines only
expl('regression',line#,..) - display selected data sets
for regression analysis

Applicability VnmrJ 3.1

Description Display or plot exponential curves resulting from t1, t2, or Kinetics analysis. Display or plot of Polynomial Curves from Diffusion or other type of analysis. No argument displays first 8 curves if that many along with the data points. Otherwise selected curves are plotted. sc, wc, sc2, and wc2 control the size of plot.

Options

'regression' signifies the beginning of generalized curve fitting. Expl displays the data in 'regression.inp' as unconnected points, and also uses 'regression.inp' to create the file 'analyze.inp', which serves as input to 'analyze' for curve fitting.

'linear', 'square', 'log' provide for plotting of the data points against the square or log of the data. The first keyword controls x-axis scale, the second keyword controls the y-axis. Default is to 'linear'.

'link' causes the data points to be connected rather than a plot of the theoretical curve.

'nocurve' produces a plot of data points only.

'tinysymbol' produces a plot with small-scale data point symbols. 'nosymbol' produces a plot of the curve only.

'noclear' does not erase the graphics screen before drawing the plot. 'oldbox' is used to plot an additional curve on an existing plot. Only the first data set in analyze.out is plotted. It causes the program to get box and scale description from expl.out in the current experiment. When the 'oldbox' option is used, a required second argument identifies the curve number and data point symbol, which will be used to represent the data.

This second argument is a number from 1 to 8.

'file' followed by a filename replaces analyze.out as the input to expl. Files: 'analyze.out' file is the data input file except for regression when it is 'regression.inp'. 'expl.out' saves certain display/plot parameters. Format for regression input, 'regression.inp': Text Line (Optional) Second text line (Optional) displayed along Y scale nsets npairs <NEXT> ху (first set, first pair) (first set, second pair) ху <NEXT> ху (second set, first pair) The optional text lines must not begin with a digit. The line beginning with 'NEXT' is inserted at the start of each data set when the number of pairs per peak is variable. In this case, set 'nsets' and 'npairs' to 0. Limits: 2048 points maximum from a data set. 2048 points maximum from all sets displayed/plotted. 8 data sets maximum displayed/plotted. 128 data sets maximum are read. Examples expl- display from the first up to the sixth curve with data points from 'analyze.out' expl(1,3,6)- display curves with indexes 1, 3, and 6 from 'analyze.out' with data points expl(1,3,6)- plot the data expl('regression')- display the data in the first up to the sixth data set in 'regression.inp' expl('regression',4,5)- display the data in the fourth and the fifth data set in 'regression.inp' See expl in the Commands Manual for the file format of

See also See expl in the Commands Manual for the file format of analyze.out

expladd Add another diffusion analysis to current display (M)

Applicability	Systems with the diffusion option.
Syntax	expladd(integral_region)
Description	Adds results of another diffusion analysis to the currently displayed results.
Arguments	integral_region specifies the number of the region whose results are to be added to the existing graph.
Examples	expladd(1)

See also	NMR Spectroscopy User Guide		
Related	expl	Display exponential or polynomial curves (C)	
	pexpl	Plot exponential or polynomial curves (C)	
	pexpladd	Add another diffusion analysis to current plot (M)	

explib Display experiment library (M)

Syntax	explib
Applicability	VnmrJ 3.1
Description	Displays the currently available experiment files. For each experiment, explib displays the name of the experiment and its subexperiments, whether an acquisition is active or its position in the acquisition queue, the current size of the experiments, the pulse sequence currently active in the experiments, and the first 50 characters of the text file in the experiment. explib also displays a message if the system is in automation mode.
See also	NMR Spectroscopy User Guide; VnmrJ Automation User Guide

explist Display current experiment chain and approx. time for each (M)

See also Displays approximate time for each experiment in a chained experiment. Related autotime Display approximate time for automation (M)

explog Display an experiment's log file

Applicability VnmrJ 3.1

Description Each acquisition generates a log file which includes when the experiment started, any acquisition errors which may have occurred, and when the experiment finished. This information may be displayed with the explog macro. This information is stored in the experiment's acqfil directory in a text file named log.

exptime Display experiment time

Syntax exptime exptime('filename') Applicability VnmrJ 3.1

exptime estimates the experiment time for the current seqfil, using
the parameters in the current experiment. "exptime('filename')"
estimates the experiment time of the specified filename.
exptime exptime('s2pul')



Agilent VnmrJ 4 Command and Parameter Reference Guide

f	Set display parameters to full spectrum
f19	Automated fluorine acquisition (M)
f19p	Process 1D fluorine spectra (M)
flcoef	Coefficient to construct F1 interferogram (P)
f2coef	Coefficient to construct F2 interferogram (P)
fastuserlogin	Gateway macro for fastuserlogin function. (M)
fattn	Fine attenuator (P)
fb	Filter bandwidth (P)
fbc	Apply baseline correction for each spectrum in an array (M)
fdm1	Set, write 1D FDM parameters, run FDM (M)
fid_scan	Start up the interactive acquisition display process
fiddc3d	3D time-domain dc correction (P)
fiddle	Perform reference deconvolution (M)
fiddle_examples	Illustrates some of the simple ways that fiddle can be used to extract worthwhile results from poor quality data
fiddled	Perform reference deconvolution subtracting alternate FIDs (C)
fiddleu	Perform reference deconvolution subtracting successive FIDs (C)
fiddle2d	Perform 2D reference deconvolution (C)
fiddle2D	Perform 2D reference deconvolution (C)
fiddle2dd	2D reference deconvolution subtracting alternate FIDs (C)
fiddle2Dd	2D reference deconvolution subtracting alternate FIDs (C)



fidmax	Find the maximum point in an FID (C)
fidpar	Add parameters for FID display in current experiment (M)
fidsave	Save data (M)
fifolpsize	FIFO loop size (P)
file	File name of parameter set (P)
files	Interactively handle files (C)
filesinfo	Return file information for files display (C)
filtfile	File of FIR digital filter coefficients (P)
findxmlmenu	Find an xml menu (M)
fitspec	Perform spectrum deconvolution (C, U)
fixgrd	Convert gauss/cm value to DAC (M)
fixpar	Correct parameter characteristics in experiment (M)
fixpar3rf	Create parameters for third rf channel (M)
fixpar4rf	Create parameters for fourth rf channel (M)
fixpar5rf	Create parameters for fifth rf channel (M)
fixgrdR	Converts Gradient Strength to DAC values
fixup	Adjust parameter values selected by setup macros (M)
fixpsg	Update psg libraries (M)
flashc	Convert compressed 2D data to standard 2D format (C)
flipflop	Set up parameters for FLIPFLOP pulse sequence (M)
Fluorine	Set up parameters for 19F experiment (M)
flush	Write out data in memory (C)
fn	Fourier number in directly detected dimension (P)
fn1	Fourier number in 1st indirectly detected dimension (P)
fn2	Fourier number in 2nd indirectly detected dimension (P)
fn2D	Fourier number to build up 2D DOSY display in freq. domain (P)
focus	Send keyboard focus to input window (C)

foldcc	Fold INADEQUATE data about two-quantum axis (C)
foldj	Fold J-resolved 2D spectrum about $f_1=0$ axis (C)
foldt	Fold COSY-like spectrum along diagonal axis (C)
fontselect	Open FontSelect window (C)
format	Format a real number or convert a string for output (C)
fp	Find peak heights or phases (C)
fpi	Report integral values from arrayed spectra. (M)
fpmult	First point multiplier for np FID data (P)
fpmult	First point multiplier for "np" FID data
fpmult1	First point multiplier for ni interferogram data (P)
fpmult2	First point multiplier for ni2 interferogram data (P)
fr	Full recall of a display parameter set (M)
framecmd	Create a new frame of image, text, and inset with 'new' option
fread	Read parameters from file and load them into a tree (C)
fsave	Save parameters from a tree to a file (C)
fsq	Frequency-shifted quadrature detection (P)
ft	Fourier transform 1D data (C)
ft1d	Fourier transform along f_2 dimension (C)
ft1da	Fourier transform phase-sensitive data (M)
ft1dac	Combine arrayed 2D FID matrices (M)
ft2d	Fourier transform 2D data (C)
ft2da	Fourier transform phase-sensitive data (M)
ft2dac	Combine arrayed 2D FID matrices (M)
ft3d	Perform a 3D Fourier transform on a 3D FID data set (M,U)
ftargs	Macro to create parameters
full	Set display limits for a full screen (C)
fullsq	Display largest square 2D display (M)
fullt	Set display limits for a full screen with room for traces (C)

Set display parameters to full spectrum

Syntax	f
Applicability	VnmrJ 3.1
Description	This commands sets the display parameters "sp" and "wp" up for a full display of a 1D spectrum. If an FID is displayed, the parameters "sf" and "wf" will be set for a full display. In multi-dimensional data sets, the parameters for both displayed dimensions will be set up. For 2D data sets, the parameters "sp", "wp", "sp1", and "wp1" would be set. For planes of higher dimensional data sets, the appropriate two groups
	of sp wp, sp1 wp1, and sp2 wp2, parameter pairs will be set.

£19 Automated fluorine acquisition (M)

Syntax	f19<	(solvent)	>
--------	------	-----------	---

Description Prepares parameters for automatically acquiring a standard ¹⁹F spectrum. The parameter wexp is set to 'procplot' for standard processing. If f19 is used as the command for automation via the enter program, then the macro au is supplied automatically and should not be entered on the MACRO line of the enter program. However, it is possible to customize the standard f19 macro on the MACRO line by following it with additional commands and parameters. For example, f19 nt=1 uses the standard f19 setup but with only one transient.
Arguments solvent is the name of the solvent. In automation mode, the solvent is supplied by the enter program. The default is 'CDC13'
Examples f19

f19('DMSO')

See also NMR Spectroscopy User Guide

Related	au	Submit experiment to acquisition and process data (M)
	enter	Enter sample information for automation run (C)
	f19p	Process 1D fluorine spectra (M)
	proc1d	Processing macro for simple (non-arrayed) 1D spectra
		(M)
	procplot	Automatically process FIDs (M)
	wexp	When experiment completes (P)

f19p Process 1D fluorine spectra (M)

Description Processes non-arrayed 1D fluorine spectra using a set of standard macros. f19p is called by procld, but can also be used directly. Fully automatic processing (up to a point where a spectrum could be plotted) is provided: Fourier transformation (using preset weighting functions),

f

	automatic phasing (aphx macro), select integral regions (hregions macro), adjust integral size (integrate macro), vertical scale adjustment (vsadjc macro), avoiding excessive noise (noislm macro), threshold adjustment (if required, thadj macro), and referencing to the TMS signal, if present (tmsref macro).		
See also	NMR Spectroscopy User Guide		
Related	aphx	Perform optimized automatic phasing (M)	
	£19	Automated fluorine acquisition (M)	
	hregions	Select integral regions for proton spectra (M)	
	integrate	Automatically integrate 1D spectrum (M)	
	noislm	Avoids excessive noise (M)	
	proc1d	Processing macro for simple (non-arrayed) 1D spectra	
		(M)	
	thadj	Adjust threshold (M)	
	tmsref	Reference spectrum to TMS line (M)	
	vsadjh	Adjust vertical scale for proton spectra (M)	

flcoef Coefficient to construct F1 interferogram (P)

Description	Holds the coefficient to construct an F1 interferogram for 2D and 3D		
	transformation. Coefficients are used by the ft2da and ft3d macros.		
	If flcoef has a null value, ft2da uses the "standard" coeffic		
	flcoef is created by the par2d macro.		
Values	lues Series of coefficients, separated by spaces (not a comma), ar		
	as a string va	ariable. For example, the coefficient for standard	
	States-Hypercomplex data set is flcoef='1 0 0 0 0 0 -1 0'.		
See also	NMR Spectroscopy User Guide		
Related	f2coef	Coefficient to construct F2 interferogram (P)	
	ft2da	Fourier transform phase-sensitive data (M)	
	ft3d	Perform a 3D Fourier transform on a 3D FID data set	
		(M,U)	
	make3dcoef	Make 3D coefficients file from 2D coefficients (M)	
	par2d	Create 2D acquisition, processing, display parameters	
		(M)	

f2coef Coefficient to construct F2 interferogram (P)

Description	Holds the coefficient to construct an F2 interferogram for 2D and 3D
	transformation. Coefficients are used by the ft2da('ni2') and ft3d
	macros. If f2coef has a null value, ft2da('ni2') uses the
	"standard" coefficients. f2coef is created by the par3d macro.
Values	Series of coefficients, separated by spaces (not a comma), and stored as a string variable. For example, the coefficient for standard States-Hypercomplex data set is $f2coef='1 \ 0 \ 0 \ 0 \ 0 \ -1 \ 0'$.

fastuserlogin Gateway macro for fastuserlogin function (M)

Syntax	
Applicability	VnmrJ 3.1
Description	On systems with VnmrJ 3.1 and above, this macro manages the FASTuser switch located in VnmrJ-User Preferences. Enabling the FASTuser switch allows users to quickly login and logout of VnmrJ.
Arguments	
Examples	
See also	

fattn Fine attenuator (P)

Description	Configuration parameter for whether the current rf channel has a fine attenuator. The value is set using the label Fine Attenuator in the Spectrometer Configuration window (opened from config).		
Values	 0 specifies the fine attenuator is not present on the channel (Not Present choice in Spectrometer Configuration window). 4095 specifies the fine attenuator is present on the channel (Present choice in Spectrometer Configuration window). 		
See also	VnmrJ Installation and Administration; User Guide: Solids; CP/MAS Installation		
Related	config	Display current configuration and possibly change it (M)	
	dpwrf	First decoupler fine power (P)	
	tpwrf	Observe transmitter fine power (P)	

fb

Filter bandwidth (P)

Description Sets the bandwidth of the audio filters, which prevents noise of higher frequency than the spectral limits from "folding in" to the spectrum. Because the transmitter is in the center of the spectrum, the range of audio frequencies that must be filtered out is half the spectral width sw (e.g., for a spectral width of 4000 Hz, frequencies higher than ±2000 Hz should be filtered out). The audio filters have some attenuation at frequencies lower than their nominal cutoff frequency, which is the frequency at which signals have been attenuated by 3 dB (50%). This impacts on quantitative accuracy near the edges of the spectrum so that the standard value of fb is 10% more than half of sw.

fb is automatically changed whenever the spectral width sw is changed and thus is normally not a user-entered parameter. For example, typing sw=4000 automatically sets fb=2200, which is 10% more than 2000 Hz. After changing the value of sw, fb can be changed.

Values	if <mark>sw</mark> i	s 500,000 or less: 1000 to 256000 Hz, 1000-Hz steps.
	if <mark>sw</mark> i	s greater than 500,000: 256 kHz, 1 MHz.
See also	NMR S	Spectroscopy User Guide
Related	SW	Spectral width in directly detected dimension (P)
	mrfb	Set the filter bandwidths for multiple receivers (P)

fbc Applies 'bc' type baseline correction to all the spectra in an array

Syntax	fbc	
Applicability	VnmrJ 3.1	
Description	The macro fbc applies 'bc' type baseline correction to all the spectra in an array. The partial integral mode should be used to set integral regions to include all significant signals, while leaving as large an area of baseline as possible blank. This minimises systematic errors in diffusion coefficient fits caused by baseline errors.	

fdm1 Set, write 1D FDM parameters, run FDM (M)

Syntax	fdm1<(filename<	,n1, v1<, n2, v2<>>>)>
	fdm1 (i) for the i	i-th trace
Description	Sets 1D Filter Diagonalization Method (FDM) parameters to the default values, writes the parameters to the curexp/datdir/fdm1.inparm file, and runs a stand-alone C++ program (/vnmr/bin/fdm1d).	
Arguments	filename is the FID file; the default is curexp+'acqfil/fid'.	
	n1, n2 is one arbitrary):	or more following variable names (the order is
	axis cheat cheatmore error fidfmt fdm fn_Sp1D	 -1 (default) to reverse the spec. No cheat if cheat=1, lines are narrower if cheat<1. No cheatmore if cheatmore=0. Error threshold for throwing away poles. FID format: VnmrJ or ASCII. 1 for FDM; -1 for Digital or Discrete Fourier Transform. Spectrum file; default is
	Gamm Gcut idat	curexp/datadir/fdm1.parm. Smoothing width (line broadening). Maximum width for a pole. Data type of ASCII FID file –4 for complex data, ignored if
	i_fid kcoef	<pre>data is in VnmrJ format. The i-th trace of the FID. If kcoef > 0, use 'complicated' dk(k)1 is</pre>
	Nb Nbc	always preferred. Number of basis functions in a single window. Number of coarse basis vectors.

F

	Npower Nsig Nskip par rho specfmt spectyp ssw t0 theta wmax wmin	Number of spectrum data points. Number of points to use. Number of points to skip. Line list file; default is curexp/datadir/fdm1.parm rho=1 is optimal. Spec format: VnmrJ or ASCII. Spectrum type: complex (default), real imag, or abs. A test parameter. Delay of the first point. Overall phase of FID (rp in radians). Maximum spectrum frequency in hertz. Minimum spectrum frequency in hertz.
	v1, v2 is th	e value for the variable(s).
Examples	fdm1('cheat', 0.8)	
~ .	LUMIL (INSIG , 5	
See also	NMR Spectroscop	by User Guide

fid_scan Start up the interactive acquisition display process

Syntax fid_scan

Applicability VnmrJ 3.1

Description The interactive acquisition display process allows interactive shimming on the FID or spectrum. The pulse sequence and parameter set for the FID / spectrum display is whatever is set in the current experiment / workspace. The normal interactive tools for FID and spectral displays (df and ds commands) are available in this interactive mode. Automatic locking, shimming, steady states, and robot control are turned off by passing the 'fidscan' argument to the au command.

The mechanism used for this interactive display is based on the au / wbs tools. The fid_scan macro does an au with the bsclear and fidscan arguments and sets wbs='fid_display'. The fid_display macro does the actual data display at block size intervals. The fidscanmode parameter controls the type of display to use. It is a list of flag characters to select various options. Possible values for the "fidscanmode" parameter include:

- \mathbf{r}' displays the reals (as a trace, not in "filled" mode)
- 'i' displays the imaginaries
- •'ri' displays both the reals and the imaginaries
- •'f' displays the FID in "filled" mode. In this mode, the 'envelope' and 'dots' mode (see dotflag parameter) are not available.
- •'rf' display the "reals" in "filled" mode.
- $\bullet\,{}^{\prime}{\rm s}^{\prime}\,$ displays the spectrum
- $\bullet\,{}^{\prime}\mathrm{e}^{\prime}\,$ displays the envelope

By default, a block size of 1 is used for fidscan mode. However, this can be changed by creating and setting a 'fidshimnt' parameter. Setting the fidshimnt=1 has the special effect of turning automatic phase

cycling (i.e., oph) off. Setting fidshimnt=8, for example, will average 8 scans before the result is displayed.

Related ft3d Perform a 3D FT on a 3D FID data set

fiddc3d Flag for 3D time-domain DC correction

Syntax	fiddc3d
Applicability	VnmrJ 3.1
Description	fiddc3d is a flag whose default value is 'nnn'. fiddc3d is created by the macro `par3d` if the former does not already exist. The first character of fiddc3d in the 3-character string refers to the F3 dimension (sw,np,fn); the second character, to the F1 dimension (sw1,ni,fn1); and the third character, to the F2 dimension (sw2,ni2,fn2). Each character may take one of two values: 'n', for no time-domain DC correction along the relevant dimension. The time-domain DC correction occurs immediately after any LP (linear prediction) operations and before all other operations on the time-domain data.

Related ft3d Perform a 3D FT on a 3D FID data set

fiddle Perform reference deconvolution

Syntax	<pre>fiddle('option'[,'filename',][,'option',['filename']][,start</pre>
	no][,finishno][,increment])

Applicability VnmrJ 3.1

Description This program performs reference deconvolution, using a reference signal with known characteristics to correct instrumental errors in experimental 1D or 2D spectra. The commands can take multiple string and numeric arguments, in the format described under OPTIONS below.

Reference deconvolution of 1D spectra

Only spectra that contain a well-resolved reference signal dominated by a single component (i.e. not a simple multiplet) are suitable for reference deconvolution. Fourier transform the raw fid with ft, preferably having zero filled (i.e. set fn $\geq 2^{n}$ p). (If there are sinc wiggles, use wft with gf = at*0.6.) Set the reference line to the chosen signal using the rl command, and then use two cursors either side of the line to define a region of spectrum which includes all of the reference signal plus a little clear baseline but no other signals. This reference region will be used to define the instrumental lineshape.

Next, decide what lineshape you would like to convert the instrumental lineshape to, and set the weighting parameters accordingly. Thus if you want a 1 Hz wide Lorentzian, set lb to 1 and all other weighting

parameters to 'n'. Bear in mind the signal-to-noise ratio penalty for resolution enhancement: if the experimental line is 2 Hz wide and you set 1b=0, you get an infinitely sharp line with infinitely poor S/N. For most purposes a sensible strategy is to set lb to _minus_ the expected _natural_ linewidth, and choose gf to give reasonable S/N; this should convert the instrumental lineshape to Gaussian. Where the signals of interest are broader than those of the reference, resolution enhancement can easily be obtained by making lb more negative. Once you have set the weighting parameters, the command fiddle will carry out the reference deconvolution and display the corrected spectrum. The integral should remain unchanged, so any resolution enhancement will result in an increase in the amplitude of both signal and noise. To save the corrected data it is necessary to use the option 'writefid' when doing the reference deconvolution, e.g. fiddle('writefid','correctedfid') will store the file 'correctedfid.fid' in the current working directory.

The options 'writefid','<filename>' and 'readcf','<filename>' will write and read the correction function respectively. Thus performing reference deconvolution on one fid using fiddle with the 'writecf' option and then using fiddle with 'readcf' to process another fid will use the first correction function to correct the second fid. This can be useful for heteronuclear lineshape correction (provided that the spectral widths for the two nuclei are in the ratio of the respective magnetogyric ratios), or for correcting spectra in which a reference signal has been suppressed (e.g. an INADEQUATE spectrum could be corrected for lineshape errors using a correction function derived from the normal carbon spectrum).

To correct a series of spectra in an arrayed or 2D experiment, use numeric arguments just as with ft: fiddle(1) will correct spectrum 1, fiddle(2,3) spectra 2 and 3, and so on.

Many reference signals have satellites; for example as well as the familiar one-bond carbon-13 satellites, TMS has singlet satellite signals from coupling to silicon-29 and quartet satellites (normally unresolved) from three-bond coupling to carbon-13. For most purposes carbon-13 satellites are small enough to be ignored, but where high accuracy is required or there are stronger (e.g. silicon-29) satellites, satellite signals can be included in the specified form of the ideal reference signal by invoking the 'satellites' option. The directory/vnmr/satellites contains a file TMS which contains details of the TMS satellite signals; the command fiddle ('satellites', 'TMS') will allow for the satellite signals when deconvoluting using TMS as a reference. For information on how to construct satellite files for other reference signals, see the file /vnmr/satellites/README.

To perform corrected difference spectroscopy, use fiddled to produce the corrected difference between successive spectra (this will halve arraydim). Since the main aim of reference deconvolution here is to optimise the purity of the difference spectrum, the target lineshape would normally be chosen to give the best possible S/N; this corresponds to choosing a target lineshape approximately twice the width of the raw experimental signals of interest. The command fiddleu produces corrected differences between successive fids and the first fid.

Reference deconvolution of 2D spectra

The commands fiddle2d/fiddle2D and fiddle2dd/fiddle2Dd function in just the same way as the parent fiddle program. Since the principal objective in 2D reference deconvolution is usually the reduction of t1-noise, ideal lineshape parameters are normally chosen for optimum S/N ratio rather than resolution enhancement. To perform 2D reference deconvolution, choose fn (preferably $\geq 2^{n}$ p) and fn1, then ft the raw data (as mentioned earlier, if there is significant signal left at the end of at it may be necessary to use wft with qf set). Display the first increment with ds(1), adjust the phase of the reference signal, and use r1 to select the reference signal. In earlier versions, it was necessary to create a parameter phinc to anticipate the changes in the reference signal phase with increasing evolution time, but the current algorithm adjusts the phase automatically (unless the option 'noaph' is selected). The deconvolution will set the reference signal phase as a function of t1 so as to place the reference signal at frequency rfp1 in f1, so remember to set rfl1 and rfp1 before using fiddle2D or the f1 frequencies may change unexpectedly.

Define the reference region with the two cursors as usual, then type the command fiddle2D('writefid', '<filename>') (or fiddle2D if a 2D difference spectrum is required, as with corrected HMBC). The 'writefid' option is essential, as fiddle2D on its own does not store the corrected time-domain data. If phase-sensitive gradient-enhanced 2D data are to be processed, alternate fids will have opposite phase modulations (i.e. the experimental array will alternate N-type and P-type pathways), and the option 'alternate' should be used.

Once the deconvolution is complete, the corrected 2D fid data can be read into an experiment and processed as normal (though if fiddle2D has been used, arraydim will no longer match the arrays set and it may be necessary to set the arguments to wft2d explicitly rather than using wft2da, or adjust the parameters manually).

Arguments The options available are as follows:

- alternate: Alternate reference phase +- (for phase sensitive gradient 2D data)
- autophase: Automatically adjust phase
- displaycf: Stop at display of correction function
- fittedbaseline: Use cubic spline baseline correction defined by the choice of integral regions
- invert: Invert the corrected difference spectrum/spectra
- noaph: Do not automatically adjust zero order phase of reference region
- nodc: Do not use dc correction of reference region
- nohilbert: Do not use Hilbert transform algorithm; use extrapolated dispersion mode reference signal unless option ...

- normalise: Keep the corrected spectrum integrals equal to that of the first spectrum
- readcf: Read correction function from file '<filename>'; the argument 'filename' must immediately follow 'readcf'
- satellites: Use satellites defined in '<filename>' in ideal reference region; '<filename>' should be in /vnmr/satellites
- stop1: Stop at display of experimental reference fid
- stop2: Stop at display of correction function
- stop3: Stop at display of corrected fid
- stop4: Stop at display of first corrected fid
- verbose: Display information about the course of the processing in the main window
- writecf: Write correction function to file '<filename>'; the argument 'filename' must immediately follow 'writecf'
- writefid: Write out corrected fid to '<filename>'; if '<filename>' does not begin with / it is assumed to be in the current working directory
- See also J. Taquin, Rev. Physique App., 14 669 (1979).

G.A. Morris, JMR 80 547 (1988).

G.A. Morris & D. Cowburn, MRC 27 1085 (1989).

A. Gibbs & G.A. Morris JMR 91 77 (1991).

A. Gibbs, G.A. Morris, A.G. Swanson and D. Cowburn, J.Magn.Reson. 101, 351-356 (1993).

G.A. Morris, in Chapter 16 of "Signal Treatment and Signal Analysis in NMR", ed. D.N. Rutledge, Elsevier, 1997.

G.A. Morris, H. Barjat and T.J. Horne, Prog. NMR Spectrosc., 31, 197 (1997).

RelatedfiddledPerform subtracting alternate fidsfiddleuPerform subtracting successive fids from the firstfiddle2DPerform 2D reference deconvolutionfiddle2ddPerform 2D reference deconvolution subtracting
alternate fids

fiddle_examplesIllustrates some of the simple ways that fiddle can be used to extract worthwhile results from poor quality data

ApplicabilityVnmrJ 3.1DescriptionThis is a small collection of fids recorded on an old XL300 and
converted to Vnmr format, and illustrates some of the simple ways that
fiddle can be used to extract worthwhile results from poor quality
data. The three files are:
mixture: a mixture of acetone and ethanol in CDCl3, with very poor
shimming and severe spinning sidebands ODCB: a (folded) spectrum of a sample containing ODCB and TMS, again recorded with very poor shimming and severe spinning sidebands NOED: an arrayed pair of fids for an NOE difference experiment with gated irradiation [see Magn.Reson. Chem. 27, 1085-1089 (1989)], this time with OK shimming (but nasty decoupler spikes) To try out fiddle with these files, simply load them into an experiment, type text and follow the instructions displayed.

fiddled Perform reference deconvolution subtracting alternate FIDs (C)

Description	Produces the corrected difference between successive spectra. Refer to		
	the descrip	tion of fiddle for details.	
See also	NMR Spectroscopy User Guide		
Related	fiddle	Perform reference deconvolution	

fiddleu Perform reference deconvolution subtracting successive FIDs (C)

Description	Produces corrected differences between successive FIDs and the first
	FID. Refer to the description of fiddle for details.
See also	NMR Spectroscopy User Guide

Related fiddle Perform reference deconvolution

fiddle2d Perform 2D reference deconvolution (C)

DescriptionFunctions the same as the fiddle program except fiddle2d performs
2D reference deconvolution. Refer to the description of fiddle for
details.See alsoNMR Spectroscopy User Guide
Related fiddleRelatedfiddlePerform reference deconvolution

fiddle2D Perform 2D reference deconvolution (C)

Description Functions the same as the fiddle program except fiddle2D performs 2D reference deconvolution. Refer to the description of fiddle for details.

See alsoNMR Spectroscopy User GuideRelatedfiddlePerform reference deconvolution

fiddle2Dd 2D reference deconvolution subtracting alternate FIDs (C)

Description	Functions the same as the fiddle program except fiddle2Dd	
	performs 2D reference deconvolution. Refer to the description of	
	fiddle for details.	
See also	NMR Spectroscopy User Guide	
Related	fiddle Perform reference deconvolution	

fidmax Find the maximum point in an FID

Syntax	fidmax<(trace)>:\$max		
Applicability	VnmrJ 3.1		
Description	fidmax finds the absolute maximum value in an FID. With no arguments, fidmax uses the currently active FID, selected by df or select. A FID index may be supplied as an optional argument. For data collected using nf > 1, if cf is active, then the maximum of only that cf element will be returned. If the cf parameter is "off", then the maximum of all cf elements will be returned. Note that the maximum value returned by fidmax is divided by the value of 'ct'.		
Examples	fidmax:\$max		
	fidmax(1):\$max		
	fidmax(arraydim):\$max		

fidpar Add parameters for FID display in the current experiment

Syntax	fidpar		
Applicability	VnmrJ 3.1		
Description	All new parameter sets have the FID display parameters dotflag,		
	axisf, vpf, vpfi, crf, and deltaf defined. Old parameter sets may		
	not have these parameters defined. The macro fidpar is provided to		
	create all these FID display parameters in the current experiment.		

fidsave Save data (M)

Description Macro to save data. It uses svfdir and svfname to construct the data filename.

fifolpsize FIFO loop size (P)

Description Configuration parameter for the size of the FIFO loop. The size depends on which controller board is present on the system—the Output board, the Acquisition Controller board, or the Pulse Sequence Controller board (refer to the description of the acquire statement in the manual User Programming for information on identifying the boards). The value is set using the label Fifo Loop Size in the Spectrometer Configuration window (opened by config).
 Values 2048
 See also VnmrJ Installation and Administration
 Related config Display current configuration and possibly change it (M)

file File name of parameter set (P)

- Description Contains the file name of the parameter set returned by a rt or rtp command. This parameter is reset when the go command is issued. If the system is not in automation mode (auto='n'), file is reset to the 'exp' value. If the system is in automation mode (auto='y'), file is set to the path of the directory where the data is stored.
 - See also NMR Spectroscopy User Guide

RelatedautoAutomation mode active (P)goSubmit experiment to acquisition (C)rtRetrieve FID (C)rtpRetrieve parameters (C)

files Interactively handle files (C)

Syntax files<(files_menu)>

Description Brings up the interactive file handling program. With this program, the mouse and keyboard are used to copy, delete, rename, change directories, and load and save experiment data. The files command uses the graphics window to display file names. A mouse clicked on a file name selects it and the file name is displayed in reverse video. Various operations can be conducted on one or more selected files. The

	menus used for the files program are placed in the standard		
	menulib directories. Refer to the manual <i>NMR Spectroscopy User Guide</i> for more information on using menus, and refer to the manual		
	User Program	<i>nming</i> for information on programming menus.	
Arguments	files_menu is the files menu to control the menu buttons; the		
	default menu	is 'files_main' or the last active files menu.	
Examples	files		
	files('fil	es_dir')	
See also	User Programming		
Related	filesinfo	Return files display information (C)	
	tape	Control tape options of files program (P)	

filesinfo Return file information for files display (C)

Syntax	 (1) filesinfo('number'):\$number_files (2) filesinfo('name'<, file_number>):\$file (3) filesinfo('redisplay') 		
Description	Allows access to the list of files selected from the files interactive display. filesinfo is normally used only by the macros that implement the menu functions of the file system and not entered from the keyboard. The command will not execute unless the files program is active.		
Arguments	'number' is a keyword to return the number of files selected in the files display, or 0 if no files have been selected.		
	\$number_files is the return variable when 'number' is used.		
	'name' is a keyword to return a list of file names selected in the files display.		
	file_number is a number following the 'name' keyword to return only the file name in the list given by file_number.		
	<pre>\$file is a string variable that returns the file name when 'name' is used.</pre>		
	'redisplay' is a keyword that causes the current contents of the directory to be displayed. This display is useful after making changes in the directory, such as deleting or creating a file.		
See also	User Programming		
Related	files Interactively handle files (C)		

filtfile File of FIR digital filter coefficients (P)

Description Specifies name of a file of FIR (finite impulse response) digital filter coefficients. This file is a text file with one real filter coefficient per line (complex filters are not supported). If the parameter filtfile

does not exist in the current experiment, enter addpar('downsamp')
or addpar('oversamp') to add it. Entering addpar('downsamp')
creates the digital filtering and downsampling parameters downsamp,
dscoef, dsfb, dslsfrq, and filtfile. Similarly, entering
addpar('oversamp') creates digital filtering and oversampling
parameters def_osfilt, filtfile, oscoef, osfb, osfilt,
oslsfrq, and oversamp.

Values File name. The file must be in the user's vnmrsys/filtlib directory.

Related	addpar	Add selected parameters to current experiment (M)
	def_osfilt	Default value of osfilt (P)
	downsamp	Downsampling factor applied after digital filtering (P)
	dscoef	Digital filter coefficients for downsampling (P)
	dsfb	Digital filter bandwidth for downsampling (P)
	dslsfrq	Bandpass filter offset for downsampling (P)
	oscoef	Digital filter coefficients for oversampling (P)
	osfb	Digital filter bandwidth for oversampling (P)
	osfilt	Oversampling filter for real-time DSP (P)
	oslsfrq	Bandpass filter offset for oversampling (P)
	oversamp	Oversampling factor for acquisition (P)
	pards	Create additional parameters used for downsampling
		(M)
	paros	Create additional parameters used for oversampling
		(M)

findxmlmenu Find an xml menu (M)

Description Find an xml menu. Used by the menu system to find and display VnmrJ menus.

fitspec Spectrum deconvolution

Syntax	fitspec<('option')>
Applicability	VnmrJ 3.1
Description	Fit lorentzian and/or gaussian curves to experimental data. fitspec uses input from a text file "fitspec.inpar", which describes the starting values for a number of lines, which should be fitted to an experimental spectrum, and creates an output file "fitspec.outpar", which contains the fitted values for these lines. Furthermore, the resulting line frequencies are also stored in the parameter "slfreq", and the resulting amplitudes in "sla". The files "fitspec.inpar" and "fitspec.outpar" contain the following
	information for each line:

frequency intensity line width gaussian fraction

A * after any of the numbers indicates, that that parameter should not be fitted.

The command fitspec in VNMR actually prepares a file "fitspec.indata", which contains the spectral data (as a text file), to which the data should be fit, and then executes the external program "fitspec", which is stored in "/vnmr/bin". This program uses as an input the files "fitspec.inpar" and "fitspec.indata", and produces after completion the output file "fitspec.outpar". This file is is then read by VNMR and uses to set "slfreq" and "sla".

fitspec('usell')

The file "fitspec.inpar" can be prepared from a line listing automatically with the command fitspec('usell'). This option of "fitspec" uses the information from the last line listing (stored in "llfrq" and "llamp"), and the parameters "slw", "vs", "rfl" and "rfp" to prepare that file. All lines are set to the same line width "slw" and the gaussian fraction is set to 0. If other starting values are required, this file should be edited.

fitspec('setslfreq')

If the output data from a spectrum deconvolution has to be used in a spin simulation, this can be done automatically, if first the spin system is defined and then the deconvolution is done, because fitspec saves it's results in "slfreq" and "slamp", which serve as input for the iterative spin simulation. If the spin system is defined after the deconvolution is complete, the contents of "slfreq" and "sla" is lost, but the result of the deconvolution is still available in "fitspec.outpar". In this case, the option "fitspec('setslfreq')" just copies the information from "fitspec.outpar" back into "slfreq" and "sla".

fixgrd Convert gauss/cm value to DAC (M)

Syntax	fixgrd(gradient_value):parameter	
Description	Uses the gcal value in the probe table to return the DAC value for a specified gradient strength.	
Arguments	gradient_value is the required gradient strength in gauss/cm.	
	parameter is any local variable or VnmrJ variable.	
Examples	fixgrd(20):gzlvl	
Related	gcal Gradient calibration constant (P)	

fixpar Correct parameter characteristics in experiment (M)

Applicability	VnmrJ 3.1
Description	After bringing parameters into the current experiment with convert,
	rt, rtp, or rtv, fixpar is automatically executed. fixpar updates

F

old parameter characteristics and reconciles parameter differences due to the hardware on the spectrometer. If a macro userfixpar exists, fixpar runs it also. This allows an easy mechanism to customize parameter sets.

Related	convert	Convert data set from a VXR-style system (C)
	fixpar3rf	Create parameters for third rf channel (M)
	fixpar4rf	Create parameters for fourth rf channel (M)
	parfix	Update parameter set (M)
	parversion	Version of parameter set (P)
	rt	Retrieve FIDs (C)
	rtp	Retrieve parameters (C)
	rtv	Retrieve individual parameters (C)
	updatepars	Update all parameter sets saved in a directory (M)
	userfixpar	Macro called by fixpar (M)

fixpar3rf Create parameters for third rf channel (M)

Applicability Systems with a second decoupler.

Description Checks for the existence of all acquisition parameters related to the second decoupler. Any parameters found to be absent are created, characterized, and initialized by the macro. fixpar3rf is run as a part of the standard fixpar macro if the system configuration parameter numrfch is greater than 2 (i.e., the number of rf channels on the system is set at 3 or more).

fixpar4rf Create parameters for fourth rf channel (M)

Applicability Systems with a third decoupler.

Description Checks for the existence of all acquisition parameters related to the third decoupler. Any parameters found to be absent are created, characterized, and initialized. fixpar4rf is run as a part of the standard fixpar macro if the system configuration parameter numrfch is greater than 3 (i.e., the number of rf channels on the system is set at 4).

fixpar5rf Create parameters for fifth rf channel (M)

ApplicabilitySystems with a deuterium decoupler channel as the fourth decoupler.DescriptionChecks for the existence of all acquisition parameters related to the
fourth decoupler. Any parameters found to be absent are created,
characterized, and initialized. fixpar5rf is run as a part of the

standard fixpar macro if the system configuration parameter numrfch is greater than 4 (i.e., the number of rf channels on the system is set at 5).

fixgrdR Converts Gradient Strength to DAC values

Description	Converts a given DAC value to gradient strength based on the value of		
	gcal in the probe file. This is the reverse of fixrgd.		
Syntax	fixgrdR(gradientstrength):\$DAC_value		
Description	fixgrdR(3):\$DAC_value		
Related	fixrgd		

fixup Adjust parameter values selected by setup macros (M)

Description Called by the experiment setup macros h1, c13, hc, hcapt, capt, and hcosy. As provided, the text of fixup is all in quotes so that it does nothing. It is intended to provide each user with a mechanism to make adjustments to values selected by the setup macros.

fixpsg Update psg libraries (M)

Description Used by patchinstall to recompile the psg files and create new psg libraries libpsglib.so in /vnmr/lib.

flashc Convert compressed 2D data to standard 2D format

Syntax	<pre>flashc<(<'nf'><,'ms' 'mi' 'rare'<,traces><,echoes>)></pre>		
Applicability	VnmrJ 3.1		
Description	Rearranges 2D "fid" data files from compressed formats to standard format or from standard format to compressed format. Compressed data is taken using the "nf" parameter to specify the number of fids in the second dimension of a 2D experiment. In other words compressed data is acquired as one large uninterrupted "multifid" acquisition.		
	Before the 6.0 release, arrayed or multislice compressed images (seqcon='nscnn'), had to be reformatted to a standard 2D format, using "flashc" before a "ft2d" could be performed on the data. Now using "ft2d('nf', <index>)" this is no longer necessary, and processing time may even be enhanced by reformatting data from the standard format (seqcon='ncsnn') to the compressed format. However for</index>		

compressed compressed 2D (seqcon='nccnn'), "flashc(...)" or "flashc('nf',...)" must be run.

For 3D data sets "flashc" is not needed. The "ft3d" routine will handle standard, compressed (seqcon='nncsn'), or compressed-compressed (seqcon='nnccn') 3D data.

The flashc command reads the file "fid" in the "acqfil" subdirectory of the current experiment. The data is reordered and written back out to the same "fid" file. Thus, the original "fid" file is lost. Precautions are taken so that in the event of an error during processing, the original "fid" file will be preserved. Also, before running a simple check is done by flashc to prevent it from being executed more than once in an experiment on the same data set. The simple check against multiple executions of flashc looks for the parameter 'flash_converted' which flashc creates when it is run. To rerun flashc the parameter can be removed with the following commands: destroy('flash_converted')

destroy('flash_converted','processed')

Compressed-compressed or Standard to Compressed Format

Using "ft2d ('nf',<index>)", flashc really only has to be used to convert a completely compressed multislice, multiecho, or multi-image sequence. However, for a large standard multi-slice experiment (seqcon='ncsnn') a performance benefit may be achieved in converting the data to a compressed format. When converting to a compressed format the first argument must always be 'nf'. When converting completely compressed or "rare" type sequences, the first argument is a string defining the type of compression. This string can either be 'ms' for multislice, 'mi' for multi-image, or 'rare' for multi-echo "rare" type fast imaging data sets. The second argument defines the number of images slices or array elements to retain.

The values of four VNMR parameters are changed by flashc.

- "ni" is set to 1 if no argument is provided.
- "nf" is set to the value of "nf" divided by the multislice "ms" or multi-image "mi" value.
- "arraydim" is set to the product of its original value and the value of the "traces" argument.
- "arrayelemts" is set to 1 if no parameters were arrayed during data acquisition or to 2 if any parameter was arrayed during data acquisition.

Compressed to Standard Format

flashc can convert a completely compressed multislice, multiecho, or multi-image sequence. It can also convert a "rare" type sequence with a compressed phase-encode echo train. When converting completely compressed or "rare" type sequences, the first argument is a string defining the type of compression. This string can either be 'ms' for multislice, 'mi' for multi-image, or 'rare' for multi-echo "rare" type fast imaging data sets. The second argument defines the number of compressed traces to retain for each "ni" and "nf" will be set to this number after "flashc" has been run.

- The values of four VNMR parameters are changed by flashc.
- "nf" is set to the value of the "traces" argument, or to 1 if no argument is provided.
- "ni" is set to the value of "nf" divided by the multislice "ms" or multi-image "mi" value.
- "arraydim" is set to the product of its original value and the original value of "nf".
- "arrayelemts" is set to 1 if no parameters were arrayed during data acquisition or to 2 if any parameter was arrayed during data acquisition.

Examples Compressed-compressed or Standard to Compressed Format

- flashc('nf'): Standard to compressed
- flashc('nf','ms',ns): Compressed phase-encode and multi-slice
- flashc('nf','mi',ns): Compressed multi-image and phase-encode Compressed to Standard Format
- flashc: Simple compressed phase-encode
- flashc('ms',ns): Compressed phase-encode and multi-slice
- flashc('mi',ns): Compressed multi-image and phase-encode
- flashc('rare',ns,etl)

arraydim	Dimension of experiment (P)	
ft2d	Fourier transform 2D data (C)	
ft3d	Fourier transform 3D data (C)	
nf	Number of FIDs(P)	
ni	Number of increments in 1st indirectly detected	
	dimension(P)	
seqcon	Acquisition loop control (P)	
	arraydim ft2d ft3d nf ni seqcon	

flipflop Set up parameters for FLIPFLOP pulse sequence (M)

Applicability	Systems with solids module.
Description	Sets up a multipulse parameter set for tuning out "phase glitch" in the
	probe and pulse amplifier.
See also	User Guide: Solid-State NMR

Fluorine Set up parameters for 19F experiment (M)

Description Set Up parameters for ¹⁹F experiment.

flush Write out data in VNMR memory

Applicability VnmrJ 3.1

Description The VNMR program keeps current data and parameters in memory buffers. Normally, this information is not written to disk until one exits VNMR or joins another experiment. Use this command to write out this information. One application is if you want to access the experimental data from a program separate from the VNMR program.

fn Fourier number in directly detected dimension (P)

Description Selects the Fourier number for the Fourier transformation along the directly detected dimension. This dimension is often referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, etc.

Values 'n' or a number equal to a power of 2 (minimum is 32). If fn is not *entered* exactly as a power of 2, it is automatically rounded to the nearest higher power of 2 (e.g., setting fn=32000 gives fn=32768). fn can be less than, equal to, or greater than np, the number of directly detected data points:

- If fn is less than np, only fn points are transformed.
- If fn is greater than np, fn minus np zeros are added to the data table ("zero-filling").
- If fn='n', fn is automatically set to the power of 2 greater than or equal to np.

fn1 Fourier number in 1st indirectly detected dimension (P)

Description	Selects the Fourier number for the Fourier transformation along the first indirectly detected dimension. This dimension is often referred to as the f_1 dimension of a multi-dimensional data set. The number of increments along this dimension is controlled by the parameter ni.		
Values	fn1 is set in a manner analogous to the parameter fn, with np being substituted by $2*ni$.		
See also	NMR Spectroscopy User Guide		
Related	fnFourier number in directly detected dimension (P)fn2Fourier number in 2nd indirectly detected dimension (P)niNumber of increments in 1st indirectly detected dimension (P)npNumber of data points (P)		

fn2 Fourier number in 2nd indirectly detected dimension (P)

- Description Selects the Fourier number for the Fourier transformation along the second indirectly detected dimension. This dimension is often referred to as the f_2 dimension of a multidimensional data set. The number of increments along this dimension is controlled by the parameter ni2. fn2 is set in a manner analogous to the parameter fn, with np being substituted by 2*ni2.
 - See also NMR Spectroscopy User Guide Related fn Fourier number in directly detected dimension (P)
 - fn1 Fourier number in 1st indirectly detected dimension (P)
 ni2 Number of increments in 2nd indirectly detected dimension (P)
 np Number of data points (P)

fn2D Fourier number to build up 2D DOSY display in freq. domain (P)

Description	In 2D DO	SY sequences (Dbppste, DgcsteSL, Doneshot, Dbppsteinept),	
	replaces	fn when setting up the 2D display.	
See also	NMR Spectroscopy User Guide		
Related	ddif	Synthesize and display DOSY plot (C)	
	dosy	Process DOSY experiments (M)	

focus Send keyboard focus to input window (C)

DescriptionSends keyboard focus to the input window. This is only useful for
macro programming.See alsoUser Programming

foldcc Fold INADEQUATE data about 2-quantum axis

1
symmetrizes 2D INADEQUATE data along the P-type uantum axis and applies an automatic DC baseline correction. nand foldcc functions for both hypercomplex and comlex 2D
1 r

foldj Fold J-resolved 2D spectrum about the F1=0 axis

Applicability VnmrJ 3.1

Description foldj symmetrizes heteronuclear 2D-J or rotated homonuclear 2D-J experiments about the F1=0 axis and functions with both complex and hypercomplex 2D data.

foldt Fold COSY-like spectrum along diagonal axis

Syntax foldt(<sym_op>)

Applicability VnmrJ 3.1

- Description foldt (<sym_op>) folds COSY-like correlation spectra about the diagonal. The 2D spectrum must exhibit a P-type diagonal in order for foldt to work properly. [A P-type diagonal is one which goes from the bottom left-hand side to the top right-hand side of the contour display.] The argument sym_op can take three string values: 'symm', 'triang' and 'covar'. The default value is 'symm'.
- Arguments If sym_op = 'symm', the folding process performs a symmetrization of the data by replacing every two symmetry-related points with the one point therein which is the smallest in magnitude. If sym_op = 'triang', the folding process performs a triangularization of the data by replacing every two symmetry-related points with their geometric mean.

If sym_op = 'covar', for "covariance NMR", the folding process answers the question of whether the two symmetry-related points are correlated. If the product of the two points (a and b) is greater than 0.0, the two points are each replaced with the sqrt(a*b). Otherwise, the two points are set to 0.0. The command foldt functions for both hypercomplex and complex 2D data but requires that fn=fn1.

fontselect Open FontSelect window (C)

Description Opens the FontSelect window for defining fonts in window panes created by setgrid. A different font can be selected for every window pane combination of rows and columns. Separate fonts can also be selected for a large or small overall graphic window.

See also NMR Spectroscopy User Guide

Related	curwin	Current window (P)
	jwin	Activate current window (M)
	mapwin	List of experiment numbers (P)
	setgrid	Activate selected window (M)
	setwin	Activate selected window (C)

format

Description	Formats a real number into a nice string for output/converts a string into upper case or lower case for output/tests a string to determine if it can represent a real number/interconverts string representations of real numbers and real numbers		
Syntax	Two arguments: format(stringvar,'upper'):stringvar format(stringvar,'lower'):stringvar format(stringvar,'isreal'):ans		
Syntax	Three arguments: format(realvar,n,m):\$sval format(realvar,n,m):\$rval format(stringvar,n,m):\$sval format(stringvar,n,m):\$rval where realvar is a variable of real type. n is the length, m is the precision (number to the right of the decimal point. stringvar is a string variable. \$sval is a string return value. \$rval is a real return value.		
Applicability	VnmrJ 3.1		
Description	format can be used for the following:		
	• formats a real number into a nice string for output		
	• converts a string into upper case or lower case for output		
	• tests a string to determine if it can represent a real number		
	• interconverts string representations of real numbers and real numbers		
Arguments	If the command is given two arguments, the first argument may be a string or real variable and the action depends on the value of the second argument. If the second argument is 'upper', this command will convert the first argument to all upper case characters. If the second argument is 'lower', this command will convert the first argument to all lower case characters. If the second argument is 'isreal', this command will test the first argument to see if it satisfies the rules for a real number. It will return a 1 in the first argument can represent a real number and a 0 otherwise.		
	If the command is given three arguments, the first argument must be a real number or string holding a real number. If it is a string variable, it must satisfy the rules for a real number. The 'isreal' option above can be used for this purpose. This command will format it into either a string with length n and precision m or another real number of length n and precision m. If you want to return the value into a string, if it is a temporary dollar parameter (e.g., \$sval), the parameter will need to be initialized as a string by first setting it to a string (e.g., \$sval=").		
Examples	<pre>format(a,5,2):sa If a=24.1264 then string sa='24.13' format(solvent,'lower'):n1 If solvent='CDCl3' then n1='cdcl3' format(\$1,'isreal'):\$a Will set \$a to 1 if \$1 represents a number. \$sval='' "Initialize \$sval to a string variable"\$snum</pre>		

	= '143.92' \$rnum = 32.75		
Examples	<pre>Format real value \$rnum = format(\$rnum,3,1):\$sval format(\$rnum,3,1):\$rval</pre>	32.75 Will set \$sval to Will set \$rval to	the string '32.8' the number 32.8
Examples	<pre>Format string value \$snum format(\$snum,3,1):\$sval format(\$snum,3,1):\$rval</pre>	<pre>= '143.92' Will set \$sval to Will set \$rval to</pre>	the string '143.9' the number 143.9

fp

Find peak heights or phases (C)

Syntax	fp<(<'phase',> <index1,index2,>)></index1,index2,>		
Applicability	VnmrJ 3.1		
Description	Following a line listing (either dll or nll), fp measures the peak height of each peak in an array of spectra. The results of the analysis are written to a text file fp.out in the current experiment directory. If the npoint parameter is defined in the current parameter set and this parameter is "on," it determines the range of data points over which a maximum is searched when determining peak heights. The possible values of npoint are 1 to fn/4. The default is 2.		
Arguments	'phase' is a keyword to measure the phase of each peak instead of height.		
	index1, index2, restricts measuring peak heights or phases to the lines listed.		
Examples	<pre>fp fp(1,3) fp('phase')</pre>		
See also	NMR Spectroscopy User Guide		
Related	dll Display listed line frequencies and intensities (C)		
	fn Fourier number in directly detected dimension (P)		
	get11 Get line frequency and intensity from line list (C)		
	nl Position cursor at the nearest line (C)		
	nllFind line frequencies and intensities (C)		
	npoint Number of points for fp peak search (P)		

fpi Report integral values from arrayed spectra.

```
Syntax fpi<('bc')>
fpi<('dc')>
fpi<('tl')>
Applicability VnmrJ 3.1
```

F

Description Following the definition of integral regions (either by hand, or using the region command), "fpi" measures the height of each integral in an array of spectra. If the keyword 'bc' or 'dc' is specified, one of commands is used to flatten the baseline or remove any baseline offset prior to evaluating the integrals. The results of the analysis are written into the text file "fpi.out " in the current experiment directory.

> "fpi" always works on the entire spectrum, i.e., it will produce a report on all defined integral regions. "fpi" will indicate the integration limits in ppm units if "axis='p" - if you prefer Hz units, set "axis='h" prior to calling "fpi".

> The resulting output, "curexp+'/fpi.out" does NOT comply with the VNMR commands for T1 analysis etc. - however, if an argument 't1' is used, "fpi" and creates a file "curexp+'/fp.out" which can be used for T1, T2 etc. analysis (note that in this case the "line positions" marked in this file are mid-points of the respective integral region).

Arguments 'bc' - optional baseline correction on each spectrum

'dc' - optional offset/drift correction on each spectrum

<code>'t1'</code> - optional creation of <code>"curexp+'/fp.out'"</code> which is compatible with <code>"t1"</code> and related commands

The 'bc' and 'dc' arguments cannot be combined.

Examples fpi

```
axis='h' fpi axis='p'
fpi('dc')
fpi('bc','t1')
fpi('t1','dc')
fpi('t1')
```

See also

Related fp

Find peak heights or phases (C)

fpmult First point multiplier for np FID data (P)

- Applicability VnmrJ 3.1
- Description Allows error correction if the first point of an FID is misadjusted. In a 1D experiment, this adjustment influences the overall integral of the spectrum. For *n*-dimensional experiments, if the correction is not made, "ridges" can appear. In 2D experiments, the ridges appear as "f₂ ridges." In 3D experiments, the ridges appear as "f₃ ridges." These ridges can clearly be seen in the noise region on the top and bottom of a 2D spectrum (when trace='f1') as a low-intensity profile of the diagonal. The sign and intensity of the ridges is controlled by the magnitude of fpmult.

It has been recognized that the first point of a FID that is sampled at exactly time equal to zero must be multiplied by 0.5 for the Fourier

transform to function properly. The fpmult parameter gives you a method to fine-tune the actual correction factor.
Values Default is 1.0, except that if the processing involves backward extension of the time-domain data with linear prediction, the default changes to 0.5. If fpmult is set to 'n', fpmult takes on its default value.
See also NMR Spectroscopy User Guide
Related fpmult1 First point multiplier for ni interferogram data (P)

- fpmult2First point multiplier for ni2 interferogram data (P)npNumber of data points (P)
 - trace Mode for *n*-dimensional data display (P)
 - wft2da Weight and Fourier transform phase-sensitive data (M)

fpmult First point multiplier for "np" FID data

Applicability	VnmrJ	3.1
---------------	-------	-----

Description For 2D experiments such as NOESY, TOCSY, or ROESY, one should run cfpmult on the transformed first increment, prior to typing wft2da, to minimize "F2 ridges" in the final 2D spectrum. This macro calculates an fpmult value for the dataset (which will then be used by wft2da).

One may do this manually for a 2D dataset by typing fpmult=1.0 wft(1) cdc

in the VNMR command line and noting whether the spectrum (essentially the baseline) moves up or down when "dc" is typed. One should vary fpmult until the "dc" correction (jump in the baseline) is as small as possible. With care, one can set fpmult to two decimal places. Typical values for fpmult are 1.00-2.00. The default value for fpmult is 1.0.

This only needs to be performed for cosine-type experiments, such as NOESY, ROESY, or TOCSY where both the t_2 FID and the t_1 interferogram decay. The macro (cfpmult) might give incorrect values for first increments of experiments having baseline distortions (i.e. water suppression with 11-echo or 1331); in such cases manual optimization of fpmult is more suitable.

Why should you bother adjusting fpmult? If the first point in t1 of a 2D spectrum is misadjusted, the result will be the appearance of a series of "F2 ridges". These ridges can clearly be seen in the noise region on the top and bottom of a 2D spectrum (when trace = 'f1') as a low-intensity profile of the diagonal. The sign and intensity of the ridges is controlled by the magnitude of misset of fpmult.

It has been recognised that the first point of a FID which is sampled at exactly $t_2 = 0$ must be multiplied by 0.5 for the Fourier Transform to function properly. The first point of a FID influences the overall integral of the resulting spectrum. The fpmult parameter gives one a way to fine-tune the actual correction factor.

NOTE: When processing 2D data, unless "lp" is approximately 0,

FPMULT will affect both the DC offset and the curvature of the spectrum.

See also alfa and calfa

fpmult1 First point multiplier for "ni" interferogram data

Applicability VnmrJ 3.1

Description fpmult1 and fpmult2 operate on "ni" and "ni2" hypercomplex or complex interferogram data, respectively, in a manner analogous to fpmult. In many 2D and 3D experiments, the t1 (ni) and t2 (ni2) values are adjusted so that there is no first-order phasing in the F1 and F2 dimensions. In this case, fpmult1 and fpmult2 should be 0.5, the default value. If the t1 and t2 values are adjusted so that there is a 180-degree first-order phase correction, fpmult1 and fpmult2 should be 1.0.

fpmult2 First point multiplier for "ni2" interferogram data

Applicability VnmrJ 3.1

Description fpmult1 and fpmult2 operate on "ni" and "ni2" hypercomplex or complex interferogram data, respectively, in a manner analogous to fpmult. In many 2D and 3D experiments, the t1 (ni) and t2 (ni2) values are adjusted so that there is no first-order phasing in the F1 and F2 dimensions. In this case, fpmult1 and fpmult2 should be 0.5, the default value. If the t1 and t2 values are adjusted so that there is a 180-degree first-order phase correction, fpmult1 and fpmult2 should be 1.0.

fr Recall all display parameters from set #n, n=1..9 (n)

Syntax	fr(n)
Applicability	VnmrJ 3.1
Description	fr(n) performs a full recall of the display parameter set, setting all current display parameters to those values.
Arguments	A second argument can be given to these commands. It prevents them from causing the automatic update of interactive programs that may be displayed.

framecmd Create a new frame

Syntax framecmd('new','image',x,y,width,height,'imagefilepath'):\$id framecmd('new','text',x,y,width,height,'textfilepath'<, color,font,fontsize>):\$id framecmd('new','inset',x,y,width,height<,cr,delta<,cr1, delta1>>):\$id framecmd('delete',\$id) framecmd('hide',\$id) framecmd('show',\$id)

Description framecmd will create a new frame of image, text, and inset with 'new' option. The type of image can be GIF, PNG, JPEG, or other image format supported by 'convert' program.

ArgumentsThe range of x is 0 at the left edge of the chart and wcmax at the right
edge of the chart.
The range of y is 0 at the bottom edge of the chart and wc2max at the
top edge of the chart.
The range of width is 0 to wcmax.
The range of height is 0 to wc2max.
The color, font, and fontsize can be adjusted with text editor in VnmrJ
window.

fread Read in variables from a file and load them in a tree

Syntax fread(filename[,tree[,'reset', 'value', 'newonly']]) filename is a valid file with proper variable format. tree can be current, global, processed, systemglobal, or usertree. 'reset' keyword can only be used if tree is specified.

Applicability VnmrJ 3.1

Description This command reads in vnmr variables from a file and loads them into a tree. The variable trees are 'current', 'global', 'processed', 'systemglobal', and 'usertree'. It can read from any file that has variables stored in the correct vnmr format. The default tree is 'current'.

Arguments A "reset" option causes the variable tree to first be cleared before the new variable file is read. Without this option, variables read from a file are added to the existing preloaded variables. In order to use the 'reset' option, the tree must also be specified. A "value" option causes only the values of the variables in the file to be loaded. If a preloaded variable does not already exist, a new one is not created. Parameter attributes are not changed. Enumerated values are not changed. In order to use the 'value' option, the tree must also be specified. A "newonly" option causes only those variables in the file which do not already exist in the tree to be loaded. In order to use the 'newonly' option, the tree must also be specified.

The 'reset', 'newonly', and 'value' options are mutually exclusive.
NOTE: if variables are read into the 'global' tree, certain parameters
will not be loaded. These are important system parameters that should
not be changed. These parameters are: userdir, systemdir, curexp,
autodir, auto, operator, vnmraddr, and acqaddr.
The 'usertree' is available for use. By default, it has no parameters
stored in it. It would typically be used by a macro for temporary
parameter storage. All of the parameter utility commands, such as
setlimit, setprotect, setvalue, getvalue, fsave, etc. will work with
'usertree' as the optional tree argument. A special incantation of fread
with a empty string as the filename will clear parameters from
'usertree'. That is, fread(",'usertree') clears 'usertree'.
Note that passing an empty string as the filename with other parameter
trees generates an error. For example, fread(",'current') is an error. As
with all the parameter utility commands, the other arguments also
work with 'usertree'. In the case of fread, this means that the 'reset',
'value', and 'newonly' options are valid for 'usertree'.

Examples fread('var1'): read in variables from file var1 into current tree. fread('sampvar','global'): read in variables from file sampvar into global tree.

fsave Save parameters from a tree to a file (C)

Syntax fsave(file<,tree>)

ApplicabilityVnmrJ 3.1

Description Writes parameters from a parameter tree to a file.

Arguments file is the name of the file, which can be any valid file for which the user has write permission. If the file already exists, it will be overwritten.

tree is one of the keywords 'global', 'current', 'processed', or 'systemglobal'. The default is 'current'. Refer to the create command for more information on types of trees.

- Examples fsave('var1')
 fsave('sampvar','global')
- See also User Programming

Related	create	Create new parameter in a parameter tree (C)
	destroy	Destroy a parameter (C)
	display	Display parameters and their attributes (C)
	fread	Read parameters from file and load them into a tree (C)
	svp	Save parameters from current experiment (C)

fsq Frequency-shifted quadrature detection (P)

Description Selects whether to use frequency-shifted quadrature detection. When fsq is turned on, if dsp is on, the observe frequency is offset by oslsfrq, and the digital filter is also offset by oslsfrq. The default value of oslsfrq is 1.25*sw.

The effect of fsq is to offset only the digital filter by oslsfrq. The observe frequency must be offset by oslsfrq by modifying the pulse sequence as described in the manual *NMR Spectroscopy User Guide*.

Values 'n' turns frequency-shifted quadrature detection off. 'y' turns it on.

See also NMR Spectroscopy User Guide

dsp	Type of DSP for data acquisition (P)
oslsfrq	Bandpass filter offset for oversampling (P)
oversamp	Oversampling factor for acquisition (P)
SW	Spectral width in directly detected dimension (P)
	dsp oslsfrq oversamp sw

Fourier transform 1D data (C)

> The wft and ft commands perform a Fourier transform on one or more 1D FID's with or without weighting applied to the FID, respectively. Both commands execute a left-shift, zero-order phase rotation, and a frequency shift (first-order phase rotation) according to the parameters lsfid, phfid, and lsfrq, respectively, on the time-domain data prior to the weighting (if appropriate) and Fourier transformation. All string arguments supplied to these two commands must precede the numeric arguments, for example,

> ft('nodc', 'noft',1,10,2) The type of Fourier transformation is determined by the parameter "proc" Solvent suppression is turned on or off with the parameters ssfilter and ssorder.

Arguments String Arguments: options can be any of the following (all string arguments must precede the numeric arguments):

ft('acq') checks if any elements of a multi-FID experiment have already been transformed. If so, these previously transformed elements will not be re-transformed.

ft('nodc') does not perform the fid drift correction. ft('dodc') does perform the fid drift correction. The global parameter dc1d determines the default if neither 'nodc' nor 'dodc' is used. If the global parameter dc1d does not exists, or it exists and is set to 'y', then fid drift correction is performed. If the dc1d parameter exists and is set to 'n', fid drift correction is not performed.

ft('nods') prevents an automatic spectral display (ds) from occurring. This is useful for various plotting macros.

ft

ft('noft') skips the actual ft step, thereby allowing to use all spectral manipulation and plotting commands on FID's.

ft('zero') zeroes the imaginary channel of the FID prior to the Fourier transform. This zeroing occurs after any FID phasing. Its use will be generally limited to wideline solids applications.

ft('nf') allows a single FID element containing 'nf' traces to be transformed as if it were 'nf' separate FID elements.

ft('ftargs') provides additional parameter-based FID processing. These parameters control amplitude, phase, frequency, sampling window, and frequency shifting corrections of individual FIDs. There is a mechanism to do ECC correction of the FIDs, based on a reference FID.

The FIDs can also be combined using a set of coefficients. These are described in the "FID parameters" section below.

Numeric arguments:

For arrayed data sets, both of these commands will Fourier transform all of the array elements. To Fourier transform selected elements of the array, wft and ft can be passed numeric arguments. Passing a single numeric argument will transform only that element. For example, wft(3) will transform only array element 3. Passing two numeric arguments will transform the inclusive array elements. For example, wft(3,7) will transform array elements 3, 4, 5, 6, and 7. Passing three numeric arguments is similar to passing two arguments with the addition that the third argument is used as the increment between successive elements that are to be transformed. For example, wft(2,10,2) will transform elements 2, 4, 6, 8, and 10. This use of numeric parameters is identical to the scheme used for displaying spectra with the dss command and other related commands.

If the string argument 'nf' precedes the list of numeric arguments, the rules for interpreting the numeric arguments change slightly. Passing no numeric arguments results in the transformation of all 'nf' traces in the first FID element. Passing a single numeric argument results in the transformation of all 'nf' traces in the requested FID element. For example, ft('nf',3) transforms all 'nf' traces for element 3. Regardless of the requested FID element, the resulting spectra are labeled as 1 to nf since multiple elements cannot be transformed using ft('nf'). Subsequent numeric arguments are interpreted as previously described.

FID parameters:

There are a set of parameters that are used to process individual FIDs prior to the FID weighting step. In each case, if the parameter does not exist or is set to "Not active", the function associated with the parameter will be omitted. The order of parameter descriptions below corresponds to the order in which any additional FID processing is done.

fidinfo

This parameter does not control any processing. It is the pathname of a file where the details of the following processing will be written.

fidecc

Pathname of the phase values of a reference data set. These phase values need to be precalculated using the calcECC program. The ECC correction involves measuring the phase of each data point in the FID and adjusting it so that the difference on phase between that data point and the first data point is the same as the corresponding phase difference in the reference data set.

fideccls

The ECC left-shift value. These parameter is ignored if the fidecc processing is omitted. This may be used to redefine the "first point" in the reference FID. Its default value is 0.

fidfreq

Frequency shift individual FIDs using the values from this parameter. The shifts are in Hertz. If the number of elements of this arrayed parameter is less than the number of FIDs, the element that will be used for a specific FID will be (FID number - 1) modulo (parameter array size) + 1.

fidautofreq

This is a special case of frequency shifting individual FIDs. Instead of specifying a frequency shift for each FID, this parameter consists of only two values. The first is an initial offset and the second is a increment for subsequent FIDs. For example, if fidautofreq=10,100, the first FID will be offset by 10, the second by (10 + 100), the third by (10 + 200), etc.

fidautophase

Phase individual FIDs, relative to the first FID. The fidautophase parameter consists of four numbers. The first number specifies the number if FID points to be used to determine of phase of that FID. Maximum value is 500. The second number is the number of points to skip from the beginning of the FID when determining the phase. The third number is the number of points at the end of the FID to use to determine the RMS noise of the FID. Maximum value is np/8. The fourth and last number is a multiplier for the RMS noise value. For a point to be considered when determining the phase, its magnitude (sqrt(re*re + im*im)) must be greater than the RMS noise time the multiplier.

fidautophasepar

The parameter is used in conjunction with fidautophase. It will hold the results of the phases calculated when fidautophase is selected.

fidphase

Phase individual FIDs using the values from this parameter. The phase angles are in degrees. If the number of elements of this arrayed parameter is less than the number of FIDs, the element that will be used for a specific FID will be (FID number - 1) modulo (parameter array size) + 1.

fidsa

Sampling window adjust individual FIDs using the values from this parameter. FID data points on either side of the shifted sampling window will be set to zero. This value is entered as the number of complex data points in the sampling window. If the number of elements of this arrayed parameter is less than the number of FIDs, the element that will be used for a specific FID will be (FID number - 1) modulo (parameter array size) + 1.

fidsas

Sampling window shift adjusts the sampling window of individual FIDs. FID data points on either side of the shifted sampling window will be set to zero. This value is entered as the number of complex data points to shift the sampling window. If the number of elements of this arrayed parameter is less than the number of FIDs, the element that will be used for a specific FID will be (FID number - 1) modulo (parameter array size) + 1.

fidamp

Amplitude adjust individual FIDs using the values from this parameter. If the number of elements of this arrayed parameter is less than the number of FIDs, the element that will be used for a specific FID will be (FID number - 1) modulo (parameter array size) + 1.

fidshift

Shift individual FIDs using the values from this parameter. The shifts are in number of complex points. Positive values shift data to the right. Negative values shift data to the left. This can be used with fidadd and fidsa to combine sections of a set of FIDs into a single FID, as is done when processing "pureshift" data sets. If the number of elements of this arrayed parameter is less than the number of FIDs, the element that will be used for a specific FID will be (FID number - 1) modulo (parameter array size) + 1.

fidautoshift

This is a special case of frequency shifting individual FIDs. Instead of specifying a frequency shift for each FID, this parameter consists of only two values. The first is an initial number of complex point to shift and the second is a increment for subsequent FIDs. For example, if fidautoshift=10,100, the first FID will be offset by 10, the second by (10 + 100), the third by (10 + 200), etc. fidadd

Individual FIDs can be combined. The fidadd parameter specifies how many FIDs to combine into a single one. If fidadd=0, this is a special value to indicate all the FIDs should be combined into a single FID. This is equivalent to fidadd=arraydim. If, for example, fidadd=3, fta and wfta will combine FIDs in groups of three. The fidadd parameter can have either a single value, as just described, or it can have another four optional values. These values are used as the coefficients to combine the FIDs. The default values of rr, ir, ri, ii are 1,0,0,1. These coefficients multiply the real(r) and imaginary(i) components of the FID, as in

rr * (input real point) + ir * (input imag point) =>
output real point

ri * (input real point) + ii * (input imag point) =>
output imag point

These coefficients are still active even if the first value of fidadd=1, that is, one input FID produces one output FID. For example, fidadd=1,1,0,0,0 will zero the imaginary channel of the resulting FID. fidadd=1,1,0,0,-1 will negate the imaginary channel, resulting in a frequency reversal.

Inverse Fourier Transformation:

ft('inverse', expnum, expansion_factor) performs an inverse FT, storing the resulting fid in the experiment defined by the second argument (first numeric argument). The expansion_factor defines the expansion of the spectrum before the inverse FT is performed. This argument is equivalent to a multiplier for the "fn" parameter, must lie between 1 and 32, and is rounded up internally to the nearest power of 2. Note that this command performs an inverse FT of the entire spectrum. Vnmr does not currently support the inverse FT of arrayed 1D or 2D data sets.

ftarg ftarg is a macro to create the parameters used by
ft('ftargs') and wft('ftargs').

calcECC(infile, outfile)

The calcECC command requires two arguments. The first is a pathname to a reference data set to be used to do the ECC corrections. The first argument should be the name of a ".fid" directory, containing a data set saved by VnmrJ. The second argument is a filename where to place the results. A typical value would be curexp+'/eccref', as in

calcECC(userdir+'/data/waterref.fid',curexp+'/eccref')

In this case, the parameter fidecc=curexp+'/eccref' accesses this information for the ft('ftargs') and wft('ftargs') commands. The calcECC commands calculated the phase angle for each data point in the FID and writes it to the output file.

Examples	ft	
	ft(1)	
	ft(3,7)	
	ft(2,10,2)	
	ft('nf',3)	
See also	NMR Spectro	scopy User Guide
Related	calcECC	Calculate ECC corrections (C)
	dcrmv	Remove dc offsets from FIDs in special cases (P)
	fn	Fourier number in directly detected dimension (P)
	ftargs	Macro to create parameters.
	lsfid	Number of points to left-shift the np FID (P)
	lsfrq	Frequency shift of the fn spectrum in Hz (P)
	nf	Number of FIDs (P)
	phfid	Zero-order phasing constant for np FID (P)
	proc	Type of processing on the np FID (P)
	ssfilter	Full bandwidth of digital filter to yield a filtered FID (P)
	ssorder	Order of polynomial to fit digitally filtered FID (P)
	wft	Weight and Fourier transform 1D data (C)

ft1d Fourier transform along f₂ dimension (C)

Syntax	<pre>(1) ftld(element_number) (2) ftld<('nf',element_number) (3) ftld<(coptions,><coefficients>)></coefficients></pre>			
Applicability	VnmrJ 3.1			
Description	For Performs the first Fourier transformation along the f_2 dimension without weighting, and matrix transposition. ft1d allows the dis of t_1 interferograms with the dcon and dconi commands. For an 2D FID data, a single array element can be weighted and transfor using syntax 1 or 2. The keyword 'nf' is used in syntax 2 to sp that the 2D data is collected in the compressed form using 'nf Complex and hypercomplex interferograms can be constructed explicitly by supplying a series of options and coefficients using sy 3.			
	For information on real as opposed to complex Fourier transforms, see the descriptions of the proc, proc1, and proc2 parameters. For information about Hadamard transforms, see the description of the proc1 parameter and the <i>VnmrJ NMR Liquids</i> user guide. For information on left-shifting, zero-order phase rotation, and frequency shifting of the FID and interferogram time-domain data during the 2D Fourier transformation, see the descriptions of the parameters lsfid, lsfid1, lsfid2, phfid, phfid1, phfid2, lsfrq, lsfrq1, and lsfrq2, as appropriate. For information on the lfs (low-frequency suppression) and zfs (zero-frequency suppression) solvent suppression			

options, see the description of the parameters ssfilter and ssorder, and the macro parfidss.

element_number is a single array element to be weighted and Arguments transformed.

options can be the keywords 'ptype' or 'ntype' but neither serve a useful function because the differential effect of these arguments is applied only during the course of the second Fourier transformation. The default is 'ntype'.

coefficients are a series of coefficients according to the following scheme: RR1 is the coefficient used to multiply the real part (first R) of spectra set 1 before it is added to the real part (second R) of the interferogram. IR2 would thus represent the contribution from the imaginary part of spectra set 2 to the real part of the interferogram, and so on. The scheme is depicted below.

```
ft1d(RR1,IR1,RR2,IR2,...,RI1,II1,RI2,II2,...)
```

where:

```
RR1*REAL(w2,element=1) -> REAL(t1)
IR1*IMAG(w2,element=1) -> + REAL(t1)
RR2*REAL(w2, element=2) \rightarrow + REAL(t1)
IR2*IMAG(w2,element=2) -> + REAL(t1)
  . .
RI1*REAL(w2,element=1) -> IMAG(t1)
II1*IMAG(w2,element=1) -> + IMAG(t1)
RI2*REAL(w2, element=2) \rightarrow + IMAG(t1)
II2*IMAG(w2,element=2) -> + IMAG(t1)
. . .
```

See also NMR Spectroscopy User Guide

Related	dconi	Interactive 2D data display (C)
	ft2d	Fourier transform 2D data (C)
	lsfid	Number of complex points to left-shift np FID (P)
	lsfid1	Number of complex points to left-shift ni
		interferogram (P)
	lsfid2	Number of complex points to left-shift ni2
		interferogram (P)
	lsfrq	Frequency shift of the fn spectrum (P)
lsfrq1 Frequency shift of the		Frequency shift of the fn1 spectrum (P)
	lsfrq2	Frequency shift of the fn2 spectrum (P)
	parfidss	Create parameters for time-domain solvent subtraction
		(M)
	phfid	Zero-order phasing constant for np FID (P)
	phfid1	Zero-order phasing constant for ni interferogram (P)
	phfid2	Zero-order phasing constant for ni interferogram (P)
	proc	Type of processing on np FID (P)
	proc1	Type of processing on ni interferogram (P)
	proc2	Type of processing on ni2 interferogram (P)
	pmode	Processing mode for 2D data (P)
	ssorder	Order of polynomial to fit digitally filtered FID (P)

ssfilter	Full bandwidth of digital filter to yield a filtered FID
	(P)
wft2d	Weight and Fourier transform 2D data (C)

wftld(coefficients) Weight and Fourier transform F2 of 2D data

Syntax	
Applicability	VnmrJ 3.1
Description	wftld and ftld perform the first Fourier transformation along the F2 dimension, with and without weighting respectively, and matrix transposition. This allows the display of t1 interferograms with the
	"dcon" and "dconi" commands.

ftlda Fourier transform phase-sensitive data (M)

Syntax	ft1da	(<arg1></arg1>	,	<arg2>)</arg2>
Dynuar	LCTUU	(\arg_r/	1	~urgz~/

Applicability VnmrJ 3.1

Description	Performs the first (f_2) transform of a 2D transform or the first part of a 3D transform. Otherwise, ftlda has the same functionality as the ft2da command. See the description of ft2da for further information. For information about Hadamard transforms, see the description of the proc1 parameter and the <i>VnmrJ NMR Liquids</i> user guide.		
Arguments	options are the same as used with ft2da. See ft2da for details.		
See also	NMR Spectro	scopy User Guide	
Related	ft2d	Fourier transform 2D data (C)	
	ft2da	Fourier transform phase-sensitive data (M)	
	wft1da	Weight and Fourier transform phase-sensitive data (M)	
	wft2da	Weight and Fourier transform phase-sensitive data (M)	

ftldac Combine arrayed 2D FID matrices (M)

Syntax	ft1dac<(<mult1><,mult2>,<,multn>)></mult1>
Applicability	VnmrJ 3.1
Description	Allows ready combination of 2D FID matrices within the framework of the 2D Fourier transformation program. No weighting is performed. ftldac requires that the data be acquired either without f_1 quadrature or with f_1 quadrature using the TPPI method. This macro is used for TOCSY (with multiple mixing times).

Arguments	mult1,mult2,,multn are multiplicative coefficients. The nth argument is a real number and specifies the multiplicative coefficient for the nth 2D FID matrix.		
Related	ft2dac Tocsy wft1da wft1dac	Combine arrayed 2D FID matrices (M) Set up parameters for TOCSY pulse sequence (M) Weight and Fourier transform phase-sensitive data (M) Combine arrayed 2D FID matrices (M)	

ftldac and wftldac Help file for wftldc macro used to combine arrayed 2D FID matrices

Syntax	wft1dac(<mult1> , <mult2> , <mult3> ,)</mult3></mult2></mult1>
Applicability	VnmrJ 3.1
Description	This macro allows the ready combination of 2D FID matrices within the framework of the 2D-FT program.
Arguments	The nth argument is a real number and specifies the multiplicative coefficient for the nth 2D FID matrix. It currently requires that the data be acquired either without F1 quadrature or with F1 quadrature using the TPPI method. WFT1DAC functions in an analogous manner.
Examples	E.COSY and TOCSY (with multiple mixing times).

ft2d Fourier transform 2D data (C)

Syntax	<pre>(1) ft2d(array_element) (2) ft2d('nf'<array_element>) (3) ft2d<(<options,><plane_number,><coefficients>)> (4) ft2d('ni' 'ni2',element_number,increment) (5) ft2d('ni' 'ni2',increment,<coefficients>)</coefficients></coefficients></plane_number,></options,></array_element></pre>
Applicability	VnmrJ 3.1
Description	Performs the complete 2D Fourier transformation, without weighting, in both dimensions. If the first Fourier transformation has already been done using ftld, wftld, ftlda, or wftlda, the ft2d command performs only the second (t_1) transform.
	For arrayed 2D FID data, a single array element can be weighted and transformed using syntax 1. If the data is collected in "compressed" form using 'nf', syntax 2 must be used. Complex and hypercomplex interferograms can be constructed explicitly by supplying a series of coefficients using syntax 3. If an arrayed 3D data set is to be selectively processed, the format of the arguments to ft2d changes to syntax 4. For example, ft2d('ni',1,2) performs a 2D transform along np and ni of the second ni2 increment and the first element within the explicit array. This command yields a 2D np-ni frequency plane.

Arrayed 3D data sets can also be subjected to 2D processing to yield 2D absorptive spectra. If the States-Haberkorn method is used along both f_1 (ni dimension) and f_2 (ni2 dimension), there are generally 4 spectra per (ni,ni2) 3D element. In this case, using syntax 5, entering ft2d('ni2',2,<16 coefficients>) performs a 2D transform along np and ni2 of the second ni increment using the 16 coefficients to construct the 2D t₁-interferogram from appropriate combinations of the 4 spectra per (ni,ni2) 3D element.

If there are n data sets to be transformed, as in typical phase-sensitive experiments, 4*n coefficients must be supplied. The first 2*n coefficients are the contributions to the real part of the interferogram, alternating between absorptive and dispersive parts of the successive data sets. The next 2*n coefficients are the contributions to the imaginary part of the interferogram, in the same order. Thus, using the definition that the first letter refers to the source data set, the second letter refers to the interferogram, and the number identifies the source data set, we have the following cases:

Data sets Coefficient order

1	RR1,	IR1,	RI1,	II1				
2	RR1,	IR1,	RR2,	IR2,	RI1,	II1,	RI2,	II2
3	RR1,	IR1,	RR2,	IR2,	RR3,	IR3,	RI1,	II1,
	RI2,	II2,	RI3,	II3				

The coefficients are often 1, 0, or -1, but this is not always the case. Any non-integral coefficient can be used, and as many coefficients can be nonzero as is desired. Up to 32 coefficients can be supplied, which at 4 per data set allows the addition, subtraction, etc., of eight 2D data sets (e.g., 8 different phase cycles).

For information on real as opposed to complex Fourier transforms, see the descriptions of the proc, proc1, and proc2 parameters. For information about Hadamard transforms, see the description of the proc1 parameter and the *VnmrJ NMR Liquids* user guide. For information on left-shifting, zero-order phase rotation, and frequency shifting of the FID and interferogram time-domain data during the 2D Fourier transformation, see the descriptions of the parameters lsfid, lsfid1, lsfid2, phfid, phfid1, phfid2, lsfrq, lsfrq1, and lsfrq2, as appropriate. For information on the lfs (low-frequency suppression) and zfs (zero-frequency suppression) solvent suppression options, see the description of parameters ssfilter and ssorder, and macro parfidss.

Arguments array_element is a single array element to be transformed.

options can be any of the following (all string arguments must precede the numeric arguments):

- 'ptype' is a keyword to transform P-type data to yield a P-type contour display.
- 'ntype' is a keyword to transform N-type data to yield a P-type contour display. This is the default.

- 't2dc' is a keyword to apply a dc correction to each t_2 FID prior to the first Fourier transform. The last 1/16-th of the time domain data is used to calculate the dc level.
- 'tldc' is a keyword to apply a dc correction to each t_1 interferogram prior to the second Fourier transform. The last 1/16-th of the time domain data is used to calculate the dc level.
- 'f2sel' is a keyword to allow only preselected f_2 regions to be transformed along t_1 . The t_1 interferograms in the non-selected f_2 regions are zeroed but *not* transformed. The same mechanism used to select baseline regions for baseline correction (bc) is used to select the f_2 regions to be transformed along t_1 . Set intmod='partial' and partition the integral of the spectrum into several regions. The even numbered f_2 regions (e.g., 2, 4, 6) are transformed along t_1 ; the odd numbered regions are not transformed along t_1 .
- 'nf' is a keyword to transform arrayed or multi-slice 2D data that has been collected in the compressed form as single 2D FIDs with multiple (nf) traces.
- 'ni2' is a keyword to transform non-arrayed 2D data that have been collected with ni2 and sw2 (instead of ni and sw1). addpar('3d') creates the necessary processing parameters for the 'ni2' operation.
- 'noop' is a keyword to not perform any operation on the FID data. This option is used mainly to allow macros, such as wft2da, to have the same flexibility as commands.

coefficients are a series of coefficients according to the following scheme: RR1 is the coefficient used to multiply the real part (first R) of spectra set 1 before it is added to the real part (second R) of the interferogram. IR2 would thus represent the contribution from the imaginary part of spectra set 2 to the real part of the interferogram, and so forth. The scheme is depicted below.

ft2d(RR1,IR1,RR2,IR2,...,RI1,II1,RI2,II2,...)

where:

```
RR1*REAL(w2,element=1) -> REAL(t1)
IR1*IMAG(w2,element=1) -> + REAL(t1)
RR2*REAL(w2,element=2) -> + REAL(t1)
IR2*IMAG(w2,element=2) -> + REAL(t1)
. . .
RI1*REAL(w2,element=1) -> IMAG(t1)
II1*IMAG(w2,element=1) -> + IMAG(t1)
RI2*REAL(w2,element=2) -> + IMAG(t1)
II2*IMAG(w2,element=2) -> + IMAG(t1)
```

'ni' is a keyword to selectively transform a particular np-ni 2D plane within a non-arrayed 3D data set. To identify the plane, 'ni' is followed by the plane_number argument, an integer from 1 through ni2.

'ni2' is a keyword to selectively transform a particular np-ni2 2D plane within a non-arrayed 3D data set. To identify the plane, 'ni2' is followed by the plane_number argument, an integer from 1 through ni.

<code>element_number</code> is the number of an element within the explicit array when selectively processing an arrayed 3D data set; it ranges from 1 to ni2

increment is the increment within the explicit array when selectively
processing an arrayed 3D data set; it ranges 1 to
arraydim/(ni*ni2).

Related dconi Interactive 2D data display (C)

licu	aconi	interactive 2D data display (0)
	dcrmv	Remove dc offsets from FIDs in special cases (P)
	fpmult	First point multiplier for np FID data (P)
	fpmult1	First point multiplier for ni interferogram data (P)
	ft1d	Fourier transform along f_2 dimension (C)
	lsfid	Number of complex points to left-shift np FID (P)
	lsfid1	Number of complex points to left-shift ni
		interferogram (P)
	lsfid2	Number of complex points to left-shift ni2
		interferogram (P)
	lsfrq	Frequency shift of the fn spectrum (P)
	lsfrq1	Frequency shift of the fn1 spectrum (P)
	lsfrq2	Frequency shift of the fn2 spectrum (P)
	parfidss	Create parameters for time-domain solvent
		subtraction (M)
	phfid	Zero-order phasing constant for np FID (P)
	phfid1	Zero-order phasing constant for ni interferogram
		(P)
	phfid2	Zero-order phasing constant for ni2 interferogram
		(P)
	proc	Type of processing on np FID (P)
	proc1	Type of processing on ni interferogram (P)
	proc2	Type of processing on ni2 interferogram (P)
	pmode	Processing mode for 2D data (P)
	ssorder	Order of polynomial to fit digitally filtered FID (P)
	ssfilter	Full bandwidth of digital filter to yield a filtered
		FID (P)
	wft1d	Weight and Fourier transform f_2 for 2D data (C)
	wft2d	Weight and Fourier transform 2D data (C)

wft2d(coefficients) Weight and Fourier transform 2D data

Syntax Applicability VnmrJ 3.1

wft2d and ft2d perform the complete 2D Fourier transformation, with and without weighting in both dimensions respectively. For arrayed 2D FID data, a single array element can be transformed using, as an example, "ft2d(array element number)". Complex and Hypercomplex interferograms can be constructed explicitly using the following coefficient table: ft2d(rr1,ir1,rr2,ir2,...,ri1,ii1,ri2,ii2,...) where rr1 * REAL(w2, element=1) --> REAL(t1) ir1 * IMAG(w2, element=1) --> + REAL(t1) rr2 * REAL(w2, element=2) --> + REAL(t1)ir2 * IMAG(w2, element=2) --> + REAL(t1)[etc.] ri1 * REAL(w2, element=1) --> IMAG(t1) ii1 * IMAG(w2, element=1) \rightarrow + IMAG(t1) ri2 * REAL(w2, element=2) \rightarrow + IMAG(t1) ii2 * IMAG(w2, element=2) --> + IMAG(t1)[etc.]

Arrayed hypercomplex data can be transformed by supplying the array index followed by the eight coefficients needed to construct the interferograms:

ft2d(array_element_number, rr1,ir1,rr2,ir2,ri1,ii1,ri2,ii2)

This is used in the special case where phase=1,2 and phase has the highest precedence in the array parameter, as for example, array='gzlvl1,phase'.

ft2d('ptype') will transform P-type data to yield a P-type contour display. ft2d('ntype') will transform N-type data to yield a P-type contour display. The same applies to wft2d. Although ft1d and wft1d will accept the string arguments 'ptype' and 'ntype', it serves no useful function in these two commands since the differential effect of these two arguments is applied only during the course of the second Fourier transformation.

ft2d('t2dc') causes a DC correction to be applied to each t2 FID prior to the first FT; ft2d('t1dc') causes a DC correction to be applied to each t1 interferogram prior to the second FT. In both cases, the last 1/16-th of the time domain data is used to calculate the DC level.

ft2d('f2sel') allows only pre-selected F2 regions to be transformed along t1; the t1 interferograms in the non-selected F2 regions are zeroed but NOT transformed. The same mechanism used to select baseline regions for baseline correction (bc) is used to select the F2 regions which are to be transformed along t1. Set intmod='partial' and partition the integral of the spectrum into several regions. The even numbered F2 regions, e.g., 2, 4, etc., will be transformed along t1; the odd numbered ones will not be transformed along t1.

ft2d('nf') transforms a non-arrayed 2D experiment which has been collected as a single 2D FID with multiple (nf) traces. In this example, each trace of the 2D FID corresponds to t2 time domain data collected at an incremented value of t1.

ft2d('nods') and wft2d('nods') prevents the spectrum display following the transform.

The 'noft' option to ftld, wftld, ft2d, and wft2d prevents the actual Fourier transform step. ftld, ft2d ('noft') will Fourier transform the t2 time domain data but not the resulting t1 interferograms. Both axes will be treated as frequency axes. ft2d('noft') will present the FID data, interpreted as if both axes are frequency axes. Other operations, such as weighting, solvent suppression, etc., will be performed as requested. Just the actual FT step is bypassed when this option is given.

ft2d ('noop') does not perform any operation on the FID data. It is used mainly to allow macros, e.g., wft2da, to have the same flexibility as actual VNMR commands.

ft2d ('ni2') transforms non-arrayed 2D data which have been collected with ni2 and sw2 (instead of ni and sw1). par3d creates the necessary processing parameters for the ft2d('ni2') operation. ft2d('ni',#) is used to selectively transform a particular "np-ni" 2D plane within a non-arrayed 3D data set; # is an integer which can range from 1 to ni2 in this example. ft2d('ni2',#) is used to selectively transform a particular "np-ni2" 2D plane within a non-arrayed 3D data set; # is an integer which can range from 1 to ni in this example. If an arrayed 3D data set is to be selectively processed, the format of the arguments to ft2d changes. For example, ft2d('ni',#1,#2) performs a 2D transform along np and ni of the #2-th ni2 increment and the #1-th element within the explicit array. This yields a 2D "np-ni" frequency plane. #1 ranges from 1 to ni2; and #2, from 1 to [arraydim/(ni*ni2)].

Arrayed 3D data sets can also be subjected to 2D processing to yield 2D absorptive spectra. If the States-Haberkorn method is used along both F1 (ni dimension) and F2 (ni2 dimension), there will generally be 4 spectra per (ni,ni2) 3D element. In this case, the command ft2d('ni2',#1, <16 coefficients>) would perform a 2D transform along np and ni2 of the #1-th ni increment using the ensuing 16 coefficients to construct the 2D t1-interferogram from appropriate combinations of the 4 spectra per (ni,ni2) 3D element.

See also For information on real vs. complex Fourier transforms, see the manual entry for "proc#". For information on left-shifting, zero-order phase rotation, and frequency shifting of FID and/or interferogram time-domain data during the 2D FT, see manual entries for "lsfid#", "phfid#", or "lsfrq#" respectively. For information on the lfs and zfs solvent suppression options, see manual entries for "ssfilter", "ssorder", and parfidss.

For information on Hadamard transforms, see the manual entries for "ht" and "proc1".

Related	phfid	parameter
	lsfid	parameter
	phfid1	parameter
	lsfid1	parameter
	phfid2	parameter
	lsfid2	parameter

F

proc	parameter
proc1	parameter
proc2	parameter
pmode	parameter
ssorder	parameter
ssfilter	parameter
parfidss	command

ft2da Fourier transform phase-sensitive data (M)

Syntax ft2da(<arg1> , <arg2>)

Applicability VnmrJ 3.1

Description

Processes 2D FID data and 2D planes at particular t_1 or t_2 times from a 3D data set for a pure absorptive display. ft2da differs from wft2da only in that, in the case of wft1da, weighting of the time-domain data is performed prior to the FT. ft2da functions analogously to ft1da and wft1da, except that ft2da and wft2da perform only the f_2 Fourier transform. For information about Hadamard transforms, see the description of the proc1 parameter and the *VnmrJ NMR Liquids* user guide.

Macros ftlda, wftlda, ft2da, and wft2da function for hypercomplex 2D FID data (phase=1,2) and for TPPI 2D FID data (phase=3 or phase=1,4) acquired either with ni or ni2. If the data were acquired with ni, no additional arguments need be used with the macros. If the data were acquired with ni2, the keyword 'ni2' must be used.

For phase=1,2:wft2da=wft2d('ptype',1,0,0,0,0,0,1,0)

For phase=3: wft2da=wft2d(1,0,0,0)

For phase=1,4:wft2da=wft2d('ptype',1,0,0,0,0,0,1,0)

Macros ftlda, wftlda, ft2da, and wft2da support selective 2D processing within a 3D FID data set. All permutations of hyercomplex and TPPI modes of data acquisition in t_1 and t_2 can be handled. For selective f_2f_3 processing, the numeric argument immediately following the 'ni2' keyword is interpreted to be the t_1 increment number, which specifies the particular f_2f_3 plane (plane_number, see below) to be processed. For selective f_1f_3 processing, the t_2 increment number either follows the keyword 'ni', which is optional, or is associated with the first numeric argument that does not immediately follow a 'bc' keyword.

For information on real as compared to complex Fourier transformation, see the description of proc or proc1. For information on the lfs (low-frequency suppression) and zfs (zero-frequency suppression) solvent suppression options, see the description of parameters ssfilter and ssorder, and the macro parfides.

Arguments options can be any of the following (the order is not important):

to the ft2d command for a description of these options.

- 'bc' is a keyword for a baseline correction of the phase-corrected f_2 spectra prior to the f_1 Fourier transform. The baseline regions must have been previously determined. A polynomial order of 1 (a spline fit) or a higher polynomial order must be specified by inserting a numerical argument following 'bc'.
- 'dc' is a keyword for a drift correction (dc) of the f_2 spectra prior to the f_1 Fourier transformation.
- 'ni' is a keyword to selectively transform a particular np-ni 2D plane within a non-arrayed 3D data set. To identify the plane, 'ni' is followed by plane_number, an integer from 1 through ni2.
- 'ni2' is a keyword to selectively transform a particular np-ni2 2D plane within a non-arrayed 3D data set. To identify the plane, 'ni2' is followed by plane_number, an integer from 1 through ni.
- 'old' is a keyword to allow data acquired before the February 25, 1988, software release to be processed correctly. 'old' does not function for selective 2D processing within 3D data sets. If no ni2 or ni plane_number is given, it is assumed that the data set is only 2D in either ni2 or ni, respectively.

See also NMR Spectroscopy User Guide

Related	flcoef	Coefficient to construct F1 interferogram (P)
	f2coef	Coefficient to construct F2 interferogram (P)
	ft1da	Fourier transform phase-sensitive data (M)
	parfidss	Create parameters for time-domain solvent subtraction
		(M)
	phase	Phase selection (P)
	proc	Type of processing on the np FID (P)
	proc1	Type of processing on the ni interferogram (P)
	ssorder	Order of polynomial to fit digitally filtered FID (P)
	ssfilter	Full bandwidth of digital filter to yield a filtered FID
		(P)
	wft1da	Weight and Fourier transform phase-sensitive data (M)
	wft2da	Weight and Fourier transform phase-sensitive data (M)

ft2dac Combine arrayed 2D FID matrices (M)

Syntax	ft2dac<(<mult1><,mult2>,<,multn>)></mult1>		
Applicability	VnmrJ 3.1		
Description	Allows ready combination of 2D FID matrices within the framework of the 2D FT program. No weighting is performed. Data must be acquired		
	either without f_1 quadrature or with f_1 quadrature using the TPPI		
	method. ft2dac is used with TOCSY (with multiple mixing times).		

F
Arguments	nents mult1,mult2,,multn are multiplicative coefficients. The nt argument is a real number and specifies the coefficient for the FID matrix.	
Related	ft1dac	Combine arrayed 2D FID matrices (M)
	Tocsy	Set up parameters for a TOCSY pulse sequence (M)
	wft1dac	Combine arrayed 2D FID matrices (M)
	wft2dac	Combine arrayed 2D FID matrices (M)

ft2dac and wft2dac Help file for wft2dc macro used to combine arrayed 2D FID matrices

Syntax	wft2dac(<mult1> , <mult2> , <mult3> ,)</mult3></mult2></mult1>
Applicability	VnmrJ 3.1
Description	This macro allows the ready combination of 2D FID matrices within the framework of the 2D-FT program. The nth argument is a real number and specifies the multiplicative coefficient for the nth 2D FID matrix. It currently requires that the data be acquired either without F1 quadrature or with F1 quadrature using the TPPI method. WFT2DAC functions in an analogous manner.
Examples	E.COSY and TOCSY (with multiple mixing times)

ft3d Perform a 3D FT on a 3D FID data set

Syntax	ft3d(<>)
Applicability	VnmrJ 3.1
Description	ft3d is a macro which executes the program ft3d in the VNMR system `bin` directory (\$vnmrsystem/bin). The environmental parameter PATH specifies the list of directories through which UNIX searches until it finds an executable ft3d program.
Arguments	The first string argument which is a non-keyword is 'datadir'. 'datadir' (without the /data subdirectory appended) is an optional argument which specifies the output directory for the 3D spectral data file(s). The default directory for the 3D spectral data is curexp/datadir3D. nfiles (an integer) is an optional argument which specifies the number of 3D data files (data1 to data`nfiles`) used to store the transformed 3D data. nfiles must be <= 32. If nfiles is entered, distributed F1F2 processing will be performed by the ft3d program if possible.
	If the optional keyword 'nocoef' is submitted as an argument to the ft3d macro, VNMR will not create a 3D coefficient file prior to invoking the ft3d program. This is useful if one has modified an existing 3D coefficient file and does not want it to be overwritten prior to the 3D transform. By default, ft3d calls the make3dcoef macro to create a coefficient file using flcoef and f2coef string parameter values.

The 't1t2' and 't2t1' are optional arguments to explicitly define the order of t1 and t2 arrays (other than ni and ni2). By default the macro looks at array parameter to make a decision and in that case if any parameter other than phase and phase2 are arrayed the macro aborts.

The next set of optional keywords for ft3d pertain to plane extraction following the complete 3D FT. 'xall' indicates that all three 2D plane types, F1F3, F2F3, and F1F2, are to be automatically extracted at the end of the 3D FT. The output directory for the extracted 2D planes is the same as that for the 3D spectral data except that the former uses the /extr subdirectory whereas the latter uses the /data subdirectory. 'f1f3', 'f2f3', and 'f1f2' can be used to select any combination of plane types to be extracted. The ft3d macro allows the user to submit any of these keywords more than once. The program getplane, however, will display an error and abort if any one plane type is multiply defined for extraction.

The 3D FID data must be loaded into the experiment in which the ft3d macro is to be run. The ft3d program is started up in background mode by this macro so that VNMR remains free for interactive processing. In other words, one can start a 3D transform from within exp4 and, at the same time, continue with any 1D or 2D processing of the 3D FID data within the same experiment using VNMR. If the 'fg' argument is given to ft3d, then the processing is done if foreground. No additional processing will be possible until the ft3d program has finished.

The optional 'noft' argument is similar to the 'noft' arguments to ft2d and ft. The Fourier transform step will be skipped in all three dimensions. In contradistinction to the 1D and 2D analogs, the 'noft' argument to ft3d causes all processing to be skipped; no weighting, phasing, etc. are performed. All axes will be treated as frequency axes. Within the /data 3D data subdirectory, there are the following files and

further subdirectories:

- data1 through data#: These are the actual binary 3D spectral data files. The number of data files depends upon the size of the largest 2D plane and the value for the UNIX environmental parameter `memsize` if nfiles is not entered.
- info: This is a directory which stores the 3D coefficient text file (coef), the binary information file (procdat), the 3D parameter set (procpar3d), and the automation file (auto). The first three files are created by the set3dproc() command within VNMR. The last file is created by the ft3d program.
- log: This is a directory which stores the log files produced by the ft3d program. f3 contains all log output for the F3 transform. For the F2 and F1 transforms, there are two log file for each data file, one for the F2 transform (f2.#) and one for the F1 (f1.#). The master one for the F2 transform and one for the F1. The file contains all the log output produced by the master ft3d program.

		e
Related	set3dproc	command
	killft3d	macro
	getplane	macro
	make3dcoef	macro
	fiddc3d	parameter
	specdc3d	parameter
	ptspec3d	parameter
	ssfilter	parameter
	ssorder	parameter
	ntype3d	parameter
	flcoef	parameter
	f2coef	parameter

The order of the arguments to the ft3d macro is not important.

ftargs Macro to create parameters

Description ftargs is a macro to create the parameters used by ft('ftargs') and wft('ftargs').

full Set display limits for a full screen (C)

Applicability V	VnmrJ	3.1
-----------------	-------	-----

Description Sets the horizontal control parameters (sc and wc) and the vertical control parameters (sc2 and wc2) to produce a display (and subsequent plot) on the entire screen (and page). For 2D data, space is left for the scales.

Related	center	Set display limits for center of screen (C)
	fullt	Set display limits for full screen with room for traces (C)
	left	Set display limits for left half of screen (C)
	right	Set display limits for right half of screen (C)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	WC	Width of chart (P)
	wc2	Width of chart in second direction (P)

fullsq Display largest square 2D display (M)

Description Adjusts sc, sc2, wc, and wc2 parameters to show the largest possible square 2D display.

Related	full	Set display limits for a full screen (C)
	fullt	Set display limits for a full screen with room for
		traces (C)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	WC	Width of chart (P)
	wc2	Width of chart in second direction (P)

fullt Set display limits for a full screen with room for traces (C)

Applicability VnmrJ 3.1

Description Sets the horizontal control parameters (sc and wc) and the vertical control parameters (sc2 and wc2) to produce a display (and subsequent plot) in the entire screen (and page) with room for traces (dconi). For 2D data, space is left for the scales.

Related	center	Set display limits for center of screen (C)
	full	Set display limits for a full screen (C)
	left	Set display limits for left half of screen (C)
	right	Set display limits for right half of screen (C)



Agilent VnmrJ 4 Command and Parameter Reference Guide

g2pul_ecc	Setup macro for eddy current compensation parameters (M)
ga	Submit experiment to acquisition and FT the result (M)
gain	Receiver gain (P)
gap	Find gap in the current spectrum (M)
gaussian	Set up unshifted Gaussian window function (M)
gcal_	Local value of the conversion factor between gradient in DAC points and gradient in G/cm (P)
gcal	Gradient calibration constant (P)
gcoil	Current gradient coil (P)
Gcosy	Convert the parameter to a gradient COSY experiment (M)
gdiff	Diffusion gradient level (P)
Gdqcosy	Convert the parameter to a gradient DQCOSY experiment (M)
get1d	Select a 1D experiment for processing (M)
get2d	Select a 2D experiment for processing (M)
getdim	Return dimensionality of experiment (M)
getemailaddr	Get email addresses from a file
geterror	Return or display an acquisition error
getfile	Get information about directories and files (C)
getgamma	Retrieves Gamma from /vnmr/nuctabref
getht	Retrieve/Save a Hadamard frequency list from a file
getlcdata	An LC-NMR communications macro
getlimit	Get the limits of a variable in a tree (C)
getll	Get intensity and line frequency of line (C)



getmodule	Get module (C)
getoffset	Sets offset based on current reference parameters
getparam	Retrieve parameter from probe file (M)
getplane	Extract planes from a 3D spectral data set (M)
getplottertype	Retrieves plotter information
getppm	Returns Cursor Value in ppm
getreg	Get frequency limits of a specified region (C)
getsampglobal	Loads sample global parameters
getshimmethods	Get proshim methods list (M)
getsn	Get signal-to-noise estimate of a spectrum (M)
gettoken	Utility macro to separate a string into tokens (M)
gettxt	Get text file from VnmrJ data file (C)
gettype	Get the type of a variable (C)
getvalue	Get value of parameter in a tree (C)
gf	Prepare parameters for FID/spectrum display in acqi (M)
gf	Gaussian function in directly detected dimension (P)
gf1	Gaussian function in 1st indirectly detected dimension (P)
gf2	Gaussian function in 2nd indirectly detected dimension (P)
gflow	Flow encoding gradient level (P)
gfs	Gaussian shift const. in directly detected dimension (P)
gfs1	Gaussian shift const. in 1st indirectly detected dimension (P)
gfs2	Gaussian shift const. in 2nd indirectly detected dimension (P)
Ghmbc	Convert the parameter to a gradient HMBC experiment (M)
ghmqc	Set up a PFG HMQC pulse sequence (M)
Ghmqc	Convert the parameter to a gradient HMQC experiment (M)
gHMQC15	Set up parameters for ¹⁵ N gHMQC experiment (M)

gHMQC_d2	Set up parameters for ¹⁵ N gHMQC experiment using dec. 2 (M)
gHMQC_d213	Set up parameters for ¹³ C gHMQC experiment using dec. 2 (M)
ghmqcps	Set up a PFG HMQC phase-sensitive pulse sequence (M)
ghsqc	Set up a PFG HSQC pulse sequence (M)
Ghsqc	Convert the parameter to a gradient HSQC experiment (M)
gHSQC15	Set up parameters for ¹⁵ N gHSQC experiment (M)
gHSQC_d2	Set up parameters for ¹⁵ N gHSQC experiment using dec. 2 (M)
gHSQC_d213	Set up parameters for ¹³ C gHSQC experiment using dec. 2 (M)
Ghsqctoxy	Convert parameters for gradient HSQCTOXY experiment (M)
gilson	Open the Gilson Liquid Handler window (C)
gilson	Allow starting the Gilson Liquid Handler GUI (M)
gin	Return current mouse position and button values (C)
globalauto	Automation directory name (P)
glue	Create a pseudo-2D dataset (M)
gmapshim	Start gradient autoshimming (M)
gmapshim_au	Start acquisition with gradient shimming (M)
gmapspin	Enable or disable spinning during gradient shimming (P)
gmapsys	Run gradient autoshimming, set parameters, map shims (M)
gmapz	Get parameters and files for gmapz pulse sequence (M)
gmap_findtof	Gradient shimming flag to first find tof (P)
gmap_z1z4	Gradient shimming flag to first shim z1-z4 (P)
gmax	Maximum gradient strength (P)
gmqcosy	Set up PFG absolute-value MQF COSY parameter set (M)
gnoesy	Set up a PFG NOESY parameter set (M)
go_ <pslabel></pslabel>	Experiment-Specific Runtime Macro

go	Submit experiment to acquisition (M)
gradfit	Calculates fit coefficients describing the variation of gradient strength with position in calibration of non-uniform pulsed field gradients
go_	Pulse sequence setup macro called by go, ga, and au (M)
gpat-gpat3	Gradient shape (P)
gplan	Start interactive image planning (C)
gradientdisable	Disable PFG gradients (P)
gradientshaping	Activate shaping on the gradient pulses (P)
gradstepsz	Gradient step size (P)
gradtype	Gradients for X, Y, and Z axes (P)
graphis	Return the current graphics display status (C)
grayctr	Gray level window adjustment (P)
graysl	Gray level slope (contrast) adjustment (P)
grecovery	Eddy current testing (M)
grid	Draw a grid on a 2D display (M)
groupcopy	Copy parameters of group from one tree to another (C)
gspoil	Spoiler gradient level (P)
gsspat	Slice-select gradient shape (P)
gtnnoesy	Set up a PFG TNNOESY parameter set (M)
gtnroesy	Set up a PFG absolute-value ROESY parameter set (M)
gtotlimit	Gradient total limit (P)
gtrim	Trim gradient level (P)
gxmax,gymax,gzmax	Maximum gradient strength for each axis (P)
gzlvl	Pulsed field gradient strength (P)
gzsize	Number of z-axis shims used by gradient shimming (P)
gzwin	Spectral width percentage used for gradient shimming (P)

g2pul_ecc Setup macro for eddy current compensation parameters (M)

Applicability Systems with Agilent Cold Probes

Description Setup macro for pulse sequence used to determine the eddy current compensation parameters.

ga Submit experiment to acquisition and FT the result (M)

Syntax ga<(<'nocheck'><,'next'><,'wait'>)>

Description Performs experiment described by the current acquisition parameters, checking parameters loc, spin, gain, wshim, load, and method to determine the necessity to perform various actions in addition to simple data acquisition. This may involve a single FID or multiple FIDs, as in the case of arrays or 2D experiments. ga causes the data to be automatically weighted and Fourier transformed (wft) at the end of each FID data acquisition.

> Before starting the experiment, ga executes two user-created macros if they exist. The first is usergo, a macro that allows the user to set up general conditions for the experiment. The second is a macro whose name is formed by go_ followed by the name of the pulse sequence (from seqfil) to be used (e.g., go_s2pul, go_dept). The second macro allows a user to set up experiment conditions suited to a particular sequence.

Arguments 'nocheck' is a keyword to override checking if there is insufficient free disk space for the complete 1D or 2D FID data set to be acquired. 'next' is a keyword to put the experiment started with ga('next')

at the head of the queue of experiments to be submitted to acquisition.

'wait' is a keyword to stop submission of experiments to acquisition until wexp processing of the experiment, started with ga('wait'), is finished.

See also NMR Spectroscopy User Guide

Related	au	Submit experiment to acquisition and process data (M)
	change	Submit a change sample experiment to acquisition (M)
	gain	Receiver gain (P)
	go	Submit experiment to acquisition (M)
	go_	Pulse sequence setup macro called by go, ga, and au
		(M)
	load	Load status of displayed shims (P)
	loc	Location of sample in tray (P)
	lock	Submit an Autolock experiment to acquisition (C)
	method	Autoshim method (P)
	sample	Submit change sample, Autoshim experiment to acquisition (M)

seqfil	Pulse sequence name (P)
shim	Submit an Autoshim experiment to acquisition (C)
spin	Submit a spin setup experiment to acquisition (C)
spin	Sample spin rate (P)
su	Submit a setup experiment to acquisition (M)
usergo	Experiment setup macro called by go, ga, and au (M)
wft	Weight and Fourier transform 1D data (C)
wshim	Conditions when shimming is performed (P)

gain Receiver gain (P)

Description Sets receiver gain or, by setting gain='n', enables Autogain for automatic adjustment of gain. Low gain in multiline, high-dynamic-range samples can cause a number of problems, including intermodulation distortions and extra lines in the spectrum. Too high a gain, on the other hand, can cause receiver overload and consequent baseline distortions. Autogain capability allows the observe channel to be set optimally for detecting and digitizing NMR signals from a wide variety of samples.

Autogain adjusts the observe channel gain such that the NMR signal takes about 50 percent of the maximum range of the ADC. This setting allows a comfortable leeway for variations in signal. The program begins acquisition in the normal manner but the first transient (after any requested steady state transients) is examined for signal level. If the intensity is too low or too high, the gain is changed and the process is repeated until the intensity is within the proper range, and then normal acquisition commences. The final gain value used for the experiment is stored and when the experiment is finished, setting gain='y' results in the value being displayed in the dgs parameter group.

If the gain is reduced by the Autogain procedure such that the noise does not trigger the least significant 1 or 2 bits in the ADC and the signal still overloads either the receiver or ADC, the system stops and displays a message indicating Autogain failure.

Values 0 to 60, in steps of 2 dB (60 represents highest possible receiver gain and 0 lowest). On 500-750-MHz systems, low-band gain is limited from 18 to 60.

'n' enables Autogain, in which the gain is automatically adjusted at the start of acquisition for an optimum value. After the acquisition is finished, setting gain='y' then allows the value of gain to be read. gain='n' may not be used for arrayed experiments.

See also NMR Spectroscopy User Guide

Related dgsDisplay group of special/automation parameters (M)gfPrepare parameters for FID/spectrum display in acqi (M)

gap Find gap in the current spectrum (M)

Syntax gap(gap,height):found,position,width

Description Looks for a gap between the lines of the currently displayed spectrum. It can be used to automatically place inserts, parameter printouts, trace labels, etc. The search starts on the left side (low-field end) of the spectrum.

Arguments gap is the width of the desired gap.

height is the starting height (same as the lower limit for the insert).

found is a return value that is set to 1 if the search is successful, or set to 0 if unsuccessful.

position is a return value that is set to the distance from the left edge of the chart (not the plot) to the left end of the gap (3 mm from the nearest peak to the left, positioning with "left gravity") if the search is successful, or set to the position (no spacing to the nearest line) of the largest gap found if unsuccessful.

width is a return value set to the total width of the first gap if the search is successful, or set to the width of largest gap found if unsuccessful.

Examplesgap(120,80);\$1,\$2,\$3See alsoUser Programming

gaussian Set up unshifted Gaussian window function (M)

Syntax gaussian<(<t1_inc><,t2_inc>)>

Description	Sets up an unshifted Gaussian window function in 1, 2, or 3 dimensions. The macro checks whether the data is 1D, 2D, and 3D.		
Arguments	t1_inc is the number of t1 increments. The default is ni. t2_inc is the number of t2 increments. The default is ni2.		
See also	NMR Spectroscopy User Guide		
Related	ni	Number of increments in 1st indirectly detected dimension (P)	
	ni2	Number of increments in 2nd indirectly detected dimension (P)	
	pi3ssbsq	Set up pi/3 shifted sinebell-squared window function (M)	
	pi4ssbsq	Set up pi/4 shifted sinebell-squared window function (M)	
	sqcosine	Set up unshifted cosine-squared window function (M)	
	sqsinebell	Set up unshifted sinebell-squared window function (M)	

gcal_ Local value of the conversion factor between gradient in DAC points and gradient in G/cm

Syntax	gcal_
Applicability	VnmrJ 3.1
Description	gcal_ is a local copy of the conversion factor from DAC points to G/cm for the probe used. gcal_ is set equal either to the value in the current probe file, if available, or to the global value gcal, by the macro makedosyparams invoked when a DOSY pulse sequence is run, and does not normally need to be set manually.
See also	gcal

gcal Gradient calibration constant (P)

Applicability	Systems with the pulsed field gradient or the imaging module.
Description	Stores the proportionality constant between the parameter values (DAC units) controlling the desired gradient and the intensity of the gradient expressed in gauss/cm. The gradients generated in the magnet require calibration of the gain on the gradient compensation board so that coordinate data, slice positions, and the field of view can be set up accurately. gcal should be located in each user's vnmrsys/global file.
Values	Number that is probe dependent, in gauss/cm-DAC unit. On the Performa I PFG module, 0.00028 to 0.00055 gauss/cm-DAC unit is nominal; On the Performa II, 0.0014 to 0.0025 gauss/cm-DAC unit is nominal.
See also	VnmrJ Imaging NMR
Related	setgcal Set gradient calibration constant (M)

gcoil Current gradient coil (P)

Description Reserved parameter that specifies which physical gradient set is currently installed. This allows convenient updating of important gradient characteristics when one gradient set is interchanged for another. When set, gcoil reads the gradient table file of the same name in /vnmr/imaging/gradtables and sets the gradient calibration parameters.

gcoil is local to each individual experiment. It is normally set the same as sysgcoil for acquiring new data, but can be set to other gradient names when working with saved data or data from another instrument. Each possible gradient name should have an associated file of that name located in the directory /vnmr/imaging/gradtables. Look at any file in this directory for an example of the proper

gradtable format, or use the macro creategtable to make new gradtables entries.

If the parameter gcoil does not exist in a parameter set and a user wants to create it, you must set the protection bit that causes the macro _gcoil to be executed when the value for gcoil is changed. There are two ways to create gcoil:

- Use the macro updtgcoil, which will create the gcoil parameter if it does not exist and set the correct protection bits.
- Enter the following commands:

create('gcoil','string') setprotect('gcoil','set',9)

gcoil and the associated gradient calibration parameter gmax is updated with the values listed in the table on the right each time a parameter set is retrieved, or when an experiment is joined. In the rare case that a gradtables file is

			-
	hh.	I A	•
- 1 6	aD.	IC.	т.

Variable Name	Value
boresize	22.50 cm
qmax	5.00 gauss/cm

modified, but the value of gcoil is not changed, manually force an update of the calibration parameters. Updating may be accomplished either by setting gcoil to itself, for example, gcoil=gcoil, or by using the macro _gcoil.

Be aware that if an old dataset is returned and processed, gradient parameters associated with that dataset will replace any new gcoil parameters.

	The table is a grad coil name: asg33) f	ient table (gradient for a horizontal	Table 2:	
	to the same maximum gradient strength.		Variable Name	Value
			boresize	5.10 cm
	On the right is a gradient table		trise	0.000200 sec
	(gradient coil name three-axis gradient maximum gradient	e: tc203) for a set with unequal strength.	gxmax	29.00 gauss/cm
lee also	User Programming	1		
Related	gmax	Maximum gradient s	strength (P)	
	setgcoil .	Assign sysgcoil conf	iguration paran	neter (M)

setgcoil	Assign sysgcoil configuration parameter (I
sysgcoil	System gradient coil (P)
updtgcoil	Update gradient coil (M)

Convert the parameter to a gradient COSY experiment (M) Gcosy

Applicability Systems with the pulsed field gradient or the imaging module.

See

gdiff Diffusion gradient level (P)

Description Predefined parameter available for use in setting a diffusion gradient level, often paired with the timing parameters tdiff or tdelta.

Gdqcosy Convert the parameter to a gradient DQCOSY experiment (M)

Description Convert the parameter to a gradient Dqcosy experiment

get1d Select a 1D experiment for processing (M)

Syntax	get1d<(experiment)>		
Description	In nonautomation mode, the macros hcosy, hcapt, capt, hcdept, and cdept all acquire two or more data sets in the experiment in which the macro was executed. These data sets are stored, complete with Fourier transformed data. The data sets are also stored directly in the experiment. The get1d macro is used to select which data set should be active for processing in that experiment. After get1d is executed, data can be stored in the conventional way with the svf command (e.g., when hcosy completes, get1d can be used to process the 1D data set).		
Arguments	experiment is the 1D data set to be used for processing. The default is the 'H1' experiment.		
Examples	get1d get1d('apt')		
See also	NMR Spectroscopy User Guide		
Related	capt Automated carbon and APT acquisition (M)		
	cdept Automated carbon and DEPT acquisition (M)		
	get2d Select a 2D experiment for processing (M)		
	hcapt Automated proton, carbon, and APT acquisition (M)		
	hcdept Automated proton, carbon, and DEPT acquisition (M)		
	hcosy Automated proton and COSY acquisition (M)		
	svf Save FIDs in current experiment (C)		

get2d Select a 2D experiment for processing (M)

Syntax get2d<(experiment)>

- Description In nonautomation mode, the macros hcosy, hcapt, capt, hcdept, and cdept all acquire two or more data sets in the experiment in which the macro was executed. These data sets are stored complete with Fourier transformed data. The data sets are also stored directly in the experiment. The get2d macro is used to select which data set should be active for processing in that experiment. After entering get2d, data may be stored in the conventional way with the svf command. For example, following completion of hcosy, get2d can be used to process the 2D data set.
- Arguments experiment is the 2D data set that should be used for processing. The default is the 'relayh' experiment.
- Examples get2d('hetcor')
- See also NMR Spectroscopy User Guide
 - Relatedget1dSelect a 1D experiment for processing (M)svfSave FIDs in current experiment (C)

getdim Return dimensionality of experiment (M)

Syntax getdim:dimensions

- Description Used in other macros to determine the number of dimensions of the current data set. Many macros make decisions based on whether a data set is multidimensional or 1D. getdim makes it easier to access this information.
- Arguments dimensions is a return variable giving the number of dimensions of the data. If ni3 is 2 or greater, dimensions is set to 4; if ni2 is 2 or greater, dimensions is set to 3; if ni is 2 or greater, dimensions is set to 2; and if ni is less than 2 or undefined, dimensions is 1.

Examples getdim:r1

- See also NMR Spectroscopy User Guide
- Related ni Number of increments in 1st indirectly detected dimension (P)
 - ni2 Number of increments in 2nd indirectly detected dimension (P)
 - ni3 Number of increments in 3rd indirectly detected dimension (P)

getemailaddr Get email addresses from a file

For a given operator, this macro will use emails found in a comma separated ".csv" file or a space separated ".txt" file stored in the /vnmr directory.
The file must be named "emailaddress.csv" or "emailaddress.txt". Email addresses in the space separated .csv file should appear as follows:
"krish krish@agilent.com
lydia lydia@agilent.com
dave dave@agilent.com"
Email addresses in the comma separated .txt file should appear as follows:
"krish,kris@agilent.com
lydia,lydia@agilent.com
dave,dave@agilent.com"
VnmrJ 3.1

geterror Return or display an acquisition error

Syntax	geterror:\$str geterror(errorNumber):\$str
Applicability	VnmrJ 3.1
Description	"geterror" will translate an error number into a descriptive string. With no argument, geterror will use acqstatus[2], which is the parameter that holds any acquisition related error. Alternatively, an error number may be supplied as an argument. If a return value is used, the error string is return to the calling macro. Otherwise, the error string is displayed.
Arguments	The optional errorNumber is an integer representing an error.
Examples	geterror:\$res
Related	acq_errors manual entry mapping error numbers to descriptive text.

getfile Get information about directories and files (C)

Syntax	 getfile(directory):\$number_files
	(2) getfile(directory,file_index):\$file,\$extension
Description	Returns information about the number of files in a directory or about a particular file in a directory

Arguments	directory is	the	name	of the	directory	for	which	information	is
	desired.								

number_files is the number of files in the directory, with dot files (e.g., .login) ignored.

file_index is the number of file for which information is desired (the order is UNIX-dependent).

file is the name of the file, excluding any extension, identified by the index (see examples below).

extension is the extension of the file name identified by the file_index. For example, if file_index points to the file named s2pul.fid, getfile returns the string s2pul to \$file and the string fid to \$extension. If the file name pointed to has no extension (e.g., dummy), no value is returned to \$extension. If the file name has more than one extension, only the last extension is returned to \$extension (e.g., the file fid.tmp.par returns fid.tmp to \$file and par to \$extension).

Complete paths (full file names) can be reconstructed like this:

```
getfile('dir',i):$filename,$ext
if ($ext='') then $path='dir'+'/'+$filename
else $path='dir'+'/'+$filename+'.'+$ext
endif
```

Paths for the rt command can be reconstructed like this:

\$path='dir'+'/'+\$filename.

```
Examples getfile('dir'):$entries
   $temp = 0
   while ($temp < $entries)
    $temp = $temp + 1
    getfile('dir',$temp):$filename,$ext
    ...
   endwhile
   See also User Programming</pre>
```

getgamma Retrieves Gamma from /vnmr/nuctabref

Description	Retrieves value of gamma for a nucleus from /vnmr/nuctabref.
Syntax	getgamma('nucleus')
See also	getgamma(tn)

getht Retrieve/Save a Hadamard frequency list from a file.

Syntax	<pre>getht(<'htfrq1' <, 'htbw1'>>)</pre>
	getht(<'save' <,'htfrq1'>>)
Applicability	VnmrJ 3.1

File format:

The format of the file is the same as the Line List display in the Edit HT Freq dialog. The first line is an optional title, specifying:

frequency [units] bandwidth [units]

frequency units are Hz or ppm.

Units of Hz are measured from center of spectrum for Hadamard frequencies. The units label is set to [Hz from center]. Units of ppm are referenced to the current spectrum in the experiment. In a 1D, it is referenced to the direct acquisition dimension. In a 2D, it is referenced to F1. bandwidth units are assumed to be in Hz.

The second and subsequent lines are a list of frequencies and bandwidths. The bandwidth column is optional, and assumed to be 20 Hz (or the current value of htbw1) if not specified.

Arguments Usage for retrieving:

getht(<'htfrq1' <,'htbw1'>>)

If there is no first argument, htfrq1 is used as the Hadamard frequency parameter. If there is no second argument, htbw1 is used as the Hadamard bandwidth, if the bandwidth is specified in the file.

In this usage, the macro retrieves the Hadamard frequency line list from a file in the current workspace directory, and sets the parameter values. It also shows the parameters in the Line List display in the Edit HT Freq dialog (editht macro), if open. The file to be copied is in currexp, e.g.

/export/home/vnmr1/vnmrsys/exp1/htfrq1.11

Usage for saving:

getht(<'save' <,'htfrq1'>>)

If the first argument is 'save', the Hadamard frequency list is copied from the Edit HT Freq line list display to the current workspace directory. If there is no second argument, htfrq1 is used as the Hadamard frequency parameter name. If a second argument is specified, it is used as the Hadamard frequency parameter name for the save file, e.g. getht('save','htfrq2') saves the file curexp + '/htfrq2.ll'.

Arguments htfrq1 - Hadamard frequency list in indirect dimension, in ppm or Hz from center of spectrum.

htbw1 - Hadamard bandwidth in indirect dimension, in Hz. It may be a single value or a list of values for each element in the htfrq1 list.

tn - nucleus used for frequency list.

Examples Example #1:

freq [Hz from center] 1172.37

G

327.69 - 346.37

- 1292.10

In Example #1, the Hadamard frequencies are in Hz from the center of the spectrum.

Example #2:

freq	[ppm]		bw	[Hz]
7.930		20		
5.819		16		
4.134		20		
1.770		20		

In Example #2, the Hadamard frequencies are in ppm, referenced to the current spectrum. The frequency bandwidth is set to 20 Hz for most of the frequencies, except for the second frequency, which is set to 16 Hz.

See also

ht

```
HsqcHT
tocsyHT
editht
sethtfrq1
htfrqdisp
dl1
```

getlcdata An LC-NMR communications macro

Applicability VnmrJ 3.1

Description This macro starts the LC data file listener (/vnmr/tcl/bin/fileListen) so that when the LC system sends a data file it is received and transferred to the appropriate experiment or automation directory. It is not necessary to use getlcdata in normal operation as the LC data file listener is automatically started when the start LC NMR run button is pressed. As described in the text above, getlcdata may be desirable for the transfer of the LC data after runs using the analyte collector where the original LC run and the NMR analysis are well separated in time.

getlimit get the limits of a variable in a tree (C)

Syntax	getlimit(name[,tree]):\$max,\$min,\$step,\$index
Description	getlimit displays or returns the limits of a variable in a tree.
	The returned values are the max value, min. value, step size, and index
	The fourth argument will return a 0 if the parameter is not using an

indexed table lookup for the maximum, minimum, and step size. If the parameter is using the table lookup mechanism, the fourth argument will be set to the index for that table.

The variable trees are current (the default), global, processed, or systemglobal.

Arguments name – the name of the variable

tree - the variable tree: current (the default), global, processed, or systemglobal.

Examples	getlimit('np'):\$max,\$min,\$step,\$index	
	sets \$max to 128000, \$min to 32, \$step to 2 and \$index to	o 0
	getlimit('lockfreq','systemglobal'):\$max	
	sets \$max to 160	
	getlimit('dpwr'):\$max,\$min,\$step,\$index	
	sets max to 49, min to 0 $step$ to 1 and max to 9	
Related	setlimit Set limits of a parameter in a tree (C)	
	setprotect Set protection mode of a parameter (C)	

get11 Get intensity and line frequency of line (C)

Syntax	getll(line_r	<pre>number)<:height,frequency></pre>		
Description	Finds the height and frequency of line from a line listing. It assumes a previous line list using dll.			
Arguments	line_number is the number of the line in the line list.			
	height is the intensity of the specified line.			
	frequency is the line frequency with units defined by the parameter $\ensuremath{\mathtt{axis}}.$			
See also	User Program	ming		
Related	axis .	Axis label for displays and plots (P)		
	dll l	Display listed line frequencies and intensities (C)		
	fp l	Find peak heights (C)		
	nll l	Find line frequencies and intensities (C)		

getmodule Gets module (C)

Syntax getmodule('modulename'<,dirpath<,tree<,parameter>>>)<:\$retva lue> Arguments arg1 - modulename arg2 - (optional) pathname where the module should be read from default is studydir/dirinfo/modules. If arg2 is an empty string, it is set to default.arg2='cp' is a keyword for curexp (auto='n') or autodir (auto='y') arg3 - (optional) (read to which) tree (default is current)arg4 - (optional) specific parameter - in this case the value of the

parameter is read into the tree or returned the calling macro, but not both. Consequently, if a return argument is supplied arg3 is ignored.

getoffset Sets offset based on current reference parameters

Description Sets offset based on current reference parameters rather than output of setref macro. The input argument is Hz. Syntax getoffset('frequency') See also getoffset(320)

getparam Retrieve parameter from probe file (M)

getparam(param<,nucleus>):\$value			
Retrieves the value of a parameter from the current probe file. The name of the probe file is referenced from the parameter probe.			
param is the name of the parameter to be retrieved.			
nucleus is the is the current v	nucleus is the nucleus to be retrieved from the probe file. The default is the current value of the parameter tn		
value is a retu	rn variable with the value of the retrieved parameter.		
getparam('tpwr'):tpwr getparam('dmf','H1'):\$dmf			
NMR Spectroscopy User Guide			
addnucleus	Add new nucleus to existing probe file (M)		
addparams	Add parameter to current probe file (M)		
addprobe	Create new probe directory and probe file (M)		
probe	Probe type (P)		
setparams	Write parameter to current probe file (M)		
tn	Nucleus for the observe transmitter (P)		
updateprobe	Update probe file (M)		
	getparam(para Retrieves the var name of the pro- param is the nar nucleus is the is the current v value is a retur getparam('tpv getparam('dmf <i>NMR Spectrosco</i> addnucleus addparams addprobe probe setparams tn updateprobe		

getplane Extract planes from a 3D spectral data set (M)

Syntaxgetplane<(<data_dir><,plane_dir><,plane_type>)>DescriptionExecutes the program getplane in the VnmrJ system bin directory
(\$vnmrsystem/bin). getplane checks whether there is sufficient
file space on the disk partition to accommodate the extracted planes.
If space is insufficient, getplane writes an error to the VnmrJ text
window and aborts. getplane does not delete the output plane

directory if it is run multiple times to individually extract different plane types.

Arguments data_dir specifies the directory (without the /data subdirectory) containing the input 3D spectral data. The first non-keyword argument to getplane is always taken to be data dir.

plane_dir specifies the directory (without the /extr subdirectory)
in which the extracted planes are to be stored. The second
non-keyword argument to getplane is always taken to be plane_dir.
If plane_dir is not specified, data_dir also specifies the output
plane directory. If both data_dir and plane_dir are not specified,
the input data directory and the output plane directory are set to
curexp/datadir3d. The parameter plane is always set equal to the
output plane directory.

plane_type can be any of the following keywords:

- 'xall' is a keyword to extract all three 2D plane types: f1f3, f2f3, f1f2.
- •'f1f3', 'f2f3', 'f1f2' are keywords to extract their respective 2D planes.

• Any of these keywords can be submitted more than once to the getplane macro, but the getplane program displays an error and aborts if any one plane type is defined for extraction more than once.

Examples getplane

getplane('data3d.inp,'data3d.planes','f1f3','f2f3')

See also NMR Spectroscopy User Guide

Related	dplane	Display a 3D plane (M)
	dproj	Display a 3D plane projection (M)
	dsplanes	Display a series of 3D planes (M)
	ft3d	Perform a 3D Fourier transform (M)
	nextpl	Display the next 3D plane (M)
	path3d	Path to currently displayed 2D planes from a 3D data set (P)
	plane	Currently displayed 3D plane type (P)
	plplanes	Plot a series of 3D planes (M)
	prevpl	Display the previous 3D plane (M)

getplottertype The getplottertype command retrieves plotter information.

Syntax getplottertype:\$rasterValue,\$plotterType
getplottertype(plotter):\$rasterValue,\$plotterType
getplottertype(plotter,'osname'):\$osname
getplottertype(plotter,'attr'):\$attr

- Description The getplottertype command retrieves plotter information. With zero or one argument, it will return the "raster" value from the devicetable file and the "Type" value from the devicenames file. With no arguments, it uses the value of the plotter parameter. The returned raster values are:
 - •0 Plotters which use the HPGL language.
 - •1 Plotters which use the PCL language and are in portrait mode.
 - $\bullet\,2\,$ Plotters which use the PCL language and are in landscape mode.
 - $\bullet 3$ Plotters which use the PostScript language and are in portrait mode.
 - •4 Plotters which use the PostScript language and are in landscape mode.
- Arguments The VnmrJ name for a plotter does not need to be the same name that the computer operating system (OS) uses for the plotter / printer. The getplottertype with two arguments, where the first argument is the VnmrJ plotter name and the second argument is the 'osname' keyword, will return the plotter / printer name used by the OS.

If a plotter name is given as the first argument and a plotter attribute, as listed in the devicetable, is the second argument, getplottertype will return the value of the attribute. If the attribute does not exist, a null string is returned.

Example:

getplottertpye(plotter,'papersize'):\$psize

getppm Returns Cursor Value in ppm

DescriptionReturns the value of the current cursor position in ppm.Syntaxgetppm:\$valueExamplesgetppm:r1

getreg Get frequency limits of a specified region (C)

Syntax	getreg(region_number)<:minimum,maximum>
Description	Returns the frequency limits of a region. The spectrum should have been previously divided into regions with the region command.
Arguments	region_number specifies the number of the region.
	minimum,maximum are return values set to the frequency limits, in Hz, of the specified region.
Examples	<pre>getreg(1):\$a,\$b getreg(\$4):cr,\$lo getreg(R1-1):r2,r3</pre>

See also	User Program	nming
Related	CZ	Clear integral reset points (C)
	ds	Display a spectrum (C)
	numreg	Return the number of regions in a spectrum (C)
	region	Divide spectrum into regions (C)
	Z	Add integral reset point at cursor position (C)

getsampglobalLoads sample global parameters

Description	Loads sample global parameters in the current workspace from the study directory.
See also	getsampglobalt
Related	getsampglobal, resetsampglobal, savesampglobal, mvsampglobal, showsampglobal

getshimmethods Get proshim methods list (M)

Applicability	VnmrJ 3.2	
Description	Scan the proshimmethods and shimmethods directories in all active	
	appdirs. Make a sorted list of all the methods. This is used by the	
	VnmrJ interface to provide a selection mechanism for shim methods.	

getsn Get signal-to-noise estimate of a spectrum (M)

Syntax getsn:current_sn,predicted_sn

Description Estimates spectrum signal-to-noise using the following algorithm:

- Measures four adjacent 5-percent portions at the left edge of the spectrum, finding the root-mean-square noise, and taking the smallest of the four values. By measuring four different values and finding root-mean- square noise instead of peak noise, the result should be reliable even if several signals are present in the selected regions.
- Next, estimates the signal level using the vertical scale adjustment macros: vsadjh for proton, vsadjc for carbon, and vsadj for other nuclei. For carbon spectra, this algorithm ignores solvent lines and TMS. For proton spectra, in addition to ignoring the largest line in the spectrum, if the tallest line is greater than three times the height of the second tallest line, the second highest line is be used instead. For other nuclei, getsn uses the tallest line in the spectrum.
- Finally, estimates the signal-to-noise at the end of the experiment by a simple extrapolation (multiplying by the square root of nt/ct).

Arguments current_sn is a return value set to the current signal-to-noise level.

predicted_sn is a return value set to the predicted signal-to-noise level at the end of the experiment.

See also	NMR Spectroscopy User Guide	
Related	ct	Completed transients (P)
	nt	Number of transients (P)
	testsn	Test signal-to-noise ratio (M)
	vsadj	Adjust vertical scale (M)
	vsadjc	Adjust vertical scale for carbon spectra (M)
	vsadjh	Adjust vertical scale for proton spectra (M)

gettoken Utility macro to separate a string into tokens (M)

- Description Gets the first occurrence of a substring in input_string which is delimited by delimiter, or by the default delimiter '\$'. The substring is returned in output_string. The next location in the string after the second delimiter is returned as a real in next_location. If there are not both one occurrence of each of the beginning delimiter and the second delimiter - in other words, if the delimiters are not paired an empty string is returned in output_string, and -1 is returned in next_location. If the delimited substring is the last substring in input_string, then the substring is returned as expected, but next_location returns -1.
- Arguments input_string
 The string to be tokenized delimiter is the delimiter for the tokens
 (default is \$)
 Examples gettoken(\$mydirname):\$mytoken, \$next_location
 gettoken(\$mydirname,'%'):\$mytoken, \$next_location
 Related reqpartest Tests whether required parameters are set (M)

gettxt Get text file from VnmrJ data file (C)

Syntax	gettxt(file)		
Description	Copies text from a data file to the current experiment.		
Arguments	file is the name of a VnmrJ data file saved from an experiment (i.e. a directory with a .fid or .par suffix). Do not include the file name suffix.		
Examples	gettxt('/vnmr/fidlib/fid1d')		
See also	NMR Spectroscopy User Guide		
Related	puttxt Put text file into another file (C)		

gettype Get the type of a variable (C)

Syntax gettype(name[,tree])<:index,name>

Description Displays or returns the type of an existing variable.

Arguments A "string" variable can return type 'string' or 'flag'. A "real" variable can return type 'real', 'delay', 'frequency', 'pulse', or 'integer'. gettype returns one or two values to a macro. The first value is an integer corresponding to the parameter type. The second value is the name of the parameter type. name can be used in commands such as settype and create.

An optional tree argument can be given. The variable trees are 'current', 'global'. 'processed' and 'systemglobal'.

The default is to search for the parameter in the 'current', 'global', and 'systemglobal' trees, in that order.

The integer values and names of the parameter types are:

- 0 "undefined"
- 1 "real"
- 2 "string"
- 3 "delay"
- 4 "flag"
- 5 "frequency"
- 6 "pulse"
- 7 "integer"

Examples gettype('dmm'):\$int,\$name sets \$int to 4 and \$name to 'flag'.
See also gettype('pw'):\$int,\$name sets \$int to 6 and \$name to 'pulse'.

getvalue Get value of parameter in a tree (C)

- Syntax getvalue(name [,index] [,tree])<:\$val>
 getvalue(name ,'size' [,tree])<:\$num>
- Description Gets the value of any parameter in a tree. The value of most parameters can be accessed simply by using their name in an expression. For example, sw? or r1=np accesses the value of sw and np, respectively. However, parameters in the processed tree cannot be accessed that way; getvalue can be used to get the value of a parameter in the processed tree.

Single elements of an arrayed parameter can be retrieved by suppling an optional "index". "index" defaults to 1. If the second argument is the keyword 'size', then the number of elements of the parameter can retrieved. If the parameter does not exist, a zero (0) will be returned.

Arguments parameter is the name of an existing parameter.

index is the number of a single element in an arrayed parameter. Default is 1.

tree is one of the keywords 'global', 'current', 'processed', or 'systemglobal'. The default is 'processed'. Refer to the create command for more information on the types of parameter trees.

If the second argument is the keyword 'size', then the number of parameter elements can be retrieved. If the parameter does not exist, a zero (0) will be returned.

tree

Examples getvalue('arraydim'):\$val

getvalue('phase','size'):\$num

See also User Programming

Related	create	Create new parameter in a parameter tree (C)
	display	Display parameters and their attributes (C)
	setgroup	Set group of a parameter in a tree (C)
	setlimit	Set limits of a parameter in a tree (C)
	setprotect	Set protection mode of a parameter (C)
	settype	Change type of a parameter (C)
	setvalue	Set value of any parameter in a tree (C)

gf Prepare parameters for FID/spectrum display in acqi (M)

Description Provided as a model for preparing parameters for the FID and spectrum display in acqi. The unmodified version of this macro turns off phase cycling, autoshimming, autolocking, spin control, temperature control, sample changer control, and autogain. It also selects the current pulse sequence and parameter set by issuing the command go('acqi') and the command acqi('par'). The automation parameters cp, wshim, alock, spin, temp, loc, and gain are then reset to their original values. Users can customize gf by copying it into their private maclib directory and editing that version to suit their needs.

See also NMR Sp	ectroscopy User	Guide
-----------------	-----------------	-------

Related	acqi	Interactive acquisition display process (C)
	alock	Automatic lock status (P)
	ср	Cycle phase (P)
	dmgf	Absolute-value display of FID data and spectrum in acqi
		(P)
	gain	Receiver gain (P)
	go	Submit an experiment to acquisition (C)
	loc	Location of sample in tray (P)
	spin	Sample spin rate (P)
	temp	Sample temperature (P)
	wshim	Conditions when shimming performed (P)

gf Gaussian function in directly detected dimension (P)

Description	Defines a Gaussian time constant of the form $\exp(-(t/gf)2)$ along the directly detected dimension. This dimension is referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, etc.	
Values	Number, in seconds. Typical value is gf='n'.	
See also	NMR Spectroscopy User Guide	
Related	gf1 Gaussian function in 1st indirectly detected dimension (P)	
	gf2 Gaussian function in 2nd indirectly detected dimension (P)	
	gfs Gaussian shift constant in directly detected dimension (P)	

gf1 Gaussian function in 1st indirectly detected dimension (P)

Description	Defines a Gaussian time constant of the form $\exp(-(t/gfl)2)$ along
	the first indirectly detected dimension. This dimension is referred to
	as the f_1 dimension of a multidimensional data set. gf1 works
	analogously to the parameter gf. The "conventional" parameters, such
	as 1b and gf, operate on the detected FIDs, while this "2D" parameter
	is used during processing of the interferograms.
Values	Number, in seconds.
See also	NMR Spectroscopy User Guide

Related gf Gaussian function in directly detected dimension (P)

gf2 Gaussian function in 2nd indirectly detected dimension (P)

Description	Defines a Gau the second in to as the f_2 of analogously to gf2 on the 2	assian time constant of the form $\exp(-(t/gf2)2)$ along adjrectly detected dimension. This dimension is referred dimension of a multidimensional data set. $gf2$ works to the parameter gf . The wti program can be used to set D interferogram data.
Values	Number, in seconds.	
See also	NMR Spectroscopy User Guide	
Related	gf	Gaussian function in directly detected dimension (P)
	wti	Interactive weighting (C)

gflow Flow encoding gradient level (P)

DescriptionPredefined parameter available for use in setting a flow encoding
gradient level, often paired with the timing parameter tflow.See alsoVnmrJ Imaging NMR

gfs Gaussian shift const. in directly detected dimension (P)

- Description Working in combination with the gf parameter, gfs allows shifting the center of the Gaussian function $\exp(-((t-gfs)/gf)2)$ along the directly detected dimension. This dimension is referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, etc. Typical value is gfs='n'.
 - See also NMR Spectroscopy User Guide

Related	gf	Gaussian function in directly detected dimension (P)
	gfs1	Gaussian shift const. in 1st indirectly detected dimension
		(P)
	gfs2	Gaussian shift const. in 2nd indirectly detected dimension
		(P)

gfs1 Gaussian shift const. in 1st indirectly detected dimension (P)

- DescriptionWorking in combination with the gfl parameter, gfsl allows shifting
the center of the Gaussian function $\exp(-((t-gfsl)/gfl)2)$ along
the first indirectly detected dimension. This dimension is referred to
as the f1 dimension in multidimensional data sets. gfsl works
analogously to the parameter gfs. The "conventional" parameters (i.e.,
lb, gf, etc.) operate on the detected FIDs, while this "2D" parameter
is used during processing of the interferograms.See alsoNMR Spectroscopy User Guide
Gaussian function in directly detected dimension (P)
 - gf1Gaussian function in 1st indirectly detected dimension (P)gfsGaussian shift const. in directly detected dimension (P)

gfs2 Gaussian shift const. in 2nd indirectly detected dimension (P)

Description Working in combination with the gf2 parameter, gfs2 allows shifting the center of the Gaussian function exp(-((t-gfs2)/gf2)2) along

See also	the second indirectly detected dimension. This dimension is referred to as the f_2 dimension in multidimensional data sets. gfs2 works analogously to the parameter gfs. The wti program can be used to set gfs2 on the 2D interferogram data.
Related	gf Gaussian function in directly detected dimension (P) gf2 Gaussian function in 2nd indirectly detected dimension (P) gfs Gaussian shift const. in directly detected dimension (P) wti Interactive weighting (C)
Ghmbc	Convert the parameter to a gradient HMBC experiment (M)
Applicability Description Arguments	Systems with a pulsed field gradient module. Prepares an experiment for a PFG (pulsed field gradient) HMQC. NMR Spectroscopy User Guide
ghmqc	Set up a PFG HMQC pulse sequence (M)
Applicability Description Arguments	Systems with a pulsed field gradient module. Prepares an experiment for a PFG (pulsed field gradient) HMQC using the sequence GHMQC. The sequence sets three gradients, all separately. NMR Spectroscopy User Guide
Ghmqc	Convert the parameter to a gradient HMQC experiment (M)
Description	Convert the parameter to a gradient HMQC experiment
gHMQC15	Set up parameters for ¹⁵ N gHMQC experiment (M)
Description	Converts the current parameter set to a gHMQC experiment for $^{15}\mathrm{N}.$
gHMQC_d2	Set up parameters for ¹⁵ N gHMQC experiment using dec. 2 (M)

M)

gHMQC_d213 Set up parameters for ¹³C gHMQC experiment using dec. 2 (M)

Description Converts the current parameter set to a gHMQC experiment for ${}^{13}C$ with decoupler 2 as ${}^{13}C$.

ghmqcps Set up a PFG HMQC phase-sensitive pulse sequence (M)

Applicability Systems with a pulsed field gradient module.

Description Prepares an experiment for a PFG (pulsed field gradient) HMQC, phase-sensitive version.

See also NMR Spectroscopy User Guide

ghsqc Set up a PFG HSQC pulse sequence (M)

Applicability	Systems with a pulsed field gradient module.		
Syntax	ghsqc<(nucleus)>		
Description	Converts a 1D standard two-pulse sequence parameter set into a parameter set ready to run a PFG (pulsed field gradient) HSQC experiment, either absolute value or phase sensitive.		
Arguments	nucleus is 13C or 15N. The default is 13C.		
See also	NMR Spectroscopy User Guide		

Ghsqc Convert the parameter to a gradient HSQC experiment (M)

Description Convert the parameter to a gradient HSQC experiment.

gHSQC15 Set up parameters for ¹⁵N gHSQC experiment (M)

Description Converts the current parameter set to a gHSQC experiment for ¹⁵N.

gHSQC_d2 Set up parameters for ¹⁵N gHSQC experiment using dec. 2 (M)

Description Converts the current parameter set to a gHSQC experiment for ${}^{15}N$ with decoupler 2 as ${}^{15}N$.

gHSQC_d213 Set up parameters for ¹³C gHSQC experiment using dec. 2 (M)

Description Converts the current parameter set to a gHSQC experiment for ${}^{13}C$ with decoupler 2 as ${}^{13}C$.

Ghsqctoxy Convert parameters for gradient HSQCTOXY experiment (M)

Description Convert the parameter to a gradient HSQCTOXY experiment

gilson Open the Gilson Liquid Handler window (C)

Syntax	gilson	
Description	Opens the Gilson Liquid Handler window, which enables setup, configuration, and operation of the VAST automatic sampler change	
See also	NMR Spectroscopy User Guide	
	$\mathbf{I} = \mathbf{I}$	

gilson Allow starting the Gilson Liquid Handler GUI

Applicability	VnmrJ 3.1		
Description	When the "gilson" macro is invoked, a window appears on the screet		
	and users then can select appropriate item in it to run the Gilson		
	Liquid Handler. If an argument is passed to gilson, for example,		
	gilson(1), then the gilson window appears and allows users to edit		
	the details of inserting and removing samples with the Gilson. However,		
	direct communication with the Gilson sample changer is not available.		

gin Return current mouse position and button values (C)

Applicability	All		
Syntax	gin<(Bn_ <press><release>)>:\$x,\$y,\$b1,\$b2,\$b3</release></press>		
Description	The gin command reports the pointer position in relationship to the graphics window and is often used with the move and draw command. The variables x and y are the x and y positions hold the pointer is millimeters. The variables $b1$, $b2$, and $b3$ hold the values for the state of the left, middle, and right mouse buttons.		
Values	x is the value in the <i>x</i> direction, in millimeters, of the pointer. The range of x is 0 at the left edge of the chart and wcmax at the right edge. A value of -1 is returned if the pointer position is outside the graphics window along the x axis.		
	$\$_y$ is the position of the pointer along the y axis. The range of y is -20 at the bottom of the chart to wc2max at the top. A value of 10000 is returned if the pointer position is outside the graphics window along the y axis.		
	\$b1 is the state of left button; returns the value 0 if released and 1 if pressed.		
	\$b2 is the of middle button; returns the value 0 if released and 1 if pressed.		
	b3 is the of right button; returns the value 0 if released and 1 if pressed.		
Arguments	no argument, returns current mouse positions and button values.		
	Bn_{press} , n=a,1,2, or 3. Wait for mouse button (any, 1, 2, or 3) or any key to be pressed.		
	Bn_release, n=a,1,2, or 3. Wait for mouse button (any, 1, 2, or 3) to be released or any key to be pressed.		
Examples	gin('B3_press'):\$x,\$y,\$b1,\$b2,\$b3 wait until button 3or any key is pressed		
	gin('Ba_press'):\$x,\$y,\$b1,\$b2,\$b3 wait until any button or any key is pressed		
	gin('B1_release'):\$x,\$y,\$b1,\$b2,\$b3 wait until button 1 is released or any key pressed		
	gin('B2_release'):\$x,\$y,\$b1,\$b2,\$b3 wait until button 2 is released or any key pressed		
See also	User Programming		
Related	box Draw a box on a plotter or graphics display (C)		
	drawDraw line from current location to another location (C)moveMove to an absolute location to start a line (C)		

globalauto Automation directory name (P)

Applicability	VnmrJ Walkup and systems with automation such as sample handling.	
Description	A global parameter that specifies the name of a directory in which the daily automation directories or study directories are saved. This parameter is created and used by the walkup macro and the VnmrJ Walkup interface	
	walkup interface.	
See also	NMR Spectroscopy User Guide; VnmrJ Automation User Guide	
Related	cqinit Initialize liquids study queue (M)	
	walkup Walkup automation (M)	

glue Create a pseudo-2D dataset (M)

Applicability	Systems with the LC-NMR accessory.			
Syntax	glue<(num_scans)>			
Description	Steps through the series of FIDs, putting them into exp5 one by one as an array, and then jumps to exp5 and changes the parameters arraydim, ni, and fn1, so that the data appear to the user to be a 2D experiment, which can then be processed and displayed with standard 2D commands (wft2d, dconi, etc.). The parameter savefile should exist and should contain the base file name to which a series of FIDs have been saved as savefile.001, savefile.002, etc.			
Arguments	num_scans is the number of FIDs copied into the exp5 array. Typically, num_scans is used if the experiment was aborted prematurely, so that the complete num_scans worth of FIDs were not actually acquired.			
See also	NMR Spectroscopy User Guide			
Related	savefile Base file name for saving FIDs or data sets (P)			

Applicability VnmrJ 3.1

gmapshim Start gradient autoshimming (M)

Applicability	Systems with gradient shimming installed.		
Syntax	gmapshim<('files' 'mapname' 'quit')>		
Description	Starts gradient autoshimming if no arguments are used. It can also retrieve a shimmap file or quit gradient autoshimming. When the gmapshim macro is done, it automatically exits, and the previous data set is retrieved.		
Arguments	'files' is a keyword to enter the gradient autoshimming files menu.		
	'mapname' is a keyword to display the current mapname.		

'quit' is a keyword to exit from gradient autoshimming and retrieve the previous data set.
See also NMR Spectroscopy User Guide
Related gmapsys Run gradient autoshimming, set parameters, map shims

(M) gmapz Get parameters and files for gmapz pulse sequence (M)

gmapshim_au Start acquisition with gradient shimming (M)

Applicability Systems with gradient shimming installed.

Description If wshim is not set to 'n', gmapshim_au checks the probe file for a lock gradient map name. If the name exists, gmapshim_au executes gmapshim('glideau') to start gradient shimming followed by acquisition. If the map name does not exist, gmapshim_au starts acquisition by running au('wait').

gmapspin Enable or disable spinning during gradient shimming (P)

Description Specifies whether or not sample spinning during gradient shimming is enabled. If spinning is enabled during gradient shimming, the pulses and delays *must* also be synchronized with the rotor period.

Values 'n' disable spinning during gradient shimming. 'y' enable spinning during gradient shimming.

spin

Related gmapz Get parameters and files for gmapz pulse sequence (M) gmapsys Run gradient autoshimming, set parameters, map shims (M) gzsize Number of z-axis shims used by gradient shimming (P)

gmapsys Run gradient autoshimming, set parameters, map shims (M)

Sample spin rate (P)

Applicability	Systems with gradient shimming installed.	
Syntax	(1) gmapsys<(option)>(2) gmapsys('shimmap'<, shimmap_option>)	
Description	Enters the Gradient Shimming Setup panel for setting parameters, mapping the shims, and performing autoshimming. This is the onl entry point to the gradient shimming Setup panel.	
	If the gmapz pulse sequence is not loaded, retrieve parameters from the last shimmap used (or current mapname) or from gmapz.par if no shimmap exists.	

G

Arguments option is one of the following keywords:

- 'addpar' adds gradient shimming parameters to the current parameter set.
- 'findgzlvl' runs an experiment to calibrate gzlvl, gzwin, and tof to optimize the spectral window.
- 'findgzwin' runs an experiment to calibrate gzwin and tof to optimize the spectral window.
- 'findtof' runs an experiment to center tof to optimize the spectral window.
- 'rec' displays the record of shim adjustments from the previous gradient shimming run.
- 'shim' start autoshimming (same as Gradient Autoshim on Z button).
- 'vi' edits the file gshim.list, which is used for editing shim offsets, mapname, or selecting coarse and fine shims.
- 'writeb0' displays the b0 plot calculated from the first two array elements.

'shimmap' is a keyword to run a shim mapping experiment and save the results (same as Make Shimmap button).

shimmap_option is one of the following values:

- 'auto' is a keyword to calibrate gzwin and then make a shimmap (same as Automake Shimmap button).
- 'manual' is a keyword to use shim offset values set manually from the file gshim.list and not the default values to make a shimmap.
- 'overwrite' is a keyword to make a shimmap and overwrite the current mapname if it exists.
- mapname is the prefix of the shimmap file name. The default is the user is queried for mapname before running the experiment.

See also NMR Spectroscopy User Guide

Related	gmapshim	Start gradient autoshimming (M)
	gmapz	Get parameters and files for $\ensuremath{\mathtt{gmapz}}$ pulse sequence (M)
	gradtype	Gradients for X, Y, Z axes (P)
	gzwin	Spectral width percentage used for gradient shimming
		(P)
	seqfil	Pulse sequence name (P)
	gmap_z1z4	Gradient shimming flag to first shim z1-z4 (P)
	gzsize	Number of z-axis shims used by gradient shimming $\left(P\right)$

gmapz Get parameters and files for gmapz pulse sequence (M)

Applicability Systems with gradient shimming installed. Syntax gmapz<(mapname)>
Description	Retrieves gra experiment.	dient shimming parameters to set up a gradient shimming
Arguments	mapname is the name of a gradient shimmap file that must exist in the shimmaps directory. gmapz retrieves parameters and loads the shimmap file from mapname. The default is to retrieve standard gradient shimming parameters from the file gmapz.par.	
See also	NMR Spectroscopy User Guide	
Related	gmapshim gmapsys	Start gradient autoshimming (M) Run gradient autoshimming, set parameters, map shims (M) Gradient shimming flag to first shim z1-z4 (P)
	guide Tarat	

gmap_findtofGradient shimming flag to first find tof (P)

Applicability	Systems wi	th gradient shimming installed.
Description	When the f calibration recommend different so	flag is set to $'y'$, gradient shimming first performs a to find tof before the start of shimming. This action is led for only homospoil deuterium gradient shimming with plyents. The default value is 'n'.
Values	'y' turns 'n' turns	on the flag. off the flag.
See also	NMR Spectroscopy User Guide	
Related	gmapshim	Start gradient autoshimming (M)
	gmapsys	Run gradient autoshimming, set parameters, map shims (M)
	gmapz	Get parameters and files for gmapz pulse sequence (M)
	tof	Frequency offset for observe transmitter (P)

gmap_z1z4 Gradient shimming flag to first shim z1-z4 (P)

Applicability	Systems wit	h gradient shimming installed.
Description	When the fl shimming fi gzsize. Wh gzsize are	ag is set to 'y', if gzsize is greater than 4, gradient rst shims on z1-z4, and then uses all shims specified by nen the flag is set to 'n'(default), all shims specified by used.
Values	'y' turns o 'n' turns o	n the flag. ff the flag.
See also	NMR Spectroscopy User Guide	
Related	gmapshim gmapsys gmapz	Start gradient autoshimming (M) Run gradient autoshimming, set parameters, map shims (M) Get parameters and files for gmapz pulse sequence (M) Number of z-axis shims used by gradient shimming (P)
	yzarze	Number of 2-axis similis used by gradient similing (1)

gmaxMaximum gradient strength (P)DescriptionThe allowed maximum gradient level (absolute value) in gauss/cm.
gmax is one of the calibration entries in a gradtables file. gxmax,
gymax, and gzmax are used when the maximum gradient level is
different for each axis in gauss/cm, which is the case for triple-axis
PFG coils.See alsoVnmrJ Installation and Administration; VnmrJ Imaging NMR
Current gradient coil (P)
gxmax,gymax,gzmax

gmqcosy Set up PFG absolute-value MQF COSY parameter set (M)

(P)

System gradient coil (P)

Applicability	Systems with the pulsed field gradient module.
Description	Converts a 1D standard two-pulse sequence parameter set into a
	parameter set ready to run a PFG (pulsed field gradient)
	absolute-value MQF COSY experiment.
See also	NMR Spectroscopy User Guide

gnoesy Set up a PFG NOESY parameter set (M)

sysgcoil

Applicability	Systems with the pulsed field gradient module.
Description	Converts a 1D standard two-pulse sequence parameter set into a parameter set ready to run a PFG (pulsed field gradient) NOESY experiment, either absolute value or phase sensitive.
See also	NMR Spectroscopy User Guide

go_<pslabel>Experiment-Specific Runtime Macro

See also The go_<pslabel> macro, if it exists, is executed when acquisition begins on a pslabel-specific basis.

go Submit experiment to acquisition (M)

Description Performs the experiment described by the current acquisition parameters, checking parameters loc, spin, gain, wshim, load, and method to determine the necessity to perform various actions in addition to data acquisition. This may involve a single FID or multiple FIDs, as in the case of arrays or 2D experiments. go acquires the FID and performs no processing. If free disk space is insufficient for the complete 1D or 2D FID data set to be acquired, go prompts the user with an appropriate message and aborts the acquisition initiation process.

> Before starting the experiment, go executes two user-created macros if they exist. The first is usergo, a macro that allows the user to set up general conditions for the experiment. The second is a macro whose name is formed by go_ followed by the name of the pulse sequence (from seqfil) to be used (e.g., go_s2pul, go_dept). The second macro allows a user to set up experiment conditions suited to a particular sequence.

Arguments 'acqi' is a keyword to submit an experiment for display by the acqi program. All operations explained above are performed, except acquisition of data is not initiated. The instructions to control data acquisition are stored so that acqi can acquire the data when the FID button is clicked. The gf macro is recommended instead of running go('acqi') directly. Using gf prevents certain acquisition events from occurring, such as spin control and temperature change. See the description of gf for more information.

> 'nocheck' is a keyword to override checking if there is not enough free disk space for the complete 1D or 2D FID data set to be acquired.

'nosafe' is a keyword to disable probe protection during the experiment.

'next' is a keyword to put the experiment started with go('next') at the head of the queue of experiments to be submitted to the acquisition system. If go('next') is entered, the go macro remains active until the experiment is submitted to the acquisition system, and no other VnmrJ commands are processed until the go macro finishes.

'sync' is a keyword in nonautomation mode that accomplishes the same effect as go('next') in synchronizing VnmrJ command execution with the submission of experiments to the acquisition system. The difference is that 'sync' does not put the experiment at the head of the queue.

'wait' is a keyword to stop submission of experiments to acquisition until wexp processing of the experiment, started with go('wait'), is finished.

 Examples
 go

 go('nosafe')
 go('next')

 See also
 NMR Spectroscopy User Guide

 Related
 acqi

 au
 Interactive acquisition display process (C)

 submit experiment to acquisition and process data

change	Submit a change sample experiment to acquisition (M)
gain	Receiver gain (P)
ga	Submit experiment to acquisition and FT the result (C)
gf	Prepare parameters for FID/spectrum display in acqi (M)
go_	Pulse sequence setup macro called by go, ga, and au (M)
load	Load status of displayed shims (P)
loc	Location of sample in tray (P)
lock	Submit an Autolock experiment to acquisition (C)
method	Autoshim method (P)
probe_protection	Probe protection control (P)
sample	Submit change sample, Autoshim exp. to acquisition (M)
seqfil	Pulse sequence name (P)
shim	Submit an Autoshim experiment to acquisition (C)
spin	Submit a spin setup experiment to acquisition (C)
spin	Sample spin rate (P)
su	Submit a setup experiment to acquisition (M)
usergo	Experiment setup macro called by go, ga, and au (M)
vnmrjcmd()	Commands to invoke the GUI popup (C)
wshim	Conditions when shimming is performed (P)

go_

Pulse sequence setup macro called by go, ga, and au (M)

Syntax	go_macro
Description	Called by the macros go , ga , or au before starting an experiment. The user typically creates this macro to set up general experiment conditions. The name of the macro is formed by combining $go_$ with the name of the pulse sequence macro (from seqfil) to be used.
Examples	go_dept go_noesy go_s2pul
See also	NMR Spectroscopy User Guide
Related	auSubmit experiment to acquisition and process data (M)gaSubmit experiment to acquisition and FT the result (M)goSubmit experiment to acquisition (M)seqfilPulse sequence name (P)usergoExperimental setup macro called by go, ga, and au (M)

gpat-gpat3 Gradient shape (P)

DescriptionPredefined string parameters available to specify gradient shapes.See alsoVnmrJ Imaging NMR

gplan Start interactive image planning (C)

Syntax	<pre>gplan(function_name, arg1, arg2,)</pre>
Description	In VnmrJ, starts an image planning session.
Arguments	'function_name',path is the name of an image planning function surrounded by single quotation marks.
	arg1, arg2, are arguments for the function, if relevant.
Examples	gplan 'clearStacks()' get 'PrevStacks()'
See also	NMR Spectroscopy User Guide

x

Multiplier for gradient pulses on alternating scans (P)

Syntax	<pre>create('gradalt','real')</pre>
Applicability	VnmrJ 3.2
Description The zgradpulse and rgradient pulse elements use the gradalt to multiply the gradient amplitude.	
	No changes are made if:
	• the local (curpar) parameter gradalt does not exist
	• the local (curpar) parameter gradalt is set to "Not Used"
	• the local (curpar) parameter gradalt is set to "1"
See also	User Programming Guide

gradfit calculates fit coefficients describing the variation of gradient strength with position in calibration of non-uniform pulsed field gradients

Syntax	gradfit(lowfrq,highfrq,D)
	gradfit(lowfrq,highfrq,D,ncoef)
Applicability	VnmrJ 3.1
Description	gradfit calculates the coefficients of a power series to fit the measured variation of gradient strength with position during the calibration of non-uniform pulsed field gradients.

Arguments gradfit takes 3 or 4 arguments: lowfrq is the lower frequency limit of the signal profile, highfrq the high frequency limit, D the diffusion coefficient of the calibrant, and ncoef is the number of coefficients in the power series (default is 8).

Examples

```
See also nugcalib
nugcal
powerfit
```

gradientdisable Disable PFG gradients (P)

Description gradientdisable is an optional global parameter for disabling the gradient pulses. If gradientdisable parameter is set to 'y', the psg software sets the gradient dac values to 0. The gradient parameters in VnmrJ and pulse sequence are not altered. This feature works in both C psg and SpinCAD Jpsg.

To use this feature, create gradientdisable as a global parameter of type 'flag'. If gradientdisable is set to 'y', the gradient amplitude values will be set to 0; if set to 'n' the gradient amplitudes will be the expected values determined by the gradient parameters and pulse sequence calculations. This feature is typically used in experiments involving Cold Probes. This feature is only effective for gradient configurations, gradtypes of 'l', 'p', and 't'.

RelatedpfgonPulsed field gradient amplifiers on/off control (P)gradtypeGradients for X, Y, and Z axes (P)

gradientshapingActivate shaping on the gradient pulses (P)

Applicability	Systems with Varian, Inc. Cold Probes
Description	Activate shaping on the gradient pulses in the pulse sequence without changing the pulse sequence source program. This feature works only the Z gradient pulses, specified using the $zgradpulse()$ PSG statement. gradientshaping is a global parameter.
Values	gradientshaping='y' enables this feature and produces a WURST shaping of gradient amplitudes. gradientshaping='n' or destroy the parameter disables this feature and produces rectangular gradients amplitudes.

gradstepsz Gradient step size (P)

Description	The maximum gradient DAC value. gradstepsz determines the type
	of gradient DAC board used in the system: 12-bit or 16-bit. It is used
	internally to convert gauss/cm gradient levels to the proper hardware
	DAC level.
Values	Systems with 12-bit DACs (older SISCO spectrometers without gradient waveform capabilities): -2047 to $+2047$ units, in integer steps.
	Systems with 16-bit DACs (SISCO spectrometers with gradient waveform capabilities): -32767 to +32767 units, in integer steps.
See also	VnmrJ Installation and Administration; VnmrJ Imaging NMR

gradtype Gradients for X, Y, and Z axes (P)

Applicability Systems with pulsed field gradient (PFG) or imaging capability.

- Description Configuration parameter for systems with optional gradients for axes. The value is set using the label X Axis, Y Axis, Z Axis in the Spectrometer Configuration window (opened from config). The values available for each axis are None, WFG + GCU, Performa I, Performa II/III, Performa II/III + WFG, Performa XYZ, Performa XYZ + WFG, SIS (12 bit), Homospoil, and Shim DAC. WFG stands for the waveform generator; GCU stands for the gradient compensation unit; and Performa I, II, III, and XYZ are types of PFG modules. For the Z Axis, Performa D is also available.
 - Values String of three characters (e.g., 'nnp'). The first character is the gradient for the X axis, second for the Y axis, and third for the Z axis. Each axis has value 'n' (None choice in Spectrometer Configuration window), 'w' (WFG+GCU), 'l' (Performa I), 'p' (Performa II/III and Performa D), 'q' (Performa II/III + WFG), 't' (Performa XYZ), 'u' (Performa XYZ + WFG), 's' (SIS (12 bit), or 'h' (Homospoil). Homospoil is functional only for the Z axis.
 - See also VnmrJ Installation and Administration; NMR Spectroscopy User Guide
 - Related config Display current configuration and possibly change it (M) pfgon PFG amplifiers on/off control (P)

graphis Return the current graphics display status (C)

Syntax (1) graphis:\$display_command (2) graphis(command):\$yes_no Description Determines what command currently controls the graphics window.

Arguments	\$display_command is a return value set to the name of the currently controlling command.
	command is the name of a command to be checked.
	\$yes_no is a return value set to 1 if the command name given by the command argument is controlling the graphics window, or set to 0 if it is not controlling the window.
Examples	graphis:\$display if (\$display='ds') then
	 endif
	graphis('ds'):\$ds_on if (\$ds_on) then
	 endif
See also	User Programming
Related	textis Return the current text display status (C)

grayctr Gray level window adjustment (P)

Description	Controls the	grayscale display available in dcon. In the dconi
	program, the	center mouse button controls the grayscale bar, which
	changes the	mean gray level and hence the value of grayctr. The
	grayctr pa	cameter (along with the parameter grays1) records the
	current setti	ngs of the gray bar as the interaction changes; the value
	can also be s	et directly. The right mouse button controls the data level
	of the maxin	num data intensity. To create grayctr, enter
	<pre>create('grayctr','real') setgroup('grayctr','display')</pre>	
	<pre>setlimit('</pre>	grayctr',64,0,1).
	To create the	e set of imaging parameters grayctr, dcrmv and
	graysl, and	in the current experiment, enter addpar('image').
Values	0 to 64 (typi	cally 32)
See also	NMR Spectroscopy User Guide	
Related	addpar	Add selected parameters to the current experiment (M)
	dcon	Display noninteractive color intensity map (C)
	dconi	Interactive 2D contour display (C)
	graysl	Gray level slope (contrast) adjustment (P)

grays1 Gray level slope (contrast) adjustment (P)

Description Controls the grayscale display available in dcon. In the dconi program, the center mouse button controls the grayscale slope as applied to the data changes and hence the value of grays1. Negative values of grays1 will invert black and white; however, negative values can be set only from the keyboard. graysl (along with the parameter grayctr) records the current settings of the gray bar as the interaction changes; the value can also be set directly. The right mouse button controls the data level of the maximum data intensity. To create graysl, enter the following command:

create('graysl','real') setgroup('graysl','display')
setlimit('graysl',10,-10,0.1)

To create the set of imaging parameters graysl, dcrmv, and grayctr in the current experiment, enter addpar('image').

Values -10 to +10 (-100 to +100, typically 1)

See also NMR Spectroscopy User Guide

RelatedaddparAdd selected parameters to the current experiment (M)dconDisplay noninteractive color intensity map (C)dconiInteractive 2D contour display (C)grayctrGray level window adjustment (P)

grecovery Eddy current testing (M)

Applicability	Systems with pulsed field gradient.
Description	Conditions an experiment for eddy current testing so that it is compatible with standard installation procedures.
See also	Pulsed Field Gradient Modules Installation, NMR Spectroscopy User Guide

grid Draw a grid on a 2D display (M)

- Syntax (1) grid<(<spacing><,><color>)>
 (2) grid<(start_f2,incr_f2,start_f1,incr_f1<,color>)>
- Description Draws grid lines over a 2D display. Grid lines are drawn on the graphics screen in the XOR mode—entering a second grid command with identical arguments erases (not redraws) the grid displayed by the first command.
- Arguments spacing specifies the approximate spacing of the grid lines, in cm. The default is intervals of approximately 1 cm, rounded so that the intervals fall at a multiple of 1, 2, or 5 (in Hz), or 1p, 2p, or 5p (in ppm).

color specifies the color of the grid lines and is one of the following keywords: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', or 'white'. The default is 'blue'.

start_f2, incr_f2, start_f1, incr_f1 define a grid by supplying the starting and increment frequencies for f2 and f1. Add the p suffix to a value to enter it in ppm (see third example below). Examples grid grid(1.5,'red') grid(1p,0.5p,3p,0.5p) See also NMR Spectroscopy User Guide Related plgrid Plot a grid on a 2D plot (M)

groupcopy Copy parameters of group from one tree to another (C)

Syntax	groupcopy(from_tree,to_tree,group)		
Description	Copies a set of ganother.	parameters of a group from one parameter tree to	
Arguments	from_tree, to_ the one of the k Refer to the cre	rom_tree, to_tree are two different parameter trees, each given by ne one of the keywords 'global', 'current', or 'processed'. efer to the create command for more information on trees.	
	group is the set 'all', 'sample	of parameters to be copied and is one of the keywords e', 'acquisition', 'processing', and 'display'.	
Examples	groupcopy('pr	ocessed','current','acquisition')	
See also	User Programming		
Related	create destroy destroygroup display setgroup	Create new parameter in a parameter tree (C) Destroy a parameter (C) Destroy parameters of a group in a tree (C) Display parameters and their attributes (C) Set group of a parameter in a tree (C)	

gspoil Spoiler gradient level (P)

Description Predefined parameter to set a spoiler gradient level.

gsspat Slice-select gradient shape (P)

Description Predefined string parameter to specify a slice-select gradient shape.

gtnnoesy Set up a PFG TNNOESY parameter set (M)

ApplicabilitySystems with the pulsed field gradient (PFG) module.DescriptionConverts a 1D standard two-pulse sequence parameter set into a
parameter set ready to run a PFG NOESY experiment (either absolute
value or phase sensitive) or a gtnnoesy experiment.

gtnroesy Set up a PFG absolute-value ROESY parameter set (M)

Applicability Systems with the pulsed field gradient (PFG) module.

Description Converts a 1D standard two-pulse sequence parameter set into a parameter set ready to run a PFG absolute-value ROESY experiment or a gtnroesy experiment.

gtotlimit Gradient total limit (P)

Applicability Systems with three-axis gradients

Description Sets the gradient limit, in gauss/cm, of the x, y, and z axes, summed together. This parameter is taken from an entry of the same name in a gradient table and should only exist if a gradient amplifier limits the combined output of all three gradient axis.

Related gcoil Read data from gradient calibration tables (P)

gtrim Trim gradient level (P)

Description Predefined parameter to set a trim gradient level.

gxmax, gymax, gzmaxMaximum gradient strength for each axis (P)

Applicability	Systems with three-axis gradients.	
Description	Defines the maximum gradient strength, in gauss/cm, for each gradient axis. These values are read in from the selected system gradient table whenever the parameter set is retrieved or the gradient coil defined by gcoil has changed. When the values are read in, gmax is set to the lowest value of the three.	
	The parameters $gxmax$, $gymax$, and $gzmax$ are used instead of $gmax$ when the gradients strengths are not equal for each axis. Unequal gradient strengths per axis are generally true for systems with three-axis PFG coils, which have a strong z gradient, and can be true for microimaging systems. Horizontal-bore imaging systems usually have gradients set to the same maximum value, and $gmax$ can be used.	
See also	NMR Spectroscopy User Guide; User Programming, VnmrJ Imaging NMR	
Related	gcoilRead data from gradient calibration tables (P)gmaxMaximum gradient strength (P)	

gzlv1 Pulsed field gradient strength (P)

Applicability	Systems with gradient shimming installed.	
Description	Specifies the pulsed field gradient DAC value.	
Values	Range from +2047 to -2048 for 12-bit gradient module, and from +32767 to -32768 for a 16-bit gradient module.	
Related	gzsize	Number of z-axis shims used by gradient shimming (P)
	gzwin	Spectral window percentage used for gradient shimming (P)

gzsize Number of z-axis shims used by gradient shimming (P)

Applicability	Systems with gradient shimming installed.	
Description	Specifies the number of z-axis shims used by gradient shimming. For example, gzsize set to 4 means that gradient shimming uses shims z1 to z4. By default, coarse shims are used if present, as determined by the shimset value	
Values	Integer from 1 to 8.	
Related	gmapshim	Start gradient autoshimming (M)
	gmapsys	Run gradient autoshimming, set parameters, map shims (M)
	gmapz	Get parameters and files for gmapz pulse sequence (M)
	gzlvl	Pulsed field gradient strength (P)
	gzwin	Spectral width percentage used by gradient shimming (P)
	shimset	Type of shimset (P)
	gmap_z1z4	Gradient shimming flag to first shim z1-z4 (P)

gzwin Spectral width percentage used for gradient shimming (P)

Applicability	Systems with gradient shimming installed.	
Description	Specifies the percentage of the spectral width sw used by gradient shimming for shimmap calculations. The value is set automatically with the buttons Find gzlvl/gzwin and Find gzwin in the gradient shimming system menu opened by gmapsys.	
Values	A real number between 0 and 100. The typical value is 50.	
Related	gmapshim gmapsys gmapz	Start gradient autoshimming (M) Run gradient autoshimming, set parameters, map shims (M) Get parameters and files for gmapz pulse sequence (M)

Pulsed field gradient strength (P)
Number of z-axis shims used by gradient shimming (P)
Spectral width in directly detected dimension (P)
Frequency offset for observe transmitter (P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

h1	Automated proton acquisition (M)
hlfreq	Proton frequency of spectrometer (P)
hlp	Process 1D proton spectra (M)
h2cal	Calculate strength of the decoupler field (C)
halt	Abort acquisition with no error (C)
hc	Automated proton and carbon acquisition (M)
hcapt	Automated proton, carbon, and APT acquisition (M)
hcchtocsy	Set up parameters for HCCHTOCSY pulse sequence (M)
hccorr	Automated proton, carbon, and HETCOR acquisition (M)
hcdept	Automated proton, carbon, and DEPT acquisition (M)
hcosy	Automated proton and COSY acquisition (M)
hdmf	Modulation frequency for the band selective homonuclear decoupling (P)
hcmult	Execute protocol actions of apptype hcmult (M)
hdof	Frequency offset for homodecoupling (P)
hdpwr	Power level for homodecoupling (P)
hdpwrf	Homodecoupling fine power (optional) (P)
hdres	Sets the tip angle resolution (P)
hdseq	Sets the decoupler waveform filename (P)
hdwshim	Hardware shimming (P)
hdwshimlist	List of shims for hardware shimming (P)
help	Display current help file
HELP	Help file for this tool
het2dj	Set up parameters for HET2DJ pulse sequence (M)
HETCOR	Change parameters for HETCOR experiment (M)



hetcor	Set up parameters for HETCOR pulse sequence (M)
hetcorcp1	Set up parameters for solids HETCOR pulse sequence (M)
hetcorps	Set up parameters for HETCORPS pulse sequence (M)
hetero2d	Execute protocol actions of apptype hetero2d (M)
hidecommand	Execute macro instead of command with same name (C)
hipwrampenable	High Power Amplifier Enable (P)
Hmbc	Convert the parameter to a HMBC experiment (M)
Hmqc	Convert the parameter to a HMQC experiment (M)
HMQC15	Set up parameters for ¹⁵ N HMQC experiment (M)
HMQC_d2	Set up parameters for ¹⁵ N HMQC experiment using dec. 2 (M)
HMQC_d213	Set up parameters for ¹³ C HMQC experiment using dec. 2 (M)
hmqcr	Set up parameters for HMQCR pulse sequence (M)
Hmqctoxy	Convert the parameter to a HMQCTOXY experiment (M)
HMQCTOXY15	Set up parameters for ¹⁵ N HMQCTOXY experiment (M)
HMQCTOXY_d2	Set up parameters for ¹⁵ N HMQCTOXY using decoupler 2 (M)
HMQCTOXY_d213	Set up parameters for ¹³ C HMQCTOXY using decoupler 2 (M)
hmqctoxy3d	Set up parameters for HMQC-TOCSY 3D pulse sequence (M)
ho	Horizontal offset (P)
hom2dj	Set up parameters for HOM2DJ pulse sequence (M)
homo	Homodecoupling control for the observe channel (P)
HOMODEC	Change parameters for HOMODEC experiment (M)
homo2d	Execute protocol actions of apptype homo2d (M)
homorof1	Delay before turning on homo decoupling rf (P)
homorof2	Delay after blanking the amp and setting T/R to receive (P)
homorof3	Delay between setting T/R switch to receive and gating the recvr on (P)
hoult	Set parameters alfa and rof2 according to Hoult (M)

hpa	Plot parameters on special preprinted chart paper (C)
Hprescan	Proton prescan (P))
hregions	Select integral regions in proton spectrum (M)
hs	Homospoil pulses (P)
Hsqc	Convert the parameter to a HSQC experiment (M)
HSQC15	Set up parameters for ¹⁵ N HSQC experiment (M)
HSQC_d2	Set up parameters for ¹⁵ N HSQC experiment using dec. 2 (M)
HSQC_d213	Set up parameters for ¹³ C HSQC experiment using dec. 2 (M)
HsqcHT	Set up the hsqcHT experiment (M)
Asqctoxy	Convert parameters to a HSQCTOXY experiment (M)
ISQCTOXY15	Set up parameters for ¹⁵ N HSQCTOXY experiment (M)
HSQCTOXY_d2	Set up parameters for ¹⁵ N HSQCTOXY using decoupler 2 (M)
HSQCTOXY_d213	Set up parameters for ¹³ C HSQCTOXY using decoupler 2 (M)
nsqctoxySE	Set up parameters for HSQC-TOCSY 3D pulse sequence (M)
nsrotor	Display rotor speed for solids operation (P)
nst	Homospoil time (P)
nt	Setting up and processing Hadamard experiments
ntbitrev	Hadamard bit reversal flag (P)
ntbw1	Hadamard pulse excitation bandwidth in ni (P)
ntcal1	RF calibration flag for Hadamard waveforms in ni (P)
ntfrq1	Hadamard frequency list in ni (P)
ntfrqdisp	Read, write, and display Hadamard frequencies
ntofs1	Hadamard offset in ni (P)
ntpwr1	Power level for RF calibration of Hadamard waveforms in ni (P)
ntss1	Stepsize for Hadamard waveforms in ni (P)
nzmm	Scaling factor for plots (P)
nztomm	Convert locations from Hz or ppm to plotter units (C)

h1 Automated proton acquisition (M)

Syntax h1<(solvent)>

Prepares parameters for automatically acquiring a standard ${}^{1}\text{H}$ spectrum. The parameter wexp is set to 'procplot' for standard processing. If h1 is used as the command for automation via the enter command, then au is supplied automatically and should not be entered on the MACRO line of the enter program. However, it is possible to customize h1 on the MACRO line by following it with additional commands and parameters. (e.g., entering h1 nt=1 uses the standard h1 setup but with only one transient).		
solvent is the name of the solvent. In automation mode, the solvent is supplied by the enter program. The default is 'CDCl3'.		
h1 h1('DMSO')		
NMR Spectroscopy User Guide		
au enter h1p procplot wexp	Submit experiment to acquisition and process data (M) Enter sample information for automation run (C) Process 1D proton spectra (M) Automatically process FIDs (M) When experiment completes (P)	
	Prepares pars spectrum. The processing. If command, the on the MACE customize h1 commands are h1 setup but solvent is t is supplied b h1 h1('DMSO') NMR Spectron au enter h1p procplot wexp	

hlfreq Proton frequency of spectrometer (P)

Description	Configuration parameter for the resonance frequency of ¹ H as
	determined by the field strength of the magnet. The value is set using
	the label Proton Frequency in the Spectrometer Configuration window.
Values	085, 100, 200, 300, 400, 500, 600, 700, 750, 800, 900 (in MHz); 3T, 4T.
See also	VnmrJ Installation and Administration
Related	config Display current configuration and possibly change it
	(M)

h1p Process 1D proton spectra (M)

Description Processes non-arrayed 1D proton spectra using standard macros. h1p is called by proc1d, but can also be used directly. Fully automatic processing (up to a point where a spectrum could be plotted) is provided: Fourier transformation (using preset weighting functions), automatic phasing (aphx macro), select integral regions (hregions macro), adjust integral size (integrate macro), vertical scale adjustment (vsadjc macro), avoiding excessive noise (noislm macro), threshold adjustment (if required, thadj macro), and referencing to the TMS signal if present (setref macro, then tmsref macro).

See also	NMR Spectroscopy User Guide		
Related	aphx	Perform optimized automatic phasing (M)	
	h1	Automated proton acquisition (M)	
	hregions	Select integral regions for proton spectra (M)	
	integrate	Automatically integrate 1D spectrum (M)	
	noislm	Avoids excessive noise (M)	
	proc1d	Processing macro for simple (non-arrayed) spectra (M)	
	setref	Set frequency referencing for proton spectra (M)	
	thadj	Adjust threshold (M)	
	tmsref	Reference spectrum to TMS line (M)	
	vsadjh	Adjust vertical scale for proton spectra (M)	

h2cal Calculate strength of the decoupler field (C)

Syntax	h2cal<(j1r,j2r<,j0>)><:gammah2,pw90,frequency>	
Description	Calculates the strength of the decoupler field. It uses the results from two experiments: one with the decoupler off-resonance at a lower frequency and the other with the decoupler off-resonance at a higher frequency than the frequency of the peak being decoupled.	
Arguments	jlr is the frequency of the decoupler during these two experiments;. The default is that h2cal prompts for a value. If the parameter dof is arrayed and has two values, h2cal assumes these two values represent the decoupler frequencies; if dof is arrayed and has more than two values, h2cal prompts for the two decoupler frequencies.	
	j2r is the reduced coupling constants from the two experiments. The default is that h2cal prompts for a value $% \lambda =0.011$	
	j0 is the full coupling constant that results when no decoupling is done. The default is a value of 142 Hz, the constant for the standard sample dioxane, or 15 Hz for the methyl iodide sample.	
	gammah2 is a return value set to the strength of the decoupler field.	
	pw90 is a return value set to the pulse width of a 90° pulse from the decoupler. It is related to the value of parameter dmf through the equation $dmf=1/pw90$.	
	frequency is a return value set to the coalescence point (i.e., frequency at which single-frequency decoupling would collapse the dioxane to a singlet).	
See also	NMR Spectroscopy User Guide	
Related	dmfDecoupler modulation frequency for first decoupler (P)dofFrequency offset for first decoupler (P)	

н

halt Abort acquisition with no error (C)

Syntax halt

Description Aborts an experiment that has been submitted to acquisition. If the experiment is active, it is aborted immediately, all data is discarded, and the experiment is interpreted as complete. Any data collected from an earlier block size transfer is retained. If any wexp processing is defined, that processing then occurs, followed by any queued experiments. The login name, and the FID directory path in file are used as keys to find the proper experiment to abort.

Under some circumstances, there is a delay between the time go is entered and the acquisition is started. During this time, instructions based on the selected pulse sequence are being generated. This is signified by the letters "PSG" appearing in the upper left corner of the status window. A halt command issued under these circumstances reports that no acquisition is active but it instead stops the instruction generation process and displays "PSG aborted".

See also NMR Spectroscopy User Guide

Related	aa	Abort acquisition with error (C)
	file	File name of parameter set (P)
	go	Submit experiment to acquisition (C)
	wexp	Specify action when experiment completes (C)
	wexp	When experiment completes (P)

hc Automated proton and carbon acquisition (M)

Syntax	hc<(solvent)>	
Description	Combines the operation of the h1 and c13 macros. In non-automation mode, both spectra are acquired in the experiment in which the hc macro was entered. After the completion of the acquisition, rttmp can be used for further processing of the two spectra.	
Arguments	solvent is the solvent name In automation mode, the enter program supplies the value. In non-automation mode, the default is 'cdcl3'.	
Examples	hc hc('dmso')	
See also	NMR Spectroscopy User Guide	
Related	c13Automatic carbon acquisition (M)enterEnter sample information for automation run (M,U)h1Automated proton acquisition (M)rttmpRetrieve experiment data from experiment subfile (M)	

hcapt Automated proton, carbon, and APT acquisition (M)

Syntax	hcapt<(solvent)>	
Description	Combines the operation of the h1 and c13 macros and the APT experiment. In non-automation mode, all spectra are acquired in the experiment in which the hcapt macro was entered. After acquisition completes, rttmp can be used for further processing of the three spectra.	
Arguments	solvent is the solvent name. In automation mode, the enter program supplies the value. In non-automation mode, the default is 'cdcl3'.	
Examples	hcapt hcapt('dmso')	
See also	NMR Spectroscopy User Guide	
Related	AptSet up parameters for APT experiment (M)c13Automatic carbon acquisition (M)enterEnter sample information for automation run (M,U)h1Automated proton acquisition (M)rttmpRetrieve experiment data from experiment subfile (M)	

hcchtocsy Set up parameters for HCCHTOCSY pulse sequence (M)

Description	Used for sidechain assignments in fully ¹³ C-enriched molecules.
See also	NMR Spectroscopy User Guide

hccorr Automated proton, carbon, and HETCOR acquisition (M)

Syntax	hccorr<(solvent)>	
Description	Combines the operation of the h1 and c13 macros and the HETCOR experiment. In non-automation mode, all spectra are acquired in the experiment in which hccorr is entered. After acquisition completes, rttmp can be used for further processing of the three spectra.	
Arguments	solvent is the solvent name. In automation mode, the enter program supplies the value. In non-automation mode, the default is 'cdcl3'.	
Examples	hccorr hccorr('dmso')	
See also	NMR Spectroscopy User Guide	
Related	c13 Automated carbon acquisition (M)	
	enter Enter sample information for automation run (M,U)	
	h1 Automated proton acquisition (M)	

hetcor	Set up parameters for HETCOR experiment (M)
rttmp	Retrieve experiment data from experiment subfile
	(M)

hcdept Automated proton, carbon, and DEPT acquisition (M)

Syntax	hcdept<(solvent)>	
Description	Combines the operation of the h1 and c13 macros and the DEPT experiment. In non-automation mode, all spectra are acquired in the experiment in which hcdept was entered. After the completion of the acquisition, rttmp can be used for further processing of the three spectra.	
Arguments	solvent is the solvent name. In automation mode, the enter program supplies the value. In non-automation mode, the default is 'cdcl3'.	
Examples	hcdept hcdept('dmso')	
See also	NMR Spectroscopy User Guide	
Related	c13Automatic carbon acquisition (M)DeptSet up parameters for DEPT experiment (M)enterEnter sample information for automation run (M,U)h1Automated proton acquisition (M)rttmpRetrieve experiment data from experiment subfile (M)	

hcosy Automated proton and COSY acquisition (M)

Syntax	hcosy<(solvent)>	
Description	Combines the operation of the h1 macro and the COSY experiment. In non-automation mode, both spectra are acquired in the experiment in which hcosy is entered. After acquisition completes, rttmp can be used for further processing of the two spectra.	
Arguments	solvent is the solvent name. In automation mode, the enter program supplies the value. In non-automation mode, the default is 'cdcl3'.	
Examples	hcosy hcosy('dmso')	
See also	NMR Spectroscopy User Guide	
Related	enter h1 rttmp	Enter sample information for automation run (C) Automated proton acquisition (M) Retrieve experiment data from experiment subfile (M)

hdmf Modulation frequency for homonuclear decoupling (P)

Applicability VNMRS liquids, 400 MR

Syntax hdmf=<value>

Description Sets the modulation frequency for the band selective homonuclear decoupling. The parameter specifies 1/pw90 at the power value, hdpwr, used for homonuclear decoupling. The parameter is not used with single frequency homonuclear decoupling.

Related	dutyc	The rf duty cycle fraction for homonuclear decoupling
		(P)
	hdof	Frequency offset for homodecoupling (P)
	hdpwr	Sets the rf attenuator to control the power for
		homonuclear decoupling (P)
	hdpwrf	Sets the rf linear modulator fine power for
		homonuclear decoupling (P)
	hdres	Sets the tip angle resolution (P)
	hdseq	Sets the decoupler waveform filename (P)
	homo	Homodecoupling control for observe channel (P)
	homorof1	Delay before turning on homo decoupling rf (P)
	homorof2	Delay after blanking the amplifier and setting T/R
		switch to receive (P)
	homorof3	Delay between setting T/R switch to receive gating on
		the receiver (P)
	tn	Nucleus for observe transmitter (P)

hcmult Execute protocol actions of apptype hcmult (M)

Description	This macro is used to e apptype.	execute the protocol actions of the hcmult	
Examples	<pre>hcmult('setup') - execute hcmult experimental setup hcmult('process') - execute hcmult processing hcmult('plot') - execute hcmult plotting</pre>		
See also	NMR Spectroscopy User	r Guide, VnmrJ Automation User Guide	
Related	apptype A execpars Se	pplication type (P) et up the exec parameters (M)	

hdof Frequency offset for homodecoupling (P)

Applicability VNMRS systems Syntax hodf=<value> Н

Description	Sets the irr similar to h is not used	radiation frequency offset for homonuclear decoupling and now tof, and dof determine the frequency. The parameter l if hdseq is set to a filename.
Values	-100000 to	100000 Hz in steps of 0.1 Hz.
Related	dutyc hdmf	The rf duty cycle fraction for homonuclear decoupling (P) modulation frequency for the band selective homonuclear decoupling (P)
	hdpwr	Sets the rf attenuator to control the power for homonuclear decoupling (P)
	hdpwrf	Homodecoupling fine power (optional) (P)
	hdres	Sets the tip angle resolution (P)
	hdseq	Sets the decoupler waveform filename (P)
	homo	Homodecoupling control for observe channel (P)
	homorof1	Delay before turning on homo decoupling rf (P)
	homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)
	homorof3	Delay between setting T/R switch to receive gating on the receiver (P)
	tn	Nucleus for observe transmitter (P)

hdpwr Power level for homodecoupling (P)

Applicability	VNMRS systems, 400 MR
Syntax	hdpwr= <value></value>
Description	Sets the rf attenuator to control the power for homonuclear decoupling. The dutyc parameter must be accounted for when setting hdpwr.
Values	-16 to 50 dB

CAUTION

ON Homodecoupling power greater than 2 watts in a switchable probe will damage the probe. Always carefully calibrate homodecoupling to avoid exceeding 2 watts. The maximum value for hdpwr is set to 49, corresponding to about 2 watts of power. The actual power delivered depends on the CW duty cycle. Before using close to the maximum value of power or duty cycle, ensure safe operation by measuring the output power.

Related	dutyc	The rf duty cycle fraction for homonuclear decoupling (P)
	hdmf	modulation frequency for the band selective homonuclear
		decoupling (P)
	hdof	Frequency offset for homodecoupling (P)
	hdpwrf	Homodecoupling fine power (optional) (P)
	hdres	Sets the tip angle resolution (P)
	hdseq	Sets the decoupler waveform filename (P)

homo	Homodecoupling control for observe channel (P)		
homorof1	Delay before turning on homo decoupling rf (P)		
homorof2	Delay after blanking the amplifier and setting T/R switch		
	to receive (P)		
homorof3	Delay between setting T/R switch to receive gating on the		
	receiver (P)		
tn	Nucleus for observe transmitter (P)		

hdpwrf Homodecoupling fine power (optional) (P)

Applicability	VNMRS liquids systems		
Syntax	hdpwrf= <value></value>		
Description	Sets the rf linear modulator fine power for homonuclear decoupling. The default is 4095 if the variable does not exist. Attenuation is added to the attenuation set by hdpwr.		
Values	0-4095		
Related	dutyc	The rf duty cycle fraction for homonuclear decoupling (P)	
	hdmf	Modulation frequency for the band selective homonuclear decoupling (P)	
	hdof	Frequency offset for homodecoupling (P)	
hdpwr Sets the rf attenuator to control the power for homonuclear decoupling (P)		Sets the rf attenuator to control the power for homonuclear decoupling (P)	
hdres Sets the tip angle resolution (P)		Sets the tip angle resolution (P)	
hdseq Sets the decoupler waveform filename		Sets the decoupler waveform filename (P)	
	homo	Homodecoupling control for observe channel (P)	
	homorof1	Delay before turning on homo decoupling rf (P)	
	homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)	
	homorof3	Delay between setting T/R switch to receive gating on the receiver (P)	
	tn	Nucleus for observe transmitter (P)	

hdres Sets the tip angle resolution (P)

VNMRS liquids systems
hdres= <value></value>
Sets the tip angle resolution to be used for the band selective waveform mode of homonuclear decoupling. The parameter is not used with single frequency homonuclear decoupling.
1 to 90 in units of degrees with 1 degree resolution

Related	dutyc	The rf duty cycle fraction for homonuclear decoupling (P)
	hdmf	Modulation frequency for the band selective
		homonuclear decoupling (P)
	hdof	Frequency offset for homodecoupling (P)
	hdpwr	Sets the rf attenuator to control the power for homonuclear decoupling (P)
	hdpwrf	Sets the rf linear modulator fine power for
		nomonuclear decoupling (P)
	hdseq	Sets the decoupler waveform filename (P)
	homo	Homodecoupling control for observe channel (P)
	homorof1	Delay before turning on homo decoupling rf (P)
	homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)
	homorof3	Delay between setting T/R switch to receive gating on the receiver (P)
	tn	Nucleus for observe transmitter (P)

hdseq Waveform filename for band selective decoupling (P)

Applicability	VNMRS liquids systems		
Syntax	hdseq='filename' - the file must have a.DEC. extension.		
Description	Sets the decoupler waveform filename (.DEC extension) for the band selective waveform mode. The irradiation frequency is determined by the transmitter offset last applied to the observe channel in the pulse sequence (typically tof) and any additional frequency offset from any phase modulation programmed implicitly into the waveform .DEC file		
Examples	hdseq='' or	does not exist - single frequency decoupling is used.	
Related	dutyc		
	hdmf	modulation frequency for the band selective homonuclear decoupling (P)	
	hdof	Frequency offset for homodecoupling (P)	
	hdpwr	Sets the rf attenuator to control the power for homonuclear decoupling (P)	
	hdpwrf	Sets the rf linear modulator fine power for homonuclear decoupling (P)	
	hdres	Sets the tip angle resolution (P)	
	homo	Homodecoupling control for observe channel (P)	
	homorof1	Delay before turning on homo decoupling rf (P)	
	homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)	
	homorof3	Delay between setting T/R switch to receive gating on the receiver (P)	
	tn	Nucleus for observe transmitter (P)	

hdwshim Hardware shimming (P)

Applicability	Systems with additional Z1 shimming hardware.		
Description	Allows go, su, au, etc., to turn on and off shimming hardware. Hardware shimming is automatically suspended during software autoshimming. Hardware shimming is only active during acquisition (go, ga, au). hdwshim is a global parameter, so it affects all experiments.		
Values	'y' turns hardware shimming on.		
	'p' turns hardware shimming on during presaturation pulse (power level change followed by pulse).		
	'n' turns shimming off.		
See also	NMR Spectroscopy User Guide		
Related	au Submit experiment to acquisition and process data (C)		
	go Submit experiment to acquisition (C)		
	su Submit a setup experiment to acquisition (M)		

ga Submit experiment to acquisition and FT the result (M)

hdwshimlist List of shims for hardware shimming (P)

Description	A global parameter that sets the shims to use during hardware shimming. If it does not exist, hardware shimming uses $z1$ by default	
	To create the create ('hd	e parameter, use wshimlist','string','global').
Values	Any string composed of z1, z1c, z2, z2c, x1, y1. Commas and blank space are ignored. Shimming is done in the order z1, z2, x1, y1, regardless of the order in the string.	
Examples	hdwshimlist='z1' hdwshimlist='z1z2x1y1'	
See also	NMR Spectroscopy User Guide	
Related	create hdwshim	Create new parameter in a parameter tree (C) Hardware shimming (P)

help Display current help file

Syntax	help		
Applicability	VnmrJ 3.1		
Description	This command displays help information that explains the functions of		
	the buttons and current utility active. This information is displayed		
	in the text window. The permanent help button on the Sun executes		
	the help command.		
	The help information that is displayed is from a file located in		

directory /vnmr/help. The name of the file matches the name of the currently active menu.

HELP Help File for this Tool

Syntax HELP Applicability VnmrJ 3.1

Description By default the help file for the current experiment defined by the pslabel parameter is shown. For all of the experiments that are found in the experiment selector items are listed under the Tab names. Thus, for example, there is a help file called J1CHTab which is general information for all of the supplied experiments located in that tab. Similarly there is an entry for Homo2DTab with general information for that group of experiments. For convenience the menu order of this help file arranged by the Tab names. If one opens Seq.Help the manual-select menu will be positioned so that all of the similar experiments for that group are nearby. All of help files are in alphabetical order.

There are two menus. One to select a specific experiment's help file and the other to conveniently view a help file describing all experiments in a given tab in the experiment selector. Both experiment and Tab help is available under the experiment menu.

het2dj Set up parameters for HET2DJ pulse sequence (M)

Description	Sets up a HET2DJ (heteronuclear 2D-J) experiment.
See also	NMR Spectroscopy User Guide
Related	foldj Fold J-resolved 2D spectrum about $f1=0$ axis (C)

HETCOR Change parameters for HETCOR experiment (M)

Description Converts the current parameter set to a HETCOR experiment. This is a phase-sensitive, multiplicity-selected experiment.

hetcor Set up parameters for HETCOR pulse sequence (M)

Syntax	hetcor<(exp_number)>
Description	Sets up a HETCOR (heteronuclear chemical shift correlation)
	experiment.

Arguments	exp_number is the number of the experiment, from 1 to 9, in which		
	a proton spe	ctrum of the sample already exists.	
See also	NMR Spectroscopy User Guide		
Related	plhxcor	Plot X,H-correlation 2D spectrum (M)	
	ppcal	Proton decoupler pulse calibration (M)	

hetcorcp1 Set up parameters for solids HETCOR pulse sequence (M)

Applicability	Systems with the solids module.	
Description	Sets up a parameter set, obtained with XPOLAR1, for HETCORCP1, the solid-state heteronuclear correlation experiment.	
See also	User Guide: Solid-State NMR	
Related	xpolar1	Set up parameters for XPOLAR1 pulse sequence (M)

hetcorps Set up parameters for HETCORPS pulse sequence (M)

Description	Sets up parameters for a heteronuclear chemical shift correlation
	experiment (absolute value and phase sensitive).
See also	NMR Spectroscopy User Guide

hetero2d Execute protocol actions of apptype hetero2d (M)

Applicability	Liquids	
Description	Perform the actions for Homonuclear 2D protocols to set up, process, and plot experiments.	
Examples	hetero2d('setup')execute hetero2d experimental setuphetero2d('process')execute hetero2d processinghetero2d('plot')execute hetero2d plotting	
See also	NMR Spectroscopy User Guide, VnmrJ Automation User Guide	
Related	apptypeApplication type (P)execparsSet up the exec parameters (M)	

hidecommand Execute macro instead of command with same name (C)

Syntax (1) hidecommand(command_name)<:\$new_name>
 (2) hidecommand('?')

Description	Renames (or hides) a built-in VnmrJ command so that a macro with the same name as the built-in command is executed instead of the built-in command.	
Arguments	command_name is the name of the command to be renamed. To reset the built-in command back to its original name, enter hidecommand with the hidden name as the argument.	
	<pre>\$new_name returns the new name of the built-in command. By using this new name, access is still available to the built-in command.</pre>	
	'?' is a keyword to display a list of all of the renamed built-in commands and their original names.	
Examples	hidecommand('sys'):\$newname hidecommand('Sys') hidecommand('?')	
See also	System Administration; User Programming	
Related	which Display which macro or command is used (M)	

hipwrampenable High Power Amplifier Enable (P)

Applicability	VNMRS solids and systems with high power amplifiers.		
Description	This parameter controls the High/Low Power Relay. If the parameter does not exist low power is used. If the parameter exists and the field corresponding to the physical channel is 'n' then low power is used. If the parameter exists and the field corresponding to the physical channel is ' y ' then high power is used. The parameter is created in the current tree as a flag with create('hipwrampenable', 'flag').		
Values	'y' Enable high power 'n' Enable low power and disable high power		
Examples	hipwrampenable='yny'		
	Physical channel 1 and 3 are high power enabled. Physical channel 2 is low power.		

Hmbc Convert the parameter to a HMBC experiment (M)

Description Convert the parameter to a HMBC experiment. See also NMR Spectroscopy User Guide

Hmqc Convert the parameter to a HMQC experiment (M)

Description Convert the parameter to a HMQC experiment.

Description Converts the current parameter set to a HMQC experiment for ¹⁵N.

HMQC_d2 Set up parameters for ¹⁵N HMQC experiment using dec. 2 (M)

Description Converts the current parameter set to a HMQC experiment for 15 N with decoupler 2 as 15 N.

HMQC_d213 Set up parameters for ¹³C HMQC experiment using dec. 2 (M)

Description Converts the current parameter set to a HMQC experiment for 13 C with decoupler 2 as 13 C.

hmgcr Set up parameters for HMQCR pulse sequence (M)

Applicability Not needed in current systems. Normally was used in systems with a ¹H only decoupler.
 Description Sets up a HMQC (heteronuclear multiple-quantum coherence) experiment with "reverse" configuration.
 See also NMR Spectroscopy User Guide

Hmgctoxy Convert the parameter to a HMQCTOXY experiment (M)

Description Convert the parameter to a HMQCTOXY experiment.

HMQCTOXY15 Set up parameters for ¹⁵N HMQCTOXY experiment (M)

Description Converts the current parameter set to a HMQCTOXY experiment for ${\rm ^{15}N}.$

HMQCTOXY_d2 Set up parameters for ¹⁵N HMQCTOXY using decoupler 2 (M)

Description Converts the current parameter set to a HMQCTOXY experiment for 15 N with decoupler 2 as 15 N.

HMQCTOXY_d213Set up parameters for ¹³C HMQCTOXY using decoupler 2 (M)

Description Converts the current parameter set to a HMQCTOXY experiment for ${}^{13}C$ with decoupler 2 as ${}^{13}C$.

hmqctoxy3d Set up parameters for HMQC-TOCSY 3D pulse sequence (M)

Description Sets up parameters for a HMQC-TOCSY 3D experiment with a presaturation option.

ho Horizontal offset (P)

- Description Horizontal offset of the each spectrum in a "stacked display" with respect to the previous spectrum,. For 1D data sets, the parameter vo sets the vertical offset. For 2D data sets, the parameter wc2 sets the vertical distance (in mm) between the first and last traces.
 - Values Number, in mm, for offset size. For a "left-to-right" presentation, ho is typically negative; for "bottom-to-top" presentation, vo or wc2 is positive.

hom2dj Set up parameters for HOM2DJ pulse sequence (M)

Description Sets up a HOM2DJ (homonuclear J-resolved 2D) experiment. See also NMR Spectroscopy User Guide

homo Homodecoupling control for the observe channel (P)

Applicability VNMRS liquids systems

Description	Homonuclear decoupling irradiates a single frequency if hdseq = ''
	(or if hdseq does not exist) or a frequency band if hdseq
	='filename'. Pulse sequences do not require explicit homonuclear
	gating commands (homo function is similar to dm). A single RF channel,
	the observe channel, is used. The homo='y' setting cannot be used
	with pulse sequences containing explicit acquire commands.
Syntax	homo=<'y' or 'n'>

- Values 'y' homonuclear decoupling rf and receiver gating is turned on during the acquisition time. Provides single frequency or band selective (hdseq = 'filename') decoupling.
 - 'n' homonuclear decoupling rf and receiver gating is turned off.

Related	hdof	Frequency offset for homodecoupling (P)
	hdpwr	Power level for homodecoupling (P)
	hdpwrf	Homodecoupling fine power (P)
	dutyc	Duty cycle for homodecoupling (optional) (P)
	tn	Nucleus for observe transmitter (P)
	homorof1	Delay before turning on homo decoupling rf (P)
	homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)
	homorof3	Delay between setting T/R switch to receive gating on the receiver (P)

HOMODEC Change parameters for HOMODEC experiment (M)

Description Converts the current parameter set to a HOMODEC experiment. A 1D proton spectrum is displayed to do peak selection.

homo2d Execute protocol actions of apptype homo2d (M)

Applicability	Liquids
Description	Perform the actions for Heteronuclear 2D protocols to set up, process, and plot experiments.
Examples	homo2d('setup')execute homo2d experimental setuphomo2d('process')execute homo2d processinghomo2d('plot')execute homo2d plotting
See also	NMR Spectroscopy User Guide, VnmrJ Automation User Guide
Related	apptype Application type (P)
	execpars Set up the exec parameters (M)

Applicability VNMRS liquids systems

Description Optional parameter for delay before turning on homonuclear decoupling after gating the receiver off. The amplifier is un-blanked and T/R switch set to transmit mode during homorof1 delay (in μ sec. units). A default delay of 2 μ sec. is used if the parameter does not exist.

Values 2 to 5 µsec. are typical.

Related	dutyc	The rf duty cycle fraction for homonuclear decoupling (P)
	hdmf	Modulation frequency for the band selective homonuclear decoupling (P)
	hdof	Frequency offset for homodecoupling (P)
	hdpwr	Sets the rf attenuator to control the power for homonuclear decoupling (P)
	hdpwrf	Sets the rf linear modulator fine power for homonuclear decoupling (P)
	hdseq	Sets the decoupler waveform filename (P)
	hdres	Sets the tip angle resolution (P)
	homo	Homodecoupling control for observe channel (P)
	homorof2	Delay after blanking the amplifier and setting T/R switch to receive (P)
	homorof3	Delay between setting T/R switch to receive gating on the receiver (P)
	tn	Nucleus for observe transmitter (P)

homorof2 Delay after blanking the amp and setting T/R switch to recv (P)

Applicability	VNMRS liquids systems		
Description	Optional parameter for delay after the transmitter is gated off, the amplifier is blanked, and before the T/R switch is set to receive. A default delay of 2 μ sec. is used if the parameter does not exist.		
Values	2 to 5 µsec. are typical.		
Related	dutyc hdmf hdof hdpwr	The rf duty cycle fraction for homonuclear decoupling (P) Modulation frequency for the band selective homonuclear decoupling (P) Frequency offset for homodecoupling (P) Sets the rf attenuator to control the power for homonuclear decoupling (P)	
	hdpwrf	Sets the rf linear modulator fine power for homonuclear decoupling (P)	
	hdseq	Sets the decoupler waveform filename (P)	
	hdres	Sets the tip angle resolution (P)	

homo	Homodecoupling control for observe channel (P)
homorof1	Delay before turning on homo decoupling rf (P)
homorof3	Delay between setting T/R switch to receive gating on the
	receiver (P)
tn	Nucleus for observe transmitter (P)

homorof3 Delay between setting T/R to receive and gating the recvr on (P)

Applicability	VNMRS liquids systems		
Description	Optional pa before the if the param	trameter for delay after the T/R switch is set to receive and receiver gate is gated on. A default delay of 2 $\mu sec.$ is used meter does not exist.	
Values	2 to 5 µsec. are typical		
Related	dutyc	The rf duty cycle fraction for homonuclear decoupling (P)	
	hdmf	Modulation frequency for the band selective homonuclear decoupling (P)	
	hdof	Frequency offset for homodecoupling (P)	
	hdpwr	Sets the rf attenuator to control the power for	
		homonuclear decoupling (P)	
	hdpwrf	Sets the rf linear modulator fine power for homonuclear decoupling (P)	
	hdseq	Sets the decoupler waveform filename (P)	
	hdres	Sets the tip angle resolution (P)	
	homo	Homodecoupling control for observe channel (P)	
	homorof1	Delay before turning on homo decoupling rf (P)	
	homorof2	Delay after blanking the amplifier and setting T/R switch	
		to receive (P)	
	tn	Nucleus for observe transmitter (P)	

hoult Set parameters alfa and rof2 according to Hoult (M)

Sets the values of alfa and rof2 according to a prescription advanced by D. I. Hoult (<i>J. Magn. Reson.</i> 51 , 110 (1983)). These parameters set the times that follow the final pulse, which can be important where the flatness of the baseline is of concern.		
NMR Spectroscopy User Guide		
alfa calfa rof2	Set alfa delay before acquisition (P) Recalculate alfa so that first-order phase is zero (M) Receiver gating time following pulse (P)	
	by D. I. Hould the times tha the flatness of <i>NMR Spectro</i> alfa calfa rof2	

н

Description	Plots a predetermined list of parameters by "filling in the blanks" a the bottom of the preprinted chart paper available for Hewlett-Packar 7475- and 7550-series plotters.		
See also	NMR Spectroscopy User Guide		
Related	apa	Plot parameters automatically (M)	
	x 0	X-zero position of HP plotter or Postscript device (P)	
	У0	Y-zero position of HP plotter or Postscript device (P)	

Hprescan Proton prescan (P))

Applicability	VnmrJ Walkup
Description	This parameter is used to keep track of the type and status of the Proton prescan. It is used for Proton, Presat, Wet1d, and Minsw protocols.
See also	VnmrJ Automation User Guide
Related	xmHprescan Set up and process Proton prescans (M)

hregions Select integral regions in proton spectrum (M)

- Description Selects integral regions, a critical step in automatic processing of proton spectra. It is critical not only because of aesthetic reasons (some people like many small integrals, others prefer a few large regions), but also because other commands, such as bc, depend on the correct integration: bc can either fail or it can make broad, unintegrated lines disappear from the spectrum. hregions was specifically designed for proton spectra and should not be used for other types of spectra. The result of hregions also depends on the lineshape and the signal-to-noise ratio of a spectrum
 - See also NMR Spectroscopy User Guide

Relatedbc1D and 2D baseline correction (C)integrateAutomatically integrate 1D spectrum (M)

hs Homospoil pulses (P)

Description Turns on homospoil pulses at various times in different pulse sequences. Homospoil is a process by which the homogeneity is temporarily made very bad ("spoiled") to cause any transverse
	magnetizations present at that time to decay rapidly to zero. hst controls the length of any homospoil pulse.
Values	In a standard two-pulse sequence, homospoil pulses can be inserted during periods A and B (delays d1 and d2): $hs='yn'$ gives a homospoil pulse at the beginning of d1, $hs='ny'$ gives a pulse during d2, and $hs='yy'$ gives homospoil pulses during both d1 and d2. The desired value is generally $hs='nn'$.
See also	NMR Spectroscopy User Guide
Related	d1First delay (P)d2Incremented delay in 1st indirectly detected dimension (P)hstHomospoil time (P)
Hsqc	Convert the parameter to a HSQC experiment (M)
Description	Convert the parameter to a HSQC experiment.
HSQC15	Set up parameters for ¹⁵ N HSQC experiment (M)
Description	Converts the current parameter set to a HSQC experiment for $^{15}\mathrm{N}.$
HSQC_d2	Set up parameters for ¹⁵ N HSQC experiment using dec. 2 (M)
Description	Converts the current parameter set to a HSQC experiment for 15 N with decoupler 2 as 15 N.
HSQC_d213	Set up parameters for ¹³ C HSQC experiment using dec. 2 (M)
Description	Converts the current parameter set to a HSQC experiment for 13 C with decoupler 2 as 13 C.
hsqcHT	Set up the hsqcHT experiment (M)
Description	Sets up parameters for a Hadamard-encoded hsqc experiment.

See also	NMR Spectroscopy User Guide	
Related	htofs1	Hadamard frequency list in ni (P)
	htfrq1	Hadamard offset in ni (P)
	fn1	Fourier number in 1st indirectly detected dimension
		(P)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	ft2d	Fourier transform 2D data (C)
	sethtfrq1	Set a Hadamard frequency list from a line list (M)
	Hsqc	Set up parameters for HSQC experiment (M)

Hsqctoxy Convert parameters to a HSQCTOXY experiment (M)

Description Convert the parameter to a HSQCTOXY experiment.

HSQCTOXY15 Set up parameters for ¹⁵N HSQCTOXY experiment (M)

Description Converts the current parameter set to a HSQCTOXY experiment for ${\rm ^{15}N}.$

HSQCTOXY_d2 Set up parameters for ¹⁵N HSQCTOXY using decoupler 2 (M)

Description Converts the current parameter set to a HSQCTOXY experiment for ${}^{15}N$ with decoupler 2 as ${}^{15}N$.

HSQCTOXY_d213Set up parameters for ¹³C HSQCTOXY using decoupler 2 (M)

Description Converts the current parameter set to a HSQCTOXY experiment for ${}^{13}C$ with decoupler 2 as ${}^{13}C$.

hsqctoxySE Set up parameters for HSQC-TOCSY 3D pulse sequence (M)

Description Sets up parameters for a HSQC - TOCSY 3D experiment.

hsrotor Display rotor speed for solids operation (P)

Applicability Systems equipped with the rotor synchronization module.

Description Controls display of rotor speed. Depending on whether the rotor synchronization module is present (set by the Rotor Synchronization label in the Spectrometer Configuration window, parameter rotorsync is set to 1 or 0. The xpolar1 macro in turn uses this to create hsrotor, which is set to 'y' if rotor synchronization is present. If the parameter srate exists, it is updated to the spin speed of the rotor at the end of the experiment. The interlock function specified by parameter in also changes. If hsrotor='y' and in='y', the experiment is terminated if rotor speed deviates more than 100 Hz.

hst Homospoil time (P)

Description	Controls pulse length if homospoil is activated by the hs parameter.		
Values	0 to 20 ms (limited by hardware).		
Values	'n' makes srate unmodified by acquisition and turns off the rotor speed display in Acqstat.		
	'y' makes th board update status display	he hardware information from the rotor synchronization srate and displays the rotor speed in the Acqstat 7.	
See also	User Guide: 1	Solid-State NMR	
Related	Acqstat	Bring up the acquisition status display (U)	
	config	Display current configuration and possibly change it	
		(M)	
	in	Interlock (P)	
	rotorsync	Rotor synchronization (P)	
	srate	Spinning speed (P)	
	xpolar1	Set up parameters for XPOLAR1 pulse sequence (M)	

ht Setting up and processing Hadamard experiments.

Syntax	ht		
Applicability	VnmrJ 3.1		
Description	To set up a Hadamard experiment, do the following.		
	1. First run a Proton, Carbon, or other 1D experiment.		
	2. When the acquisition is finished, process and phase the spectrum.		
	3. Run the editht macro to open the Edit HT Freq popup. Create a Hadamard frequency list for the nucleus of interest. Save the frequency list.		

4. For a heteronuclear Hadamard experiment, run a Proton experiment, and adjust spectral width and decoupling as desired.

5. Load the desired Hadamard experiment. Check the Hadamard frequency list and other parameters.

6. Start the acquisition of the Hadamard experiment.

7. When acquisition is complete, process with proc1='ht' wft2da. Parameters used:

htfrq1: Hadamard frequency list in indirect dimension.

ni: Number of increments in indirect dimension. Typically set to the size of htfrq1 plus htofs1.

htofs1: Offset in Hadamard processing (number of increments to skip).

proc1: Type of processing in indirect dimension. Set to 'ht'.

fn1: Fourier number in indirect dimension. It must be larger than the number of Hadamard frequencies, and larger than the minimum difference between Hadamard frequencies.

Hadamard Spectroscopy

Hadamard spectroscopy is a technique for acquiring multidimensional data sets using a small number of transients, and reconstructing the nD spectrum using a Hadamard transform. It is based on selective excitation of a predetermined set of frequencies.

Acquisition.

A list of frequencies to selectively excite is determined from a 1D spectrum, or other means. A series of shaped pulses is created from the frequency list, using a Hadamard matrix to selectively excite or invert the signals of interest. The matrix size must be greater than the number of frequencies in the list. A typical 8x8 Hadamard matrix is shown below.

+++++++ +++--++ ++---++ +-+-+++ +-++-++ +--++++ +--++++

Processing.

Hadamard processing in the indirect dimension is done by summing, adding or subtracting the acquired data increments in combinations according to the Hadamard matrix elements. Each sum gives a trace corresponding to one of the frequencies in the list, and is placed at the appropriate frequency in the indirect dimension. The areas between the frequencies in the list are zero filled. The direct dimension is Fourier transformed, giving the 2D spectrum.

```
See also editht
getht
HsqcHT
tocsyHT
sethtfrq1
ft2d
ft2da
E Kunce and
```

E. Kupce and R. Freeman, "Two-dimensional Hadamard spectroscopy," J. Magn. Reson. 162 (2003), pp. 300-310.

htbitrev Hadamard bit reversal flag (P)

Description	A flag to enable or disable bit reversal of the Hadamard matrix. The flag should be the same for both acquisition and processing for the Hadamard transform to be successful.
Values	'y' enable Hadamard bit reversal 'n' disable Hadamard bit reversal Default value is 'n'.
See also	NMR Spectroscopy User Guide
Related	htfrq1 Hadamard frequency list in ni (P)

htbw1 Hadamard pulse excitation bandwidth in ni (P)

Description	The excitation bandwidth used to generate the frequencies contained in the shaped pulses used by the Hadamard matrix. If a single value is specified, the same bandwidth is used for all frequencies. If the parameter is arrayed, the bandwidth array element is used by the corresponding array element in htfrq1.		
Values	Default value is 20.0 if the parameter does not exist.		
See also	NMR Spectroscopy User Guide		
Related	htfrq1	Hadamard frequency list in ni (P)	
	ni	Number of increments in 1st indirectly detected dimension (P)	

htcal1 RF calibration flag for Hadamard waveforms in ni (P)

- Description A flag to allow power optimization of Hadamard waveforms in the 1st indirect dimension.
 Values 0 power optimization using htpwr1 is disallowed
 >0 power optimization using htpwr1 is allowed
 - Default value is 0.

See also	NMR Sp	ectroscopy User Guide
Related	htfrq1	Hadamard frequency list in ni (P)
	htpwr1	Power level for rf calibration of Hadamard waveforms in ni
	nı	Number of increments in 1st indirectly detected dimension
		(P)

htfrq1 Hadamard frequency list in ni (P)

Description	A list of frequencies used in Hadamard spectroscopy, used for creating the Hadamard pulse shapes, and for placing the transformed traces at the correct frequencies in the indirect dimension.		
Values	Typical values are an arrayed set of frequencies between $-{\tt sw1/2}$ and ${\tt sw1/2}.$		
See also	NMR Spectroscopy User Guide		
Related	htofs1	Hadamard offset in ni (P)	
	fn1	Fourier number in 1st indirectly detected dimension (P)	
	ni	Number of increments in 1st indirectly detected	
		dimension (P)	
	sethtfrq1	Set Hadamard frequency list from a line list (M)	
	proc1	Type of processing on ni interferogram (P)	
	sw1	Spectral width in 1st indirectly detected dimension (P)	

htfrqdisp Read, write, and display Hadamard frequencies.

Syntax	htfrqdisp		
Applicability	VnmrJ 3.1		
Description	The htfrqdisp macro is used by the Edit HT Freq dialog for setting and displaying Hadamard frequencies. It is not usually used from the command line.		
See also	editht getht ht HsqcHT tocsyHT sethtfrq1 ft2d ft2da		

htofs1 Hadamard offset in ni (P)

- Description The number of array elements to skip in ni when doing the Hadamard transform. The first element of the Hadamard matrix typically has all positive values (++++), and is usually not useful in constructing the Hadamard data.
 - Values Default value is 0. Typical values are 1 or 2.

See also NMR Spectroscopy User Guide

htpwr1 Power level for RF calibration of Hadamard waveforms in ni (P)

Description	Power level for optimizing Hadamard waveforms in the 1st indirect dimension.		
Values	-16 to 63 dB in steps of 1 dB.		
See also	NMR Spectroscopy User Guide		
Related	htfrq1Hadamard frequency list in ni (P)htcal1RF calibration flag for Hadamard waveforms in ni (P)niNumber of increments in 1st indirectly detected dimension (P)		

htss1 Stepsize for Hadamard waveforms in ni (P)

Description	Sets the ste parameter i	psize during Hadamard waveform creation. Typically, this s not needed, and a default stepsize is used.
Values	Does not exist - default stepsize is used. 0 default stepsize is used. >0 stepsize in microseconds.	
See also	NMR Spectr	roscopy User Guide
Related	htfrq1 ni	Hadamard frequency list in ni (P) Number of increments in 1st indirectly detected dimension (P)

hzmm Scaling factor for plots (P)

Description	Contains the quotient of wp divided by wc, a scaling factor useful for		
	plotting. hzmm applies to 1D only.		
See also	NMR Spectroscopy User Guide		
Related	wc Width of chart (P)		
	WP Width of plot (P)		

hztomm Convert locations from Hz or ppm to plotter units (C)

Syntax	<pre>(1) hztomm(x_position)<:xmm> (2) hztomm(x_position,y_position)<:xmm,ymm> (3) hztomm(<'box',><'plotter' 'graphics',>x_left, x right,y bottom,y top)<:x1mm,x2mm,y1mm,y2mm></pre>		
Description	Converts locations from Hz, or ppm, to plotter units.		
Arguments	x_position in syntax 1 is a location along the 1D axis, in Hz or ppm, to be converted to plotter units using the current values of parameters sp and wp. Plotter units are mm on most plots and are scaled for graphics display. For ppm entries, use the p suffix following numerical values (see first example below).		
	x_position, y_position in syntax 2 is a coordinate, in Hz or ppm, on a 2D plot to be converted to plotter units, using the parameters sp and wp to convert the horizontal position and the parameters $sp1$ and $wp1$ to convert the vertical position.		
	x_left,x_right,y_bottom,y_top in syntax 3 are box edges, in Hz or ppm, on a 2D plot to be converted to plotter units, using the parameters sp and wp to convert the left and right edges, and parameters sp1 and wp1 to convert the top and bottom edges.		
	'box' is a keyword to draw a box and to make the first two return arguments, if supplied, give the location of the upper left corner of the box, in plotter units.		
	'plotter' is a keyword to select the plotter. The default is 'graphics'.		
	'graphics' is a keyword to select the graphics screen. This is the default.		
	x1mm, x2mm, y1mm, y2mm are return arguments giving values in plotter units. If return arguments are not supplied, the results are displayed instead.		
Examples	hztomm(20p) hztomm(xpos,ypos):xmm,ymm hztomm('box','plotter',20,50,10,30)		
See also	NMR Spectroscopy User Guide		
Related	boxDraw a box on a plotter or graphics display (C)spStart of plot in directly detected dimension (P)		

sp1	Start of plot in 1st indirectly detected dimension
	(P)
wp	Width of plot in directly detected dimension (P)
wp1	Width of plot in 1st indirectly detected dimension
	(P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

i	Insert sample (M)
ihwinfo	Hardware status of console (U)
il	Interleave arrayed and 2D experiments (P)
ilfid	Interleave FIDs during data processing (C)
imagefile	Display an image file (M)
imagemath	Fit images to an specified function (M)
imageprint	Plot non interactive gray scale image (M)
imconi	Display 2D data in interactive grayscale mode (M)
import1Dspec	Import ASCII Spectrum into VnmrJ / VNMR (M)
in	Lock and spin interlock (P)
inadqt	Set up parameters for INADEQUATE pulse sequence (M)
index2	Projection or 3D plane index selected (P)
inept	Set up parameters for INEPT pulse sequence (M)
initialize_iterat	Set iterate string to contain relevant parameters (M)
input	Receive input from keyboard (C)
ins	Integral normalization scale (P)
ins2	2D volume value (P)
insref	Fourier number scaled value of an integral (P)
ins2ref	Fourier number scaled volume of a peak (P)
insert	Insert sample (M)
inset	Display an inset spectrum (C)
integ	Find largest integral in a specified region (C)
integrate	Automatically integrate 1D spectrum (M)
int_flg	determines whether dosy uses integrals or peak heights for DOSY fitting (P)



Agilent Technologies

intmod	Integral display mode (P)
intvast	Produces a text file of integral regions (M)
intvast	Produce a text file containing the integral of the partial regions (M)
io	Integral offset (P)
is	Integral scale (P)
isadj	Automatic integral scale adjustment (M)
isadj2	Automatic integral scale adjustment by powers of two (M)
isCSIMode	Determine if graphics area is split for CSI mode (C)
isiin	System global parameter for ISI interlock
isreal	Utility macro to determine a parameter type (M)
isstring	Utility macro to determine a parameter type (M)
isvnmrj	Identifies the interface that is in use, either Vnmr or VnmrJ
iterate	Parameters to be iterated (P)

i Insert sample (M)

DescriptionTurns off the eject air, waits for sample to slowly drop, and then turns
off the slow drop air. The macro insert functions the same as i.See alsoNMR Spectroscopy User GuideRelatedeEject sample (M)ejectEject sample (M)insertInsert sample (M)

ihwinfo Hardware status of console (U)

Syntax	(From UNIX) ihwinfo('startup' 'abort')	
Description	Displays status of digital hardware in the console. The output is intended for service personnel and probably not meaningful to users.	
Arguments	'startup' is a keyword to display the status at the conclusion of t last console startup (powerup, reboot, etc.).	
	'abort' is a keyword to display the status the last time an acquisition was aborted or the console rebooted from the host computer (abortallacqs). In this context, exiting from either the FID display	

or lock display of acqi counts as an abort. Only the status from the last abort can be displayed. Examples ihwinfo('startup')

	ihwinfo('abo	rt')
See also	NMR Spectrosco	opy User Guide
Related	abortallacqs	Reset acquisition computer in a drastic situation (C)
	showconsole	Show console configuration parameters (U)

il Interleave arrayed and 2D experiments (P)

Applicability Interleaving is not currently is implemented for VNMRS, 400MR and their DD2 analogs.

Description Experimental interleaving, which is turned on with il="y", applies only to arrayed experiments. bs transients are performed for each member of the array, followed by bs more transients for each member of the array, so on until nt transients have been collected for each member of the array. Thus il will only be of relevance if bs < nt.

Values: "y","n"

There are some restrictions on the pulse sequence statements that work correctly with the interleave feature on VNMRS architecture systems such as VNMRS, 400MR and their DD2 analogs.

For interleave to work correctly on the above systems, the phase cycling and other real-time calculations must be based on ct or ssctr real time variables. Pulse sequence statements that do a relative modification of real-time variables such as incr(), decr(), dbl(), hlv(), etc may not work correctly with il='y' setting. All phase calculations and other real time calculations must be defined starting with operations on ct or ssctr variable.

In addition the initialization function initval() may also not work correctly. Instead of initval() use the variant $F_{initval}$ for this purpose.

ilfid Interleave FIDs during data processing (C)

Description Converts a multiple FID element into a single FID. It is possible to effectively extend the Nyquist frequency (i.e., increase the effective spectral width sw) by acquiring a number of FIDs with different *tau2* values and then reprocessing the data. ilfid does the necessary processing of time-domain data to achieve this extension, assuming that a pulse sequence (not supplied) has been written to generate the required data.

When invoked in an experiment of nf FIDs, each of np points, ilfid sorts the data into a single FID of np*nf points that can then be

transformed. The interleaving takes the first complex point of each of the nf FIDs and places them in sequential order in the new FID. It then takes the second complex point from each of the nf FIDs and appends them sequentially to the new FID. This operation is repeated for all complex points. Although ilfid adjusts np and nf, it does not alter other parameters such as sw.

CAUTION

Because ilfid alters the data irrevocably, it is strongly recommended that you save the FID before using ilfid.

Examples	Illustrated below is the interleaving of an FID with nf=3 and np=4. Each point is represented by two digits. The first digit is the nf number and the second digit is the sequential point for that nf value. Data before the ilfid command:		
	11, 12, 13, 14; 21, 22, 23, 24; 31, 32, 33, 34		
	Data after the ilfid command:		
	11, 21, 31, 12, 22, 32, 13, 23, 33, 14, 24, 34		
See also	NMR Spectroscopy User Guide		
Related	nf Number of FIDs (P)		
	np Number of data points (P)		
	sw Spectral width in directly detected dimension (P)		

imagefile Display an image file (M)

Applicability	Imaging		
Syntax	<pre>imagefile('output_option','imagefile'<,x,y,w,h,'mol'>)</pre>		
Description	Display or plot an imagefile at default location and size or, optionally, at location and size specified by: x (x-position), y (y-position), w (width), h (height), and mol if it is an image file of a molecular structure. Display all, plot all, or clear all images for the current experiment.		
Arguments	output_option choices are:		
	clear, clear all images for the current experiment display, display imagefile displayall, displays all images for the current experiment plot, plot imagefile plotall, plot all images for the current experiment		
	imagefile, name of image file to display or plot		
	x, x position		
	y, y position		
	w, width		
	h, height		

mol molecular structure image file

Examples imagefile('clear') clear all images for the current experiment. imagefile('displayall') display all images for the current experiment.

imagemath Fit images to an specified function (M)

Syntax imagemath(fit_type,fit_var,dir_flag)

Description Calls standalone Linux program to fit data to the specified function (fit_type), either T2, or DIFF for a T2 map or diffusion calculation.

Data is fitted to a single exponential with the ADC or T2 options. The output is given in two images:

A computed S(0) image (filename S0)

A map of either ADC or T2 (filenameADC or filenameT2).

The diffcalc linux program is invoked with the DIFF option. The output depends on the number of diffusion directions applied.

The argument dir_flag (if supplied) or the parameter aipData (if dir_flag is not supplied), determines where the program reads and writes data; if aipData or dir_flag = 'saved', it uses the parameter file to determine the input directory (e.g., sems_01.img), and appends the name of the fit type to the directory name (e.g., sems_01_ADC.img) for the output directory; if aipData or dir_flag = 'processed', it uses curexp/recon as the input directory and curexp/<fit_type> as the output directory. Calling imagemath from the Current viewport, using the current data, reads the data from/written to curexp.

See the *VnmrJ Imaging User's Guide* manual for information on the image math programs fdffit or diffcalc .

Arguments

- fit_type 'ADC, 'T2', or 'DIFF'; default is 'ADC'
- fit_var Name of the parameter that holds the independent variable.
 Defaults to:
 'bvalue' for ADC fit
 'te' for T2 fit
 blank string for DIFF fit
 optional string argument that mimics aipSave.
 The macro imagement looks at aipSave if no dirflag
 argument is given.

Examples

imagemath('ADC', 'bvalue', 'saved')
imagemath('T2', 'te')
imagemath('DIFF')
imagemath('DIFF', '', 'saved')

imageprint Plot non interactive gray scale image (M)

Description	Sends to the plotter a dcon color intensity map with linear instead of		
	logarithmic increments and with grayscale instead of colors.		
See also	NMR Spectroscopy User Guide		
Related	dcon Display noninteractive color intensity map (C)		

imconi Display 2D data in interactive grayscale mode (M)

Description Calls the dconi program with the arguments required for grayscale image display: dconi('dcon','gray','linear').

import1Dspec Import ASCII Spectrum into VnmrJ / VNMR (M)

Syntax	<pre>Syntax import1Dspec('ascii_file'<,'av'>)</pre>		
Applicability VnmrJ 3.1			
Description	"import1Dspec" imports a 1D ASCII spectrum into VnmrJ / VNMR. For the conversion, the ASCII file (Y Y or X,Y X,Y data,one Y value per line) is read in, and a UNIX utility with the same name ("bin/import1Dspec", a C program) is used to build the binary files "datdir/data" and "datdir/phasefile" in the current experiment. "vs" is set to 100.		
	Note that the imaginary part of the data set consists of zeroes only, i.e., the data cannot be phased - worse than that, setting "lp" to values other than 0 in "ph" mode would cause serious intensity distortions across the spectrum. To prevent such problems, "import1Dspec" offers two options:		
	• By default, the macro sets lp=0 rp=0 and sets the parameter protections bits such that these two parameters cannot be changed. Note that this may cause cause VnmrJ / VNMR to produce errors when the user (intentionally or inadvertently) attempts to change the phase within "ds".		
	• Alternatively, you can specify 'av' as second argument. In this case, the macro switches to "av" mode, and "dmg" is locked, such that the user cannot switch to "ph" mode. This has the disadvantage that negative parts of the spectrum are flipped to positive - but at least you will not have the usual problem with the line broadening (from the imaginary dispersion tails) that 'av' has with "normal" spectra. The (minor) advantage of this method is that "lp" and "rp" can be used to store the original "rp" and "lp" values.		

I.

If the current experiment contains nD or arrayed 1D data, "import1Dspec" first does a "setup('H1', 'CDC13')", then imports the spectrum.

Limitation: Only works for simple, non-arrayed 1D spectra. The values of "rp" and "lp" are meaningless, the only meaningful parameter that is set from the imported data set is "fn". The other parameters (notably any applicable acquisition parameter, such as "sw", "tn", "sfrq", "nt", "ct", "np", "at", as well as possibly processing parameters such as weighting etc.) need to be set AFTER THE IMPORTING, along the following scheme:

\$ct=ct "remember 'ct'" tn=.. sw=.. nt=.. at=.. ... setvalue('ct',\$ct)"restore 'ct'" 1b=.. sb=.. groupcopy('current','processed','acquisition') groupcopy('current','processed','processing')

If you have a series of similar 1D ASCII spectra to process (e.g., a series of c13 spectra acquired with the same standard parameters) it is very helpful first to prepare such parameters according to the scheme above, then, for the importing, FIRST to retrieve these parameters, then to call "import1Dspec".

Arguments "ascii_file": Path to a pure ASCII file with either Y data (such as from "writetrace") or X,Y data (such as from "writexy"). The optional second argument 'av' selects and locks the spectrum in absolute value mode (see above); default is 'ph' mode.

import1Dspec('spectrum.xy') Examples import1Dspec('spectrum.xy','av') import1Dspec('spectrum.txt') import1Dspec('spectrum.txt','av') import1Dspec('trace.1') import1Dspec('trace.1','av') import1Dspec('xytrace.1') import1Dspec('xytrace.1','av') Related writetrace

Create ascii file from phasefile (f1 or f2) trace (M) Create x,y ascii file from phasefile (f1 or f2) trace writexy (\mathbf{M}) import1Dspec Create phasefile and data from ASCII spectrum (U)

import1DspecCreate phasefile and data from ASCII spectrum (U)

Syntax import1Dspec <- fn ##> <- vs #. #> ascii file <phase file <data>> Applicability VnmrJ 3.1 Description "import1Dspec" imports a 1D ASCII spectrum into VnmrJ / VNMR.

Arguments	"ascii_file": Path "writetrace") o "source/import1D cc -O -o /vnmr/h or (for a local in cc -O -o ~/bin/in	to a pure ASCII file with either Y data (such as from r X,Y data (such as from "writexy"). Pspec.c" is a C program that can be compiled with pin/import1Dspec import1Dspec.c -m32 stallation) mport1Dspec import1Dspec.c -m32
	"phasefile": Optic be imported into output file uses t and ".xy" stripped	onal path to a binary "phasefile" that can afterwards VNMR / VnmrJ using "import1Dspec"; the default he same name as the ASCII file (with extensions ".txt" d off) but with ".phf" extension.
	" data ": Optional j importing "phase phasefile, but wi	path to a binary "data" file that is required when file"; the default data file uses the same name as the th ".dat" extension.
	"- fn ##": Options points (fn/2 in V file is somehow t "zerofill" (add does not contain MUST be numeric be rounded UP to or its next highe in the ASCII file, "-vs ##": Optiona	al, creates a phasefile with the specified number of /NMR!!!); should NOT be necessary, unless the ASCII truncated; by default, "import1Dspec" will flat baseline at the high-field end) if the ASCII file a power of 2 in points; the argument following "-fn" c; if the specified number is NOT a power of 2, it will b the next higher power of 2. If the specified number r power of 2 are smaller than the number of points , the spectrum is truncated at the high-field end.
	writing spectra ir out Y values in m the value of "vs" spectrum. Also h and positive. The	a, "ai" (absolute intensity) mode, "writetrace" writes am (spectrum multiplied by "vs"); specifying "-vs" with from VNMR permits recreating the original ("ai") ere, the argument following "-vs" MUST be numeric e default "vs" value (downscaling factor) is 1.0.
Examples	import1Dspec spectrum.txt import1Dspec spectrum.xy import1Dspec spectrum.txt phasefile import1Dspec spectrum.txt phasefile data import1Dspec spectrum.xy phasefile data import1Dspec -fn 64000 spectrum.xy datdir/phasefile datdir/data import1Dspec -vs 327.54 spectrum.xy datdir/phasefile datdir/data import1Dspec -vs 327.54 -fn 32000 spectrum.xy	
Related	import1Dspec writetrace writexy	Import ASCII spectrum into VnmrJ / VNMR (M) Create ascii file from phasefile (f1 or f2) trace (M) Create x,y ascii file from phasefile (f1 or f2) trace

in Lock and spin interlock (P)

Description Controls error handling based on lock level and spin speed, and specifies action based on lock level failure or spinner failure. The

(M)

I

action can be to generate an error and halt acquisition, or to generate a warning and continue acquisition.

- Values Can be set to one or two characters:
 - If set to two characters, the first character specifies the action for lock failure and the second character specifies the action for spinner failure.
 - If set to only one character, that character specifies the same action for either lock or spinner failure.

'n' stops any system checking so that acquisition continues regardless of the lock level or spin speed.

'w' makes the system check the lock level and the spin speed. A warning message is added to the log file if the lock level falls below a preset hardware level (about 20 on the lock meter) or if spin is set to a particular value and the spin speed goes out of regulation; however, acquisition is not stopped.

'y' makes the system check the lock level and spin speed. Acquisition is halted if the lock level falls below a preset hardware level (about 20 on the lock meter) or if spin is set to a particular value and the spin speed goes out of regulation.

See also NMR Spectroscopy User Guide

Related spin Sample spin rate (P)

inadgt Set up parameters for INADEQUATE pulse sequence (M)

Description	Sets up parameters for 2D INADEQUATE (Incredible Natural		
	Abundance Double-Quantum Transfer Experiment).		
See also	NMR Spectroscopy User Guide		
Related	foldcc Fold INADEQUATE data about 2-quantum axis (C)		

index2 Projection or 3D plane index selected (P)

Stores whether a projection or 3D plane index is selected. It shows the current status only and cannot be used to select a plane or projection. This parameter is also displayed in the Status window below "Index."		
0 indicates a projection is selected.		
1 to the half the Fourier number of the normal axis indicates a 3D plane is selected; the number is the index of the 3D plane.		
NMR Spectroscopy User Guide		
dplaneDisplay a 3D plane (M)dprojDisplay a 3D plane projection (M)nextplDisplay the next 3D plane (M)		

Т

prevpl	Display the previous 3D plane (M)
select	Select a spectrum or 2D plane without displaying it (C)

inept Set up parameters for INEPT pulse sequence (M)

Description	Sets up parar	Sets up parameters for the INEPT (Insensitive Nuclei Enhanced			by	
	Polarization T	'ransfer) experiment.				
See also	NMR Spectros	scopy User Guide				
Related	ppcal	Proton decoupler pu	lse calibratio	on (M)		

Description	Takes the current spin system (contained in spinsys) and derives	
	from it relevant parameters. This can be used to control which	
	parameters are iterated during a spin simulation iteration (e.g., for an	
	ABC spin system, iterate is set to 'A, JAB, JAC, B, JBC, C').	
See also	NMR Spectroscopy User Guide	
Related	iterate Parameters to be iterated (P)	

input Receive input from keyboard (C)

Syntax	<pre>input<(<prompt><,delimiter>)>:var1,var2,</prompt></pre>		
Description	Receives fields of characters from the keyboard and stores them into one or more variables.		
Arguments	prompt is a string displayed on the command line.		
	delimiter is a character separating input fields. The default is a comma.		
	<pre>var1,var2, are return values. input stores the values into as many of these arguments as given and ignores the rest of the input line.</pre>		
Examples	<pre>input:\$b input('Enter pulse width:'):pw input('x and y coordinates'):cr,cr1 input('Enter lastname:firstname',':'):\$last,\$first</pre>		
See also	User Programming		
Related	string Create a string variable (C)		

ins Integral normalization scale (P)

Description	Sets the integral value, independent of is and vs. Reported integral values are scaled by fn; that is, the reported integral of a given region is independent of fn. The insref parameter is also used to determine a reference integral value. The setint macro sets integral value.		
See also	NMR Spectroscopy User Guide		
Related	dlni	Display list of normalized integrals (M)	
	fn	Fourier number in directly detected dimension (P)	
	is	Integral scale (P)	
	insref	Fourier number scaled value of an integral (P)	
	mark	Determine intensity of spectrum at a point (C)	
	setint	Set value of an integral (M)	
	vs	Vertical scale (P)	

ins2 2D volume value (P)

Description	Adjusts the 2D volume value, independent of is and vs. The volume is scaled by Fourier numbers for the two dimensions.				
See also	NMR Spectroscopy User Guide				
Related	is	is Integral scale (P			
	ins2ref	Fourier number scaled volume of a peak (P)			
	112d	Automatic and interactive 2D peak peaking (C)			
	VS	Vertical scale (P)			

insref Fourier number scaled value of an integral (P)

- Description Set to the Fourier number scaled value of a selected integral. The reported integral values will be *(integral value)*ins/insref/fn.* If insref is "not used", the sum of all integrals will be ins. The "not used" mode is the equivalent of the normalized integral mode. If insref is zero or not defined, the reported integrals will be *(integral value)*ins/fn.*
 - See also NMR Spectroscopy User Guide

Related	fn	Fourier number in directly detected dimension (P)
	ins	Integral normalization scale (P)
	liamp	Amplitudes of integral reset points (P)
	setint	Set value of an integral (M)

Description Set to the Fourier number scaled volume of the selected peak. The reported volume is *volume*ins2/ins2ref/fn/fn1*. If ins2ref is "not used", sum of all volumes is ins2. The "not used" mode is equivalent to a normalized volume mode. If ins2ref is zero or not defined, the reported volume is *volume*ins2/fn/fn1*.
See also NMR Spectroscopy User Guide
Related fn Fourier number in directly detected dimension (P)

Related	tn	Fourier number in directly detected dimension (P)
	fn1	Fourier number in first indirectly detected dimension
		(P)
	ins2	2D volume value (P)
	112d	Automation and interactive 2D peak picking (C)

insert Insert sample (M)

Description Turns off the eject air, waits for the sample to slowly drop, and then
turns off the slow drop air. The macro i is identical in function to
insert.
See also NMR Spectroscopy User Guide
Related e Eject sample (M)
eject Eject sample (M)

ejectEject sample (M)iInsert sample (M)

inset Display an inset spectrum (C)

DescriptionDisplays the part of the spectrum between the two cursors as an inset.
Before entering inset, run the ds command and display two cursors.
The vertical position is shifted up about one-quarter of the height of
the whole display canvas. The old spectrum remains on the screen, but
the parameters shown at the bottom are relevant to the new display.
If present, the integral trace is duplicated. The scale is also duplicated
if it is present. After running inset, you can shift the displayed
spectrum, expand it, or even contract it with the left and right mouse
buttons.See alsoNMR Spectroscopy User Guide

Related ds Display a spectrum FID (C)

integ Find largest integral in a specified region (C)

Syntax integ<(highfield,lowfield)><:size,value>

т

Description	Finds the largest absolute-value integral in the specified region, or the total integral if no reset points are present between the specified limits.		
Arguments	highfield and lowfield are the limits of the region. The defau values are the parameters sp and $sp+wp$, respectively.		
	size is a return value with the size of the largest integral. The size depends on the value of the parameter is and can be positive or negative.		
	value is a re value depend	eturn argument with the value of the largest integral. This s on ins, insref, and fn, and is independent of is.	
Examples	integ:r1,r2 integ(500,1000):\$height integ(100+sp,300+sp):\$ht,\$val		
See also	User Programming		
Related	fn	Fourier number in directly detected dimension (P)	
	ins	Integral normalization scale (P)	
	insref	Fourier number scaled value of an integral (P)	
	is	Integral scale (P)	
	rp	Zero-order phase in directly detected dimension (P)	
	sp	Start of plot in directly detected dimension (P)	
	wp	Width of plot in directly detected dimension (P)	

integrate Automatically integrate 1D spectrum (M)

Description	A universal macro for selecting integral regions and adjusting the				
	integrals in size and offset. Only if regions are not already selected,				
	and if intmod is set to 'partial', will integrate call region to				
	select integral regions. For proton spectra, the selection is done				
	through the hregions macro; for ¹⁹ F and ³¹ P spectra (for wide				
spectral windows, multiplet spectra), region is called with					
	arguments, and for other nuclei (mostly decoupled, single-line spe				
	other optimized parameters are used with region, such that lir				
	consisting of a few data points only are recognized.				
See also	NMR Spectroscopy User Guide				
Related	hregions Select integral regions in proton spectrum (M)				
	internal display mode (D)				

intmod	Integral display mode (P)
isadj	Adjust integral scale (M)
region	Automatically select integral regions (C)

int_flg

Syntax int_flg Applicability VnmrJ 3.1 Description int_flg determines whether dosy uses integrals or peak heights for DOSY fitting. int_flg='y' requires that valid integral resets be defined. Arguments in_flg='y' invokes fitting of peak integrals in_flg='n' invokes fitting of peak heights See also dosy

intmod Integral display mode (P)

Description	Controls display and plotting of the spectral integral.		
Values	'off' indicates that no integrals are displayed or plotted.		
	'full' indicates that all integral regions are displayed or plotted.		
	'partial' indicates that every other integral region is plotted (typically used to display integrals of only peaks and not of the baseline region).		
See also	NMR Spectroscopy User Guide		
Related	plc	Plot carbon spectrum (M)	
	plh	Plot proton spectrum (M)	
	plp	Plot phosphorus spectrum (M)	

intvast Produce a text file of integral regions (M)

Applicability	Systems with VAST accessory.
Syntax	intvast (last)
Description	intvast produces a text file, integ.out in the current experiment, containing the integrals of the partial regions of each spectra from wells 0 to last.
Arguments	last is the number last sample well. The default is 96.
See also	NMR Spectroscopy User Guide
Related	pintvast Plot the integrals (M)

intvast Produce a text file containing the integral of the partial regions

Applicability	VnmrJ 3.1
Description	The intvast macro produce a text file containing the integral of the partial regions. The integral regions of the spectra need to be preset. The resulting file, called integ.out, is placed in the local experiment directory.
Examples	intvast(<number data="" of="" wells="">)</number>

I.

```
See also dsvast
dsvast2d
plvast
plvast2d
intvast
pintvast
plateglue
vastglue
vastget
```

io Integral offset (P)

DescriptionOffset of the integral with respect to the spectrum.Values0 to 200, in mm.See alsoNMR Spectroscopy User Guide

is Integral scale (P)

Description	Multiplier that adjusts height of the displayed integral trace. Note that the ins parameter controls integral value, and that is has no effect on integral value.		
Values	1 to 1e9		
See also	NMR Spectroscopy User Guide		
Related	ins	Integral normalization scale (P)	
	ins2	2D volume value (P)	
	insref	Fourier number scaled value of an integral (P)	
	integ	Find largest integral in a specified region (C)	

isadj Automatic integral scale adjustment (M)

Syntax	isadj<(height<,neg_height>)>
Description	Adjusts the height of the integrals in a display to make the tallest integral fit the paper. Optionally, the height of the maximum integral can be specified by an argument. Negative integrals, if present, are given a limit of 10 mm if parameter io is less than 10; otherwise, they are set so they end 5 mm above the spectrum. Negative integrals can also be given a height. Whichever part of the integrals (positive or negative) runs into the given limit will be used to scale is.
Arguments	height is the size, in mm, of the maximum integral on display. The default is the height that makes the tallest integral fit the paper.

	neg_height is the desired height, in mm, of the largest negative integral. If io is less than 10, the default is 10; otherwise, the default height is 5 mm above the spectrum.		
Examples	isadj		
	isadj(100)		
	isadj(100,100)		
See also	NMR Spectroscopy User Guide		
Related	io Integral offset (P)		
	is Integral scale (P)		
	isadj2 Automatic integral scale adjustment by powers of two		
	(M)		

isadj2 Automatic integral scale adjustment by powers of two (M)

Syntax	isadj2<(height<,neg_height>)>:scaling_factor		
Description	Functionally the same as <i>isadj</i> except that <i>isadj2</i> adjusts the integral height by powers of two and returns the scaling factor to the calling macro.		
Arguments	height is the size, in mm, of the maximum integral on display.		
	neg_height is the desired height, in mm, of the maximum negative integral on display.		
	<pre>scaling_factor is a return value giving the ratio of the new integral size to the old value (new_is/old_is).</pre>		
Examples	isadj2 isadj2(100) isadj2(100,100) isadj2(50):r1		
See also	NMR Spectroscopy User Guide		
Related	is Integral scale (P) isadj Automatic integral scale adjustment (M)		

isCSIMode Determine if graphics area is split for CSI mode

Syntax	$\texttt{isCSIMode:}\$ - return 1 if graphics area is split for CSI display, 0 if not
Description	In CSI display mode, the graphics area is split to two. Left side is used to display images or/and spectra of CSI or arrayed 1D data in grid layout; right side is used to display and interact with selected trace(s). Spectra in the grid will be updated when selected trace is manipulated.
	Vertical and horizontal are special cases of grid layout.
	The graphic area can be split explicitly with command jFunc(88,1) aipSetState(11), and merged by jFunc(88,0) aipSetState(1).

L

isiin System global parameter for ISI interlock

Description The systemglobal parameter isiin controls the safety interlock for the high power pulse field gradient. This is accessed in the config window under the Z Axis Gradient as the Performa D selection. The isiin parameter is set to 'y' when the Performa D choice is selected. For all other choices, the isiin parameter is set to n.

isreal Utility macro to determine a parameter type (M)

Syntax isreal(paramname<, tree>)

Description Returns 1 if and only if paramname is a real type. It returns 0 if paramname is a string type. If there is an error, the error is reported and the macro also returns 0. The value of tree is 'current', 'global', 'processed' or 'systemglobal' and the default is 'current'.

> There is some unfortunate ambiguity and vagueness in regard to vnmr parameters and their types. The meaning of real and string vary slightly depending upon context. There are seven types altogether. The macro gettype returns a unique integer value when operating on the parameter. Of the seven types, two can be broadly categorized as string, and five can be broadly categorized as real. Since one of the string category types is 'string' and one of the real category types is 'real', this is where the ambiguity arises. The return values for gettype are:

category	type	gettype returns
string	'string'	2
U	'flag'	4
real	'real'	1
	'delay'	3
	'frequency'	5
	'pulse'	6
	'integer'	7

The isreal function returns 0 for the string category and 1 for the real category. This function is consistent with the typeof() operator. The typeof() operator is primarily intended to ascertain the type of the input argument to a macro, so using it for other purposes is not recommended. Also, it does not take a tree argument. Note that typeof() returns 0 for reals and 1 for strings, the opposite of this macro, but it should be clear from the name what is intended. A sister macro isstring returns the same value as typeof().

Related isstring Utility macro to determine a parameter type (M) typeof Return identifier for argument type (O)

isstring Utility macro to determine a parameter type (M)

Syntax isstring(paramname<, tree>)

Description Returns 1 if and only if paramname is a string type. It returns 0 if paramname is a real type. If there is an error, the error is reported and the macro also returns 0. The value of tree is 'current', 'global', 'processed' or 'systemglobal' and the default is 'current'.

> There is some unfortunate ambiguity and vagueness in regard to vnmr parameters and their types. The meaning of real and string vary slightly depending upon context. There are seven types altogether. The macro gettype returns a unique integer value when operating on the parameter. Of the seven types, two can be broadly categorized as string, and five can be broadly categorized as real. Since one of the string category types is 'string' and one of the real category types is 'real', this is where the ambiguity arises. The return values for gettype are:

category	type	gettype returns
string	'string'	2
-	'flag'	4
real	'real'	1
	'delay'	3
	'frequency'	5
	'pulse'	6
	'integer'	7

The function isstring returns 0 for the real category and 1 for the string category. This function is consistent with the typeof() operator. The typeof() operator is primarily intended to ascertain the type of the input argument to a macro, so using it for other purposes is not recommended. Also, it does not take a tree argument. Note that typeof() returns 0 for reals and 1 for strings, the opposite of this macro, but it should be clear from the name what is intended. A sister macro isstring returns the same value as typeof().

Related	isreal	Utility macro to determine a parameter type (M)
	typeof	Return identifier for argument type (O)

isvnmrj Identifies the interface is use, either Vnmr or VnmrJ

Syntax	isvnmrj:\$val
Applicability	VnmrJ 3.1
Description	The isvnmrj command identifies which interface is in use, either vnmr or vnmrj. This command would typically only be used in macros.
Arguments	The command returns a 1 if the interface is vnmrj, otherwise it returns a 0.
Examples	isvnmrj:\$ans

iterate Parameters to be iterated (P)

Description Contains parameters to be iterated during iterative spin simulations. If the Set Params button is used in setting up spin simulation parameters, iterate is initialized to a string containing all parameters appropriate to the current spin system.

- Values List of parameters, separated by commas (e.g., iterate='A,B,JAB').
- See also NMR Spectroscopy User Guide

Related initialize_iterate Set iterate string to contain relevant parameters (M)



Agilent VnmrJ 4 Command and Parameter Reference Guide

jaddsub	Join the add/subtract experiment
jcurwin	Work space numbers of all viewports (P)
jdesign	Start Plot Designer Program (M)
jexp	Join existing experiment (C)
jexp1-jexp9999	Join existing experiment and display new parameters (M)
jexpn	Join experiment n, where n is a number between 1 and 9
jnewexp	Experiment-Specific Runtime Macro
jplot	Plot from Plot Designer program (C)
jplotscale	Scale plot parameters (M)
jplotunscale	Restore current experiment parameters (M)
jprint	Prints the selected images to a printer or file (M)
jpublish	Macro to archive and/or copy to system a local protocol (M)
jumpret	Set up parameters for JUMPRET pulse sequence (M)
jviewport	Work space numbers of the current viewports (P)
jviewportlabel	Work space labels for all viewport buttons (P)
jviewports	Viewport layout (P)
jwin	Activate and record activity in current window (M)

jaddsub

Join the add/subtract experiment

Applicability VnmrJ 3.1

Description

jaddsub joins the add-subtract experiment, as defined by the global addsubexp parameter. jaddsub will create this parameter if it does not exist, and set it to a default value of 5. jaddsub with an argument,



as in jaddsub('silent') will not clear the graphics, text window, or menu system. It does not matter what the argument is. The last displayed or selected FID is added to ("add") or subtracted from ("sub") the current contents of the add/subtract experiment. An optional argument allows the FID to be first multiplied by a 'multiplier'. The FID data are divided by the number of time averages of the data, reflected in the parameter ct. To get unscaled data, use a multiplier of ct. The parameters lsfid and phfid may be used to shift or phase rotate the selected FID before it is combined with the data in the add/subtract experiment. A multi-fid add/subtract experiment may be created with the add or sub command. The optional argument 'new' will create a new FID element in the add/subtract experiment. For example, the commands clradd select(1) add from some experiment will create the add/subtract experiment with a single FID in it. If the next commands typed are select(2) add, then a single FID which is the sum of the original FIDs one and two will be made in the add/subtract experiment. If, on the other hand, the commands select(2)add ('new') were typed, then the add/subtract experiment will contain an array of two FIDs corresponding to the original FIDs one and two, espectively. One detail is that the arraydim parameter may need to be updated after constructing a multi-fid add/subtract experiment. The recipe for doing this is to join the add/subtract experiment (jaddsub) and enter setvalue('arraydim', <num>, 'processed') where <num> is replaced by the number of FIDs in that experiment. For example, if twelve FIDs were put into the add/subtract experiment, one would enter setvalue('arraydim', 12, 'processed') Individual FIDs in a multi-fid add/subtract experiment may subsequently be added to and subtracted from. The add and sub command without a 'trace' argument will add or subtract from the first FID in the add/subtract experiment. Adding the 'trace' argument followed by a required index number will select another FID to be the target of the add/subtract. For example, select(4) add('trace',6) will take the fourth FID from the current experiment and add it to the sixth FID in the add/subtract experiment. When using the 'trace' argument, that FID must already exist in the add/subtract experiment by using an appropriate number of add('new') or sub('new') commands. silent

Arguments siler new

jcurwin Work space numbers of all viewports (P)

Description	An arrayed global parameter, set to the work space numbers used by
	all viewports.
See also	NMR Spectroscopy User Guide, VnmrJ Walkup
Related	curwin Current window (P)

J

jviewport	Work space numbers of the current viewports (P)
jviewportlabel	Work space labels for all viewport buttons (P)

jdesign Start Plot Designer Program (M)

Syntax	jdesign
e e	5 5

Description	Opens the Plot Designer program, which provides mechanisms for		
	positioning spectra, parameters, axes, and other plot output on a page		
	Text annotation and drawing features are available.		
See also	NMR Spectroscopy User Guide		
Related	jplot	Plot from Plot Designer program (C)	

jexp Join existing experiment (C)

Syntax	<pre>(1) jexp(exp_number)</pre>		
	(2) jexp:\$current_exp_number,\$current_exp_name		
Description	Joins an existing experiment (syntax 1) or returns the current experiment number and experiment name (syntax 2). After entering this command, until another "join experiment" command or macro is entered, all actions (including changes of parameters, acquisition of data, and display of data) apply to the parameters and data of the experiment joined.		
	The jexp command does not refresh the display or display new experiment parameters. Use one of the macros jexp1, jexp2, etc. to join an experiment and have the screen refreshed and new parameters displayed.		
Arguments	ts exp_number is a number from 1 to 9999 for existing experiment be joined.		
	\$current_exp number.	_number is a return value with the current experiment	
	\$current_ex <u>r</u> name.	name is a return value with the current experiment	
Examples	jexp(3) jexp:\$expp jexp:r1,n1		
See also	NMR Spectroscopy User Guide; VnmrJ Walkup		
Related	cexp	Create an experiment (M)	
	delexp	Delete an experiment (M)	
	jexp1-jexp9	Join existing experiment and display new parameters (M)	
	unlock	Remove inactive lock and join experiment (C)	

jexp1-jexp9999Join existing experiment and display new parameters (M)

jexp1, jexp2, jexp3,,jexp9999		
Joins an existing experiment, refreshes the screen, and displays the main menu and the new experiment parameters. After entering this macro, until another "join experiment" command or macro is entered, all actions (including changes of parameters, acquisition of data, and display of data) apply to the parameters and data of the experiment joined.		
To join an experiment without refreshing the screen and displaying new parameters, use the $jexp$ command.		
jexp8 jexp354		
NMR Spectroscopy User Guide		
cexp delexp jexp unlock	Create an experiment (M) Delete an experiment (M) Join existing experiment (C) Remove inactive lock and join experiment (C)	
	jexp1, jexp Joins an exis main menu a macro, until all actions (i display of da joined. To join an ex new paramet jexp8 jexp354 <i>NMR Spectro</i> cexp delexp jexp unlock	

jexpn Join experiment n, where n is a number between 1 and 9

Syntax	jexpn
Applicability	VnmrJ 3.1
Description	Join experiment n, where n is a number from 1 to 9 describing an existing experiment. After this command, all actions including changes of parameters, acquisition of data, display of data, etc. will apply to the parameter and data of experiment n, until the next "jexp" command is executed. Without an argument, jexp:\$num returns the current experiment number in the variable \$num and jexp:\$num, \$name returns both the experiment number and experiment name to the variables \$num and \$name.

jnewexp Creates and Joins a New Experiment

Description Creates and joins a new experiment. Syntax jnewexp Plot from Plot Designer program (C)

Syntax	jplot<(<'-setup'><,template)>		
Description	Starts plotting from the Plot Designer program to the current plotter.		
Arguments	'-setup' is a keyword to start jdesign, the Plot Designer program, to allow interactive design and plotting.		
	template is the name of a file that will be used to make a plot of the current experiment. The default is a saved file chosen by the user.		
Examples	jplot jplot('t1')		
See also	NMR Spectroscopy User Guide		
Related	jdesign	Start Plot Designer program (M)	
	jplotscale	Scale plot parameters (M)	
	jplotunscale	Restore current experiment parameters (M)	

jplotscale Scale plot parameters (M)

jplot

Applicability	Plot Designer program		
Description	Scales parameters of plotting area and an imported plot. When a region is drawn in Plot Designer, jplotscale automatically changes the plotting area parameters wcmax and wc2max. The parameters io, is, vs, wc, and wc2 of a plot imported into a region are adjusted according to wcmax and wc2max.		
See also	NMR Spectroscopy User Guide		
Related	jplot	Plot from Plot Designer program (C)	
	jplotunscale	Restore current experiment parameters (M)	

jplotunscaleRestore current experiment parameters (M)

Applicability	Plot Designer program	
Description	Restores the current experiment parameters (io, is, vs, wc, and wc2) to a plot within a region that was created in Plot Designer. For example, entering jplotunscale jexp2 jplotscale restores the parameters of experiment 2 to a plot and then jplotscale applies the adjusted parameters to the plot.	
See also	NMR Spectroscopy User Guide	
Related	jplot jplotscale	Plot from Plot Designer program (C) Scale plot parameters (M)

jprint Prints the selected images to a printer or file (M)

Description The jprint macro takes the value of the parameters printregion, printsend, printfile, printlayout, printformat, printsize.

jpublish Macro to archive and/or copy to system a local protocol (M)

Syntax jpublish

Applicability VnmrJ 3.1

Description Assembles all elements of the experiment protocol in the current experiment into a archive protocolname_proto.tar.Z into a User_Protocols directory of the local vnmrsys directory. If the user is the VnmrJ admin, the protocol is installed into the VnmrJ system for all users. Note that the file protocolListWalkup.xml in /vnmr/adm/users would need to be edited to add the new protocol to the experiment chooser in a walkup-style account. The user is prompted for all needed information and the addition of one additional setup support macro is allowed. By default, jpublish will assign the local protocol name and seqfil as being associated. This is almost always the case.

jumpret Set up parameters for JUMPRET pulse sequence (M)

DescriptionSets up parameters for a jump-and-return water suppression sequence.See alsoNMR Spectroscopy User Guide

jviewport Work space numbers of the current viewports (P)

Description	A global parameter, set to the work space number that the current viewport is joined to. The parameter is set when the viewport starts. Each viewport may be joined to a different work space.	
See also	NMR Spectroscopy L	lser Guide, VnmrJ Walkup
Related	curwin	Current window (P)
	jcurwin	Work space numbers of all viewports (P)
	jviewports	Viewport layout (P)
	jviewportlabel	Work space labels for all viewport buttons (P)
jviewportlabel Work space labels for all viewport buttons (P)

Description	An arrayed global parameter, set to the labels on the toolbar buttons		
	used to switch viewports. It is used by the viewport editor under ${\bf Edit}$		
	-> Viewports.		
See also	NMR Spectroscopy User Guide, VnmrJ Walkup		
Related	jviewport Work space numbers of the current viewports (P)		

jviewports Viewport layout (P) vpaction Set initial state for multiple viewports (M)

jviewports Viewport layout (P)

Description An arrayed global parameter, used to keep track of the viewport layout. It is used by the viewport editor under **Edit -> Viewports** to change the viewport layout.

Related	jcurwin	Work space numbers of all viewports (P)
	jviewport	Work space numbers of the current viewports (P)
	jviewportlabel	Work space labels for all viewport buttons (P)
	vpaction	Set initial state for multiple viewports (M)
	vpset3def	Set the viewport state to three default viewports
		(M)
	vpsetup	Set new viewports (M)

jwin Activate and record activity in current window (M)

Syntax	jwin(pane_number)	
Description	Activates and records the activity in a specific window pane, created by setgrid, in the VnmrJ graphics window. jwin is executed when you double-click the left mouse button in a multiple-paned graphics window.	
Arguments	pane_number is the number of the pane to join.	
Examples	jwin(2)	
See also	NMR Spectroscopy User Guide	
Related	curwin	Current window (P)
	fontselect	Open FontSelect window (C)
	mapwin	List of experiment numbers (P)
	setgrid	Activate selected window (M)
	setwin	Activate selected window (C)



Agilent VnmrJ 4 Command and Parameter Reference Guide

killft3d	Terminate any ft3d process started in an experiment (M,U)
killplot	Stop plot jobs and remove from plot queue (M)
killprint	Stop print jobs and remove from print queue (M)
kind	Kinetics analysis, decreasing intensity (M)
kinds	Kinetics analysis, decreasing intensity, short form (M)
kini	Kinetics analysis, increasing intensity (M)
kinis	Kinetics analysis, increasing intensity, short form (M)

killft3d Terminate any ft3d process started in an experiment (M,U)

Syntax	killft3d(exp_number)	
Description	Terminates any ft3d program that has been started in the specified VnmrJ experiment. killft3d can be executed from any experiment For each ft3d process terminated, the relevant 3D data subdirector is also deleted. Remote ft3d processes, denoted by the call name ftr3d in the process table (displayed by the UNIX command ps -azx), are not directly terminated by killft3d but die of their own accord due to the deletion of the 3D data subdirectory.	
	The killft3d command can also be run as a shellscript from UNIX Its function is analogous to the associated VnmrJ macro.	
Arguments	exp_number is a number from 1 to 9 that identifies the experiment that started the $ft3d$ program.	
Examples	killft3d(4)	
See also	NMR Spectroscopy User Guide	
Related	ft3d Perform a 3D Fourier transform (M,U)	



killplot Stop plot jobs and remove from plot queue (M)

Description Kills all current plot jobs in the plot queue for the active plotter in VnmrJ, then removes the jobs from the plot queue. Unless the user executing killplot is root, only that user's plot jobs are deleted from the plot queue. To kill a plot that is in progress (i.e., a plot in which you have not entered page), use the page('clear') command. The plotter may have to be reinitialized after killplot is executed. To reinitialize the plotter, turn it off and then back on after a few seconds. Hewlett-Packard (HP) pen plotters appear to be more susceptible to this problem than the other HP output devices supported by VnmrJ. If one port is configured to be both a printer and a plotter, killplot can cause both plot and print jobs to that port to be deleted. For example, if printer='LaserJet_300', plotter='LaserJet_300R', and a plot command pl pscale page is followed by a print command ptext (vnmruser+'/psglib/noesy.c'), entering killplot deletes both jobs. See also NMR Spectroscopy User Guide Related killprint Stop print jobs and remove from print queue (M)

		~
page	Move plotter forward one or more pages (C)	
pl	Plot spectra (C)	
pscale	Plot scale below spectrum or FID (C)	
ptext	Print out a text file (M)	
showplotq	Display plot jobs in plot queue (M)	

killprint Stop print jobs and remove from print queue (M)

Description Kills all current print jobs in the print queue for the active printer in VnmrJ, then removes the jobs from the print queue. Unless the user executing killprint is root, only that user's print job is deleted from the print queue. It is slightly possible that the printer may have to be reinitialized after the execution of this macro. To reinitalize the printer, turn it off, wait a few seconds, and then turn it back on. If one port is configured to be both a printer and a plotter, killprint can cause both print and plot jobs to that port to be deleted. For example, if printer='LaserJet_300', plotter='LaserJet_300R', and a plot command pl pscale page is followed by a print command ptext(vnmruser+'/psglib/noesy.c'), entering killprint deletes both jobs. See also NMR Spectroscopy User Guide Related killplot Stop plot jobs and remove from plot queue (M)

ptextPrint out a text file (M)showprintqDisplay print jobs in print queue (M)

kind Kinetics analysis, decreasing intensity (M)

Description If the signal decreases exponentially toward a limit, the output is matched by I = A1 * EXP(-T/TAU) + A3. This macro supplies the necessary keywords to the analyze command, which uses the output of fp (i.e., the file fp.out) as input. The results can be displayed with expl. See also NMR Spectroscopy User Guide

Related	analyze	Generalized curve fitting (C)
	expl	Display exponential/polynomial curves (C)
	fp	Find peak heights (C)
	kinds	Kinetic analysis, decreasing intensity, short form
		(M)
	kini	Kinetics analysis, increasing intensity (M)
	kinis	Kinetic analysis, increasing intensity, short form
		(M)

kinds Kinetics analysis, decreasing intensity, short form (M)

Description	Produces a summary of the results from kind.		
See also	NMR Spectroscopy User Guide		
Related	kind Kinetics analysis, decreasing intensity (M)		

kini Kinetics analysis, increasing intensity (M)

Description	If the signal matched by	increases exponentially toward a limit, the output is
	I = -A1 * EX	P(-T/TAU) + A3 - A1. This macro supplies the necessary
	keywords to the file fp.c	the analyze command, which uses the output of fp (i.e., but) as input. The results can be displayed with expl.
See also	NMR Spectroscopy User Guide	
Related	kind	Kinetics analysis, decreasing intensity (M)
	kinis	Kinetic analysis, increasing intensity, short form
		(M)

kinis Kinetics analysis, increasing intensity, short form (M)

Description	Produces a summary of the results from kini.		
See also	NMR Spectroscopy User Guide		
Related	kind	Kinetics analysis, decreasing intensity (M)	
	kini	Kinetics analysis, increasing intensity (M)	

K



Agilent VnmrJ 4 Command and Parameter Reference Guide

laser	SVS adiabatic localization
lastlk	Last lock solvent used (P)
lastmenu	Menu to display when Return button is selected (P)
latch	Frequency synthesizer latching (P)
lb	Line broadening in directly detected dimension (P)
1b1	Line broadening in 1st indirectly detected dimension (P)
1b2	Line broadening in 2nd indirectly detected dimension (P)
lc1d	Pulse sequence for LC-NMR (M)
lcdatast	An LC-NMR plotting and display macro (M)
lcpar2d	Create 2D LC-NMR acquisition parameters (M)
lcpeak	Peak number (P)
lcplot	Plot LC-NMR data (M)
lcpsgset	Set up parameters for various LC-NMR pulse sequences (M)
lcset2d	General setup for 2D LC-NMR experiments (M)
left	Set display limits to left half of screen (C)
legrelay	Independent control of magnet leg relay (P)
length	Determine length of a string (C)
1f	List files in directory (C)
lgcp	X Lee-Goldburg cross polarization (CP) between protons and X with a choice of SPINAL or TPPM decoupling
liamp	Amplitudes of integral reset points (P)
lifrq	Frequencies of integral reset points (P)



liMMap	Calculate csi map of integrals for a peak (C)
liqbear	Liquids Bearing Air Level (P)
listenoff	Disable receipt of messages from send2Vnmr (M)
listenon	Enable receipt of messages from send2Vnmr (M)
listparam	List parameters in simple format (UNIX)
lkof	Track changes in lock frequency (P)
112d	Automatic and interactive 2D peak picking (C)
112dbackup	Copy current II2d peak file to another file (M)
112dmode	Control display of peaks picked by II2d (P)
llamp	List of line amplitudes (P)
llfrq	List of line frequencies (P)
llMMap	Calculate csi map of peak height for a peak defined by cs (C)
ln	Find natural logarithm of a number (C)
load	Load status of displayed shims (P)
loadcolors	Load colors for graphics window and plotters (M)
loaduserprefs	Load Operator Preferences
loc	Location of sample in tray (P)
locaction	Locator action (M)
lock	Submit an Autolock experiment to acquisition (C)
lockacqtc	Lock loop time constant during acquisition (P)
lockfreq	Lock frequency (P)
lockgain	Lock gain (P)
lockphase	Lock phase (P)
lockpower	Lock power (P)
locktc	Lock time constant (P)
log	
logate	Transmitter local oscillator gate (P)
lookup	Look up words and lines from a text file (C)
locprotoexec	Execute a protocol from the locator (M)
lp	First-order phase in directly detected dimension (P)

lp1	First-order phase in 1st indirectly detected dimension (P)
1p2	First-order phase in 2nd indirectly detected dimension (P)
lpalg	LP algorithm in np dimension (P)
lpalg1	LP algorithm in ni dimension (P)
lpalg2	LP algorithm in ni2 dimension (P)
lpext	LP data extension in np dimension (P)
lpext1	LP data extension in ni dimension (P)
lpext2	LP data extension in ni2 dimension (P)
lpfilt	LP coefficients to calculate in np dimension (P)
lpfilt1	LP coefficients to calculate in ni dimension (P)
lpfilt2	LP coefficients to calculate in ni2 dimension (P)
lpnupts	LP number of data points in np dimension (P)
lpnupts1	LP number of data points in ni dimension (P)
lpnupts2	LP number of data points in ni2 dimension (P)
lpopt	LP algorithm data extension in np dimension (P)
lpopt1	LP algorithm data extension in ni dimension (P)
lpopt2	LP algorithm data extension in ni2 dimension (P)
lpprint	LP print output for np dimension (P)
lpprint1	LP print output for ni dimension (P)
lpprint2	LP print output for ni2 dimension (P)
lptrace	LP output spectrum in np dimension (P)
lptrace1	LP output spectrum in ni dimension (P)
lptrace2	LP output spectrum in ni2 dimension (P)
ls	List files in directory (C)
lsfid	Number of complex points to left-shift the np FID (P)
lsfid1	Number of complex points to left-shift ni interferogram (P)
lsfid2	Number of complex points to left-shift ni2 interferogram (P)
lsfrq	Frequency shift of the fn spectrum (P)
lsfrq1	Frequency shift of the fn1 spectrum (P)

lsfrq2	Frequency shift of the fn2 spectrum (P)
lvl	Zero-order baseline correction (P)
lvltlt	Control sensitivity of IvI and tlt adjustments (P)

Syntax Applicability VnmrJ 3.1 Description To set all frequencies: Click on the "Set All Freq (Hz) " Button Phase cycle: autoph='n', only phase cycles up to nt autoph='y' goes through nt*array pcflag='n' - turns off the phase cycle entirely Central (Base) Frequency=resto-restol (or H1offset-restol) Restol (Local offset) is a small offset 0 to ca. 20 Hz from the global frequency of the reference. If after clicking on "Set All Freq (Hz)", Local offset appears to be large, then H1offset is not calibrated correctly. For water suppression optimization: sglarray=1; sglpower=0; For RF pulse tpwr array: sglpower=1

lastlk Last lock solvent used (P)

Description Contains the name of the last lock solvent. Intended for use with the optional sample changer, this parameter is a user global variable (stored in the user's global file) and is not accessible to multiple users simultaneously. On a multiuser automation run, you should preferably access the last lock solvent from the file /vnmr/acqqueue/lastlk.

- Values String containing the name of the solvent.
- See also NMR Spectroscopy User Guide

Related solvent Lock solvent (P)

lastmenu Menu to display when Return button is selected (P)

Description Contains the name of the menu to display when the Return button is clicked on certain menus. For example, if the Phase F2 button in the 2D Processing menu (controlled by the file process_2D) is clicked, lastmenu is set to 'process_2D', the ft and aph commands are executed, the ds window is opened, and the Interactive 1D Spectrum Display menu (ds_1 file) is displayed. Appearing in this menu is a Return button. Because lastmenu is still set to 'process_2D',

	clicking on th lastmenu is	he Return button redisplays the 2D Processing menu. stored in the \$vnmrsys/global file.
Values	String containing the name of a menu (e.g., 'process_2D').	
See also	User Program	nming
Related	menu	Change status of menu system (C) Select a menu without immediate activation (C)
	newmenu	Select a menu without mimeulate activation (0)

latch Frequency synthesizer latching (P)

Description	Configuration parameter for whether the PTS frequency synthesizer has latching capabilities (all digits of the frequency value are sent to the synthesizer at once). The value for each channel is by the Latching label in the Spectrometer Configuration window.
Values	<pre>'n' indicates the synthesizers do not have latching capabilities (Not Present choice from the Spectrometer Configuration window). 'y' indicates the synthesizers have latching capabilities (Present choice from the Spectrometer Configuration window).</pre>
See also	VnmrJ Installation and Administration
Related	configDisplay current configuration and possibly change it (M)

1b Line broadening in directly detected dimension (P)

Description	Sets line broadening and exponential weighting along the directly detected dimension. This dimension is often referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, etc.	
Values	A positive value gives the desired line broadening, in Hz, which is the used to calculate a decaying exponential function of the form $\exp(-t^*\pi^*lb)$.	
	A negative value gives a resolution enhancement function (increasing exponential) of the form $\exp(-t*\pi*lb)$.	
	'n' turns off line broadening and exponential weighting.	
See also	NMR Spectroscopy User Guide	
Related	expFind exponential value of a number (C)1b1Line broadening in 1st indirectly detected dimension (P)1b2Line broadening in 2nd indirectly detected dimension (P)	

1b1 Line broadening in 1st indirectly detected dimension (P)

Description	Sets line broadening and exponential weighting along the first indirectly detected dimension. This dimension is often referred to as the f_1 dimension in multidimensional data sets. 1b1 works analogously to the parameter 1b. The "conventional" parameters (1b, gf, etc.) operate on the detected FIDs, while this "2D" parameter is used during processing of the interferograms.	
Values	s A positive value gives the desired line broadening, in Hz, which is the used to calculate a decaying exponential function of the form $\exp(-t^*\pi^*lb1)$. A typical value is between 0.0001 to 1000 Hz.	
	A negative value gives a resolution enhancement function (increasing exponential) of the form $\exp(-t*p*lb1)$.	
	'n' turns off line broadening and exponential weighting.	
See also	NMR Spectroscopy User Guide	
Related	exp Find exponential value of a number (C)	
	1b Line broadening in directly detected dimension (P)	
	1b2 Line broadening in 2nd indirectly detected dimension (P)	

1b2 Line broadening in 2nd indirectly detected dimension (P)

Description	Sets line broadening and exponential weighting along the second indirectly detected dimension. This dimension is often referred to as the f_2 dimension in multidimensional data sets. 1b2 works analogously to the parameter 1b. 1b2 can be set with wti on the 2D interferogram data.
Values	A positive value gives the desired line broadening, in Hz, which is then used to calculate a decaying exponential function of the form $\exp(-t^*\pi^*lb2)$.
	A negative value gives a resolution enhancement function (increasing exponential) of the form $\exp(-t^{\pi}\pi^{1}b^{2})$.
	'n' turns off line broadening and exponential weighting.
See also	NMR Spectroscopy User Guide
Related	EXP Find exponential value of a number (C)
	Line broadening in directly detected dimension (P)
	wti Interactive weighting (C)

1c1d Pulse sequence for LC-NMR (M)

ApplicabilitySystems with LC-NMR accessory.DescriptionCreates parameters to set up a pulse sequence that can be used to
start an LC-NMR run, including triggering the injection of a sample,

L

and can be used also to obtain multiple solvent-suppressed spectra using multi frequency Shifted Laminar Pulses (SLP) and gradients. The sequence is coded without a d2 variable, thus allowing ni to be used to obtain a series of spectra without resulting in any delay in the sequence being incremented.

The sequence requires a phase table, lc1d, to be found in the tablib directory. Phases of the selective pulses, the observe pulse, and the receiver and separately controlled by phase variables.

Note that the lcld sequence uses power scaling of shaped pulses, which is supported starting in VnmrJ 5.2. Because of this feature, this sequence *will not run* in earlier versions of VnmrJ.

1cdatast An LC-NMR plotting and display macro

Applicability	VnmrJ 3.1
Description	The engine for display and plotting of LC detector data. The default mode is a horizontal display of detector 1. Stop codes are marked if encountered in the LC data.
Arguments	The following arguments are recognized and any number can be entered in any order. plot sends output to plotter. side activates vertical display on the side of the NMR data. In this mode the LC data are positioned between wc and wcmax and scaled appropriately to fit. In the stopped -flow mode, the side option also places the stopped-flow NMR data at a position so that it is time-aligned with the relevant LC peak. det2 displays or plots the outputs of detectors one and two. full The detector data is displayed vertically at sc.
Examples	<pre>lcdatast(<arguments>)</arguments></pre>

1cpar2d Create 2D LC-NMR acquisition parameters (M)

Applicability	Systems with LC-NMR accessory.	
Description	Creates the acquisition parameters ni, sw1, and phase, which can be used to acquire a 2D LC-NMR data set. lcpar2d is functionally the same as addpar('2d').	
Related	addpar lcset2d	Add selected parameters to current experiment (M) General setup for 2D LC-NMR experiments (M)

lcpeak Peak number (P)

Applicability Systems with LC-NMR accessory.

LC-NMR

Description Contains the number of the peak being sensed or the loop being flushed.

lcplot Plot LC-NMR data (M)

Applicability	Systems with LC-NMR accessory.
Syntax	lcplot
Description	Plots LC-NMR data. This macro is executed with the Plot
	button on the Spare pane when LC-NMR is active.

lcpsgset Set up parameters for various LC-NMR pulse sequences (M)

Applicability	Systems with LC-NMR accessory.
Syntax	<pre>lcpsgset(file,parameter1,parameter2,,parameterN)</pre>
Description	Sets up parameters for various LC-NMR pulse sequences using information in a parlib file. Rather than returning the entire parameter file, lcpsgset returns the parameters listed. lcpsgset, in general, is never entered from the keyboard but is used as part of experiment setup macros.
Arguments	file is the file from the user or system parlib that provides information on setting up parameters listed. The parameters seqfil and pslabel are set to the supplied file name.
	parameter1, parameter1,, parameterN are 1 to 11 parameters to be returned from the parlib file.
Examples	lcpsgset('lccosy','ds','ap','ss','d1','axis','phase')

lcset2d General setup for 2D LC-NMR experiments (M)

Applicability	Systems with LC-NMR accessory.
Syntax	lcset2d(experiment<,F2_dig_res<,F1_dig_res>>)
Description	Runs the macro lcpar2d to create new parameters needed for 2D LC-NMR experiments, then selects starting values for a number of parameters. The lcset2d macro is "internal" and not normally entered directly by the user.
Arguments	experiment is the name of a 2D LC-NMR experiment.
	<code>F2_dig_res</code> is the f_2 digital resolution desired, in Hz/pt.
	<code>F1_dig_res</code> is the f_1 digital resolution desired, in Hz/pt.
Examples	lcset2d('lcnoesy')

left Set display limits to left half of screen (C)

Description Sets the horizontal control parameters sc and wc to produce a display (and subsequent plot) in the left half of a screen (and page). For 2D data, space is left for the scales.

Related	center	Set display limits for center of screen (C)
	full	Set display limits for a full screen (C)
	fullt	Set display limits for full screen with room for traces
		(C)
	right	Set display limits for right half of screen (C)

legrelay Independent control of magnet leg relay (P)

Description	Gives override capability over the magnetic leg high and low (broad band rf signal routing. This parameter does not normally exist but ca be created by the user with the command create('legrelay','string').		
	The legrelay override is operational only on standard systems shipped starting in November 1990 and on certain special systems shipped before that date. A system includes the override capability if it uses N-type connectors instead by BNC connectors on the magnet leg.		
Values	'n' indicates normal logic is used to set the leg relay.		
	'h' indicates the leg relay is set to the high band.		
	'l' indicates the leg relay is set to the low (broad) band.		
	Any other value results in an error message and an abort of pulse sequence generation.		
See also	User Programming		
Related	create Create new parameter in a parameter tree (C)		

length Determine length of a string (C)

Syntax	length(string):\$string_length		
Description	Returns the length in characters of a specified string.		
Arguments	string is zero or more characters enclosed in single quotes.		
	string_length is the number of characters (a real number) in string.		
Examples	length('abc'):r1 length(solvent):\$len		
See also	User Programming		
Related	substr Select a substring from a string (C)		

L

1f List files in directory (C)

Syntax	lf<(directory)>		
Description	Lists the files in a directory, with output on the text output window. Directories are suffixed by "/", executable files by "*", and links by "@"		
Arguments	directory is the name of a directory. The default is the current working directory. lf is equivalent to the UNIX command ls $-F$ and uses the same options (e.g., -1 for a long listing such as $lf('-1 *.fid')$).		
Examples	lf lf('data')) lf('-l *.fid')		
See also	NMR Spectroscopy User Guide		
Related	dir List files in directory (C)		
	List files in directory (C)		

1gcpX Lee-Goldburg cross polarization (CP) between protons
and X with a choice of SPINAL or TPPM decoupling

Syntax	
Applicability	VnmrJ 3.1
Description	X Lee-Goldburg cross polarization (CP) between protons and X with a choice of SPINAL or TPPM decoupling. Used for selective CP with suppression of homonuclear dipolar interactions and for setup of Lee-Goldburg HETCOR.
	Setup:
	Load a calibrated data set and select the protocol Lgcp. For a new nucleus calibrate CP with Onepul and Tancpx and then select Lgcp.
	Select the desired decoupling method, TPPM or SPINAL. The manual file onepul describes calibration of decoupling.
	Before running Lee-Goldburg CP use Tancpx to calibrate aHhx with a known field strength. This can be done by calibrating CP with aH90 = aHhx and then determining pwH90. The proton field strength is ?B1H = $1/(4.0*pwH90)$. Match the Hartmann Hahn condition by varying aXhx as needed. It is helpful to array aXhx and note the positions of the intensity maxima.
	Select the protocol Lgcp. Set ofHX = ?B1H/sqrt(2) and continue to use aHhx from the previous step. Note that of HX = - ?B1H/sqrt(2) is incorrect relative to the phase cycles of pwH90 and pwHtilt.
	Recalibrate the proton excitation pulse pwH90 to the value usually used for proton excitation if desired.
	After recalibration of pwH90 set pwHtilt = pwH90*35.3/90.0. Set Shape = 'const' and Channel = 'from'. Match the Hartmann-Hahn condition by varying aXhx as needed. It is helpful to array aXhx and note the

positions of the intensity maxima. You will note that the pattern is shifted to higher amplitude because of the offset. Choose one of the maxima for CP.

Note: the CP module only allows an offset on the channel selected in Channel. Since this must be 'from' or protons, a ramped amplitude cannot be applied to X. It is possible to use a ramp on protons, though that is not a usual practice for Lee-Goldburg CP.

Array the contact time tXhx for the Lee-Goldburg CP. You will find that non-protonated X nuclei cross polarize weakly for all contact times and that protonated nuclei polarize to their fullest extent with a short contact time.

Lee-Goldburg CP cross polarization can only occur through an X-H dipolar mechanism and X-H-H three spin CP is suppressed. A long contact time however can increase the importance of long range X-H interactions. This distinction is important when using Lee Goldburg CP for Lee-Goldburg HETCOR. Spin diffusion is suppressed and long distance X-H correlations can be recognized.

Note that signal to noise of Lgcp is about 50% of that of Tancpx.

Parameter Groups:

90H: Module: no

Sequence: tancpx.c

Description: Provides a 90-degree pulse on dec that can be used as a preparation pulse. Also used to store the dec calibration.

Parameters: Channels Page

aH90 - the amplitude of the pulse.

pwH90 - the pulse length.

cpHX: Module: yes

Sequence: tancpx.c

Description: Implements constant, linear or tangent-ramped cross polarization from dec to obs.

Parameters: Sequence Page

shHX - 'c', constant, 'l', linear and 't', tangent shape on the channel designated with chHX.

chHX - the variable-amplitude channel, 'fr' from or 'to', to.

aHhx - median amplitude of the dec channel.

aXhx - median amplitude of the obs channel.

bHX - +/- tangent curvature (>0 to 1.0e5).

dHX - amplitude width of the ramp. A negative width sets a ramp that starts high and decreases in amplitude.

tHX - contact time

ofHX - overall offset on the variable channel

 ${\rm frHX}$ = 'dec' - channel with the initial polarization (must be set - not shown)

L

to HX = 'obs' - $\$ channel with the final polarization (must be set - not shown).

Implementation: CP hx = getcp("HX",0.0,0.0,0,1);

Underscore functions: _cp_(hx,phHhx,phXhx);

Hseq: Module: yes

Sequence: tancpx.c

Description: Chooses SPINAL or TPPM decoupling on the dec channel during acquisition.

Parameters: Sequence Page - the Hspinal and Htppm groups overlap.

Hseq - chooses the decoupling sequence, TPPM or SPINAL.

Implementation: DSEQ dec = getdseq("H"); The router implements
getspinal() or gettppm().

Underscore functions: _dseqon(dec); runs _tppm(); or _spinal(); _dseqoff(dec); runs decprgoff();

Hspinal: Module: yes

Sequence: tancpx.c

Description: Implements SPINAL decoupling on the dec channel during acquisition.

Parameters: Sequence Page

aHspinal - amplitude of the dec channel.

pwHspinal - approximate 180-degree flip angle on resonance.

phHspinal - +/- small angle phase. SPINAL64 is implemented with phases = +/- 1.0, +/- 1.5 and +/-2.0 times phHspinal.

chHspinal = 'dec' must be set (not shown).

Implementation: SPINAL dec = getspinal("H"); or DSEQ dec =
getdseq("H");

Underscore functions: _spinal(dec); and decprgoff(); or _dseqon(dec); and _dseqoff(dec);

Htppm: Module: yes

Sequence: tancpx.c

Description: Implements TPPM decoupling on the dec channel during acquisition.

Parameters: Sequence Page

aHtppm - amplitude of the dec channel.

pwHtppm - approximate 180-degree flip angle on resonance.

phHtppm - +/- small angle phase. TPPM is implemented with phases = +/- 1.0 times phHtppm for alternating pulses.

chHtppm = 'dec' must be set (not shown).

Implementation: TPPM dec = gettppm("H"); or DSEQ dec = getdseq("H");

Underscore functions: _tppmon(dec); and decprgoff(); or _dseqon(dec); and _dseqoff(dec);

liamp Amplitudes of integral reset points (P)

Description Stores the integral amplitudes at the integral reset points for a list of integrals. To display the values of liamp, enter display('liamp'). Values of liamp can also be accessed in MAGICAL macros using, for example, liamp[\$i]. Values are stored as absolute numbers (summations of data point values) and, as such, are a function of the parameter fn. The values displayed by the dli, pir, and dpir programs are related to liamp values by the relationship:

Displayed or plotted integral =
liamp[i]*is/(fn/128)*ins)

See also NMR Spectroscopy User Guide

Related	display	Display parameters and their attributes (C)
	dli	Display list of integrals (C)
	dpir	Display integral amplitudes below spectrum (C)
	fn	Fourier number in directly detected dimension (P)
	lifrq	Frequencies of integral reset points (P)
	pir	Plot integral amplitudes below spectrum (C)

lifrq Frequencies of integral reset points (P)

Description	Stores the frequencies of integral reset points for a list of integrals. The frequencies are stored in Hz and are <i>not</i> adjusted by the reference parameters rfl and rfp .	
See also	NMR Spectroscopy User Guide	
Related	liamp	Amplitudes of integral reset points (P)
	rfl	Ref. peak position in directly detected dimension (P)
	rfp	Ref. peak frequency in directly detected dimension (P)

limmap Calculate csi map of integrals for a specified peak (C)

Syntax	<pre>liMMap(cs<,fullpath>) - calculate csi map of integrals for a peak specified by chemical shift cs.</pre>	
	<pre>liMMap(freq1, freq2<,fullpath>) - calculate csi map of integrals for a peak defined by freq1, freq2.</pre>	
Description	Map is saved in fdf format, in identical format as images.	
	Default map path is xxx.csi/maps/li_ <chem_shift_ppm>. User specified map path should be fullpath with map name, but without suffix .fdf.</chem_shift_ppm>	
	cs is chemical shift of the peak, which can be specified by cr (cursor).	

L

freq1, freq2 are absolute frequencies (as in lifrq).

Parameters llfrq and lifrq are used to determine peak position and integral region.

If peak cs is given, lifrq is searched to determine integral region for peak cs. If not found, the entire spectrum region will be integrated. This gives a full integral csi map (csi image).

If freq1, freq2 are given, llfrq is search to determine peak position. If not found, peak position will be 0.5*(freq1+freq2).

Multiple fdf maps will be created for multi-slice csi data, with default naming xxx.csi/maps/li_<chem_shift_ppm>_n.fdf, where n is slice index.

A single 3D fdf map will be created for 3D csi data.

ligbear Liquids Bearing Air Level (P)

Description This global parameter is the DAC value used when the liquids spinner bearing air is turned on. If the parameter does not exist the value defaults to 0xc000. To create the parameter: create('liqbear','integer','global') setlimit('liqbear',65535,0,1,'global') Values 0 - 65535

listenoff Disable receipt of messages from send2Vnmr (M)

Description Deletes the file \$vnmruser/.talk, thereby disallowing send2Vnmr to send commands to VnmrJ See also User Programming Related listenon Enable receipt of messages from send2Vnmr (M) send2vnmr Send a command to VnmrJ (U)

listenon Enable receipt of messages from send2Vnmr (M)

Description	Writes files with the VnmrJ port number that /vnmr/bin/s	end2Vnmr
	needs to talk to VnmrJ. The command then to send comm	ands to
	VnmrJ is	
	/vnmr/bin/send2Vnmr \$vnmruser/.talk command.	
See also	User Programming	
Related	listenoff Disable receipt of messages from send2Vnmr	(M)
	send2vnmr Send a command to VnmrJ (U)	

Syntax	listparam	filename <parametergroup></parametergroup>
Applicability	ty VnmrJ 3.1	
Description	Lists parame one line per comparison p output using used togener	ters from a VNMR paramater file in a simple format using parameter value. One application of listparam is for purposes, in which case one would typically sort the the 'sort' command, see below. listparam can also be ate JCAMP-DX style parameter output.
Arguments	filename is a	VNMR parameter file, like
	\$HOME/vnmr \$HOME/vnmr \$HOME/vnmr	sys/exp1/procpar sys/exp1/curpar sys/global
	/vnmr/conp	ar
	xyz.fid/pr	ocpar
	parametergro parameter ty The following	pup is an optional argument that permits specifying the pe. By default, only acquisition parameters are dumped. g options exist (only the first two characters are relevant):
	\bullet acquisition $\ -$	list acquisition parameters (default)
	• processing -	list processing parameters only
	• display - list	display parameters only
	• spsim - list s	spin simulation parameters only
	• sample - list	sample parameters only
	•all - list ALI	a parameters (output indicates group for each parameter)
	• JCAMP - list	acquisition parameters in JCAMP-DX format.
	• Inactive para	meters are suppressed, for FID saving
	• JS - list acqu format (for s	uisition, sample & processing parameters in JCAMP-DX aving with spectra)
	• JP - list acqu without Ggro parameters)	isition, sample & processing parameters, plus parameters oup assignment in JCAMP-DX format (for saving with
Examples	Using listp	aram on single files:
		listparam vnmrsys/exp1/procpar sort
		listparam vnmrsys/exp1/curpar all
		listparam xyz.fid/procpar JCAMP sort
	Using listp	aram to compare parameters:
		listparam xyz.fid/procpar sort > xyz.pars
		listparam abc.fid/procpar sort > abc.pars
		diff xyz.pars abc.pars
Related	diffparam	report differences between parameter sets (UNIX)
	vnmr2jcamp	create JCAMP parameters from VNMR parameters (UNIX)
	svfj	save FID in JCAMP-DX format

1kof Track changes in lock frequency (P)

- Description Tracks changes in the lock frequency resulting from changes in the solvent, and minor changes caused by the magnet drifting. The frequency units for lkof are in Hz, analogous to sfrq and tof, or dfrq and dof. lkof affects two components of the system: autolock on the console and acqi on the host computer. If lkof exists, it offsets the current value of the lockfreq parameter.
 - See also NMR Spectroscopy User Guide

Related lockfreq Lock frequency (P)

112d Automatic and interactive 2D peak picking (C)

Syntax (1) 112d<(options)><:\$num>

(2) 112d('info'<, #>):\$peak_number,\$f1,\$f2,\$amplitude, \$volume,\$label,\$comment,\$FWHH1,\$FWHH2,\$f1_min, \$f1_max,\$f2_min,\$f2_max

Description Automatically finds and integrates peaks that are above the threshold th in a 2D spectrum or a 2D plane of a 3D spectrum, and writes the peak location, volume, full-width at half-height (FWHH), volume, and the boundaries of the integrated region to a file in the 112d subdirectory of the current experiment directory. For 2D spectra, the file name is peaks.bin, and for 2D planes of 3D spectra, the file name is peaks_f#f#_#.bin, where f#f# gives the plane direction (e.g., f1f3) and the final # gives the number of the plane. For easy import and export of peak data, 112d also allows insertion and deletion of peaks interactively as well as reading and writing of text peak files.

Two-dimensional volumes are scaled in a manner analogous to 1D integrals, using the parameters ins2 and ins2ref. The ins2ref parameter is the Fourier number scaled value of a selected volume. The reported value of a peak volume is (*unscaled volume*) \times ins2/ins2ref/fn/fn1. The unscaled volume of a peak can be obtained from the command 112d('info',peak#). ins2ref can be set to the unscaled value divided by fn and fn1. The report volume for that peak is then the value of ins2.

- Arguments options (syntax 1) are any of the following (dconi is not necessarily active):
 - 'adjust' is a keyword to adjust the bounds of all peaks in the displayed area so that no boundaries overlap, and then to recalculate peak volumes.
 - 'draw' is a keyword to draw the peaks, boxes, numbers, and labels on the spectrum based on the value of the parameter 112dmode.
 - 'info', 'total' displays the total number of peaks in the current peak table. If a single return value is requested, printing is suppressed and the total number of peaks is returned.

- 'peak' is a keyword to find all peaks in the displayed area above a threshold th. If dconi is active and in the box mode, 112d finds peaks only in the area defined by the cursors. The 'peak' option is the default if no arguments are entered.
- 'pos' or 'neg' keywords can be used in addition to 'peak' or 'clear' to operate only on positive or negative peaks.
- 'read'<, file > reads in a binary peak file, where file is the name of the peak file. If a full path is not specified, the file is searched for first in the current working directory and then in the ll2d subdirectory of the current experiment directory.
- 'readtext'<, file> reads in a text peak file, where file is the name of the peak file. If a full path is not specified, the file is searched for first in the current working directory and then in the 112d subdirectory of the current experiment directory.
- 'reset' is a keyword to delete all peaks in the peak table.
- 'volume' is a keyword to find the bounds of each peak in the displayed area and integrate this area.
- 'writetext'<, file> writes a peak file to a text file, where file is the name of the text file written. If a full path is not specified, the file is written in the current working directory.

options (syntax 1) can also be any of the following (dconi must be active):

- 'clear' is a keyword to delete all peaks in the displayed region if in the dconi cursor mode, or to delete all peaks within the cursors if in the dconi box mode.
- 'combine' is a keyword to combine all peaks within the area defined by the cursors into a single peak (in dconi box mode only). The center of the new peak is at the average of all combined peaks' centers, and the bounds of this peak contains the maximum extents of the combined peaks' bounds. If all combined peaks have the same label, this label is assigned to the new peak.

CAUTION

All individual peaks to be combined are deleted prior to the creation of the new combination peak, and there is no automatic way to restore the original peaks. Therefore, it is recommended that you make a backup copy of the peak file prior to using this option.

- 'comment' is a keyword to prompt for an 80-character comment. The comment is assigned to the nearest peak in the dconi cursor mode or to all peaks within the cursors in the dconi box mode.
- 'comment', text executes the 'comment' option using the string entered for text instead of prompting for a comment.
- 'label' is a keyword to prompt for a 15-character label. The label is assigned to the nearest peak in dconi cursor mode or assigned to all peaks within the cursors in dconi box mode. To erase an existing label, enter a label consisting of one or more spaces.

- 'label', text executes the 'label' option using the string entered for text instead of prompting for a label.
- 'mark' is a keyword to insert a peak at the current cursor position if in the dconi cursor mode. If in the dconi box mode, 'mark' is a keyword to integrate the area within the cursors and assign that area to all peaks within the cursors that do not have their bounds already defined. If there are no peaks within the area defined by the cursors, using 'mark' finds the highest point within this area, marks that as a peak, integrates the area within the cursors, and assigns that area to the peak. The displayed values of the volume integrals are scaled by ins2 and ins2ref and the Fourier number of the 2D experiment.
- 'unmark' is a keyword to delete the nearest peak if in dconi cursor mode. If in the dconi box mode, 'unmark' deletes all peak bounds that are completely within the area defined by the cursors. Peaks are not deleted in the box mode.

options (syntax 1) also can be any of the following (dconi does not have to be active because 112d is executed on a peak number):

- 'combine', #1, #2, ... executes the 'combine' option on the list of peak numbers that follow the 'combine' keyword. If a single return value is requested, the peak number of the new combination peak is returned.
- 'comment', text, # executes the 'comment' option on peak # using the string entered for text instead of prompting for a comment.
- 'label',text, # executes the 'label' option on peak # using the string entered for text instead of prompting for a label.
- 'unmark', # deletes peak number #.

snum (syntax 1) is a return value set to the total number of peaks that have been picked unless the arguments 'combine', #1, #2, ... are used, in which case snum is the number of the newly created combination peak.

Syntax 2 arguments are the following:

- 'info'<, #> displays information in the text window about peak number #. If no peak number is included, dconi must be active and the default is the peak nearest to the cursor. If return values are requested, the display is suppressed.
- \$peak_number is a return value set to the number of the peak, either the second argument # or, if no value is given for #, the peak nearest to the cursor in dconi.
- $\,$ \$f1 and \$f2 are return values set to the peak frequencies in f_1 and f_2 of peak <code>\$peak_number</code>.
- \$amp is a return value set to the amplitude of peak \$peak_number.
- \$vol is a return value set to the unscaled volume of \$peak_number. peak. This value can be used to set the ins2ref parameter.
- \$label is a return value set to the label of peak \$peak_number.
- \$comment is a return value set to the comment about \$peak_number.

	•\$FWHH1 and sof \$peak_number 1	\$FWHH2 are return values set to full-width at half-height nber.
	• \$f1_min, \$f1 bounds of \$p	l_max,\$f2_min,\$f2_max are return values set to the eak_number.
Examples	112d	
	112d:\$npea	S
	ll2d('volur	ne')
	112d('read	,'peaklist.inp')
	ll2d('mark	
	112d('labe]	L','Peak 1')
	112d('info	,'total'):\$npeaks
	112d('comb:	lne',3,4,5,6):\$cpn
	112d('info	,3):\$num,\$f1,\$f2,\$amp,\$vol,\$label
See also	NMR Spectro	scopy User Guide
Related	dconi	Interactive 2D contour display (C)
	ins2	2D volume value (P)
	ins2ref	Fourier number scaled volume of a peak (P)
	112dbackup	Copy current 112d peak file to another file (M)
	112dmode	Control display of peaks picked by 112d (P)
	parll2d	Create parameters for 2D peak picking (M)
	pll2d	Plot results of 2D peak picking (C)
	th	Threshold (P)
	th2d	Threshold for integrating peaks in 2D spectra (P)
	xdiag	Threshold for excluding diagonal peaks when peak
		picking (P)

112dbackup Copy current II2d peak file to another file (M)

Syntax	ll2dbackup<(file)>		
Description	Backs up the current <u>ll2d</u> peak file by copying it to a file with a different file name. The default <u>ll2d</u> peak file is peaks.bin for 2D data.		
Arguments	file is the name to be given to the backup file. If a full path is not specified, the file is written to the current working directory. If no argument is provided, the system prompts for a file name. If no file name is specified at the prompt, the default <pre>ll2d</pre> peak file name with .bck appended is used.		
See also	NMR Spectroscopy User Guide		
Related	Automatic and interactive 2D peak picking (C)		

112dmode Control display of peaks picked by II2d (P)

Description Sets the display attributes of peaks picked by the 112d command

- Values A string variable composed of 4 characters, with each character taking the value 'y' (display the peak attribute) or 'n' (do not display the attribute). The first character determines if a "+" is drawn on the screen in dconi displays to mark peaks, the second character controls the drawing of the peak number, the third character controls drawing of the peak bounds box, and the last character controls drawing of the peak label.
- See also NMR Spectroscopy User Guide Related 112d Automatic and interactive 2D peak picking (C)

11amp List of line amplitudes (P)

Description	Stores a list	of line amplitudes above the threshold set by th.
See also	NMR Spectroscopy User Guide	
Related	d11	Display listed line frequencies and intensities (C
	llfrq	List of line frequencies (P)
	th	Threshold (P)

11frq List of line frequencies (P)

Description	Stores a list of line frequencies above the threshold set by th. Frequencies are stored in Hz and are <i>not</i> adjusted by reference parameters rfl and rfp.	
See also	NMR Spectroscopy User Guide	
Related	llamp	List of line amplitudes (P)
	rfl	Ref. peak position in directly detected dimension (P)
	rfp	Ref. peak frequency in directly detected dimension (P)
	th	Threshold (P)

11MMap Calculate csi map of peak height for a peak defined by cs (C)

Syntax	llMMap(cs<,fullpath<,nearestPeak>)		
Description	tion Peak is specified by chemical shift cs, which can be specified by (cursor).		
	Map is saved in fdf format, in identical format as images.		

Default map path is xxx.csi/maps/ll_<chem_shift_ppm>. User specified map path should be fullpath with map name, but without suffix .fdf.

A third argument can be given to specify whether to optimize cs to the nearest peak max. Default is yes.

Multiple fdf maps will be created for multisllce csi data, with default naming xxx.csi/maps/ll_<chem_shift_ppm>_n.fdf, where n is slice index.

A single 3D fdf map will be created for 3D csi data.

In Find natural logarithm of a number (C)

Syntax	ln(value)<	:n>	
Description	Finds the natural logarithm (base e) of a number. To convert the value to base 10, use $log_{10}x = 0.43429*ln(x)$.		
Arguments	value is a number.		
	n is the retur display the lo	on value giving the logarithm of value. The default is to ogarithmic value in the status window.	
Examples	ln(.5) ln(val):ln_val		
See also	User Programming		
Related	atan	Find arc tangent of a number (C)	
	cos	Find cosine value of an angle (C)	
	exp	Find exponential value of a number (C)	
	sin	Find sine value of an angle (C)	
	tan	Find tangent value of an angle (C)	

Load Load status of displayed shims (P)

Description Sets whether shim values are used. load is automatically set to 'y' by the rts and is automatically set to 'n' by su, go, au, and shim. Shim DAC values are automatically loaded after the console is rebooted (the last values returned before the console was rebooted).

Values 'y' begins any noninteractive shimming process or data acquisition after loading the shim DACs with the shim values from the current experiment. It also prevents acqi from delivering shim values to that experiment.

> 'n' begins any noninteractive shimming process or data acquisition with the current values stored in the shim DACs. Shim values in the current experiment are ignored.

See also	NMR Spectroscopy User Guide	
Related	acqi	Interactive acquisition display process (C)
	au	Submit experiment to acquisition and process data (C)
	go	Submit experiment to acquisition (C)
	rts	Retrieve shim coil settings (C)
	shim	Submit an autoshim experiment to acquisition (C)
	su	Submit a setup experiment to acquisition (M)

loadcolors Load colors for graphics window and plotters (M)

Syntax	loadcolors<(color_file)>		
Description	Loads the co loadcolors of setcolor loadcolors	lor table for VnmrJ graphics window and plotters. is generated by the color program and includes a series commands. On bootup, the bootup macro calls to set the graphics and plotter colors.	
	The loadcol plotter suppo configured.	lors macro checks the value of maxpen to decide if the orts colors. If maxpen is greater than 1, a color printer is	
Arguments	color_file is the name of the file to load. loadcolors first searches for this file in the directory \$vnmruser/templates/ directory. If not found there, loadcolors then searches the user_templates/vnmr directory. The default is a color table with the same name as the value of the plotter parameter that loadcolors searches for in the same two directories.		
Examples	loadcolors loadcolors('mycolortable')		
See also	VnmrJ Imaging NMR		
Related	bootup	Macro executed automatically when VnmrJ activated (M)	
	color	Select plotting colors from a graphic interface (M)	
	maxpen	Maximum number of pens to use (P)	
	setcolor	Set colors for graphics window and for plotters (C)	

loaduserprefsLoad Operator Preferences

See also At operator login, this macro loads the operator-specific parameter values set in the Preferences/UserPrefs panel.

loc Location of sample in tray (P)

Description	Indicates whether a sample changer is present and enabled, present		
	but disabled, or not present. If the changer is present and enabled, the		
	value of loc	sets the location in the tray of the sample in use or to	
	be used. The	loc parameter is stored in the global tree. When an	
	acquisition is	started, certain global parameters, including loc, are	
	saved with th	e experiment parameters. The saveglobal parameter	
	specifies whic	ch global parameters are saved.	
	The auto_au macro controls most of the automation features, including setting the value of loc.		
Values	A number between 1 and traymax indicates the sample location.		
	0 indicates th	e changer is not present or disabled.	
See also	NMR Spectroscopy User Guide; VnmrJ Walkup		
Related	auto_au	Controlling macro for automation (M)	
	saveglobal	Save selected parameters from global tree (P)	
	traymax	Sample changer tray size (P)	

locaction Locator action (M)

Description Perform an action on an object in the locator database. The action depends on the type of object selected, the action performed, and the target selected for the action.

Related	dndfid	Retrieve and process fid data from the locator (M)
	dndjoin	Join a work space from the locator (M)
	dndpar	Retrieve a parameter set from the locator (M)
	dndshims	Retrieve a shimset set from the locator (M)
	locprotoexec	Execute a protocol from the locator (M)
	xmmakenode	Make a new study queue node (M)

lock Submit an Autolock experiment to acquisition (C)

- Description Performs an automatic locking operation using the acquisition computer, optimizing lock power, phase, and gain. If necessary, lock obtains lock through a software-controlled search. lock is the only method to automatically adjust lock phase (usually needed only after probe change or lock channel tuning). lock also sets the rf frequencies, decoupler status, and temperature.
 - See also NMR Spectroscopy User Guide
 - RelatedauSubmit experiment to acquisition and process data (C)changeSubmit a change sample experiment to acquisition (M)gaSubmit experiment to acquisition and FT the result (C)

L

go	Submit experiment to acquisition (C)
sample	Submit change sample, autoshim experiment to acquisition
	(M)
shim	Submit an Autoshim experiment to acquisition (C)
spin	Submit a spin setup experiment to acquisition (C)
su	Submit a setup experiment to acquisition (M)

lockacqtc Lock loop time constant during acquisition (P)

- Description Controls time constant of lock loop during acquisition (i.e., time constant by which the lock feedback corrects disturbances of the magnetic field).
 - Values 1, 2, 3, or 4 (where 1 sets 1.2 seconds, 2 sets 4.7 seconds, 3 sets 12 seconds, and 4 sets 48 seconds).
 If lockacqtc does not exist, it is set to 48 seconds. All systems are designed to work well with the default settings, and there should rarely be a reason to alter the lock time constant. However, to experiment

```
create('lockacqtc','integer','global')
setlimit('lockacqtc',4,1,1,'global') lockacqtc=n
```

with other values, create lockacgtc and set a new value:

where *n* is the new value.

```
See also NMR Spectroscopy User Guide
```

Related create Create new parameter in a parameter tree (C) locktc Lock time constant (P) setlimit Set limits of a parameter in a tree (C)

lockfreq Lock frequency (P)

Description Sets system lock frequency. The value is entered using the Lock Frequency label in Spectrometer Configuration window. The value of lockfreq must be set correctly in order to observe NMR signals.

lockfreq can find the lock signal or resonance. Traditionally, Varian spectrometers have used the parameter z0 for this purpose; however, using lockfreq can require less shimming when switching solvents and less adjustment to the lock phase. To use lockfreq, set z0='n'.

Values 1 to 160 (in MHz), 'n' Use the true ²H frequency. Typical values of lockfreq are shown in the chart below.

¹ H Frequency		
200	30.710	30.6976
300	46.044	46.0625
400	61.395	61.471
500	76.729	• • •

¹ H Frequency	,		
600	92.095	• • •	
750	115.250		

Refer to the manual *VnmrJ Installation and Administration* for details on finding the correct lock frequency.

The commands; go, lock, shim, and su reset the lock frequency in the console to the current value of lockfreq. Lock frequency in the console can be set with the sethw command.

lockfreq is offset by the value of lkof, if that parameter exists, but sethw directly uses its numeric argument, without any offset by lkof.

See also	VnmrJ Installation and Administration; NMR Spectroscopy User Guide	
Related	config	Display current configuration and possibly change it (M)
	go	Submit experiment to acquisition (M)
	lkof	Track changes in lock frequency (P)
	lock	Submit an Autolock experiment to acquisition (C)

sethwSet values for hardware in acquisition system (C)setlockfreqSet lock frequency (C)shimSubmit an Autoshim experiment to acquisition (C)suSubmit a setup experiment to acquisition (M)

z 0 Z0 field position (P)

lockgain Lock gain (P)

Description Contains the current lock gain value as set by computer control. The value is stored in vnmrsys/global and can be examined by typing lockgain?.Values 0 to 48 dB, in 1-dB steps.

See also NMR Spectroscopy User Guide

lockphase Lock phase (P)

Description	Contains the current lock phase. The value is stored in
	vnmrsys/global and can be examined by typing lockphase?.
Values	0 to 360, in degrees, in 1.4-degree steps.
See also	NMR Spectroscopy User Guide

lockpower Lock power (P)

Description Contains the current lock power value as set by computer control. The value is stored in vnmrsys/global and can be examined by typing lockpower?.

Values 0 to 68 dB, in 1-dB steps, 68 is full power.

See also NMR Spectroscopy User Guide

locktc Lock time constant (P)

Description	Controls lock loop time constant when system is not performing acquisition (idle, lock display, shim display, FID display, autoshim, autolock, etc.).	
Values	1, 2, 3, or 4 (where 1 corresponds to 1.2 seconds, 2 to 4.7 seconds, 3 to 12 seconds, and 4 to 48 seconds). If locktc does not exist, the system uses a value of 1, the fastest value. To experiment with other value, create locktc and set a value (e.g., create('locktc','integer','global') setlimit('locktc',4,1,'global') locktc=2).	
See also	NMR Spectroscopy User Guide	
Related	create Create new parameter in a parameter tree (C)	

createCreateCreateNew parameterIn a parameterIn tree (ClockacqtcLock acquisition time constant (P)setlimitSet limits of a parameter in a tree (C)

log

Syntax	log(base	$(x) = 0.43429 \times \ln(x)$
Applicability	VnmrJ 3.1	
Description	sin	<pre>sin(angle)<:n>, radians, n is destination parameter</pre>
	COS	<pre>cos(angle)<:n>, radians, n is destination parameter</pre>
	tan	<pre>tan(angle)<:n>, radians, n is destination parameter</pre>
	asin	<pre>asin(angle)<:n> radians, n is destination parameter</pre>
	acos	<pre>acos(angle)<:n>, radians, n is destination parameter</pre>
	atan	atan(value)<:n>, pi/2 to -pi/2n, n is destination
	parameter	
	atan2	atan2(x,y)<:n>, y/x is pi/2 to -pi/2n, n is
	destination	n parameter
	exp	<pre>exp(value)<:n>, n is destination parameter</pre>
	ln	<pre>ln(value)<:n>, n is destination parameter</pre>
	sqrt	<pre>sqrt(value)<:n>, n is destination parameter</pre>
	abs	abs(value)<:n>, n is destination parameter

Description	Specifies whether the transmitter local oscillator (L.O.) is gated with the transmitter rf output or with the transmitter I.F. (intermediate frequency).		
	The logate parameter does not exist in most parameter sets; the system internally sets it to 'l'. To use the value 's', create logate and change the value by entering: create('logate','string') setenumeral('logate',2,'l','s') logate='s'.		
Values	'l' makes the transmitter L.O. gate with the rf output, producing better signal-to-noise, usually most important in liquids NMR.		
	's' makes the transmitter L.O. gate with the I.F. signal, producing sharper pulses, especially important in solid-state NMR.		
See also	User Guide: Solid-State NMR		
Related	createCreate new parameter in a parameter tree (C)setenumeralSet values of a string variable in a tree (C)		

Lookup Look up words and lines from a text file (C)

Applicability VnmrJ 4.0

Syntax lookup('codeword',argument<,'codeword', argument<,...>>):\$n1<\$n2<,...>>

Description The 'lookup' program allows one to search a text file for a word and return to the user subsequent words or lines. In this context, a "word" is defined as any string of characters delimited by "whitespace". By default, "whitespace" includes the space character, a tab, a newline, a carriage return, and a comma. The whitespace characters may also be specified. A word may, therefore, actually be a string a digits, a string or letters, or a combination of letters and digits. Note that punctuation marks, unless they are defined as whitespace as the comma is by default, can also form words or be part of a word. A line is any string of characters from the current word to the next carriage return. A line will include all "whitespace" characters except the carriage return. Depending on the codeword, word searches and word counts can be case insensitive or case sensitive. The 'lookup' program recognizes nine special codewords when these are supplied as arguments. These codewords and their meaning are listed below.

> 'file' : this codeword specifies that the next supplied argument will be the name of the text file which will be active. If this codeword is used, it MUST be the first argument passed to 'lookup' and the file name MUST be the second argument passed to 'lookup'. The search through a text file is a top to bottom search. The 'file' codeword has the additional feature of resetting the start of a search to the top of the text file. Subsequent searches through a previously accessed text file will continue from where the previous search stopped, provided the

'file' codeword is not used. The 'file' codeword also resets the whitespace characters back to their default values. If the text file does not exist, lookup will abort with an error.

- 'seek' : this codeword causes the 'lookup' program to search the text file for words which match those supplied as arguments following the 'seek' codeword. An implicit 'seek' is initially assumed for each call to 'lookup'. The 'lookup' program maintains a pointer to the word following the last successful 'seek'. The first argument following an explicit 'seek' codeword is interpreted as a word to search for, not a potential codeword. The second or later argument following an explicit 'seek' will be interpreted as a codeword if it matches one of the nine cases. Therefore, for example, one can search for the word 'file' without having it interpreted as a codeword by having it immediately follow the 'seek' codeword in the argument list. This seek is case insensitive.
- 'seekcs' : this codeword is the case sensitive equivalent to the seek codeword. In all other respects, it is the same as 'seek'. One can alternate between case sensitive and case insensitive searches.
- 'skip' : 'skip' increments the word pointer to the next word in the text file. This codeword may optionally be followed by a number which will specify how many words to skip.
- 'read' : 'read' returns to the user the word currently being pointed to and increments the pointer to the next word in the text file. This codeword may optionally be followed by a number which will specify how many words to return to the user.
- 'readline' : 'readline' returns to the user the word currently being pointed to and all following words until the end of the current line. The pointer is incremented to the first word of the next line in the text file. This codeword may optionally be followed by a number which will specify how many lines to return to the user.
- 'count': 'count' returns to the user the number of times words in the text file match the subsequent argument. The count starts at the current word pointer and proceeds to the end of the text file. The word count is case insensitive. That is, if you use 'count' to count the instances of the word "The", it will return the number of words that match "The" and "the".
- 'countcs' : this codeword is the case sensitive equivalent to the count codeword. In all other respects, it is the same as 'count'. If you use 'countcs' keyword to count the instances of the word "The", it will return the number of words that exactly match "The".
- 'countline' : this codeword returns the number of lines in the file.
 That is, it counts the number of newline characters (\n). If the
 'mfile' keyword is used (see below), the countline will count
 from the current file position to the end of the file.

'delimiter': this codeword specifies that the next supplied argument will be a list of characters which will be used to identify the whitespace used to identify words. The newline, tab, carriage return, backslash, and single quote characters are specified by \n, \t, \r, \\, and \' respectively. The two arguments 'delimiter',' \t\n\r', will reselect the default whitespace. The 'file' codeword will also reselect the default whitespace. The distinction is that the 'file' codeword will restart the search from the beginning of the file while the 'delimiter' codeword will continue from the current search position. Following the 'delimiter' codeword and it's argument, an implicit 'seek' is assumed.

Lookup can also be used to search multiple text files and read the contents of these files. Two additional codewords are used to implement this multiple file lookup. These two codewords and their meaning are listed below.

- 'mfile' : this codeword specifies that the next supplied argument will be the filekey to select one of multiple text files being accessed. If this codeword is used, it MUST be the first argument passed to 'lookup' and the filekey MUST be the second argument passed to 'lookup'. The first time a file is selected, or to restart the search at the beginning of the file, the name of the file is used instead of the filekey. Subsequent calls to lookup on this file would use the value returned by the 'filekey' codeword as the argument following the 'mfile' codeword. The 'mfile' codeword resets the whitespace to the default values. If the text file does not exist, lookup will abort with an error.
- 'filekey' : returns to the macro the current location within the file being accessed. In combination with the 'mfile' codeword, a subsequent call to lookup will start the search at the location within the file specified by the value of the filekey. The filekey serves both as a pointer to the file and as character offset within that file. The mfile and filekey codewords can be used to access multiple text files. In addition, these codewords can be used to keep track of various locations within a single file, in order to restart the search from that location.

The mfile and file keywords can be used together. Consider the case in which a macro named macrol uses lookup with the file codeword. A second macro, named macrol, also uses lookup, but on a separate file. If macrol uses the file codeword, and then in the middle of looking up words in that file, calls macrol, the next time macrol calls lookup, the lookup command will refer to the file used by macrol. Macrol can avoid having called macros alter the file it is reading by using the mfile codeword.

Examples:

Examples using this text file as the object of the calls to 'lookup':

lookup('file',systemdir + '/manual/lookup')

Select this file for the search
lookup('user','skip',2,'read',2,'readline'):\$n1,\$n2,\$n3
,\$ret

Seek is assumed with the call to lookup. The word 'user' is found on line 6. 'skip', 2 causes the pointer to jump two words. The codeword 'read' causes the following word to be put into n1. n1therefore is set to 'or'. The argument 2 specifies two words to be read. Therefore, n2 is set to 'lines.'. Note that the value of n2 includes the period. The word pointer now points to the word 'In' on line 6. The codeword 'readline' causes the remaining characters up to the next carriage return to be placed in n3. Therefore, n3 is set to 'In this context, a "word". The pointer now points to the first word in the next line, which is "is". The variable reat is set to the number of arguments successfully returned from the text file. This value is of use when deciding if you are at the end of the text file, since the variable receiving the value from the 'readline' will be set to ". In this case, ret will be set to 3.

lookup('skip',8,'read','skip',3,'read',2,'seek','comma'
):\$n3,\$n4,\$n5

'Skip', 8 causes the pointer to jump eight words. The 'read' sets \$n3 equal to 'by'. 'Skip', 3 then jumps three words. 'Read', 2 sets \$n4 equal to "whitespace" and \$n5 equal to 'includes'. The 'seek' argument then searches for the word 'comma', which it finds on line 12. The word pointer now points to the next word 'can'. Note that this seek did not find the string 'comma.' on line 9.

lookup('delimiter',' ,\'.\n\
t"','seek','file','must','skip',6,'read'):\$n6

'delimiter' and it's argument ',\'.\n\t"' sets whitespace to be space, comma, single quote, period, newline, tab, and double quote. The explicit 'seek' selects the next argument 'file' as a search word. Note that this 'file' is not interpreted as a codeword. Since single quotes are now whitespace, file is found on line 22. The search for the word 'must' matches MUST on line 24, since the search is case insensitive. 'Skip', 6 jumps six words. 'Read' sets \$n6 equal to 'lookup'. Note that it did not set \$n6 equal to "lookup" since single quotes have been chosen to be whitespace.

lookup('seekcs','The','read'):\$n7

'seekcs' is the case sensitive search. It finds the word 'The' on line 26. The 'read' sets to 'search'. Note that the word 'the' on line 25 did not match the capitalized search word 'The'.

As a working example of the lookup command, see the 'nextplotter' macro.

The use of the mfile and filekey codewords is demonstrated by the following macro. This macro reads two files, one word at a time, and puts the words into a third file.

541

L

```
lookup('mfile','filea','filekey'):$keya
lookup('mfile','fileb','filekey'):$keyb
morea = 2
smoreb = 2
$word = ''
while (($morea=2) or $moreb=2)) do
  if (\$morea = 2) then
lookup('mfile',$keya,'read','filekey'):$word,$keya,$mor
ea
     if (\$morea = 2) then
        write('file',curexp+'/textmerge',' %s',$word)
     endif
  endif
  if (\$moreb = 2) then
lookup('mfile',$keyb,'read','filekey'):$word,$keyb,$mor
eb
     if (\$moreb = 2) then
        write('file',curexp+'/textmerge',' %s',$word)
     endif
  endif
endwhile
```

In a second example, the macro calls a second macro, named macro1, and macro1 uses lookup with the file argument to access some file. By using the mfile codeword, the lookup commands in this macro are not affected by the call to macro1.

```
lookup('mfile','file1','filekey'):$key1
$ret = 2
while $ret > 1 do
lookup('mfile',$key1,'readline','filekey'):$line,$key1,
$ret
    "do something else"
    macro1
endwhile
```

Note that the 'filekey' codeword can by placed anywhere after the first two arguments. Several 'filekeys' can be specified and be used to start searches at specific locations in a file. For example,

```
lookup('mfile','filea','filekey'):$key
lookup('mfile',$key,'seek','First','filekey','Second','
filekey'):$key1,$key2
lookup(''mfile',\$key1, ...) will start the search in filea after
the first occurrence of the word "First".
lookup(''mfile',$key2, ...) will start the search in filea after
the first occurrence of the word "Second".
```

See also	User Programming	
Related	dialog	Display a dialog box from a macro (C)
	systemdir	VnmrJ system directory (P)

locprotoexec Execute a protocol from the locator (M)

Description	When a protocol is dragged from the locator and dropped onto the
	graphics canvas, this macro adds the protocol to the end of the study
	queue, and executes the macro associated with the protocol.

Related	dndfid	Retrieve and process fid data from the locator (M)
	dndjoin	Join a work space from the locator (M)
	dndpar	Retrieve a parameter set from the locator (M)
	dndshims	Retrieve a shimset set from the locator (M)
	locaction	Locator action (M)
	xmmakenode	Make a new study queue node (M)

First-order phase in directly detected dimension (P) 1p

Description Specifies the first-order phase-correction angles along the directly detected dimension according to the formula absorption $spectrum(\omega) =$ real channel(ω) * cos θ + imaginary channel(ω) * sin θ where the phase angle θ is a function of frequency, i.e. $\theta = rp + (\omega - \omega_0)/sw *lp$ ω_0 is defined to be the right end of the spectrum (i.e., 1p has zero effect at the right edge of the spectrum and a linearly increasing effect going to the left). In multidimensional data sets, 1p controls the phase of the directly detected dimension: f_2 dimension in 2D data sets, f_3 dimension in 3D data sets, etc.

Values -3600 to +3600, in degrees. Typical values are between 0 and -180.

L

See also	NMR	Spectroscopy User Guide
Related	aph	Automatic phase adjustment of spectra (C)
	lp1	First-order phase in 1st indirectly detected dimension (P)
	lp2	First-order phase in 2nd indirectly detected dimension (P)
	rp setl	Zero-order phase in directly detected dimension (P) Set parameters for zero linear phase (M)

1p1 First-order phase in 1st indirectly detected dimension (P)

Description	Controls the first-order phase constant along the first indirectly		
	detected dimension during the process of phase-sensitive 2D		
	transformation. The first indirectly detected dimension is often		
	referred to as the f_1 dimension of a multidimensional data set.		
See also	NMR Spectroscopy User Guide		
Related	1p First-order phase in directly detected dimension (P)		
	1p2First-order phase in 2nd indirectly detected dimension (P)		
	rp1 Zero-order phase in 1st indirectly detected dimension (P)		

1p2 First-order phase in 2nd indirectly detected dimension (P)

Description Controls the first-order phase constant along the second indirectly detected dimension during a ds, dconi, or equivalent display operation on the 2D data or a 1D trace therein. The second indirectly detected dimension is often referred to as the f_2 dimension of a 3D (or higher dimensionality) data set.

	-	
Related	dconi	Interactive 2D contour display (C)
	ds	Display a spectrum (C)
	lp	First-order phase in directly detected dimension (P)
	rp2	Zero-order phase in 2nd indirectly detected dimension
		(P)

1palg LP algorithm in np dimension (P)

NMR Spectroscopy User Guide

See also

Description Specifies the linear prediction (LP) algorithm to use in the np dimension. The resulting LP coefficients are used to appropriately extend the complex time-domain data prior to a normal Fourier transform. The LP algorithms work both on complex t_2 FIDs and on hypercomplex or complex t_1 interferograms. Enter addpar('lp') to create lpalg and other np dimension LP parameters in the current experiment

Values 'lpfft' does a least-squares calculation of lpfilt complex LP coefficients using lpnupts complex time-domain data points. Eigenvalue decomposition of the least-squares matrix is done using Householder tridiagonalization followed by the QL method with implicit shifts.

> 'lparfft' does a non-least-squares calculation of lpfilt complex LP coefficients using (lpfilt+1) complex, autoregressive (AR) matrix elements. These AR matrix elements are calculated from the raw, complex time-domain data using lpnupts points.

> Note that the 'lpfft' algorithm is preferred by far. While 'lparfft' can model broad lines and can extend data sets when mostly noise exists, it cannot model narrow lines.

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to the current experiment (M)
	lpalg1	LP algorithm in ni dimension (P)
	lpalg2	LP algorithm in ni2 dimension (P)
	lpext	LP data extension in np dimension (P)
	lpfilt	LP coefficients to calculate in np dimension (P)
	lpnupts	LP number of data points in np dimension (P)
	lpopt	LP algorithm data extension in np dimension (P)
	lpprint	LP print output in np dimension (P)
	lptrace	LP output spectrum in np dimension (P)
	np	Number of data points (P)
	proc	Type of processing on np FID (P)
	strtlp	Starting point for LP calculation in np dimension (P)
	strtext	Starting point for LP data extension in np dimension
		(P)

1palg1 LP algorithm in ni dimension (P)

- Description Specifies the LP (linear prediction) algorithm to use in the ni dimension. lpalg1 functions analogously to lpalg. Enter addpar('lp',1) to create lpalg1 and other ni dimension LP parameters in the current experiment. Values 'lpfft' or 'lparfft'
 - See also NMR Spectroscopy User Guide
 - Related addpar Add selected parameters to the current experiment (M) lpalg LP algorithm in np dimension (P) ni Number of increments in 1st indirectly detected dimension (P)

1pa1g2 LP algorithm in ni2 dimension (P)

Description	Specifies the LP (linear prediction) algorithm to use in the ni2 dimension. lpalg2 functions analogously to lpalg. Enter addpar('lp',2) to create lpalg2 and other ni2 dimension LP parameters in the current experiment.	
Values	'lpfft' or 'lparfft'	
See also	NMR Spectroscopy User Guide	
Related	addparAdd selected parameters to the current experiment (MlpalgLP algorithm in np dimension (P)ni2Number of increments in 2nd indirectly detecteddimension (P)	

1pext LP data extension in np dimension (P)

Description Specifies number of complex time-domain data points for LP (linear prediction) in the np dimension by which the original data is to be extended (or altered) in either the forward or backward direction. lpext is constrained by (strtext-lpext)>= ≥0 for lpopt='b' and by (strtext+lpext-1)<=fn/2 for lpopt='f'. In the np direction, if (strtext-lpext)=0 and lpopt='b' (backwards linear prediction with calculation of the first point), fpmult defaults to the theoretical value of 0.5 instead of 1.0. Enter addpar('lp') to create lpext and other np dimension LP parameters in the current experiment.

ent (M)
')
nsion
;

1pext1 LP data extension in ni dimension (P)

Description Specifies number of complex time-domain data points for LP (linear prediction) in the ni dimension by which the original data is to be extended (or altered) in either the forward or backward direction. lpext1 functions analogously to lpext. Enter addpar('lp',1) to create lpext1 and other ni dimension LP parameters in the current experiment.

Related addpar Add selected parameters to the current experiment (M)

lpext	LP data extension in np dimension (P)
ni	Number of increments in 1st indirectly detected
	dimension (P)

1pext2 LP data extension in ni2 dimension (P)

- Description Specifies number of complex time-domain data points for LP (linear prediction) in the ni2 dimension by which the original data is to be extended (or altered) in either the forward or backward direction. lpext2 functions analogously to lpext. Enter addpar('lp',2) to create lpext2 and other ni2 dimension LP parameters in the current experiment.
 - Related addparAdd selected parameters to the current experiment (M)lpextLP data extension in np dimension (P)ni2Number of increments in 2nd indirectly detected
dimension (P)

1pfilt LP coefficients to calculate in np dimension (P)

Description Specifies number of complex LP (linear prediction) coefficients in the np dimension to be calculated from a specified region of the time-domain data. lpfilt should be greater than nsignals, where nsignals is the number of sinusoidal signals contained in that FID (or interferogram). Enter addpar('lp') to create lpfilt and other np dimension LP parameters in the current experiment.

addpar	Add selected parameters to the current experiment (M)
lpalg	LP algorithm in np dimension (P)
lpfilt1	LP coefficients to calculate in ni dimension (P)
lpfilt2	LP coefficients to calculate in ni2 dimension (P)
np	Number of data points (P)
	addpar lpalg lpfilt1 lpfilt2 np

1pfilt1 LP coefficients to calculate in ni dimension (P)

Description Specifies number of complex LP (linear prediction) coefficients in the ni dimension to be calculated from a specified region of the time-domain data. lpfilt1 functions analogously to lpfilt. Enter addpar('lp',1) to create lpfilt1 and other ni dimension LP parameters in the current experiment.
Related addpar Add selected parameters to the current experiment (M) lpfilt LP coefficients to calculate in np dimension (P) ni Number of increments in 1st indirectly detected

dimension (P)

1pfilt2 LP coefficients to calculate in ni2 dimension (P)

Description Specifies number of complex LP (linear prediction) coefficients in the ni2 dimension to be calculated from a specified region of the time-domain data. lpfilt2 functions analogously to lpfilt. Enter addpar('lp',2) to create lpfilt1 and other ni2 dimension LP parameters in the current experiment.

RelatedaddparAdd selected parameters to the current experiment (M)lpfiltLP coefficients to calculate in np dimension (P)niNumber of increments in 1st indirectly detected
dimension (P)

1pnupts LP number of data points in np dimension (P)

Description Specifies number of complex time-domain data points in the np dimension to be used in constructing the autoregressive (lpalg='lparfft') or least- squares (lpalg='lpnefft') matrix from which the complex LP (linear prediction) coefficients are calculated. Note that lpnupts greater than or equal to 2*lpfilt is required for both algorithms. Enter addpar('lp') to create lpnupts and other np dimension LP parameters in the current experiment.

RelatedaddparAdd selected parameters to the current experiment (M)lpalgLP algorithm in np dimension (P)lpfiltLP coefficients to calculate in np dimension (P)lpnupts1LP number of data points in ni dimension (P)lpnupts2LP number of data points in ni2 dimension (P)npNumber of data points (P)

1pnupts1 LP number of data points in ni dimension (P)

Description Specifies number of complex time-domain data points in the ni dimension to be used in constructing the autoregressive (lpalg1='lparfft') or least- squares (lpalg1='lpnefft') matrix from which the complex LP (linear prediction) coefficients are calculated. lpnupts1 functions analogously to lpnupts. Enter addpar('lp',1) to create lpnupts1 and other ni dimension LP parameters in the current experiment.

Related	addpar	Add selected parameters to the current experiment (M)
	lpalg1	LP algorithm in ni dimension (P)
	lpnupts	LP number of data points in np dimension (P)
	ni	Number of increments in 1st indirectly detected
		dimension (P)

1pnupts2 LP number of data points in ni2 dimension (P)

- Description Specifies number of complex time-domain data points in the ni2 dimension to be used in constructing the autoregressive (lpalg2='lparfft') or least- squares (lpalg2='lpnefft') matrix from which the complex LP (linear prediction) coefficients are calculated. lpnupts2 functions analogously to lpnupts. Enter addpar('lp',2) to create lpnupts2 and other ni2 dimension LP parameters in the current experiment.
 - RelatedaddparAdd selected parameters to the current experiment (M)lpalg2LP algorithm in ni2 dimension (P)lpnuptsLP number of data points in np dimension (P)ni2Number of increments in 2nd indirectly detected
dimension (P)

LP algorithm data extension in np dimension (P)

Description Specifies how the specific LP (linear prediction) algorithm is to extend (or alter) forward or backward the time-domain data in the np dimension. Enter addpar('lp') to create lpopt and other np dimension LP parameters in the current experiment.

Multiple LP operations, extended forward or backward, can be performed on each FID or interferogram. This is accomplished by arraying the LP processing parameters (e.g., lpopt='b', 'f', 'b'). The number of LP operations is determined by the LP processing parameter with the largest array size. LP parameters having a smaller array size are padded out with their last value. The most common use for this capability is to back-calculate the first 1 to 2 points in an FID or interferogram and subsequently to extend the length of the time-domain data by LP.

A printout can be obtained for each LP operation on an individually definable FID or interferogram. For example, if lprint=30,30 and lptrace=1,2, the text file lpanalyz.out.1 contains the LP printout for the first LP operation on FID 1 and lpanalyz.out.2 contains the LP printout for the second LP operation on FID 2.

Values 'b' indicates the LP coefficients are to be used in the back-calculation of a specified number of time-domain data points.

'f' indicates the LP coefficients are to be used in the forward extension of the time-domain data by a specified number of points. The characteristic polynomial in z space, derived from the complex LP coefficients, is set up and rooted. Any root found to lie outside the unit circle is reflected back into the unit circle. New complex LP coefficients are then calculated from these adjusted complex roots.

Related addparAdd selected parameters to the current experiment (M)lpalgLP algorithm in np dimension (P)lpopt1LP algorithm data extension for ni dimension (P)

lpopt2	LP algorithm data extension for ni2 dimension (P)
lpprint	LP print output for np dimension (P)
lptrace	LP output spectrum for np dimension (P)
np	Number of data points (P)

LP algorithm data extension in ni dimension (P)

Description	Specifies how the specific LP (linear prediction) algorithm is to extend (or alter) forward or backward the time-domain data in the ni dimension. lpopt1 functions analogously to lpopt. Enter addpar('lp',1) to create lpopt1 and other ni dimension LP parameters in the current experiment.	
Related	addpar lpopt	Add selected parameters to the current experiment (M) LP algorithm data extension for np dimension (P)
	ni	Number of increments in 1st indirectly detected dimension (P)

1popt2 LP algorithm data extension in ni2 dimension (P)

Description Specifies how the specific LP (linear prediction) algorithm is to extend (or alter) forward or backward the time-domain data in the ni2 dimension. lpopt2 functions analogously to lpopt. Enter addpar('lp',2) to create lpopt2 and other ni2 dimension LP parameters in the current experiment.

Related addparAdd selected parameters to the current experiment (M)lpoptLP algorithm data extension for np dimension (P)ni2Number of increments in 2nd indirectly detected
dimension (P)

lpprint LP print output for np dimension (P)

- Description Controls LP (linear prediction) print output for the np dimension and creates an output file in the current experiment directory (curexp) with the name lpanalyz.out.1. Enter addpar('lp') to create lpprint and other np dimension LP parameters in the current experiment.
 - Values Comprised of sum of decimal values of the following bit fields, in which each bit field controls an independent output option:
 - Bit 0 (decimal value 1) writes out the LP matrix and Y vector from which the LP coefficients are calculated.

- Bit 1 (decimal value 2) writes out the LP coefficients that have been obtained using either of the two supported algorithms.
- Bit 2 (decimal value 4) writes out the LP roots obtained from the characteristic polynomial derived from the LP coefficients; this only applies for lpalg='lpfft' and lpopt='f'.
- Bit 3 (decimal value 8) writes out the original and recalculated values for each LP extended (or altered) complex time-domain data point.
- Bit 4 (decimal value 16) writes out the internal LP parameter structure.

For example, lpprint=12 and lptrace=1 yields the following information in the file curexp/lpanalyz.out.1 for spectrum 1 along f_2 : the values for all lpfilt complex LP coefficients and the original and recalculated values for each of the lpext LP extended (or altered) complex time-domain data points.

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to the current experiment (M)
	curexp	Current experiment directory (P)
	lpalg	LP algorithm in np dimension (P)
	lpext	LP data extension in np dimension (P)
	lpfilt	LP coefficients to calculate in np dimension (P)
	lpopt	LP algorithm data extension for np dimension (P)
	lpprint1	LP print output for ni dimension (P)
	lpprint2	LP print output for ni2 dimension (P)
	lptrace	LP output spectrum in np dimension (P)
	np	Number of data points (P)

1pprint1 LP print output for ni dimension (P)

Description Controls LP (linear prediction) print output for the ni dimension and creates an output file in the current experiment directory (curexp) with the name lpanalyz1.out.1. lpprint1 functions analogously to lpprint. Enter addpar('lp',1) to create lpprint1 and other ni dimension LP parameters in the current experiment.
See also NMR Spectroscopy User Guide
Related addpar Add selected parameters to the current experiment (M) lpprint LP print output for np dimension (P)
ni Number of increments in 1st indirectly detected dimension (P)

1pprint2 LP print output for ni2 dimension (P)

Description Controls LP (linear prediction) print output for the ni2 dimension and creates an output file in the current experiment directory (curexp)

with the name lpanalyz2.out.1. lpprint2 functions analogously to lpprint. Enter addpar('lp',2) to create lpprint2 and other ni2 dimension LP parameters in the current experiment.
See also NMR Spectroscopy User Guide
Related addpar Add selected parameters to the current experiment (M) lpprint LP print output for np dimension (P) ni2 Number of increments in 2nd indirectly detected dimension (P)

1ptrace LP output spectrum in np dimension (P)

Description	Specifies for dimension is Enter addpa parameters i	which spectrum LP (linear prediction) output in the np produced in accordance with the parameter lpprint. r('lp') to create lptrace and other np dimension LP n the current experiment.
See also	NMR Spectre	pscopy User Guide
Related	addpar	Add selected parameters to the current experiment (M)
	lpalg	LP algorithm in np dimension (P)
	lpprint	LP print output in np dimension (P)
	lptrace1	LP output spectrum in ni dimension (P)
	lptrace2	LP output spectrum in ni2 dimension (P)
	np	Number of data points (P)

1ptrace1 LP output spectrum in ni dimension (P)

Description	Specifies for which spectrum or trace LP (linear prediction) output in the ni dimension is produced in accordance with the parameter lpprint1. lptrace1 functions analogously to lptrace. Enter addpar('lp',1) to create t lpprint2 and other ni dimension LP parameters in the current experiment.		
See also	NMR Spectroscopy User Guide		
Related	addpar lpprint1 lptrace ni	Add selected parameters to the current experiment (M) LP print output in ni dimension (P) LP output spectrum in np dimension (P) Number of increments in 1st indirectly detected dimension (P)	

1ptrace2 LP output spectrum in ni2 dimension (P)

Description Specifies for which spectrum or trace LP (linear prediction) output in the ni2 dimension is produced in accordance with the parameter

	<pre>lpprint2. lptrace2 functions analogously to lptrace. Enter addpar('lp',2) to create lptrace2 and other ni2 dimension LP parameters in the current experiment.</pre>	
See also	NMR Spectro	scopy User Guide
Related	addpar	Add selected parameters to the current experiment (M)
	lpprint2	LP print output in ni2 dimension (P)
	lptrace	LP output spectrum in np dimension (P)
	ni2	Number of increments in 2nd indirectly detected dimension (P)

1s List files in directory (C)

Syntax	ls<(directory)>	
Description	Lists the names of files in a directory on the text output window. 1s is identical to dir and lf .	
Arguments	directory is the name of a directory. The default is the current working directory. 1s is equivalent to the UNIX command 1s and uses the same options (e.g., -1 for a long listing such as $ls('-1 *.fid')$).	
Examples	ls ls('data') ls('-l *.fid')	
Related	dirList files in directory (C)lfList files in directory (C)	

1sfid Number of complex points to left-shift the np FID (P)

Description	Specifies is to be e right shift parameter referred t dimension is proper	number of complex points (not real points) that the np FID either left-shifted (lsfid>0) or right-shifted (lsfid<0). A t adds zeros to the front of the FID. lsfid (and related rs phfid and lsfrq) operate on complex np FID data, to as the t_2 dimension in a 2D experiment or as the t_3 in in a 3D experiment. lsfid is in the processing group and by handled by a wti operation (display).
Values	-fn/2 to	np/2 (or $-fn/2$ to $fn/2$ if $fn < np$), 'n'
Related	dfid	Display a single FID (C)
	ds	Display a spectrum FID (C)
	fn	Fourier number in directly detected dimension (P)
	ft	Fourier transform 1D data (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ft2d	Fourier transform 2D data (C)
	lsfid1	Number of complex points to left-shift ni interferogram(P)
	lsfid2	Number of complex points to left-shift ni2 interferogram (P)

lsfrq	Frequency shift of the fn spectrum in Hz (P)
np	Number of data points (P)
phfid	Zero-order phasing constant for the np FID (P)
wft	Weight and Fourier transform 1D data (C)
wft1d	Weight and Fourier transform f_2 of 2D data (C)
wft2d	Weight and Fourier transform 2D data (C)
wti	Interactive weighting (C)

1sfid1 Number of complex points to left-shift ni interferogram (P)

Description Specifies number of hypercomplex (for hypercomplex interferogram data) or complex (for complex interferogram data) points that the ni interferogram is to be either left-shifted (lsfid1>0) or right-shifted (lsfid1<0). A right shift adds zeros to the front of the FID. lsfid1 (and related parameters phfid1 and lsfrq1) operate on ni interferogram data, both hypercomplex and complex. ni interferogram data are referred to as the t₁ dimension in both a 2D and a 3D experiment. lsfid1 is in the processing group and is properly handled by a wti operation (display); that is, a wti operation on an ni interferogram applies the parameters phfid1, lsfid1, and lsfrq1, if selected, to the time-domain data prior to the Fourier transformation. Values -fn1/2 to ni (or -fn1/2 to fn1/2 if fn1<2*ni), 'n'

Related	fn1	Fourier number in 1st indirectly detected dimension (P)
	lsfid	Number of complex points to left-shift np FID (P)
	lsfid2	Number of complex points to left-shift ni2
		interferogram (P)
	lsfrq1	Frequency shift of the fn1 spectrum in Hz (P)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	phfid1	Zero-order phasing constant for ni interferogram (P)
	wti	Interactive weighting (C)

1sfid2 Number of complex points to left-shift ni2 interferogram (P)

Description Specifies the number of hypercomplex (for hypercomplex interferogram data) or complex (for complex interferogram data) points that the ni2 interferogram is to be either left-shifted (lsfid2>0) or right-shifted (lsfid2<0). A right shift adds zeros to the front of the FID. lsfid2 (and related parameters phfid2 and lsfrq2) operate on ni2 interferogram data, both hypercomplex and complex. ni2 interferogram data are referred to as the t₂ dimension in a 3D

experiment. lsfid2 is in the processing group and is properly handled by a wti operation (display).

Values	-fn2/2 to n	i2 (or -fn2/2 to fn2/2 if fn2<2*ni2), 'n'
Related	fn2	Fourier number in 2nd indirectly detected dimension (P)
	lsfid	Number of complex points to left-shift np FID (P)
	lsfid1	Number of complex points to left-shift ni interferogram(P)
	lsfrq2	Frequency shift of the fn2 spectrum in Hz (P)
	ni2	Number of increments in 2nd indirectly detected dimension (P)
	phfid2	Zero-order phasing constant for ni2 interferogram (P)
	wti	Interactive weighting (C)

1sfrq Frequency shift of the fn spectrum (P)

- Description Sets a frequency shift of spectral data, in Hz. lsfrq is the time-domain equivalent of lp within VnmrJ. lsfrq (and related parameters phfid and lsfid) operate on complex np FID data, referred to as the t_2 dimension in a 2D experiment or as the t_3 dimension in a 3D experiment. lsfrq is in the processing group and is properly handled by a wti operation (display).
 - Values A positive value results in peaks being shifted downfield (to the left). A negative value results in peaks being shifted upfield (to the right).

Related	dfid	Display a single FID (C)
	ds	Display a spectrum FID (C)
	fn	Fourier number in directly detected dimension (P)
	ft	Fourier transform 1D data (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ft2d	Fourier transform 2D data (C)
	lp	First-order phase in directly detected dimension (P)
	lsfid	Number of complex points to left-shift np FID (P)
	lsfrq1	Frequency shift of the fn1 spectrum in Hz (P)
	lsfrq2	Frequency shift of the fn2 spectrum in Hz (P)
	phfid	Zero-order phasing constant for np FID (P)
	wft	Weight and Fourier transform 1D data (C)
	wft1d	Weight and Fourier transform f ₂ of 2D data (C)
	wft2d	Weight and Fourier transform 2D data (C)
	wti	Interactive weighting (C)

1sfrq1 Frequency shift of the fn1 spectrum (P)

Description Sets a frequency shift of spectral data, in Hz. lsfrq1 is the time-domain equivalent of lp1 within VnmrJ. lsfrq1 (and related parameters phfid1 and lsfid1) operate on ni interferogram data, both hypercomplex and complex. ni interferogram data are referred to as the t_1 dimension in both a 2D and a 3D experiment. lsfrq1 is in the processing group and is properly handled by a wti operation (display); that is, a wti operation on an ni interferogram applies the parameters phfid1, lsfid1, and lsfrq1, if selected, to the time-domain data prior to the Fourier transformation.

Values A positive value results in peaks being shifted downfield (to the left). A negative value results in peaks being shifted upfield (to the right).

Related	fn1	Fourier number in 1st indirectly detected dimension
		(P)
	lp1	First-order phase in 1st indirectly detected dimension
		(P)
	lsfid1	Number of complex points to left-shift ni
		interferogram(P)
	lsfrq	Frequency shift of the fn spectrum in Hz (P)
	lsfrq2	Frequency shift of the fn2 spectrum in Hz (P)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	phfid1	Zero-order phasing constant for ni interferogram (P)
	wti	Interactive weighting (C)

1sfrq2 Frequency shift of the fn2 spectrum (P)

Description	Sets a frequency shift of spectral data in Hz. $1sfrq2$ is the time-domain equivalent of $1p2$ within VnmrJ. $1sfrq2$ (and related parameters phfid2 and $1sfid2$) operate on ni2 interferogram data, both hypercomplex and complex. ni2 interferogram data is referred to as the t ₂ dimension in a 3D experiment. $1sfrq2$ is in the processing group and is properly handled by a wti operation (display).		
Values	A positive value results in peaks being shifted downfield (to the left A negative value results in peaks being shifted upfield (to the right)		
Related	fn2	Fourier number in 2nd indirectly detected dimension (P)	
	lp2	First-order phase in 2nd indirectly detected dimension (P)	
	lsfid1	Number of complex points to left-shift ni interferogram (P)	
	lsfid2	Number of complex points to left-shift ni2 interferogram (P)	
	lsfrq	Frequency shift of the fn spectrum in Hz (P)	
	ni2	Number of increments in 2nd indirectly detected dimension (P)	

phfid2 Zero-order phasing constant for ni2 interferogram (P)
wti Interactive weighting (C)

1v1 Zero-order baseline correction (P)

Description When spectral display is active, the command dc turns on a linear drift correction (baseline correction). The result of this operation includes calculating a zero-order baseline correction parameter lvl. This is done by averaging of a small number of points at either end of the display and drawing a straight line baseline between them.

Related	cdc	Cancel drift correction (C)
	lvltlt	Control sensitivity of lvl and tlt adjustments (P)
	tlt	First-order baseline correction (P)

lvltlt Control sensitivity of lvl and tlt adjustments (P)

Description Controls the sensitivity of the interactive lvl and tlt adjustments. lvltlt is in the "current" parameter set and is basically a multiplier for the sensitivity. If this parameter does not exist, it can be created by commands create('lvltlt') setgroup('lvltlt', 'display'). Values The default value is 1.0. Larger values make the adjustments larger.

- Smaller values make the adjustments smaller.RelatedcreatedsCreate new parameter in a parameter tree (C)dsDisplay a spectrum (C)
 - Iv1Zero-order baseline correction (P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

macro	Macro name (P)
macrocat	Display a user macro file in text window (C)
macrocp	Copy a user macro file (C)
macrodir	List user macro files (C)
macroedit	Edit a macro with user-selectable editor (M)
macrold	Load a macro into memory (C)
macrorm	Remove a user macro (C)
macrosyscat	Display a system macro file in text window (C)
macrosyscp	Copy a system macro to become a user macro (C)
macrosysdir	List system macros (C)
macrosysrm	Remove a system macro (C)
macrovi	Edit a user macro with the vi text editor (M)
make3dcoef	Make a 3D coefficients file from 2D coefficients (M)
makedosypar ams	Create parameters for DOSY processing (M)
makefid	Make a FID element using numeric text input (C)
makeeccglob als	Create global parameters for ECC control (M)
makeslice	Synthesize 2D projection of 3D DOSY experiment (C)
makeStudy	Create and manage Study Clones. (M)
makeuser	Add a new Vnmr user account or update an existing Vnmr user account (U)
makeuserpsg	Compiles the user PSG sources and constructs the user PSG object library
man	Display online description of command or macro (M)
managedb	Update user files (U)
manualpath	Path to user's manual directory (P)



manvi	Edit online description of a command or macro (M)
mapwin	List of experiment numbers (P)
mark	Determine intensity of spectrum at a point (C)
masvt	Type of variable temperature system (P)
maxattench1 -4	Maximum limit for attenuator setting for rf channel 1-4 (P)
maxpen	Maximum number of pens to use (P)
md	Move display parameters between experiments (C)
menu	Change status of menu system (C)
menuvi	Edit a menu with vi text editor (M)
method	Autoshim method (P)
mf	Move FIDs between experiments (C)
mfblk	Copy FID block (C)
mfclose	Close memory map FID (C)
mfdata	Move FID data (C)
mfopen	Memory map open FID file (C)
mftrace	Move FID trace (C)
mht	Move Hadamard parameters from one workspace to another
minsw	Reduce spectral width to minimum required (M)
mkchsums	Make checksum(s) for a given directory or file (C)
mkCPprotoco l	Make Protocol
mkdir	Create new directory (C)
mlabel	Menu label (P)
move	Move to an absolute location to start a line (C)
movedssw	Set down sampling parameters for selected spectral region (M)
moveossw	Set over sampling parameters for selected spectral region (M)
movesw	Move spectral window according to cursors (M)
movetof	Move transmitter offset (M)
mp	Move parameters between experiments (C)
mparval	Moves a Paramter Value Between Experiments

mqcosy	Set up parameters for MQCOSY pulse sequence (M)
mrev8	Set up parameters for MREV8 pulse sequence (M)
mrfb	Set the filter bandwidths for multiple receivers (P)
mref	Set referencing based on a existing spectrum of the sample (M)
mrgain	Set the gain for multiple receivers (P)
mspec	Set up the display of multiple spectra (C)
mstat	Display memory usage statistics (C)
mstring	Menu string (P)
mtune	Tune probe using swept-tune graphical display (M)
mv	Move and/or rename a file (C)
mvsampgloba l	Moves sample global parameters
mxconst	Maximum scaling constant (P)
mz	Move Integral Reset Points to specified experiment

macro Macro name (P)

Description	A string parameter, available in each experiment, similar to the $n1$, $n2$, and $n3$ parameters. Certain macros, such as $h1p$, need to know which macro invoked them. This parameter is used to pass that information.		
See also	User Programming		
Related	h1p Process simple proton spectra from h1 macro (M)		
	n1,n2,n3	Name storage for macros (P)	

macrocat Display a user macro file in text window (C)

Syntax	<pre>macrocat(file1<,file2><,>)</pre>	
Description	Displays one of	r more user macro files in the text window.
Arguments	file1, file2, are the names of macros in the user macro library.	
Examples	macrocat('build') macrocat('dan','george')	
See also	User Programming	
Related	macrodir	List user macros (C) Display a system macro file in taxt window (C)
	macrosyscal	Display a system macro me in text window (C)

Syntax macrocp(from_file,to_file)

- Description Makes a copy of the existing user macro file and places the copy in the user's macro library. Using macrocp to make a backup copy is the recommended procedure to modify a macro but still be able to revert to the previous version if you are unsure about the modification. macrocp can also be useful for writing a new macro that is very similar to an existing macro.
- Arguments from_file is the name of an existing user macro file to be copied. The file must be in the user's macro library.

to_file is the file name to be given to the copy. This name must be different from the name of the original macro.

Examples macrocp('dan','dan.old')

See also User Programming

Related macrocat Display a user macro file in text window (C) macrodir List user macros (C) macrosyscp Copy a system macro to become a user macro (C)

macrodir List user macro files (C)

Description Lists the names of user macro files in the user's macro library. See also User Programming Related macrosysdir Lists system macros (C)

macroedit Edit a macro with user-selectable editor (M)

Syntax macroedit(file)

Description Opens a MAGICAL macro file from a user's personal macro library for editing (if you want to edit a system macro, copy it to a personal library and then use macroedit).

The default editor is vi. To select another editor, first set UNIX environmental variable vnmreditor to the name of the editor; that is, in the .login file, change the line

setenv vnmreditor old_ed

to become

setenv vnmreditor new_ed (e.g., setenv vnmreditor emacs).

Second, make sure a script with the prefix vnmr_ followed by the name of the editor is placed in the bin subdirectory of the VnmrJ system directory (e.g., vnmr_emacs).

	The script file makes adjustments for the type of graphic interface in use. Scripts provided in the software include <i>vnmr_vi</i> and <i>vnmr_textedit</i> . To create other scripts, refer to the <i>vnmr_vi</i> script for non-window editor interfaces or refer to <i>vnmr_textedit</i> for window-based editor interfaces.		
Arguments	file is the name of the macro file you wish to edit.		
Examples	macroedit('pa')		
See also	User Programming		
Related	paramedit	Edit a parameter and its attributes with user-selected editor (C)	
	paramvi	Edit a parameter and its attributes with vi editor (M)	
	edit	Edit a file with user-selectable editor (C)	
	macrovi	Edit a user macro with vi editor (M)	
	menuvi	Edit a menu with the vi editor (M)	
	textvi	Edit text file of current experiment with vi editor (M)	

macrold Load a macro into memory (C)

Syntax macrold(file)<:dummy>

Description Loads a macro, user or system, into memory. If the macro already exists in memory, it is overwritten by the new macro. Loading a macro into memory increases the execution speed of the macro. The trade-off is that the macro uses memory. The mstat command displays macros that have been loaded into memory. One or more individual macros, or all the macros loaded in memory, can be removed from memory with the purge command.

If a macro already loaded into memory is edited using macrovi or macroedit, the changed macro automatically is loaded by those macros. This overwrites the previous macro. However, if a macro is edited or created some other way (with macrocp perhaps), the changed version is not automatically loaded. If the macro already exists in memory, the previous version executes unless the user runs macrold.

Arguments file is the name of the macro file to be loaded into memory. For loading macros, the same search path is used as when deciding which macro to execute. That is, the user's private maclib directory is searched first and finally the system maclib. If an absolute path is supplied as the file argument, that macro is loaded. This allows macros not in a maclib to be loaded and executed from VnmrJ.

> dummy is any throwaway variable. Requesting a return value suppresses the message in the status window (line 3) that the macro is loaded.

Examples macrold('pa')
macrold('_sw'):\$noline3

See also User Programming

Related	macrocp	Copy a user macro file (C)
	macroedit	Edit a macro with user-selectable editor (M)
	macrovi	Edit a user macro with the vi text editor (M)
	mstat	Display memory usage statistics (C)
	purge	Remove macros from memory (C)

macrorm Remove a user macro (C)

Syntax	macrorm(file)		
Description	Removes a user macro from the user's macro directory. If the macro has already been loaded in memory, it remains in memory until a new macro of the same name is loaded or the program exits.		
Arguments	file is the name of the user macro to be removed.		
Examples	macrorm('pa')		
See also	User Programming		
Related	delcom	Delete a user macro (M)	
	macrodir	List user macros (C)	
	macrosysrm	Remove a system macro (C)	
	purge Remove all macros from memory (C)		

macrosyscat Display a system macro file in text window (C)

Syntax	<pre>macrosyscat(file1<, file2><,>)</pre>	
Description	Displays one o	r more system macro files in the text window.
Arguments	file1, file2 library.	2, are names of macros in the system macro
Examples	macrosyscat macrosyscat	('build') ('dan','george')
See also	User Program	ning
Related	macrocat	Display a user macro file in text window (C)
	macrosysdir	Lists system macros (C)

macrosyscp Copy a system macro to become a user macro (C)

<pre>macrosyscp(from_file,to_file)</pre>
Makes a copy of the existing system macro file and places the copy in
the user's macro library. This is the recommended way to modify a
system macro for personal use.

Arguments	from_file is the name of an existing system macro file to be cop The file must be in the system macro library.		
	to_file is the name of the co many cases, it i of the same na system macro.	e file name to be given to the copy. In this case, the pied macro can be the same as the original macro. In s the same, allowing the user to have a personal macro me as the system macro but which will override the	
Examples	<pre>macrosyscp(' macrosyscp('</pre>	pa','pa') pa','mypa')	
See also	User Programm	ning	
Related	macrocp	Copy a user macro file (C)	
	macrosyscat	Display a system macro file in text window (C)	
	macrosysdir	Lists system macros (C)	

macrosysdir List system macros (C)

Description Lists the names of system macros in the system macro library. See also User Programming Related macrodir List user macros (C)

macrosysrm Remove a system macro (C)

Syntax	macrosysrm(file)	
Description	Removes a system macro file from the system macro directory. If the macro has already been loaded in memory, it remains in memory until a new macro of the same name is loaded or the program exits.	
Arguments	file is the name of the system macro file to be removed.	
Examples	macrosysrm('pa')	
See also	User Programming	
Related	macrorm	Remove a user macro (C)
	macrosysdir	Lists system macros (C)
	purge	Remove all macros from memory (C)

macrovi Edit a user macro with the vi text editor (M)

Syntax macrovi(file)

Description Initiates creating a new user macro or modifying an existing user macro using the UNIX vi text editor. On the Sun workstation, a pop-up window contains the edit. On the GraphOn, the edit is done

	on the entire to a personal	terminal. To edit a system macro, first copy the macro library and then edit it using macroedit or macrovi.	
Arguments	file is the n of a new use	ame of an existing user's macro to be edited or the name r's macro to be created.	
Examples	macrovi('pa')		
See also	User Programming		
Related	macroedit	Edit a macro with a user-selectable editor (C)	
	vi	Edit text file with vi text editor (C)	

make3dcoef Make a 3D coefficients file from 2D coefficients (M)

Syntax make3dcoef<('t1t2'|'t2t1')>

Description

Makes a 3D coefficients file from 2D coefficients and writes the file in the path stored by curexp. 2D coefficients are supplied as strings in the parameters f2coef and f1coef. This macro is capable of handling 3D data collected with any number of data sets (e.g., TPPI, Hypercomplex, Rance SE, Kay SE, and phase-sensitive gradient in one or both dimensions). make3dcoef is called by the ft3d macro.

The 2D coefficients are supplied as strings in floof and floof. These coefficients are the same as found by processing with wft2d(ldcoefs). Note that wft2da (for States-Hypercomplex method) is equivalent to wft2d(l,0,0,0,0,0,0,-1,0), and that wft2d (for absolute-value mode) is equivalent to wft2d(l,0,0,-1).

Coefficients are separated by spaces and not commas. For example, if a 3D data set collected by the States-Hypercomplex method in both ni and ni2 dimensions, $flcoef='1 \ 0 \ 0 \ 0 \ 0 \ -1 \ 0'$ and $f2coef='1 \ 0 \ 0 \ 0 \ 0 \ -1 \ 0'$. And if a 3D data set collected in absolute-value mode in both ni and ni2 dimensions, $flcoef='1 \ 0 \ 0 \ -1'$ and $f2coef='1 \ 0 \ 0 \ -1'$.

The flooef and f2coef parameters are created by the par3d macro. Execution of make3dcoef when flooef and f2coef have no value or inconsistent values causes the macro to abort, which enables the user to enter these values and reexecute the macro. For example, the value of flooef when the F1 dimension can be processed with wft2da is '1 0 0 0 0 0 -1 0'. The value of f2coef when the F2 dimension can be processed with wft2d(1,0,1,0,0,-1,0,1) is '1 0 1 0 0 -1 0 1'.

The parameters flcoef and flcoef must be 2D coefficients that give proper ni and nil first planes with the same rp (assuming lp is 0 by using calfa) values. For example, processing the phase-sensitive gradient dimension should not be done with $1 \ 0 \ 0 \ 1 \ 0 \ 1 \ 0 \ 1 \ 0$ and applying 45° phase shifts to rp, but with $1 \ 0 \ 1 \ 0 \ 1 \ 0 \ -1$, or its variant, that gives the same rp value as the other dimension. This also applies to Rance-type or Kay-type sensitivity-enhanced dimensions.

Note that sensitivity-enhanced sequences (gradient or otherwise) can be processed two different ways to give "orthogonal" data sets. The coefficients must be picked so that they have the same ${\tt rp}$ as the other dimension.

This macro can also handle coefficients that are not 1s or 0s. For example, if processing requires that a data set contributes to the interferogram after a 30° phase shift, $\cos(30)$ and $\sin(30)$ can be selected as the real and imaginary contributions, respectively, during the construction of the interferogram.

Arguments 'tlt2' means array='phase,phase2' in simple hypercomplex data sets. It means array='tlrelated','t2related' with multiple sets in general.

't2t1' means array='phase2, phase' in simple hypercomplex data sets. It means array='t2related', 't1related' with multiple sets in general.

If no argument is used and if array='phase,phase2' or array=
'phase2,phase, the macro automatically decides on 't1t2' or
't2t1', respectively.

See also NMR Spectroscopy User Guide

Related	array	Parameter order and precedence (P)		
	calfa	Recalculate alfa so that first-order phase is zero (M)		
	curexp	Current experiment directory (P)		
	f1coef	Coefficient to construct F1 interferogram (P)		
	f2coef	Coefficient to construct F2 interferogram (P)		
	ft3d	Perform a 3D Fourier transform on a 3D FID data set (M)		
	lp	First-order phase in directly detected dimension (P)		
	ni	Number of increments in 1st indirectly detected dimension		
		(P)		
	ni2	Number of increments in 2nd indirectly detected dimension		
		(P)		
	ntype3	Specify whether f_1 or f_2 display expected to be N-type (P)		
	d			
	rp	Zero-order phase in directly detected dimension (P)		
	wft2d	Weight and Fourier transform 2D data (C)		
	wft2da	Weight and Fourier transform phase-sensitive data (M)		

makedosyparamsCreate parameters for DOSY processing (M)

Syntax	makedosyparams(dosytimecubed,dosyfrq)		
Description	This macro is automatically called by the Dbppste, DgcsteSL, Doneshot,		
	Dbppsteinept, Dgcstecosy, and Dgcstehmqc sequences to create the		
	parameters dosyfrq, dosygamma, and dosytimecubed, which are		
	necessary for the dosy analysis. Do not manually run		
	makedosyparams.		

See also	NMR Spectroscopy User Guide	
Related	dosy	Process DOSY experiments (M)
	dosyfrq	Larmor frequency of phase encoded nucleus in DOSY (P)
	dosygamma	Gyromagnetic constant of phase encoded nucleus in DOSY (P)
	dosytimecubed	Gyromagnetic constant of phase encoded nucleus in DOSY (P)

makefid Make a FID element using numeric text input (C)

Syntax makefid(file<,element_number<,format>)

Description Creates FID files that can be used to introduce computed data into an experiment. The number of points comes from the number of numeric values read from the input file. If the current experiment already contains a FID, you will not be able to change either the format or the number of points from that present in the FID file. Use rm(curexp+'/acqfil/fid') to remove the FID.

The makefid command does not look at parameter values when establishing the format of the data or the number of points in an element. Thus, if the FID file is not present, it is possible for makefid to write a FID file with a header that does not match the value of dp or np. Because the active value is in the processed tree, you need to use the setvalue command if any changes are required.

Arguments file is the name of the input file. It contains numeric values, two per line. The first value is assigned to the X (or real) channel; the second value on the line is assigned to the Y (or imaginary) channel.

element_number is the number of the element or FID and is any integer larger than 0. The default is the first element or FID. If the FID element already exists in the FID file, the program overwrites the old data.

format is a character string with the precision of the resulting FID file and can be specified by one of the following strings:

dp=n' dp=y' 16-bit' 32-bit'	single-pre double-pre single-pre double-pr	ecision (16-bit) data ecision (32-bit) data ecision (16-bit) data ecision (32-bit) data	
		If an FID file precision; oth	e exists, makefid uses the same format string for nerwise, the default is double-precision (32-bit) data.
		element_nu	mber and format arguments can be entered in any order.
	Examples	makfid('fi	d.in',2,'32-bit')
	See also	NMR Spectro	scopy User Guide; User Programming
	Related	cp curexp	Copy a file (C) Current experiment directory

dp	Double precision (P)
mv	Move and/or rename a file (C)
np	Number of data points (P)
rm	Delete file (C)
setvalue	Set value of any parameter in a tree (C)
writefid	Write numeric text file using a FID element (C)

makeeccglobals Create global parameters for ECC control (M)

Applicability	Systems with Varian, Inc. Cold Probes
Description	Creates the following nine global parameters required for ECC control by PSG:
	tc1z, tc2z, tc3z, tc4z, amp1z, amp2z, amp3z, amp4z, and chiliConf

Related chiliConf

makeslice Synthesize 2D projection of 3D DOSY experiment (C)

Syntax	<pre>makeslice(<option>,lowerlimit,upperlimit)</option></pre>		
Arguments	option is either 'i' or 's'.		
	'i' includes the "tails" of diffusion peaks that lie outside the range between lowerlimit and upperlimit. The default is 'i'.		
	's' only includes the integration peaks whose diffusion coefficient lies between the specified limits.		
	lowerlimit is the lower diffusion limit (in units of 10^{-10} m ² /s) to be displayed.		
	upperlimit is the upper diffusion limit (in units of 10^{-10} m ² /s) to be displayed.		
Description	Synthesizes an integral projection between specified diffusion limits of a 3D DOSY spectrum onto the frequency-frequency plane. makeslice requires the first 2D increment of the 3D DOSY data to have been transformed.		
See also	NMR Spectroscopy User Guide		
Related	dosyProcess DOSY experiments (M)showoriginalRestore first 2D spectrum in 3D DOSY spectrum (M)		

makeStudy Create and manage Study Clones.

Syntax

ApplicabilityVnmrJ 3.1DescriptionDo not use this macro from the command line.See alsoUser Guide: Automation-Clone a New Study

makeuser

Add a new Vnmr user account or update an existing Vnmr user account (U)

Syntax makeuser

Applicability VnmrJ 3.1

Description

The makeuser command is provided to create a new user account with permission to access the VNMR files and programs. The makeuser command will also install the necessary files and directories into the user's home directory.

The makeuser command can be run by the system administrator's root account or by any current user with appropriate permissions. In order to add a new user to the system, makeuser must be run by root.

When root executes makeuser, the location of the VNMR system directory, if not available from the vnmrsystem environmental parameter, will be requested. The most likely location, which is provided as the default, is /vnmr. When executed as root, the user name whose account is to be added or updated may be supplied as an argument to makeuser. If no name is supplied, one is requested. The makeuser script checks to see if the user is already defined in the system. If not, the /etc/passwd file will be updated with the new user and the user will be added to the nmr group in the /etc/group file. A home directory will also by made. The location of the home directory is encoded in the makeuser script and may be altered if desired. By convention, on Sun systems, the home directory is made in the /home directory. Also by convention, on IBM systems, the home directory is in the /u directory. Note that the makeuser script updates the /etc/passwd and /etc/group files on the local machine. If Network Information Services (formerly known as Sun Yellow Pages) is running, this may not be the correct thing to do. In this case, the script could be executed on the host which is the password server. Additional steps may be required to make the new account available over the network. Refer to Sun documentation for this information.

The above operations require root privileges to execute. After finishing these tasks, the makeuser script gives root an opportunity to exit from the script.

A user other than root can run the makeuser script. In this case, the makeuser script will only update the current user's files. If one tries to update another account, an error will be reported. If a user runs the makeuser script (or the root account decided not to exit when given the above opportunity), the following question will appear

Automatically configure the 'user' account (y or n) where user is replaced by the user's name. If this is answered with a y, then the necessary UNIX files and Vnmr files will be added to the account and various Vnmr subdirectories will be made. The next time the user logs out and then logs in, Vnmr will automatically start. After logging out and then logging in, whenever vnmr is typed, VNMR will start. If the above question is answered with an n, meaning one does not want all the files automatically updated, then two additional questions are asked.

- Automatically configure UNIX environment (.files) (y or n)
- Automatically configure Vnmr directories and global parameters (y or n)

Answering n to one or both of these questions invokes an interactive mode where one is asked

OK to update Unix_file (y or n)

for the various UNIX dotfiles (for example, .login, .cshrc, .Xdefaults, etc).

For the Vnmr related files and subdirectories, one is asked

- Create subdirectory of your VNMR user directory (y or n)
- Update your VNMR global parameters (y or n)

If one decides not to do the automatic configuration of the UNIX dotfiles, then samples of what is required for proper functioning of Vnmr is defined in the file .xlogin in the user_templates subdirectory of the Vnmr system directory. The actual UNIX dotfiles which are used for the automatic update are also stored in this same user_templates directory. he Vnmr global file is also stored there.

makeuserpsg Compiles the user PSG sources and constructs the user PSG object library

Syntax	
Applicability	VnmrJ 3.1
Description	MAKEUSERPSG is a UNIX makefile which is invoked by the shellscript PSGGEN. MAKEUSERPSG has the following attributes:
	• All compilation and library construction is performed in the user PSG directory;
	• Any additional source (*.c) and header files (*.h and *.p) and the makefile itself, unless already present, are linked from the system PSG directory into the user PSG directory via soft links;

• The three possible names for the user PSG object library are LIBPSGLIB.A, LIBPSGLIB_FPC.A, and LIBPSGLIB_FPA.A. The first name is used only for Sun 4 systems. The last two names are used for Sun 3 systems with SEQGEN_OPTION set to f68881 or ffpa, respectively.

MAKEUSERPSG currently has no error recovery. Therefore, if an error occurs, the user PSG directory will not be cleaned up, i.e., the soft

links to files in the system PSG directory will remain in this directory along with any object file previously created by the make-file.

man	Display online description of command or macro (M)
Syntax	<pre>man('file')<:\$return></pre>
	Displays a description of commands and macros from files in the applications directory. The manual file is displayed in the text window when it is retrieved by the man macro. The man macro aborts if a name is not supplied as an argument.
Arguments	file – name of a command or macro in one of the applications directories.
	: $sres$ - supply a return argument to suppress messages if the manual page does not exist.
Examples	man('mark')
	<pre>man('notAcommand'):\$res</pre>
See also	NMR Spectroscopy User Guide; User Programming
Related	manviEdit online description of a command or macro (M)manualpathPath to user's manual directory (P)

managedb Update user files (U)

Syntax	managedb update
Description	Updates VnmrJ database for the Locator.
See also	NMR Spectroscopy User Guide

manualpath Path to user's manual directory (P)

Description	Contains the absolute path to a user's directory of VnmrJ manual entries. If manualpath exists for a user, it must be defined in the		
	user's global parameter file. Enter		
	create('manualpath','string','global') to create the		
	manualpath parameter.		
See also	User Programming		
Related	man Display online description of a command or macro (M)		

manvi Edit online description of a command or macro (M)

Syntax	manvi('file')	
Description	Enables editing or creating an online description of commands and macros stored in any of the applications directories for to which the user has write permission.	
Arguments	file is the name of a command macro.	
Examples	manvi('mark')	
See also	User Programming	
Related	man Display online description of command or macro (M)	

mapwin List of experiment numbers (P)

Description Arrayed global parameter that maintains a list of experiment numbers for the window panes in the VnmrJ graphics window.

Related	curwin	Current window (P)
	fontselect	Open FontSelect window (C)
	jwin	Activate current window (M)
	setgrid	Activate selected window (M)
	setwin	Activate selected window (C)

mark Determine intensity of spectrum at a point (C)

- Syntax (1) mark<(f1_position)><:intensity>
 - (2) mark<(left_edge,region_width)><:intensity, integral>
 - (3) mark<(f1_position,f2_position)><:intensity>
 - (4) mark<(f1_start,f1_end,f2_start,f2_end)>
 <:intensity,integral,c1,c2>

 - (6) mark('reset')
- Description Find the intensity of a spectrum at a point. Either 1D or 2D operations can be performed in the cursor or box mode for a total of four separate functions: 1D operations in cursor mode (syntax 1), 1D operations in box mode (syntax 2), 2D operations in cursor mode (syntax 3) and 2D operations in box mode (syntax 4).

In the *cursor mode*, the intensity at a particular point is found. In the *box mode*, the integral over a region is calculated. The displayed integral is scaled in the same way as output from dli is scaled; that

is, by the ins and insref parameters. For 2D operations, this is the volume integral and the volume is scaled by ins2 and ins2ref. In addition, the mark command in the box mode finds the maximum intensity and the coordinate(s) of the maximum intensity.

The mark command requires that transformed data be present in the current experiment. If required, it recomputes the phase file from the complex data (i.e., it rephases the data if required); however, the mark command requires parameters from the command line if no data is displayed (i.e., if ds or dconi has not been executed).

Note that 2D operations require that 2D data be present. This not only means that ni must be larger than 1, but also that the data was transformed using ftld, ft2d or an equivalent (and not ft or its equivalents).

The mark command, as well as the MARK button of ds, writes output to a file in the current experiment. For 1D operations, the file is named mark1d.out; for 2D operations, it is mark2d.out. If this file already exits, VnmrJ appends output from the current mark operation to the end of the file. (Older versions of VnmrJ used ds.out and dconi.out as files for output from the MARK button). Either file can be read by other programs at any time between operations.

The following criteria establish the exact function. The command checks them in the following order until it determines the exact function:

- 1. Number of numeric parameters.
- 2. Number of return values called out.
- 3. Which display command (ds or dconi) was last used.

4. Nature of the data in the experiment.

The first two criteria only serve to distinguish between box mode and cursor mode. The nature of the data in the experiment and the last display command entered determines whether a 1D or a 2D operation is selected.

Arguments f1_position defines the position, in Hz, along the f_1 axis in the 1D and 2D cursor modes. The default is cr (1D) or cr1 (2D).

left_edge defines the position of the left edge of the region, in Hz, to be integrated in 1D box mode. The default is cr.

region_width defines the width, in Hz, of the region, which extends to the right of left_edge, in 1D box mode. The default is delta.

f2_position defines the position, in Hz, along the f_2 axis in the 2D cursor mode. The default is delta1.

<code>fl_start</code> and <code>fl_end</code> define region along the \mathbf{f}_1 axis in the 2D box mode.

f2_start and f2_end define region along the $\rm f_2$ axis in the 2D box mode.

'trace' is a keyword to select a 1D operation if 2D data is present. It must be either the first or the last argument (e.g.,

 ${\tt mark('trace',400)}$ determines the intensity at 400 Hz in the current trace).

Μ

'reset' is a keyword to erase the output files from the mark command. No other argument can be used with this keyword. Use rename to rename the current mark output files (e.g., rename(curexp+'/mark1d.out', curexp+'/mark.16.01.89')

intensity is a return value set to the intensity of the spectrum at the point for either 1D or 2D operations (the maximum if cursor mode was selected).

integral is a return value set to the integral of the spectrum at the point. integral is not returned in the cursor mode.

c1, c2 are return values set to the coordinates where the maximum intensity was found in 2D mode. c1 and c2 are not returned in the cursor mode.

Examples 1D data sets:

mark(cr)	cursor mode for 1D data
<pre>mark(cr,delta)</pre>	box mode for 1D data

2D data sets (2D mode): In this mode, the order of the arguments to mark is independent of the trace parameter.

<pre>mark(cr1,cr)</pre>	cursor mode for 2D data
<pre>mark(cr1,delta1,cr,delta)</pre>	box mode for 2D data

2D data sets (1D mode): In this mode, the selection of the arguments to mark is dependent on the trace parameter. If trace='f2', then cr, delta, sp, or wp are appropriate. If trace='f1', then cr1, delta1, sp1, and wp1 are appropriate.

<pre>mark('trace',cr)</pre>	cursor mode for selected 2D trace
<pre>mark('trace',cr1,delta1)</pre>	box mode for selected 2D trace

Alternate: MARK button in the ds program.

See also NMR Spectroscopy User Guide; User Programming

Related	cr	Cursor position in directly detected dimension (P)
	cr1	Cursor position in 1st indirectly detected dimension
		(P)
	curexp	Current experiment directory (P)
	dconi	Interactive 2D contour display (C)
	delta	Difference of two frequency cursors (P)
	dli	Display list of integrals (C)
	ds	Display a spectrum (C)
	ft1d	Fourier transform along f ₂ dimension (C)
	ft2d	Fourier transform 2D data (C)
	ins	Integral normalization scale (P)
	ins2	2D volume value (P)
	insref	Fourier number scaled value of an integral (P)
	ins2ref	Fourier number scaled volume of a peak (P)
	mv	Move and/or rename a file (C)
	ni	Number of increments in 1st indirectly detected
		dimension (P)

masvt Type of variable temperature system (P)

Description	Identifies the type of VT system in use: the standard Oxford V controller or the Oxford-Sorenson or solids VT controller system with the Varian VT CP/MAS probe). masvt is a global paramet is active on all of each user's experiments on a per user account The current value of the parameter can be displayed by typing m		
	Note that the VT Controller option displayed by config must be set to Present for either VT controller system to be active. If masvt does not exist, it can be created with the command create('masvt','string','global').		
	The new Highland VT controller is autosensing, making masvt superfluous for systems with this controller.		
Values	'y' indic	ates the solids VT system is in use.	
	'n', any other value but 'n' and 'y', or if masvt does not exist, indicate that the Oxford Varian VT controller, if present, is in use.		
See also	VnmrJ Installation and Administration		
Related	config	Display current configuration and possibly change values (M)	
	create	Create a new parameter in a parameter tree (C)	
	vttype	Variable temperature controller present (P)	

maxattench1-4 Maximum limit for attenuator setting for rf channel 1-4 (P)

Description maxattench1, maxattench2, maxattench3, and maxattench4, are optional global parameters for the limiting the maximum attenuator settings for rf channel 1, channel 2, channel 3, and channel 4 (respectively) from pulse sequence statements and through tpwr/dpwr/... settings on go command. If maxattench2 is present, the attenuator setting check will be carried out by SpinCAD and C psg. If the attenuator setting exceeds the limit set in maxattench2, psg aborts with error message. This command is only applicable for check during the go command.

See also SpinCAD

maxpen Maximum number of pens to use (P)

DescriptionControls the maximum number of pens that will be used.Values1 to the number of pens in the system plotter. If maxpen=x and the

'alues 1 to the number of pens in the system plotter. If maxpen=x and the software attempts to use pen x+y, it uses pen y instead.
See also	NMR Spectroscopy User Guide		
Related	pen	Select a pen or color for drawing (C)	
	setpen	Set maximum number of HP plotter pens (M)	

md Move display parameters between experiments (C)

Syntax	<pre>md(<from_exp,>to_exp)</from_exp,></pre>		
Description	Moves the saved display parameters from one experiment to another. These parameters must have been saved with the s command (e.g., $s2$).		
Arguments	from_exp specifies the number of the experiment, 1 through 9, from which the parameters are to be taken. The default is that the parameters are moved from the currently active experiment.		
	to_exp specifies to which experiment the parameters are to be moved.		
Examples	md(4) md(2,3)		
See also	NMR Spectroscopy User Guide		
Related	mf Move FIDs between experiments (C)		
	mp Move parameters between experiments (C)		
	s Save display parameters as a set (M)		

menu Change status of menu system (C)

Syntax	<pre>(1) menu(menu_name) (2) menu<('off')></pre>	
Description	The VNMR menu system allows up to eight buttons to be active at a time, enabling the user to perform most actions with the mouse rather than typing in commands. All menus are stored in the library menulik in the system directory or in the user's menulib. See menuvi to change these menus.	
	If the menu system becomes deactivated for some reason, select the Menu On button in the Permanent Menu to reactivate it. Entering menu('main') also works.	
Arguments	s menu_name is the name of the file controlling the menu (e.g., 'main' Including this argument activates the menu system and displays the menu controlled by menu_name.	
	'off' is a keyword to turn off the menu system.	
Examples	menu menu('fitspec') menu('off')	
See also	User Programming	
Related	menuvi Edit a menu with the vi text editor (M)	

mlabel	Menu label (P)
newmenu	Select a menu without immediate activation (C)

menuvi Edit a menu with vi text editor (M)

Syntax	menuvi(menu)	
Description	Edits a Classic VNMR menu file using the UNIX vi text editor. On the Sun workstation, a pop-up window contains the edit. On the GraphOn, the edit is done on the entire terminal.	
Arguments	menu is the name of file controlling a menu.	
Examples	menuvi('display_1D')	
See also	User Programming	
Related	menu	Change status of menu system (C)
	newmenu	Select a menu without immediate activation (C)
	vi	Edit text file with vi text editor (C)

method Autoshim method (P)

Description	Selects the method for automatic shimming. Refer to the manual NMR Spectroscopy User Guide for information on how to write or alter methods.	
Values	Name of file in the /vnmr/shimmethods library for one of the defined shim methods in the system. To display all available methods, enter ls('/vnmr/shimmethods'). Standard methods include 'zlz2' (selects shimming of the Z1 and Z2 gradients) and 'allzs' (selects shimming all spinning gradients, Z1 to Z4 or Z5, depending on the magnet type). Shim methods can also be stored in a user's shimmethods directory (e.g., /home/vnmr1/vnmrsys/shimmethods).	
See also	NMR Spectroscopy User Guide	
Related	ls newshm stdshm	List files in current directory (C) Interactively create a shim method with options (M) Interactively create a shim method (M)

mf Move FIDs between experiments (C)

Syntaxmf(<from_exp,>to_exp)DescriptionMoves the last acquired FID, as well as its associated parameters, from
one experiment to another. The text, the processed acquisition
parameters and the current display and processing parameters are also
moved to the specified experiment.

Arguments	from_exp specifies number of the experiment from which the FID i to be taken. The default is the FID is moved from the currently activ experiment.	
	to_exp specifies to which experiment the FID is to be moved.	
Examples	mf(4) mf(3,2)	
See also	NMR Spectroscopy User Guide	
Related	md Move display parameters between experiments (C)	
	mp Move parameters between experiments (C)	

mfblk Copy FID block (C)

Syntax	<pre>mfblk(<src_expno,>src_blk_no,dest_expno,dest_blk_no)</src_expno,></pre>		
Description	n Copies data from a source FID block specified by src_blk_no t destination FID block specified by dest_expno and dest_blk_r using memory-mapped input and output.		
	<pre>mfblk searches for the source and destination FID file in the direct \$vnmruser/expN/acqfil, where N is the requested experiment number or the current experiment number. If the FID file is not op mfblk opens the file, copies the data, and closes the file. If a num of blocks need to be copied, explicitly opening and closing the file with the commands mfopen and mfclose can significantly speed the data reformatting process.</pre>		
	mfblk can also be used to append blocks of data to a FID file by specifying that the dest_blk_no is greater than the number of blocks in a file.		
	Be aware that mfblk can modify data returned to an experiment with the rt command. To avoid modification, enter the following sequence of VnmrJ commands before running mfblk:		
	<pre>cp(curexp+'/acqfil/fid', curexp+'/acqfil/fidtmp') rm(curexp+'/acqfil/fid') mv(curexp+'/acqfil/fidtmp', curexp+'/acqfil/fid')</pre>		
Arguments	src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.		
	<pre>src_blk_no specifies the source block of data to be copied. Block numbers start at 1 and run from 1 to the number of blocks in a file.</pre>		
	dest_expno specifies the experiment number of the destination FID file.		
	dest_blk_no specifies the destination block to send the copied data.		
Examples	<pre>mfblk(1,2,1) copies current experiment, block 1 to exp 2, block 1. mfblk(3,2,6,2) copies exp 2, block 2 to exp 6, block 2.</pre>		
See also	User Programming		
Related	mfcloseMemory map close FID file (C)mfdataMove FID data (C)		

mfopen	Memory map open FID file (C)
mftrace	Move FID trace (C)

mfclose Close memory map FID (C)

Description	Closes experiment source and destination FID files that have been
	explicitly opened with mfopen.
See also	User Programming
Polatod	m fb]] t Move FID block (C)

Related	MLDIK	Move FID block (C)
	mfdata	Move FID data (C)
	mfopen	Memory map open FID file (C)
	mftrace	Move FID trace (C)
	rfblk	Reverse FID block (C)
	rfdata	Reverse FID data (C)
	rftrace	Reverse FID trace (C)

mfdata Move FID data (C)

Syntax	<pre>mfdata(<src_expno,>src_blk_no,src_start_loc, \ dest_expno,dest_blk_no,dest_start_loc,num_points)</src_expno,></pre>
Description	Copies data specified by src_start_loc from a FID block specified by src_blk_no to a destination location specified by dest_expno, dest_blk_no, and dest_start_lo, using memory-mapped input and output. The data point locations and the num_points to be copied are specified by data points corresponding to the np parameter, not bytes or complex points.
	mfdata searches for the source and destination FID file in th directory \$vnmruser/expN/acqfil, where N is the requested experiment number or the current experiment number. If the FID file is not open, mfdata opens the file, copies the data, and closes the file. If a number of blocks need to be copied, explicitly opening and closing the files with the commands mfopen and mfclose can significantly speed up the data reformatting process.
	Be aware that mfdata can modify data returned to an experiment with the rt command. To avoid modification, enter the following sequence of VnmrJ commands before running mfdata:
	<pre>cp(curexp+'/acqfil/fid',curexp+'/acqfil/fidtmp') rm(curexp+'/acqfil/fid') mv(curexp+'/acqfil/fidtmp',curexp+'/acqfil/fid')</pre>
Arguments	src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.
	<pre>src_blk_no specifies the source block of data to be copied. Block numbers start at 1 and run from 1 to the number of blocks in a file.</pre>

src_start_loc specifies the starting data location within the specified block to copy the data. Data locations start from 0 and are specified as data points corresponding to the np parameter.

 ${\tt dest_expno}$ specifies the experiment number of the destination FID file.

dest_blk_no specifies the destination block to send the copied data.

dest_start_loc specifies the starting data destination location within the specified block to send the copied data.

Examples mfdata(1,0,2,1,(nv-1)*np,np) copies np points of data from the starting location 0 of block 1 of the current experiment to the data location (nv-1)*np of block 1 of experiment 2.

See also User Programming

Related	mfblk	Move FID block (C)
	mfclose	Memory map close FID file (C)
	mfdata	Move FID data (C)
	mfopen	Memory map open FID file (C)
	mftrace	Move FID trace (C)
	rfblk	Reverse FID block (C)
	rftrace	Reverse FID trace (C)

mfopen Memory map open FID file (C)

Syntax	<pre>mfopen<(<src_expno,>dest_expno)></src_expno,></pre>
Description	Explicitly opens experiment source and destination FID files for using memory-mapped input and output. Opening a file explicitly can significantly speed up the data reformatting process.
	<pre>mfopen searches for the FID file to be opened in the directory \$vnmruser/expN/acqfil, where N is the requested experiment number or the current experiment number. Without arguments, mfopen assumes the source and destination files are the same and are in the current experiment.</pre>
	After a file is open, the data reformatting commands mfblk, mfdata, mftrace, rfblk, rfdata, and rftrace can be used for moving around data. The mfclose must be used to close the file when data reformatting has been completed.
Arguments	src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.
	dest_expno specifies the experiment number of the destination FID file.The default is the FID file of the current experiment.
	If only one argument is provided, $mfopen$ uses that as the experiment number of the destination FID file and assumes the source is the FID file of the current experiment.
Examples	<pre>mfopen mfopen(3) mfopen(1,2)</pre>

See	also	User	Programming	γ
DCC	anou	0001	1 rogramment	1

Related	mfblk	Move FID block (C)
	mfclose	Memory map close FID file (C)
	mfdata	Move FID data (C)
	mftrace	Move FID trace (C)
	rfblk	Reverse FID block (C)
	rfdata	Reverse FID data (C)
	rftrace	Reverse FID trace (C)

mftrace Move FID trace (C)

Syntax	<pre>mftrace(<src_expno,>src_blk_no,src_trace_no, \</src_expno,></pre>			
	<pre>dest_expno,dest_blk_no,dest_trace_no)</pre>			
Description	Copies FID traces specified by src_trace_no from a FID block			

specified by src_blk_no to a destination location specified by dest_expno, dest_blk_no, and dest_trace_no, using memory-mapped input and output. If a number of blocks need to be copied, explicitly opening and closing the files with the commands mfopen and mfclose can significantly speed up the data reformatting process.

mftrace searches for the source and destination FID file in the directory \$vnmruser/expN/acqfil, where N is the requested experiment number or the current experiment number. If the FID file is not open, mftrace opens the file, copies the data, and closes the file.

mftrace cannot be used to append data to a FID file. Its purpose is for moving around data.

Be aware that mftrace can modify data returned to an experiment with the rt command. To avoid modification, enter the following sequence of VnmrJ commands before running mftrace:

cp(curexp+'/acqfil/fid',curexp+'/acqfil/fidtmp')
rm(curexp+'/acqfil/fid')
mv(curexp+'/acqfil/fidtmp',curexp+'/acqfil/fid')

Arguments src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.

src_blk_no specifies the source block of data to be copied. Block
numbers start at 1 and run to the number of blocks in a file.

src_trace_no specifies the source trace of data within the specified block to be copied. Trace numbers run from 1 to number of traces in a file.

dest_expno specifies the experiment number of the destination FID file.

dest_blk_no specifies the destination block to send the copied data.

src_trace_no specifies the destination trace of data within the specified block to be copied. Trace numbers run from 1 to the number of traces in a file. mftrace(1,1,2,1,nv) copies trace 1 from block 1 of the current Examples experiment to trace nv of block 1 of experiment 2. See also User Programming Related mfblk Move FID block (C) Memory map close FID file (C) mfclose mfdata Move FID data (C) Memory map open FID file (C) mfopen rftrace Reverse FID trace (C) Reverse FID block (C) rfblk

rfdata Reverse FID data (C)

mht Move Hadamard parameters from one workspace to another

Syntax	<pre>mht(<from_exp,> to_exp)</from_exp,></pre>		
	from_exp is the workspace number to move parameters from.		
	If not specified, the current workspace is used.		
	to_exp is the workspace number to move Hadamard		
	parameters into.		
Applicability	VnmrJ 3.1		
Description	The mht macro moves Hadamard parameters from one workspace f		
	another. It transfers the following parameters: htfrq1, htbw1, sw or		
	sw1, tof or dof.		
	mht is used in the "Move HT pars to exp" entry box in the editht		
	dialog. It may also be used from the command line.		
Arguments	htfrq1 - Hadamard frequency list in indirect dimension, in Hz from center of spectrum, or ppm.		
	htbw1 - Hadamard band width in indirect dimension, in Hz. It may be a single value or a list of values for each element in the htfrq1 list.		
	tn - nucleus used for frequency list.		
	sw - spectral width in direct dimension		
	sw1 - spectral width in 1st indirect dimension		
	tof - frequency offset in direct dimension		
	dof - frequency offset in 1st indirect dimension		
Examples			
See also	ht		
	editht		
	HsqcHT		
	tocsvHT		

minsw Reduce spectral width to minimum required (M)

- DescriptionSearches the spectrum for peaks, sets new limits accordingly, and then
calls movesw to calculate a new transmitter offset tof and spectral
width sw.See alsoNMR Spectroscopy User GuideRelatedmoveswMove spectral window according to cursors (M)
movetofMove transmitter offset (M)
 - swSpectral width in directly detected dimension (P)tofFrequency offset for transmitter offset (P)

mkchsums Make checksum(s) for a given directory or file

Syntax	<pre>mkchsums(fullpath<,checksum file name>)</pre>
Description	mkchsums generates a checksum file for a given directory or file
	(fullpath required). Checksum file contains time stamp, checksums and
	other info. Command aborts if checksum file of given name or default
	name already exists.

mkCPprotocol Make Protocol

<pre>mkCPprotocol(<name,path,tabname,time,seqfil,type,setup_macro, required experiments,menu1,menu2,dialog option)></name,path,tabname,time,seqfil,type,setup_macro, </pre>		
VnmrJ 3.1		
This utility is used to create a protocol.		
<pre>mkCPprotocol('cobalt',userdir+'/templates/vnmrj/protocols' ,'Std1d',23,'s2pul','LIB','cobalt','','exotic','nucleus ','')</pre>		
arg1 - protocol name if $\# < 1$ then arg1=pslabel		
arg2 - directory where the protocol will be written		
if \$# < 2 then		
arg2=userdir+'/templates/vnmrj/protocols		
arg3 - tabname - name of the ExperimentPanel tab		
if \$# < 3 then arg2=apptype from parlib		
arg4 - default time (real number)		
if \$# < 4 then arg2=ACQtime from parlib		
arg5 - seqfil, if $\# < 5$ then seqfil from parlib entry		
or =arg1		
arg6 - type, if \$# < 6 then \$arg6='LIB'		
arg7 – macro, if \$# < 7 then \$arg7=arg1		

```
arg8 - required exp, if $# < 8 then $arg8 is not used
arg9 - menu1, if $# < 9 then $arg9 is not used
arg10 - menu2, if $# < 10 then $arg10 is not used
arg11 - dodialog option
See also
```

mkdir Create new directory (C)

- Syntax mkdir('xxx') make xxx as a subdirectory of the current directory. mkdir('/usr2/vnmr1/fidlib'):\$res make /usr2/vnmr1/fidlib an absolute pathname.
- Description Creates a new UNIX directory. The function of the VnmrJ mkdir command is similar to the UNIX mkdir command. If the first argument is '-p', the mkdir command will make parent directories as needed. The mkdir command will return a 1 for successfully making the directory. The mkdir command will return a 0 for failing to make the directory. Examples mkdir('tests') mkdir('/home/george') See also NMR Spectroscopy User Guide

```
Related rmdir Remove directory (C)
```

mlabel Menu label (P)

Description	Stores the label for a menu button. Usually this parameter is arrayed, with one label for each button in the menu. This parameter is stored in a user's global file and is set whenever a menu is called.	
See also	User Program	nming
Related	menu mstring	Change status of menu system (C) Menu string (P)

move Move to an absolute location to start a line (C)

Syntaxmove(<'graphics'|'plotter'>, x, y)DescriptionMoves the start of a line to an absolute location with the coordinates
given as an argument. move is part of a line drawing capability that
includes the pen and draw commands. pen selects the pen number of

the plotter ('pen1', 'pen2', etc.) or the color ('red', 'green', 'blue', etc.). move sets the point from which to start drawing the line. draw draws a line from that point to the point given by the draw arguments. Refer to the description of the draw command for examples of using the line drawing capability.

Arguments 'graphics' and 'plotter' are keywords selecting output to the graphics window or a plotter device. The default is 'plotter'. The output selected is passed to subsequent pen, move, or draw commands, remaining unchanged until different output is specified.

x, y are the absolute coordinates, in mm, of a point to move to. The range of x is 0 at the left edge of the chart and wcmax at the right edge of the chart. The range of y is -20 at the bottom of the chart and wc2max at the top.

See also NMR Spectroscopy User Guide

Related	draw	Draw line from current location to another location (C)
	gin	Return current mouse position and button values (C)
	pen	Select a pen or color for drawing (C)
	wcmax	Maximum width of chart (P)
	wc2max	Maximum width of chart in second direction (P)

movedssw Set downsampling parameters for selected spectral region (M)

Description Sets the parameters dslsfrq and downsamp to appropriate values for digital filtering and downsampling in a cursor-selected spectral region. To accomplish this, Fourier transform an oversampled data set, and then run the ds program. In the resulting spectral display, enclose the desired region with the cursors, and then run movedssw.

See also NMR Spectroscopy User Guide

Related	downsamp	Downsampling factor applied after digital filtering
		(P)
	ds	Display a spectrum (C)
	dslsfrq	Bandpass filter offset for downsampling (P)

moveossw Set oversampling parameters for selected spectral region (M)

Description Sets the parameters oslsfrq and sw to appropriate values for oversampling and digital filtering in a cursor-selected spectral region. To accomplish this, acquire a data set without digital filtering, and then run the ds program. In the resulting spectral display, enclose the desired region with the cursors, and then run moveossw. The value of oversamp is manually set.

 See also
 NMR Spectroscopy User Guide

 Related
 ds
 Display a spectrum (C)

 oslsfrq
 Bandpass filter offset for oversampling (P)

 oversamp
 Oversampling factor for acquisition (P)

 sw
 Spectral width in directly detected dimension (P)

movesw Move spectral window according to cursors (M)

Syntax movesw<(width)>

Description	Uses the parameters cr and delta to calculate a new transmitter offset tof and a new spectral width sw. If referencing was used, it is also adjusted. The movesw macro also sets sp and wp to display the spectral window.		
Arguments	width specifies the spectral width sw . The default is to use a value calculated from the parameter delta.		
Examples	movesw movesw(5000)		
See also	NMR Spectroscopy User Guide		
Related	cr delta minsw movetof	Cursor position in directly detected dimension (P) Cursor difference in directly detected dimension (P) Reduce spectral width to minimum required (M) Move transmitter offset (M)	
	sp	Start of plot (P)	
	SW	Spectral width in directly detected dimension (P)	
	tof	Frequency offset for observe transmitter (P)	
	wp	Width of plot (P)	

movetof Move transmitter offset (M)

Syntax movetof<(frequency)>

Description	Moves the transmitter offset parameter tof so that the current cursor position, defined by cr, becomes the center of the spectrum. If referencing was used, movetof maintains the referencing.	
Arguments	frequency specifies the transmitter frequency rather than using the cursor position to define the frequency. This provides a convenient method of moving the transmitter frequency outside the current spectral window.	
See also	NMR Spectroscopy User Guide	
Related	cr	Cursor position in directly detected dimension (P)
	minsw	Reduce spectral width to minimum required (M)
	movesw	Move spectral window according to cursors (M)
	tof	Frequency offset for observe transmitter (P)
	LUI	riequency offset for observe transmitter (1)

Move parameters between experiments (C)

Syntax	<pre>mp(<from_exp,>to_exp)</from_exp,></pre>	
Description	Moves text and the current display, processing, and acquisition parameters from one experiment to another. No FID is transferred.	
Arguments	from_exp specifies the number of the experiment from which the parameters are to be taken; default is the parameters are moved from the currently active experiment.	
	to_exp specifies to which experiment the parameters are to be moved.	
Examples	mp(4) mp(2,3)	
See also	NMR Spectroscopy User Guide	
Related	md Move display parameters between experiments (C)	
	mf Move FIDs between experiments (C)	

mparval Moves a Paramter Value Between Experiments

Description	Moves a parameter value between experiments.
Syntax	<pre>mparval(parametername, <origin>, target)</origin></pre>
Examples	mparval('sw',2,3)
Arguments	If only two arguments are supplied, the value of the first argument is moved to the workspace defined by the second argument.
Related	mf, mp, md

mgcosy Set up parameters for MQCOSY pulse sequence (M)

Syntax	mqcosy<(level)>
Description	Sets up a multiple-quantum filtered COSY experiment.
Arguments	level is the desired quantum level of filtration.
Examples	mqcosy mqcosy(3)
See also	NMR Spectroscopy User Guide

mref		Set referencing based on a existing spectrum of the sample (M)
	Syntax	<pre>mref(<source_exp,>target_exp)<:\$ret></source_exp,></pre>
		<pre>mref(source_fid)<:\$ret></pre>
]	Description	Use a primary referenced spectrum to reference a secondary spectrum acquired in another work space (or experiment) at the same

mp

temperature, using the same lock sample, and either a different or the same nucleus without adding a secondary reference sample. The primary spectrum must be properly referenced using the IUPAC recommended Ξ values. Ξ is the normalized frequency such that the ¹H signal from TMS is 100.00 MHz.

Begin with a source_exp spectrum (typically a 1 H spectrum) and reference it using an internal reference (such as TMS, see the IUPAC recommendations).

Join a different experiment and acquire a target_exp spectrum on a different (or same) nucleus. Enter

mref(<source_exp,>target_exp).

Referencing of 2D data sets using mref only applies to the directly detected dimension. The indirect dimensions is referenced using reff1 and reff2 (after using mref or after manual referencing of the observe dimension). The reference frequency for the secondary spectrum, reffrq_b, is calculated as follows:

reffrq_b = (reffrq_a / Ξ_a) * Ξ_b

mref also corrects for possible changes in the lock frequency:

reffrq_b = (reffrq_a / lockfreq_a) * lockfreq_b

mref works if the lock frequency changed between the two acquisitions, if the two spectra were acquired on different instruments, or at different field strengths.

mref calculates rfl and rfp after calculating reffrq:

rfp = 0rfl = sw/2 - (sfrq - reffrq) * 1e6

The systemglobal parameters lockfreq and hlfreq must saved in the local parameters using the saveglobal mechanism when the go command is executed. The mref macro only tracks lock frequency changes if these systemglobal parameters are saved in the local parameters.

The mref macro works with earlier data if both data sets were:

- acquired at the same lock frequency (on the same instrument).
- the lockfreq (on a data station) and (on older instruments) hlfreq parameters are set to the values used to acquire the data.

Referencing action from mref are reported the on line 3. Suppress the report by suppling a return argument, e.g.:

\$ret='' mref('myfid.fid'):\$ret

The referencing message is captured in the return argument "\$ret" and the contents of this string variable can be used to label plots with the referencing information.

Limitations: the macro works with data recalled from an archive or acquired on an other system provided the data was acquired using VNMR6.1C or newer.

Setting the global (or local) flag bioref='y' enables Bio-NMR referencing (based on nuctables/nuctabrefBio) and disables

Μ

standard IUPAC / organic chemistry referencing (based on <code>nuctables/nuctabref</code>).

See /vnmr/nuctables/nuctabref.

Arguments source_exp - experiment containing the primary referenced spectrum or the full (or relative) path and fid file name containing the primary references spectrum.

target_exp - experiment contining spectra to be referenced based upon the primary experiment referencing.

\$ret - return argument for output of mref.

Alternatively, the name of a FID file (with or without extension) can be given as a single argument; in this case, the data in the CURRENT experiment are referenced based on the referencing in the specified FID file.

Examples mref(3) – uses the current experiment as the source and applies the reference to the specified experiment as the target.

mref(1,2) – experiment 1 is the source and experiment 2 is the target.

mref('myfid')

mref('/data/fids/myfid.fid')

RelatedsetrefSet Frequency Referencing Based on Lock Signal Shift (M)setref1Set Frequency Referencing for f1 Evolution Dimension (M)setref2Set Frequency Referencing for f2 Evolution Dimension (M)reff1Reference f1 Indirect Dimension from Observe Dimension
(M)reff2Reference f2 Indirect Dimension from Observe Dimension
(M)

bioref Flag for Bio-NMR Referencing (P)

mrev8 Set up parameters for MREV8 pulse sequence (M)

Applicability	Systems with a solids module.	
Description	Converts FLIPFLOP, BR24, or S2PUL parameter set into the MREV8 multiple-pulse line narrowing sequence.	
See also	User Guide: Solid-State NMR	
Related	br24 cylmrev flipflop s2pul	Set up parameters for BR24 pulse sequence (M) Set up parameters for cycled MREV8 pulse sequence (M) Set up parameters for FLIPFLOP pulse sequence (M) Set up parameters for standard two-pulse sequence (M)

mrfb Set the filter bandwidths for multiple receivers (P)

Applicability Systems with multiple receivers

Description	An array of fb settings to apply to individual receivers in a multiple receiver system. The first element applies to the first receiver, the second to the second receiver, and so on. If mrfb exists and is active, these settings override the setting specified by the fb parameter; otherwise, fb is used as the filter bandwidth setting for all receivers. If there are fewer elements in mrfb than there are receivers, the remaining receivers are set to the fb value.
	Note that some older multiple receiver systems do not have the hardware to provide individual receiver control. In that case, the filter setting for receiver 1 is used on receivers 1 and 2 and the setting for receiver 3 is used on receivers 3 and 4.
	Also note that mrfb is not automatically set when sw is changed. Normally, you can leave mrfb inactive and let fb be used for all receivers.

Examples mrfb=fb/3,fb/2 sets the filter bandwidth of the first receiver to fb/3, the second to fb/2, and of the rest to fb.

Related fb Filter bandwidth (P)

mrgain Set the gain for multiple receivers (P)

Applicability Systems with multiple receivers

Description An array of 'gain' settings to apply to individual receivers in a multiple receiver system. If it exists and is active, these settings override the setting specified by the 'gain' parameter; otherwise, 'gain' is used as the gain setting for all receivers. Note that not all multiple receiver systems have the hardware set up to provide individual receiver control. In that case, the gain setting for receiver 1 is used on receivers 1 and 2 and the setting for receiver 3 is used on receivers 3 and 4.

Examples mrgain=30,40,20 sets the gains of receiver 1 to 30, receiver 2 to 40 and receivers 3 and 4 to 20.

Related gain Receiver gain (P)

mspec Select multiple spectra to display (C)

Description This command can be used to select multiple spectra. Once the spectra are selected, the ds command is in mspec mode, for example., it will display the selected spectra.

Note: The mspec command itself does not display the spectra.

To exit mspec mode (i.e., return to standard ds mode), use ds(n), or mspec('clear').

To turn off/on mspec mode withtout clearing the selection, use mspec('off') and mspec('on').

In mspec mode, all spectra should have the same spans (spectral width), but each spectrum may have different resolutions (fn).

1. SELECT SPECTRA BY INDEXES OF ARRAYED DATA

The following commands select traces from arrayed spectra in current experiment.

mspec(index1, index2, index3, ...<, color1, color2, ...>) or
mspec(index1, color1, index2, color2, ...)

e.g., mspec(1,3,5,'blue') to display traces 1,3,5 in blue.

mspec(1, 'blue',2, 'cyan',3, 'yellow')

mspec('all'<, color1, color2, ...>)

select all traces in arrayed data, up to 64.

mspec('first-last:step'<,color1, color2,...>)

e.g., mspec('1-') to select all traces

mspec('65-') to select 65 to last trace

mspec('1-:2') to select every other traces

mspec('10-20:2') to select traces 10,12,14,16,18,20

If color is not specified, the default spectrum color (i.e., 'spectrum') will be used.

If fewer color(s) than spectra are specified, the color(s) will be recycled.

Colors are specified by names:

```
'red','green','blue','cyan','magenta','yellow','orange'
,'black','white','pink','gray' 'spectrum','spectrum2' to
'spectrum9','cursor','integral','threshold','scale','fi
d','imaginary','parameter'.
```

2. SELECT SPECTRA BY KEYS

A spectrum key is a name given when a fdf spectra data is loaded (see savefdfspec and aipLoadSpec commands).

The following are special keys reserved for data in current experiment:

'FID' - data in current fid buffer,

'SPEC' - data in current phasefile buffer,

'BASE' - data in current baseline buffer.

comboKey is a string to combine (add/subtract/scale) spectra, or select a trace.

For examples:

key1='SPEC' -current trace in SPEC

key2='spec' -current trace in spec

key3='SPEC-spec' -difference between current traces in SPEC and spec

key3='spec:2' - second trace in spec

key4='spec*0.5' - scale current trace in spec by half.

The following commands select spectra by keys or comboKeys (see below).

mspec(key1,key2,key3,...)

key1, key2, ... are spectrum keys or comboKeys.

Note, all arguments should be string, and are interpreted as keys or comboKeys.

mspec(n,comboKey,yoff,color)

This selects one spectrum at a time.

- Note: the comboKey has to contain ':', so it will not be confused with colorNames.
- Spectrum n is specified by a given key, and will be displayed with vertical offset yoff, in a given color. yoff is specified in millimiters, color is specified by name.

for example,

```
mspec(1,'spec:1', 0, 'cyan')
mspec(2,'baseline:1', 0, 'yellow')
mspec(3,'spec:1-baseline:1', 40, 'red')
```

This will display the first trace of a spectral data, and the first trace of a baseline data at the same vertical offset, and display the difference 40mm above.

mstat Display memory usage statistics (C)

Syntax	<pre>mstat<(program_id)></pre>
Description	Displays statistics on memory usage by programs that use the procedures allocateWithId and release.
Arguments	program_id is the program ID, usually the same name as the program. The default is to display all program IDs and associated memory statistics.
Examples	mstat mstat('proc2d')
See also	User Programming

mstring Menu string (P)

Description Stores command strings to be executed when a VnmrJ menu button is clicked. Usually the mstring parameter is arrayed, with one string for each button in the menu. The string can be any string of commands that can otherwise appear in a macro or on the command line. This parameter is stored in a user's global file and is set whenever a menu is called.

See also	User Program	nming
Related	menu	Change status of menu system (C)
	mlabel	Menu label (P)

mtune Tune probe using swept-tune graphical display (M)

Description mtune replaces qtune on the Varian NMR System and/or Linux. mtune runs in the spectra screen and uses VnmrJ panels. Enter mtune to retrieve parameters and panels.

- all parameters changeable on-the-fly (exception: tune channel for the Varian NMR System).
- one or two markers are selectable to tune at the same time.
- vertical autoscale button.
- number of acquired points changeable for better resolution at large spectral widths (more points will update less often).
- quit button returns user to current experiment and returns mtune to the original frequencies.
- See also NMR Spectroscopy User Guide

RelatedtchanRF channel number used for tuning (P)tugainAmount of receiver gain used by qtune (P)tuneAssign frequencies (C)

Move and/or rename a file (C)

Syntax	<pre>mv(from_file,to_file)</pre>	
Description	Renames and/or moves a file or directory. mv functions the same as the command rename.	
Arguments	from_file is the name of the file to be moved and/or renamed.	
	to_file is t from_file a the to_file	the new name of the file and/or the new location. If the argument has an extension such as .fid or .par, be sure a argument has the same extension.
Examples	<pre>mv('/home/vnmr1/vnmrsys/seqlib/d2pul',</pre>	
See also	NMR Spectroscopy User Guide	
Related	сору	Copy a file (C)
	ср	Copy a file (C)
	delete	Delete a file, parameter directory, or FID directory
		(C)
	rename	Move and/or rename a file (C)
	rm	Delete a file (C)

mv

mvsampglobalMoves sample global parameters

Description	Loads sample global parameters into the current workspace from the designated workspace.
Syntax	mvsampglobal(origin)
Examples	mvsampglobal(3)
Related	getsampglobal, resetsampglobal, savesampglobal, mvsampglobal, showsampglobal

mxconst Maximum scaling constant (P)

Description Before the start of data acquisition, noise is sampled to determine the number of bits of noise present. This number is used to set the maximum number of scaling operations on the data that can occur (essentially relevant only if dp='n'). mxconst is used to adjust this amount of scaling.

Increasing mxconst to 1, for example, permits additional scaling operations, allowing acquisition to proceed slightly longer in single-precision mode. Decreasing mxconst to -1 allows fewer scaling operations before reaching the message "maximum transients accumulated".

One special case exists. If mxconst is set to less than -90 and single-precision acquisition is used (dp='n'), then scaling of the data is disabled. In this mode, reports of data overflowing the 16 bits is also disabled.

mxconst does not exist in standard parameter sets. If it does not exist, its value defaults to 0. To modify mxconst, first create it by entering create('mxconst','integer') and then enter the desired value.

CAUTION: Do not change mxconst unless you are fully aware of the consequences.

See also NMR Spectroscopy User Guide Related create Create new parameter in a parameter tree (C) dp Double precision (P)

mz

Move Integral Reset Points to specified experiment

Syntaxmz (<from, > to)ApplicabilityVnmrJ 3.1Descriptionmz takes the same arguments as mf. It only moves the integral reset
points (lifrq and liamp parameters) from one experiment to another.



Agilent VnmrJ 4 Command and Parameter Reference Guide

n1,n2,n3	Name storage for macros (P)
ncomp	The number of components to be used in discrete DOSY fitting (P)
newexp	Create a new VNMR experiment
newmenu	Select a menu without immediate activation (C)
newshm	Interactively create a shim method with options (M)
nextexp	Value of Next Experiment
nextlocQ	Next Available Location
nextpl	Display the next 3D plane (M)
nfni	Number of increments in 1st indirectly detected dimension (P)
ni2	Number of increments in 2nd indirectly detected dimension (P)
ni3	Number of increments in 3rd indirectly detected dimension (P)
niter	Number of iterations (P)
nimax	Maximum limit of ni (P)
nl	Position cursor at the nearest line (C)
nli	Find integral values (C)
nlivast	Produces a text file of integral regions without a sum region (M)
nlivast2	Produces a text file with normalized integral regions (M)
nlivast3	Produces a text file with normalized integral regions (M)
nll	Find line frequencies and intensities (C)
nlni	Find normalized integral values
nm	Select normalized intensity mode (C)
nml	Returns the current transmitter corresponding to the nucleus in argument 1



nm2d	Select Automatic 2D normalization (M)
Noesy	Convert the parameter to a NOESY experiment (M)
Noesyld	Convert the parameter set to a Noesy1d experiment (M)
noise	Measure noise level of FID (C)
noisemult	Control noise multiplier for automatic 2D processing (M)
noislm	Limit noise in spectrum (M)
notebook	Notebook name (P)
np	Number of data points (P)
npoint	Number of points for fp peak search (P)
nrecords	Determine number of lines in a file (M)
nt	Number of transients (P)
ntrig	Number of trigger signals to wait before acquisition (P)
ntype3d	Specify whether $f_1 \mbox{ or } f_2 \mbox{ display expected to be N-type (P)}$
nuctable	Display VNMR style nucleus table for a given H1 frequency (M)
nugcal	A parameter array containing calibration information from calibration of non-uniform field gradients (P)
nugcal_	A parameter array containing calibration information from calibration of non-uniform field gradients (P)
nugcalib	The nugcalib macro calculates the probe/pulse sequence specific coefficients from an experiment designed to map the non-uniformity (NUG) of the pulsed field gradients (M)
nugflag	Tells the macro dosy to use processing with correction for non-uniform field gradients (P)
numrcvrs	Number of receivers in the system (P)
numreg	Return the number of regions in a spectrum (C)
numrfch	Number of rf channels (P)

n1, n2, n3 Name storage for macros (P)

Description	Stores arbitrary character strings for macros. Each experiment has these three string parameters available.	
See also	User Program	nming
Related	dgs	Display group of special/automation parameters (M)
	r1-r7	Real value storage for macros (P)

ncomp The number of components to be used in discrete DOSY fitting

Syntax	ncomp
Applicability	VnmrJ 3.1
Description	ncomp determines the number of components to be used in fitting the signal decay in DOSY when the parameter <code>dosyproc='discrete'</code> .
Arguments	ncomp should be an integer >0
See also	dosy

newexp Create a new VNMR experiment (M)

Syntax	newexp<:\$num>
Applicability	VnmrJ 3.1
Description	newexp creates a new VNMR experiment which is used as a temporary work space and can hold a complete 1D, 2D, or 3D data set. The newexp macro will copy the "current" and "processed" parameter trees to the newly created experiment's curpar and procpar files. If the global 'newexpdir' parameter exists and is not the null string ("), and its value is the path name of an existing directory, the new experiment will be created in that directory. The newexp macro will return the number of the experiment it created.
Arguments	There are no arguments for newexp.
Examples	newexp newexp:\$expnum

newmenu Select a menu without immediate activation (C)

Syntax	(1) newmenu(menu_name)(2) newmenu:\$current_menu
Description	Selects a menu but does not activate it (syntax 1). This is most useful when picking which menu will be active when an interactive command exits. newmenu can also return the name of the currently active menu (syntax 2).
Arguments	<code>menu_name</code> is the name of the file controlling the menu selected. For example, the command string <code>newmenu('manipulate_1D')</code> ds causes the menu controlled by <code>manipulate_1D</code> to be displayed when the Return button in the ds menu is selected.
Examples	<pre>\$current_menu returns the file name of the currently active menu. newmenu('display_1D') newmenu:\$name1</pre>

See also	User Programming	
Related	menu	Change status of menu system (C)
	menuvi	Edit a menu with the vi text editor (M)

newshm Interactively create a shim method with options (M)

Svntax	newshm		
Description	Interactively creates a <i>method</i> string to be used in autoshimming of the magnetic field homogeneity. The string may consist of a series of shimming operations. The command dshim('method') describes method strings. Any text editor may be used to make and modify the strings.		
	newshm provides for either lock shimming or FID shimming, permitting the user to choose whichever is best. Lock shimming is much faster, but FID shimming is frequently much more effective in improving the field. With FID shimming, the FID evaluation range limits are requested. The full range is 0 to 100. Sensitivity to higher order gradients is greatly increased by setting the finish limit to about 5 or 10 with the start limit at 0.		
	newshm begins by asking for the name of the user's new shim method. If the non-spin (transverse) controls are chosen for adjustment, the spinner is turned off; otherwise, it is turned on. If uncertain about the shim criteria, the "medium to medium" choice is suitable in most circumstances. The new method is found in curexp+'//shimmethods.		
	To shim after running newshm, type method='methodname' and then enter shim or set the wshim parameter to shim before the start of acquisition. 'methodname' is the name supplied to newshm. For more information on shimming, see the manual NMR Spectroscopy User Guide.		
	Compared to stdshm, the newshm macro is more flexible and provides for a shimming time and FID evaluation limits supplied by the user. The primary difference between the macros is that stdshm provides for determining an estimated shimming time for the selected shim controls. When no time limit is supplied, autoshim continues until the exit criteria is met or the number of cycles reaches a limit.		
See also	NMR Spectroscopy User Guide		
Related	curexpCurrent experiment directory (P)dshimDisplay a shim method string (M)methodAutoshim method (P)shimSubmit an Autoshim experiment to acquisition (C)stdshmInteractively create a shim method (M)wshimConditions when shimming is performed (P)		
	vi Edit text file with vi text editor (C)		

nextexp Value of Next Experiment

Description This macro returns the value of the next highest workspace that does not exist. Syntax nextexp Examples nextexp:\$next_open_exp

nextlocQ Next Available Location

Description This utility returns the next open location in an automation tray. Syntax nextlocQ Related showtray, hidetray

nextpl Display the next 3D plane (M)

Syntax nextpl

Description Displays the 2D color map of the next 3D plane in the set of planes defined by the parameters plane and path3d. If nextpl immediately follows the command dproj, nextpl results in the display of the first 3D plane within that specified set and is therefore equivalent to the command dplane(1). For example, if dplane(40) has just been executed, nextpl results in the display of 3D plane 41 of that set. The nextpl macro is more efficient than dplane or dproj because the 3D parameter set (procpar3d) is not loaded into VnmrJ—it is assumed to have already been loaded by dplane or dproj, for example.

See also NMR Spectroscopy User Guide

Related	dplane	Display a 3D plane (M)
	dproj	Display a 3D plane projection (M)
	dsplanes	Display a series of 3D planes (M)
	getplane	Extract planes from a 3D spectral data set (M)
	path3d	Path to currently displayed 2D planes from a 3D data set
		(P)
	plane	Currently displayed 3D plane type (P)
	plplanes	Plot a series of 3D planes (M)
	prevpl	Display the previous 3D plane (M)

nfni Number of increments in 1st indirectly detected dimension (P)

Description Number of increments of the evolution time d2, and thus the number of FIDs that will comprise the first indirectly detected dimension of a multidimensional data set. To create parameters ni, phase, and sw1 to acquire a 2D data set in the current experiment, enter addpar('2d').

- Values 8 is minimum; typical values range from 32 to 512. In microimaging, ni greater than 0 is the imaging mode and ni equal to 0 is the projection mode.
- See also NMR Spectroscopy User Guide; VnmrJ Imaging NMR

Related addpar Add selected parameters to the current experiment (M) celem Completed FID elements (P)

> d2 Incremented delay in 1st indirectly detected dimension (P) ni2 Number of increments in 2nd indirectly detected dimension (P)

ni2 Number of increments in 2nd indirectly detected dimension (P)

Description Number of increments of the evolution time d3, and thus the number of FIDs that will comprise the second indirectly detected dimension of a multidimensional data set. To create parameters d3, ni2, phase2, and sw2 to acquire a 3D data set in the current experiment, enter addpar('3d').

See also NMR Spectroscopy User Guide

Related addpar Add selected parameters to the current experiment (M)

- d3 Incremented delay in 2nd indirectly detected dimension (P)
 ni Number of increments in 1st indirectly detected dimension (P)
 - par3d Create 3D acquisition, processing, and display parameters (M)
 - phase2 Phase selection for 3D acquisition (P)
 - sw2 Spectral width in 2nd indirectly detected dimension (P)

ni3 Number of increments in 3rd indirectly detected dimension (P)

Description Number of increments of the evolution time d4, and thus the number of FIDs that will comprise the third indirectly detected dimension of a multidimensional data set. To create parameters d4, ni3, phase3.

and sw3 to acquire a 4D data set in the current experiment, enter addpar('4d').

See also	NMR Spectroscopy User Guide		
Related	addpar Add selected parameters to the current experiment (M)		
	d4	Incremented delay in 3rd indirectly detected dimension (P)	
	ni	Number of increments in 1st indirectly detected dimension	
	(P)		
	ni2	Number of increments in 2nd indirectly detected dimension	
		(P)	
	par4d Create 4D acquisition parameters (M)		
	phase3 Phase selection for 4D acquisition (P)		
	sw3 Spectral width in 3rd indirectly detected dimension (P)		

niter Number of iterations (P)

Description	Sets the maximum number of iterations in an iterative simulation.
Values	1 to 9999. The value is initialized to 20 if the Set Params button is
	used in setting up spin simulation parameters.
See also	NMR Spectroscopy User Guide

nimax Maximum limit of ni (P)

Description	Maximum limit of ni. Used to prevent running an unrealistic number of Hadamard-encoded experiments.		
Values	Any positive real integer.		
See also	NMR Spectroscopy User Guide		
Related	sethtfrq1 Set a Hadamard frequency list from a line list (M)		
	ni	Number of increments in 1st indirectly detected	
		dimension (P)	
	htfrq1	Hadamard frequency in ni (P)	

nl Position cursor at the nearest line (C)

Syntax	nl<:height<,frequency>>		
Description	Moves the cursor to the nearest calculated line position.		
Arguments	height is a return value set to the height of the line.		
	frequency is a return value set to the frequency of the line.		
Examples	nl		
	nl:r1,r2		
See also	NMR Spectroscopy User Guide		

Ν

nli Find integral values (C)

Description	Equivalent to the dli command except that no screen display is produced. For a list of integrals, nli stores the reset points in the parameter lifrq and stores the amplitudes in the parameter liamp.	
See also	NMR Spectroscopy User Guide	
Related	CZ	Clear integral reset points (C)
	dli	Display list of integrals (C)
	dlni	Display list of normalized integrals (M)
	liamp	Amplitudes of integral reset points (P)
	lifrq	Frequencies of integral reset points (P)
	Z	Add integral reset point at cursor position (C)

nlivast Produces a text file of integral regions without a sum region (M)

Applicability	Systems with VAST accessory.
Syntax	nlivast(last)
Description	Using predefined integral regions from the spectra for each well, nlivast writes a text file, integ.out, containing the integrals of the regions. The file is written into the current experiment. Does not add an additional region that is the sum of all the defined regions for each well (see dlivast).
Arguments	last is the number of the last well. The default is 96.
See also	NMR Spectroscopy User Guide

nlivast2 Produces a text file with normalized integral regions (M)

Applicability Systems with VAST accessory.

Syntax nlivast(well)

Description Using predefined integral regions from the spectra for each well, nlivast2 writes a text file, integ.out, containing the integrals of the regions. The file is written into the current experiment. Integrals are normalized to the integral specified by the argument well. The macro nlivast2 does not add an additional region that is the sum of all the defined regions for each well (see dlivast). All of the spectra are integrated.

Arguments well is the number of the reference sample well. The default reference is well 96.

See also NMR Spectroscopy User Guide

nlivast3 Produces a text file with normalized integral regions (M)

Applicability Systems with VAST accessory.

Syntax nlivast(well)

Description Using predefined integral regions from the spectra for each well, nlivast3 writes a text file, integ.out, containing the integrals of the regions. The file is written into the current experiment. Integrals are referenced to the integral specified by the argument well. The integral of spectrum from the sample specified by well is set to 1000. The macro nlivast3 does not add an additional region that is the sum of all the defined regions for each well (see dlivast). All of the spectra are integrated.

Arguments well is the number of the reference sample well. Reference integral set to 1000. The default reference is well 96.

See also NMR Spectroscopy User Guide

nll Find line frequencies and intensities (C)

Syntax nll<('pos'<,noise_mult>)><:number_lines,scale>

Description Equivalent to the command dll except that the line listing is not displayed or printed. The results of this calculation are stored in llfrg and llamp. The frequencies are stored as Hz and are not referenced to rfl and rfp. Amplitudes are stored as the actual data point value; they are not scaled by vs. Arguments 'pos' is a keyword that causes only positive lines to be listed. noise_mult is a numerical value that determines the number of noise peaks listed for broad, noisy peak. The default is 3. A smaller value results in more peaks, a larger value results in fewer peaks, and a value of 0.0 results in a line listing containing all peaks above the threshold th. Negative values of noise_mult are changed to 3. number_lines is a return argument with the number of lines in the line list. scale is a return argument with a scaling factor for line amplitudes. This scaling factor accounts for vs and whether the lines are listed in absolute intensity mode or normalized mode. Examples nll:n1 nll('pos'):pn nll(2.5),sc See also User Programming Related dll Display listed line frequencies and intensities (C) List of line amplitudes (P) llamp

List of line frequencies (P)

llfrq

nlni Find normalized integral values

ApplicabilityVnmrJ 3.1Descriptionnli is the equivalent of dli except that no screen display is produced.

nm Select normalized intensity mode (C)

- Description Selects the normalized intensity mode in which spectra are scaled so that the largest peak in the spectrum is vs mm high. The alternative is the absolute intensity mode (selected by the ai command) in which the scale is kept constant from spectrum to spectrum to allow comparison of peak heights from one spectrum to another. The modes are mutually exclusive (i.e., the system is always in either nm or ai mode). Enter aig? to show which mode is currently active.
 - See also NMR Spectroscopy User Guide

Related	ai	Select absolute intensity mode (C)
	aig	Absolute intensity group (P)
	VS	Vertical scale (P)

nm1 Returns the current transmitter corresponding to the nucleus in argument 1.

Syntax	
Applicability	VnmrJ 3.1
Description	Returns the transmitter corresponding to the nucleus in argument 1.
	on the Channels screen of the Acquisition Page. If probeConnect and
	preAmpConfig are present they are used. Otherwise if rfchannel is
	present, it is used or if rfchannel is not present the defaults are set.

nm2d Select Automatic 2D normalization (M)

Syntax nm2d<(noisemult)>

Description Sets up parameters th and vs2d automatically for a 2D contour plot and color map display. nm2d measures the highest signal in the spectrum and sets vs2d so that the highest signal is in the range of the highest color level. It then calculates the noise threshold so that the number of points above the noise threshold is between 10% and 30% of all the points. At the same time, the difference between the mean value of all the points above the threshold (peak points) and the mean value of all the points under the threshold (noise points) is maximized. This noise threshold is then multiplied by the noise multiplier.

nm2d works both with absolute-value and phase-sensitive spectra. trace can be set to 'fl' or 'f2'.

Arguments noisemult specifies the noise multiplier number that multiplies the noise threshold:

- For 1 H, 19 F and 31 P (high dynamic range nuclei), and homonuclear spectra in general, the default value is 4.
- For HMQC/HSQC type spectra, the default value is also 4 but noise multipliers of 3 to 5 are often more adequate.
- For HETCOR and 2D-INADEQUATE spectra, the default value is 2.
- For "quick & dirty" COSY spectra with lots of t1 noise and other artifacts, a value of 8 and higher may be adequate for suppressing the artifacts.
- For 2D-INADEQUATE spectra, a value below 3 is appropriate to catch signals right above the noise level.
- If the multiplied noise threshold is below th=1, vs2d is scaled up; otherwise, th is increased to the desired level.
- Minimum value is 1.5 (if a lower value is entered, the value is set to 1.5).

Examples nm2d nm2d(3)

IIIIZQ(5)

See also NMR Spectroscopy User Guide

Related dconi Interactive 2D contour display (C) noisemu Control noise multiplier for automatic 2D processing (M) lt proc2d Process 2D spectra (M) th Threshold (P) trace Mode for *n*-dimensional data display (P)

vs2d Vertical scale for 2D displays (P)

Noesy Convert the parameter to a NOESY experiment (M)

Description	Convert the parameter to a NOESY experiment.
See also	NMR Spectroscopy User Guide
Related	foldt Fold COSY-like spectrum along diagonal axis (C)

Noesy1d Convert the parameter set to a Noesy1d experiment (M)

Description Convert the parameter set to a NOESY 1D experiment.

See also	NMR Spect	roscopy User Guide
Related	Proton	Set up parameters for ¹ H experiment (M).
	sel1d	Selective 1D protocols to set up (M).

noise Measure noise level of FID (C)

Description Measures the noise level of a FID. By using pw=0 so that no real signal is accumulated, one or more transients can be acquired. The value of np must be greater than 4096. noise then performs a statistical analysis of the noise, providing noise level, dc level, etc., for each channel. The noise level measurement can be repeated at various settings of gain and various settings of fb, etc., for a full system diagnosis.

Arguments excess_noise is excess noise and is used to calculate the noise figure.

last_noise is the last measured mean square noise and is used to calculate the noise figure.

- block_number is the block number. The default is 1.
- r1 returns the real dc offset.
- r2 returns the imaginary dc offset.
- r3 returns the real rms noise.
- r4 returns the imaginary rms noise.
- r5 returns the average rms noise.
- r6 returns the percentage channel imbalance.
- r7 returns the noise figure.

See also NMR Spectroscopy User Guide

Related	ddf	Display data file in current experiment (C)
	ddff	Display FID file in current experiment (C)
	ddfp	Display phase file in current experiment (C)
	fb	Filter bandwidth (P)
	gain	Receiver gain (P)
	np	Number of data points (P)
	pw	Pulse width (P)

noisemult Control noise multiplier for automatic 2D processing (M)

Syntax noisemult<(noise_multiplier)>

Description	Predetermines the noise multiplier used by the nm2d macro when starting automatic 2D experiments. This multiplier determines the threshold level in 2D spectra.	
Arguments	noise_multiplier is a noise multiplier, the same as used in the nm2d macro. The default is 8 for homonuclear 2D spectra or 4 for other spectra.	
Examples	noisemult noisemult(10)	
See also	NMR Spectroscopy User Guide	
Related	nm2dAutomatic 2D normalization (M)proc2dProcess 2D spectra (M)	

nois1m Limit noise in spectrum (M)

Syntax	noislm<(max_noise)>
Description	Limits the noise present in a spectrum by reducing the vertical scale vs . If the noise is smaller than the noise limit, vs is left untouched. The noise limit is in single root-mean-square noise size; the peak-to-peak noise (width of the noise band) is about twice that value. The noise is determined by taking the smallest value from four 5% regions at the left end of the spectrum. Any filter cutoff at the end will decrease the apparent noise in the spectrum, and therefore increase the noise limit in the central part of the spectrum. Because of the particular algorithm used in this macro, signals at the left end of the spectrum should not affect the result of noislm.	
Arguments	max_noise is the maximum root-mean-square size, in mm, of the noise. The default is 2.	
Examples	noislm noislm(5)	
See also	NMR Spectroscopy User Guide	
Related	vs vsadj vsadjc vsadjh	Vertical scale (P) Automatic vertical scale adjustment (M) Automatic vertical scale adjustment for ¹³ C spectra (M) Automatic vertical scale adjustment for ¹ H spectra (M)

notebook Notebook name (P)

Description Specifies the notebook name of a sample, which is saved with a study.

Related cqsavestudy	Macro to save study queue parameters (M)
page	Name of page (P)
samplename	Sample name (P)
studypar	Study parameters (P)

np Number of data points (P)

- Description Sets number of data points to be acquired. Generally, np is a *dependent* parameter and is calculated automatically when sw or at is changed. If a particular number of data points is desired, np can be entered, in which case at becomes the dependent parameter and is calculated based on sw and np.
 - Values np is constrained to be a multiple of 2 (Acquisition Controller or Pulse Sequence Controller board) or a multiple of 64 (Output board). (See the acquire statement in the manual *User Programming* for a description of these boards.)
 - See also NMR Spectroscopy User Guide

at	Acquisition time (P)
dp	Double precision (P)
setlimit	Set limits of a parameter in a tree (C)
SW	Spectral width in directly detected dimension (P)
	at dp setlimit sw

npoint Number of points for fp peak search (P)

If npoint is defined in the current parameter set and has a value, it Description determines the range of data points over which the fp command searches for a maximum for each peak. To create npoint and give it a value other than the default, enter create ('npoint', 'integer') npoint=x, where x is the new value. Values 1 to fn/4. The default is 2. See also NMR Spectroscopy User Guide Related create Create new parameter in a parameter tree (C) Fourier number in directly detected dimension (P) fn fp Find peak heights (C)

nrecords Determine number of lines in a file (M)

Syntax	<pre>nrecords(file):\$number_lines</pre>		
Description	Returns the number of lines (or records) in a file.		
Arguments	file is the name of the file.		
	<pre>\$number_lines returns the number of lines in the named file.</pre>		
Examples	<pre>nrecords(userdir+'/mark1d.out'):\$num</pre>		
See also	User Programming		

nt Number of transients (P)

Description	Sets the number of transients to be acquired (i.e., the number of repetitions or scans performed to make up the experiment or FID).
Values	1 to 1e9. For an indefinite acquisition, set nt to a very large number such as 1e9.

See also NMR Spectroscopy User Guide; VnmrJ Imaging NMR

ntrig Number of trigger signals to wait before acquisition (P)

Applic	cability	Systems	with LC-NMR	accessory.	
D	• • •	a	1	• • •	

Description Sets the number of trigger signals from the LC to wait for on the external gate line before beginning acquisition. If ntrig is 0 or the parameter does not exist, the external gate signal is ignored. If ntrig noes not exist, the parlc macro can create it. ntrig is not normally entered by the user.

See also NMR Spectroscopy User Guide

Related parlc Create LC-NMR parameters (M)

ntype3d Specify whether f_1 or f_2 display expected to be N-type (P)

Description	Indicates w is, opposite processing	whether the f_1 or f_2 display is expected to be N-type, that to the sense of precession defined by f_3 , under normal 3D conditions.		
Values	'yn' speci normal 3D	' specifies that f_1 is expected to have an N-type display under nal 3D processing conditions.		
	'ny' speci normal 3D	$n_{y'}$ specifies that f_2 is expected to have an N-type display under ormal 3D processing conditions.		
	'yy' speci under norr changes the portion of	fies that both f_1 and f_2 are expected to have N-type displays nal 3D processing conditions. Setting ntype3d ='yy' e sense of precession in f_1 and f_2 by negating the imaginary the t_1 and t_2 interferograms prior to Fourier transformation.		
See also	NMR Spectroscopy User Guide			
Related	fiddc3d	3D time-domain dc correction (P)		
	ft3d	Perform a 3D Fourier transform on a 3D FID data set (M,U)		
	ptspec3d	Region-selective 3D processing (P)		
	specdc3d	3D spectral drift correction (P)		
	ssfilter	Full bandwidth of digital filter to yield a filtered FID (P)		
	ssorder	Order of polynomial to fit digitally filtered FID (P)		
	rftype	Type of rf generation		

nuctable Display VNMR style nucleus table for a given H1 frequency (M)

Syntax	<pre>nuctable<(h1_freq)></pre>
Description	The VnmrJ nucleus table is a single nucleus table, /vnmr/nuctables/nuctable, which is calculated based on a proton frequency of 1000.000 MHz. nuctable can be used to reconstruct a traditional nucleus table, e.g., based on a proton frequency of 200.057 MHz, or to calculate a nucleus table for any given proton frequency.
Arguments	h1_freq (optional): proton frequency on which the calculated / displayed nucleus table will be based. Without argument, nuctable prints a nucleus table based on the proton frequency for which the current VnmrJ / VNMR installation is configured.
Examples	nuctable(200.057) nuctable:
Related	restorenuctable Calculate and (Re-)store accurate nuctable (M)

nugcalA parameter array containing calibration information from
calibration of non-uniform field gradients

Syntax Applicability	nugcal VnmrJ 3.1
Description	nugcal is a parameter array summarising the results of a calibration of non-uniform field gradients. The first value is the gradient calibration value gcal used; cl-c4 are the coefficients of a fourth order power series in the exponent of the Stejskal-Tanner equation. nugcal is a global parameter specific for a a given probe and pulse sequence. The parameter nugcal_ is a local copy that is set when a dosy experiment is run, to ensure that the correct parameters are available for subsequent processing if nugflag='y'.
See also	dosy nugcalib nugflag

nugcal_ A parameter array containing calibration information from calibration of non-uniform field gradients

Syntax	nugcal_
Applicability	VnmrJ 3.1
Description	<code>nugcal_</code> is a local copy of the parameter <code>nugcal</code> , set when a dosy
	experiment is run to ensure that the correct parameters are available
Ν

for subsequent processing if nugflag='y'. nugcal is a parameter array summarising the results of a calibration of non-uniform field gradients. The first value is the gradientcalibration value gcal used; c1-c4 are the coefficients of a fourth order power series in the exponent of the Stejskal-Tanner equation.

```
See also dosy
nugcal
nugcalib
nugflag
```

nugcalib The nugcalib macro calculates the probe/pulse sequence specific coefficients from an experiment designed to map the non-uniformity (NUG) of the pulsed field gradients.

Syntax	nugcalib
	nugcalib(calibrant,(T D),saveglobal,saveprobe)
Applicability	VnmrJ 3.1
Description	nugcalib calculates a set of four coefficients that relate the nominal gradient strength per DAC point, gcal, to the calculated diffusional signal attenuation as a function of gradient for a given probe and pulse sequence. As input, nugcalib requires:
	\bullet the calibrant used ('w' for pure water, 'd' for dilute HDO, 'o' for other;
	• the temperature (T) in Celsius if 'w' or 'd', or the diffusion coefficient (D) in units of 10**-10 m2/s if 'o';
	 decisions on whether or not to save the results in the global parameter file and/or in the current probe file.
	This information is supplied either as four arguments (see below) or by dialogue. The macro:
	• takes a set of signal profiles measured under a read gradient, performs monoexponential DOSY fitting on each point across the profile, and uses the resultant data and the known diffusion coefficient for the calibrant to obtain a map of relative gradient strength as a function of position;
	• fits this map with gradfit (C) to obtain a set of coefficients;
	• uses these coefficients to extrapolate into regions of small signal;
	• normalises the signal profile with profile_int (C);
	• takes the gradient coefficients and signal profile and uses deacy_gen to calculate the diffusional attenuation as a function of nominal gradient strength;
	• and uses powerfit (C) to fit this decay to the exponential of a power series in the Stejskal-Tanner exponent, storing the results in the array nugcal_ (and optionally in the global parameter nugcal and/or the current probe file).
Arguments	<pre>nugcalib('w',temperature,('n' 'y'),('n' 'y'))</pre>

```
nugcalib('d',temperature,('n'|'y'),('n'|'y'))
nugcalib('o',diffusion coefficient,('n'|'y'),('n'|'y'))
See also decay_gen
dosy
gcal
gcal
gcal
gcal
gradfit
nugcal
nugcal
nugcal
nugflag
powerfit
profile_int
```

nugflag Tells the macro dosy to use processing with correction for non-uniform field gradients

Syntax	nugflag='y'
	nugflag='n'
Applicability	VnmrJ 3.1
Description	When nugflag='n', DOSY processing invoked by the dosy macro uses simple mono- or multi-exponential fitting to estimate diffusion coefficients by fitting to the Stejskal-Tanner equation. When nugflag='y', a modified Stejskal-Tanner equation is used in which the exponent is replace by a power series, the coefficients for which are stored in the array nugcal. Correction for non-uniform gradients is available in both 2D and 3D DOSY, but only for discrete fitting (dosyproc='discrete') and not for CONTIN.
See also	nugcal
	nugcalib
	dosy
	dosyproc

numrcvrs Number of receivers in the system (P)

ApplicabilitySystems with multiple receivers.DescriptionAn integer giving the number of receivers installed in the system.
numrcvrs is set from the config panel by the vnmr1 user.

numreg Return the number of regions in a spectrum (C)

Syntax	numreg:number_regions	
Description	Returns the number of regions in a spectrum previously divided by the region command, by manual means using the z command, or by the Resets button in ds. A <i>region</i> is the area between two reset points in integral mode, with every other reset point designating the start of a <i>baseline</i> region and not included in the count of regions.	
Arguments	number_regions returns the number of peak regions in the spectrum	
Examples	numreg:\$num	
See also	User Programming	
Related	ds Display	a spectrum (C)
	getreg Get free	uency limits of a specified region (C)
	region Divide s	spectrum into regions (C)
	z Add int	egral reset point at cursor position (C)

numrfch Number of rf channels (P)

Description Holds the number of rf channels available. The value is set with the Number of RF Channels label in the Spectrometer Configuration window. numrfch represents the hardware in the system. For example, if the last experiment used the second decoupler, numrfch is set to 2. The software then leaves the second decoupler on if it was on and leaves it off if it was off.

CAUTION Do not reset numrfch to eliminate the use of a channel. See the description of dn2 and dn3 for the method to disable channels.

Values	The fifth channel can only be used with the deuterium decoupler channel.	
See also	VnmrJ Installation and Administration	
Related	config	Display current configuration and possibly change it (M)
	dn2	Nucleus for the second decoupler (P)
	dn3	Nucleus for the third decoupler (P)
	dn4	Nucleus for the fourth decoupler (P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

off	Make a parameter inactive (C)
on	Make a parameter active or test its state (C)
onCancel	Specify special functions and Labels for the "Cancel Command" button
operator	Operator name (P)
operatorlogin	Sets workspace and parameters for the operator (M)
opx	Open shape definition file for Pbox (M)
oscoef	Digital filter coefficients for over sampling (P)
osfb	Digital filter bandwidth for over sampling (P)
osfilt	Over sampling filter for real-time DSP (P)
oslsfrq	Bandpass filter offset for over sampling (P)
overrange	Frequency synthesizer overrange (P)
oversamp	Over sampling factor for acquisition (P)
owner	Operating system account owner (P)

off Make a parameter inactive (C)

Syntax	off(parameter<,tree>)
Description	Turns off an active parameter in any tree.
Arguments	parameter is the name of the parameter.
	tree is type of parameter tree: 'current', 'global', 'processed', or 'systemglobal'. The default is 'current'. Refer to the create command for more information on the types of trees.
Examples	off('gf') off('n','global')



Make a parameter active or test its state (C)

Syntax on(parameter<, tree>) <: \$active>

Description Turns on an inactive parameter in any tree or tests if a parameter is active. Real variables (not strings) can be turned on and off. This can be done in any tree with the commands on and off, and by entering name='y' or name='n' to change the active flag for variables in the current tree only. The variable trees are 'current', 'global', 'processed' and 'systemglobal'. The default tree is 'current'. To test the active flag of a variable, use on(...):\$x. This does not change the active flag of the variable, but sets \$x to 1, if the variable is active, or to 0, if it is not active. If the variable does not exist, a value of -1 is returned. Care should be taken if using the return value as a test for a conditional statement. For example, in the following fragment, on('var1'):\$e if \$e then

```
write('line3','if statement is true with value of
%d',$e)
endif
```

the write command will be executed if 'var1' is active, writing the message *if statement is true with value of 1* It will also be executed if 'var1' does not exist, writing the message *if statement is true with value of -1*.

To only execute the write command if the variable is active, use something like the following:

```
on('var1'):$e
if ($e > 0.5) then
    write('line3','var1 is active')
endif
```

Arguments parameter is the name of the parameter to make active or to test.

tree is type of parameter tree: 'current', 'global', 'processed', or 'systemglobal'. The default is 'current'. Refer to the create command for more information on the types of trees.

\$active is 1 if the parameter is active, or is 0 if it is not active. Adding a return argument makes on conduct only a test of whether the specified parameter is active and does *not* turn on the parameter if it is inactive.

on

See also	User Program	nming
Related	create	Create new parameter in a parameter tree (C)
	off	Make a parameter inactive (C)

onCancel Specify special functions and labels for the "Cancel" Command" button

Description The onCancel command can specify a shell command to execute when the Cancel Command button is clicked. It can also set the message displayed when the Cancel Command button is clicked. Finally, it can control whether the standard "Cancel Command" mechanism will be used. The onCancel command is only active for the duration of the macro execution, including the duration of any macros called by the initial macro. It can be called multiple times to set different behaviors in different sections of the macros.

The onCancel command is ignored if called from background or automation.

Arguments The first argument specifies a shell call to be made, that is, it is like an argument to the shell command. If this special function is selected, it is only invoked the first time the Cancel Command is clicked. After that, the standard Cancel Command function will be used. Using " as the first argument reselects the standard cancel mechanism.

The second argument specifies the message to be displayed if the Cancel Command button is clicked. The default is ", which means use the standard message. This message will be displayed until the standard cancel mechanism is used and then the standard message will be displayed. If the second argument is ' ', that is, a single space, then Cancel Command messages will be suppressed.

The third argument specifies the action of the standard Cancel Command mechanism. The value 'yes' selects the standard mechanism in addition to the cancel mechanism specified by the first argument. The value 'no' disables the standard mechanism. The cancel mechanism specified by the argument argument will be invoked the first time the Cancel Command is clicked and the standard mechanism will be invoked on subsequent clicks of the Cancel Command button.

The value 'off' disables the standard mechanism until the macro finishes. The default value is 'no'.

The onCancel call with no arguments resets everything to default behavior. It is equivalent to onCancel(",",'yes').

A typical example might be for a macro to look for a file in order to decide if it needs to quit. For this example, the following might be used.

```
shell('rm -f '+curexp+'/cancelMyMacro'):$e
onCancel('touch
'+curexp+'/cancelMyMacro','Cancelling myMacro')
```

```
$cancel=0
while (<some conditions>) and ($cancel = 0) do
    // Do macro stuff
    // Check for cancel
    exists(curexp+'/cancelMyMacro','file'):$e
    if ($e) then
        shell('rm -f '+curexp+'/cancelMyMacro'):$e
        $cancel = 1
        endif
endwhile
```

To suppress the Cancel Command button, the following could be used.

```
onCancel('','Critical section. Cannot cancel
command','off')
```

The first argument specifies no special cancel function. The second argument displays a message and the third argument turns off the standard cancel mechanism. Since the standard cancel mechanism is turned off, the supplied message will be displayed every time the Cancel Command button is clicked. To resume standard cancel mechanisms later in the macro, the onCancel command with no arguments, or onCancel(",",'yes'), or some other variant can be used.

Another example is when a macro calls external programs via the shell command.

```
exists('myProgA', 'bin'):$e,$pathMyProgA
if ($e) then
    exists('killMyProgA', 'bin'):$e,$killpath
    onCancel($killpath, 'Terminating myProgA')
    shell($pathMyProgA):$e
endif
exists('myProgB', 'bin'):$e,$pathMyProgB
if ($e) then
    exists('killMyProgB', 'bin'):$e,$killpath
    onCancel($killpath, 'Terminating myProgB')
    shell($pathMyProgB):$e
endif
onCancel // re-select deafult behavior
```

This assumes shell scripts killMyProgA and killMyProgB have been written to terminate the respective programs. The default third argument is 'no', so if myProgA is cancelled, the macro continues and myProgB will execute. If a 'yes' had been supplied to the first onCancel call, then if the Cancel Command button were clicked while myProgA was executing, killMyProgA would have been called and the standard cancel mechanism would have aborted the macro. Following completion of myProgB, the default cancel mechanism is selected.

To just change the message that is displayed when the Cancel Command button is clicked, use

```
onCancel('', 'Macro '+$0+' canceled', 'yes')
```

operator Operator name (P)

Applicability	VnmrJ Walkup	
Description	Specifies the operator name. It is set when an operator logs into the Walkup interface. Multiple operators may be defined for a single user using the VnmrJ Administrator interface.	
Related	acct	Writes records for operator login and logoff (M)
	operatorlogin	Sets workspace and parameters for the operator
		(M)
	vnmr_accounting	Open Accounting window (U)

operatorloginSets workspace and parameters for the operator (M)

Syntax Operatoriogin Operator email panetiev	Syntax	operatorlogin	operator	email	panelleve	1
--	--------	---------------	----------	-------	-----------	---

Description Sets the panel display level and other parameters for an operator when the operator logs in. It also clears the new sample area in the study queue, and disables the command line if the operator has insufficient privileges. An operator may be logged in from the Switch operator dialog in the Utilities menu.

Related	acct	Writes records for operator login and logoff (M)
	email	Email address (P)
	operator	Operator name (P)
	panellevel	Display level for VnmrJ interface pages (P)
	vnmr_accounting	Open Accounting window (U)

Open shape definition file for Pbox (M)

Syntax opx<(name<.ext>)>

Description	Opens the pulse shape/pattern definition input file shapelib/Pbox.inp for the Pbox software and writes the file heade	
Arguments	name is the name of the output shape file. ext is a file name extension that specifies the file type.	
Examples	opx opx('newfile.DEC')	
Related	Pbox Pulse shaping software (U)	

oscoef Digital filter coefficients for over sampling (P)

Description	Specifies number of coefficients used in the digital filter. Enter
	addpar('oversamp') to add oscoef to the current experiment if
	oscoef does not exist. addpar('oversamp') creates digital filtering
	and oversampling parameters def_osfilt, filtfile, oscoef, osfb,
	osfilt, oslsfrg, and oversamp.

Values The default is 7.5*oversamp for inline DSP (dsp='i'). A larger number of coefficients gives a filter with sharper cutoffs; a smaller number gives a filter with more gradual cutoffs. The value of oscoef does not need to be changed when oversamp is changed because oscoef is automatically adjusted by VnmrJ to give filter cutoffs that are the same regardless of the value of oversamp.

The number of coefficients for real-time DSP (dsp='r') is determined by the hardware and is not adjustable.

Related	addpar	Add selected parameters to current experiment (M)
	dsp	Type of DSP for data acquisition (P)
	filtfile	File of FIR digital filter coefficients (P)
	osfb	Digital filter bandwidth for oversampling (P)
	oslsfrq	Bandpass filter offset for oversampling (P)
	oversamp	Oversampling factor for acquisition (P)
	paros	Create additional parameters used by oversampling (M)

osfb Digital filter bandwidth for oversampling (P)

Description Specifies bandwidth of the digital filter used for oversampling. If osfb does not exist in the current experiment, enter addpar('oversamp') to add it. addpar('oversamp') creates digital filtering and oversampling parameters def_osfilt, filtfile, oscoef, osfilt, oslsfrg, and oversamp.

Values Number, in Hz. A value less than sw/2 rejects frequencies at the edges of the spectrum; a value greater than sw/2 aliases noise and signals at frequencies outside of $\pm sw/2$.

'n' sets the bandwidth to sw/2.

Related	addpar	Add selected parameters to current experiment (M)
	def_osfilt	Default value of osfilt (P)
	filtfile	File of FIR digital filter coefficients (P)
	oscoef	Digital filter coefficients for oversampling (P)
	osfilt	Oversampling filter for real-time DSP (P)
	oslsfrq	Bandpass filter offset for oversampling (P)
	oversamp	Oversampling factor for acquisition (P)
	paros	Create additional parameters used by oversampling
		(M)
	SW	Spectral width in directly detected dimension (P)

osfilt Oversampling filter for real-time DSP (P)

Applicability	Systems with real-time DSP.	
Description	Sets the type of real-time digital filter to be used on systems equipped with the real-time DSP hardware option. osfilt is normally set automatically by the software based on the user's global parameter def_osfilt, so that osfilt only needs to be changed if a particular experiment is to be run with a different digital filter than the default.	
Values	'a' or 'A' for the Analog $Plus^{\text{TM}}$ digital filter.	
	'b' or 'B' for the brickwall digital filter.	
	'' (null string) causes osfilt to be set to the value contained in the def_osfilt when an acquisition is initiated (with go, for example).	
Related	def_osfiltDefault value of osfilt (P)dspType of DSP for data acquisition (P)	

oslsfrq Bandpass filter offset for oversampling (P)

Description	Selects a ban frequency. In does not exis addpar ('ove oversampling	dpass filter that is not centered about the transmitter this way oslsfrq works much like lsfrq. If oslsfrq t in the current experiment, add it with ersamp'), which creates digital filtering and parameters, the same as the <u>paros</u> macro.
Values	Number, in H transmitter fr	z. A positive value selects a region upfield from the requency. A negative value selects a downfield region.
Related	addpar def_osfilt filtfile fsq lsfrq oscoef osfb	Add selected parameters to current experiment (M) Default value of osfilt (P) File of FIR digital filter coefficients (P) Frequency-shifted quadrature detection (P) Frequency shift of the fn spectrum in Hz (P) Digital filter coefficients for oversampling (P) Digital filter bandwidth for oversampling (P)

osfilt	Oversampling filter for real-time DSP (P)
oversamp	Oversampling factor for acquisition (P)
paros	Create additional parameters used for oversampling
	(M)

overrange Frequency synthesizer overrange (P)

Applicability Systems with optional version X46 of the PTS frequency synthesizer.

Description Configures whether an rf channel has version X46 of the PTS frequency synthesizer. The value for each channel is set using the label Frequency Overrange in the Spectrometer Configuration window.

Values Not Present, 10000 Hz, or 100000 Hz

Not Present indicates that this rf channel does not have the frequency overrange option.

10000 or 100000 indicate that this rf channel has the frequency overrange option. The **10000 Hz** or **100000 Hz** choices are determined by the letters H, J, or K found in the PTS Synthesizers model number. The normal value for overrange is 10000 Hz. If **Frequency Overrange** is set to 10000 Hz or 100000 Hz, the **Latching** value for that RF channel must also be set to **Present**. When set to either 10000 Hz or 100000 Hz, overrange guarantees a range of phase-continuous frequency jumps of at least 10 kHz or 100 kHz in each jump direction.

- See also VnmrJ Installation and Administration
 - Related config Display current configuration and possibly change it (M) latch Frequency synthesizer latching (P)

oversamp Oversampling factor for acquisition (P)

Description Specifies the oversampling factor for the acquisition. With inline digital filtering (dsp='i'), np*oversamp data points are acquired at a rate of sw*oversamp. The data is then transferred to the host computer, digitally filtered, and downsampled to give np points and a spectral width of sw.

With real-time digital filtering (dsp='r'), the oversampling, digital filtering, and down sampling all occur as each data point is collected, so that only np data points are ever stored in the acquisition computer memory and subsequently transferred to the host computer.

If oversamp does not exist in the current experiment, enter the command addpar('oversamp') to add it. addpar('oversamp') creates digital filtering and oversampling parameters def_osfilt, filtfile, oscoef, osfb, osfilt, oslsfrq, and oversamp.

If oversamp is set to a number, then that number represents the amount of oversampling to apply when collecting the data. The oversamp value is automatically calculated whenever sw is changed, provided oversamp is not set to 'n'. That is the distinction between oversamp='n' and oversamp=1. In both cases, no oversampling will be used. This occurs, for example, if the sw parameter is greater than half the maximum spectral width. However, if sw is reduced so that oversampling is possible, then if oversamp is set to 'n', oversamp will remain set to 'n' and oversampling will not occur. On the other hand, if oversamp is set to 1, then oversamp is recalculated and oversampling will occur. Therefore, the oversamp parameter accurately represents whether oversampling is performed for a data set. When oversamp is automatically determined based on a change to sw, it is set to the maximum possible oversampling factor. The value of oversamp can be manually reset.

Note that setting oversamp greater than 1 means oversampling is selected for the experiment. However, if the oversampling facility is not present in the system (i.e., dsp='n'), then the oversamp parameter is automatically reset to 1, indicating that no oversampling will be performed.

Two other experiment local parameters reflect whether DSP is used during the acquisition of a data set:

- fb is set to Not Active if DSP is used.
- oscoef reflects whether real-time (dsp='r') or inline (dsp='i') DSP was used. If real-time, oscoef is set to Not Active. If inline, oscoef is set to the value used by the inline algorithm.
- Values Number less than or equal to 68. For inline DSP, sw*oversamp and np*oversamp are limited by the values in the following table:

Maximum sw*oversamp	Maximum	np*oversamp
500 kHz 100 kHz	2M 128K	

The maximum np*oversamp is given for double precision data (dp='y'). For dp='n', multiply this value by 2.

'n' causes normal acquisition to be done without digital filtering.

Related	addpar	Add selected parameters to current experiment (M)
	<pre>def_osfilt</pre>	Default value of osfilt parameter (P)
	dp	Double precision (P)
	dsp	Type of DSP for data acquisition (P)
	fb	Filter bandwidth (P)
	filtfile	File of FIR digital filter coefficients (P)
	fsq	Frequency-shifted quadrature detection (P)
	np	Number of data points (P)
	oscoef	Digital filter coefficients for oversampling (P)
	osfb	Digital filter bandwidth for oversampling (P
	osfilt	Oversampling filter for real-time DSP (P)
	oslsfrq	Bandpass filter offset for oversampling (P)
	paros	Create additional parameters used by oversampling (M)
	SW	Spectral width in directly detected dimension (P)

owner Operating system account owner (P)

Description Set to the Unix or Linux account owner. It is set when VnmrJ is started.



Agilent VnmrJ 4 Command and Parameter Reference Guide

p1	Enter pulse width for p1 in degrees (C)
p1	First pulse width (P)
p2pul	Set up sequence for PFG testing (M)
p31	Automated phosphorus acquisition (M)
p31p	Process 1D phosphorus spectra (M)
pa	Set phase angle mode in directly detected dimension (C)
pa1	Set phase angle mode in 1st indirectly detected dimension (C)
pacosy	Plot automatic COSY analysis (C)
pad	Preacquisition delay (P)
padept	Perform adept analysis and plot resulting spectra (C)
page	Submit plot and change plotter page (C)
page	Name of page (P)
panellevel	Display level for VnmrJ interface pages (P)
pap	Plot out "all" parameters (C)
par2d	Create 2D acquisition, processing, and display parameters (M)
par3d	Create 3D acquisition, processing, and display parameters (M)
par3rf	Get display templates for 3rd rf channel parameters (M)
par4d	Create 4D acquisition parameters (M)
paramedit	Edit a parameter and its attributes with user-selected editor (C)
paramgroup	Create a set of new parameters in a workspace and optionally add a display string to the dg and ap parameters.



paramvi	Edit a parameter and its attributes with vi editor (M)
pardiff	Report differences between parameter sets (M)
pards	Create additional parameters used by down sampling (M)
parfidss	Create parameters for time-domain solvent subtraction (M)
parfix	Update parameter sets (M)
parlc	Create parameters for LC-NMR experiments (M)
parlist	List complete parameters in simple format (M)
parll2d	Create parameters for 2D peak picking (M)
parlp	Create parameters for linear prediction (M)
parmax	Parameter maximum values (P)
parmin	Parameter minimum values (P)
paros	Create additional parameters used by over sampling (M)
parside	Sets Up Parameters for Plotting Reference on Side
parstep	Parameter step size values (P)
partop	Sets Up Parameters for Plotting Reference on Top
parversion	Version of parameter set (P)
patchinstall	Install a VnmrJ patch
patchuninstall	Uninstall a VnmrJ patch
patchmake	Build a custom Vnmr patch
path3d	Path to currently displayed 2D planes from a 3D data set (P)
paxis	Plot horizontal LC axis (M)
Pbox	Pulse shaping software (U)
pbox_shapeinfo	Returns Pbox Shape Information
pbox_bw	Define excitation band (M)
pbox_bws	Define excitation band for solvent suppression (notch) pulses (M)
pbox_dmf	Extract dmf value from pbox.cal or Pbox shape file (M)
pbox_dres	Extract dres value from pbox.cal or Pbox shape file (M)

pbox_name	Extract name of last shape generated by Pbox from pbox.cal (M)
pbox_pw	Extract pulse length from pbox.cal or Pbox shape file (M)
pbox_pwr	Extract power level from Pbox.cal or Pbox shape file (M)
pbox_pwrf	Extract fine power level from pbox.cal or Pbox shape file (M)
pbox_rst	Reset temporary Pbox/Vnmr variables (M)
pboxget	Extract Pbox calibration data (M)
pboxpar	Add parameter definition to the Pbox.inp file (M)
pboxrst	Reset temporary Pbox variables (M)
pboxunits	Converts to Pbox default units (M)
pcmapapply	Apply Phase Correction Map to Data (C)
pcmapgen	Generate Phase Correction Map (C)
pcmapopen	Phase Correction Map Open (C)
pcon	Plot contours on a plotter (C)
pcss	Calculate and show proton chemical shifts spectrum (M)
peak	Find tallest peak in specified region (C)
peak2d	Return information about maximum in 2D data (C)
peakmin	Find the minimum point
pen	Select a pen or color for drawing (C)
pexpl	Plot exponential or polynomial curves (C)
pexpladd	Add another diffusion analysis to current plot (M)
pfgon	Pulsed field gradient amplifiers on/off control (P)
pfww	Plot FIDs in whitewash mode (C)
pge	Convert parameter set to PGE pulse sequence (M)
pge_calib	Calibrate gradient strengths for PGE pulse sequence (M)
pge_data	Extract data from single element of PGE pulse sequence (M)
pge_output	Output results from PGE pulse sequence (M)
pge_process	Automated processing of data from PGE pulse sequence (M)

pge_results	Calculate diffusion constant for integral region (M)
pge_setup	Set up gradient control parameters for PGE pulse sequence (M)
ph	Set phased mode in directly detected dimension (C)
ph1	Set phased mode in 1st indirectly detected dimension (C)
ph2	Set phased mode in 2nd indirectly detected dimension (C)
phase	Change frequency-independent phase rp (M)
phase	Phase selection (P)
phase1	Phase of first pulse (P)
phase2	Phase selection for 3D acquisition (P)
phase3	Phase selection for 4D acquisition (P)
phasing	Control update region during interactive phasing (P)
phfid	Zero-order phasing constant for the np FID (P)
phfid1	Zero-order phasing constant for ni interferogram (P)
phfid2	Zero-order phasing constant for ni2 interferogram (P)
Phosphorus	Set up parameters for ³¹ P experiment (M)
pi3ssbsq	Set up pi/3 shifted sinebell-squared window function (M)
pi4ssbsq	Set up pi/4 shifted sinebell-squared window function (M)
pin	Pneumatics Router Interlock ((P)
pintvast	Plot VAST Intergral Data in a stacked 1D-NMR matrix format
pir	Plot integral amplitudes below spectrum (C)
pirn	Plot normalized integral amplitudes below spectrum (M)
piv	Plot integral amplitudes below spectrum (M)
pivn	Plot normalized integral amplitudes below spectrum (M)
pl	Plot spectra (C)
pl2d	Plot 2D spectra in whitewash mode (C)

plt2Darg	Plot 2D arguments (P)
plane	Currently displayed 3D plane type (P)
plapt	Plot APT-type spectra automatically (M)
plarray	Plotting macro for arrayed 1D spectra (M)
plate_glue	Define a glue order for plotting and display (U)
plc	Plot a carbon spectrum (M)
pLCNMR	Plot all forms of LC-NMR data
plcosy	Plot COSY- and NOESY-type spectra automatically (M)
pldept	Plot DEPT data, edited or unedited (M)
plexpinfo	Plots Experiment Information
plfid	Plot FIDs (C)
plfit	Plot deconvolution analysis (M)
plgrid	Plot a grid on a 2D plot (M)
plh	Plot proton spectrum (M)
plhet2dj	Plot heteronuclear J-resolved 2D spectra automatically (M)
plhom2dj	Plot homonuclear J-resolved 2D spectra automatically (M)
plhxcor	Plot X,H-correlation 2D spectrum (M)
pll	Plot a line list (M)
pllogo	Plots Logo
pll2d	Plot results of 2D peak picking (C)
Plock	Sets Protection Bit for a Parameter
plockport	Port number to use to lock out multiple ProTune processes (P)
plot	Automatically plot spectra (M)
plot1d	Plotting macro for simple (non-arrayed) 1D spectra (M)
plot2D	Plot 2D spectra (M)
plotfile	Plot to a file (M)
plothiresprep	High resolution plot output preparation (M)
plotlcnmr	An LC-NMR plotting macro (M)
plotmanual	Plot manually (M)

plotlogo	Plots a logo (M)	
plotside	Plot spectrum on side (M)	
plotter	Plotter device (P)	
plottop	Plot spectrum on top (M)	
plottopside	Plot spectrum on top and side (M)	
plp	Plot phosphorus spectrum (M)	
plplanes	Plot a series of 3D planes (M)	
plt2Darg	Plot 2D arguments (P)	
pltext	Plot text file (M)	
pltmod	Plotter display mode (P)	
plvast	Plot VAST Data in a stacked 1D-NMR matrix format	
plvastget	Plot VAST spectral data in a vertical stacked plot mode	
plvast_replot	Replot VAST spectral data one spectrum per page of paper	
plvast2d	Plot VAST data in a stacked pseudo-2D format (M)	
plww	Plot spectra in whitewash mode (C)	
pmode	Processing mode for 2D data (P)	
poly0	Display mean of the data in regression.inp file (M)	
woq	Find value of a number raised to a power (C)	
powerfit	Fits the diffusional attenuation calcuated by decay_gen to the exponential of a power series in the calibration of the non-uniformity of pulsed field gradients (C)	
qq	Decoupler pulse length (P)	
ppa	Plot a parameter list in plain English (M)	
ppcal	Proton decoupler pulse calibration (M)	
ppf	Plot peak frequencies over spectrum (C)	
pph	Print pulse header (M)	
ppmm	Resolution on printers and plotters (P)	
pprofile	Plot pulse excitation profile (M)	
pps	Plot pulse sequence (C)	
prealfa	Specify a delay for longer ring down (P)	

preAmpConfig	Set the band of the preamp, high or low, connected to each transmitter channel (P)		
prep	Run prepare acquisition macro (M)		
Presat	Set up parameters for presat $^1\mathrm{H}$ experiment (M)		
prescan	Study queue prescan (P)		
presig	Preamp Signal Level Selection Parameter (parameter)		
prevpl	Display the previous 3D plane (M)		
prescan_CoilTable	Read or update the CoilTable File (M)		
prescan_tn	Return tn string for a given atomic number (M)		
printer	Printer device (P)		
printfile	Path to the print-to-file image (P)		
printformat	Format of saved-to-file image (P)		
printlayout	Layout of printed image (P)		
printoff	Stop sending text to printer and start print operation (C)		
printon	Direct text output to printer (C)		
printregion	Screen region to be printed (P)		
printsize	Size of printed image (P)		
printsend	Defines where image will print (P)		
probe	Probe type (P)		
probeConnect	Specify which nucleus can be acquired on each RF channel (P)		
Probe_edit	Edit probe for specific nucleus (U)		
probe_edit	Edit probe for specific nucleus (M)		
probe_protection	Probe protection control (P)		
proc	Type of processing on np FID (P)		
procl	Type of processing on ni interferogram (P)		
proc1d	Processing macro for simple (non-arrayed) 1D spectra (M)		
proc2	Type of processing on ni2 interferogram (P)		
proc2d	Process 2D spectra (M)		
procarray	Process arrayed 1D spectra (M)		
process	Generic automatic processing (M)		

procplot	Automatically process FIDs (M)		
profile	Set up pulse sequence for gradient calibration (M)		
profile_int	Normalise the experimental signal profile during calibration of non-uniform pulsed gradients (C)		
proj	Project 2D data (C)		
proshimhelp	Proshim help (C)		
Proton	Set up parameters for ¹ H experiment (M)		
protune	Macro to start ProTune (M)		
protune	Shell script to start ProTune operation (U)		
protunegui	Macro to start ProTune in graphical user interface (M)		
prune	Prune extra parameters from current tree (C)		
pscale	Plot scale below spectrum or FID (C)		
pseudo	Set default parameters for pseudo-echo weighting (M)		
psg	Display pulse sequence generation errors (M)		
psggen	Compile a user PSG object library (M,U)		
psgset	Set up parameters for various pulse sequences (M)		
psgupdateon	Enable update of acquisition parameters (C)		
psgupdateoff	Prevent update of acquisition parameters (C)		
pshape	Plot pulse shape or modulation pattern (M)		
pshapef	Plot the last created pulse shape (M)		
pshr	PostScript High Resolution plotting control (P)		
pslabel	Pulse sequence label (P)		
psMain	Prescan controlling macro		
pslw	PostScript Line Width control (P)		
pssl	Plot Arrayed Numbers (C)		
ptext	Print out a text file (M)		
ptspec3d	Region-selective 3D processing (P)		
ptsval	PTS frequency synthesizer value (P)		
pulseinfo	Shaped pulse information for calibration (M)		
pulsetool	RF pulse shape analysis (U)		
purge	Remove macro from memory (C)		
puttxt	Put text file into a data file (C)		

putwave	Write a wave into Pbox.inp file (M)			
pw	Enter pulse width pw in degrees (C)			
pw	Pulse width (P)			
pw90	90° pulse width (P)			
pwd	Display current working directory (C)			
pwr	Set power mode in directly detected dimension (C)			
pwr1	Set power mode in 1st indirectly detected dimension (C)			
pwr2	Set power mode in 2nd indirectly detected dimension (C)			
pwsadj	Adjust pulse interval time (M)			
pwxcal	Decoupler pulse calibration (M)			
pxbss	Bloch-Siegert shift correction during Pbox pulse generation (P)			
pxrep	Flag to set the level of Pbox reports (P)			
pxset	Assign Pbox calibration data to experimental parameters (M)			
pxshape	Generates a single-band shape file (M)			
Pxsim	Simulate Bloch profile for a shaped pulse (U)			
Pxspy	Create shape definition using Fourier coefficients (U)			
<pslabel>_plot</pslabel>	Experiment-Specific Plot Macro			
<pslabel>_process</pslabel>	Experiment-Specific Processing Macro			
<pslabel>_setup</pslabel>	Experiment-Specific Setup Macro			

p1 Enter pulse width for p1 in degrees (C)

Syntax	p1(flip_angle<,90_pulse_width>)
Description	Calculates the flip time, in μ s, given a desired flip angle and the 90° pulse. The value is entered into the pulse width parameter p1.
Arguments	flip_angle is the desired flip angle, in degrees.
	90_pulse_width is the 90° pulse, in $\mu s.$ The default is the value of parameter $pw90$ if it exists.
Examples	p1(30) p1(90,12.8)

See also	NMR Spectroscopy User Guide		
Related	ernst	Calculate the Ernst angle pulse (C)	
	p1	First pulse width (P)	
	pw90	90° pulse width (P)	

p1 First pulse width (P)

Description	Length of first pulse in the standard two-pulse sequence.		
Values	0, 0.2 μ s to 150,000 μ s, in 0.1 μ s steps 0.1 μ s to 8190 sec, smallest value possible is 0.1 μ s, finest incremen possible is 12.5 ns.		
See also	NMR Spectroscopy User Guide		
Related	p1 Enter pulse width $p1$ in degrees (C)		

plpat Shape of excitation pulse (P)

Applicability	Systems with imaging capabilities.		
Description	Specifies the shape of pulse p1 when used in imaging experiments.		
Values	'hard', 'sinc', 'gauss', 'sech', 'sine', or any shape resident i the system pulse shape library or libraries.		
See also	VnmrJ Imaging NMR		
Related	p1First pulse width (P)pwpatShape of refocusing pulse (P)		

p2pul Set up sequence for PFG testing (M)

Applicability	Systems with the pulsed field gradient (PFG) module. This sequence is
	not for NMR applications.

- Description Sets up the PFG two-pulse sequence, a system checkout sequence for PFG installation. Several modes are controlled by the cmd parameter.
 - cmd='twinkle' sequentially addresses DACs 0 through 4. On the gradient channel interface, lights become a slow binary counter.
 - cmd='pulse' makes a pulse of value gzlvl1 for a time gt1.
 - cmd='bipulse' makes a pulse of value gzlvl1 for a time gt1 followed by a pulse of value -gzlvl1 for a time gzlvl1.
 - For other modes, see the PFG installation manual.
 - $See \ also \quad Pulsed \ Field \ Gradient \ Modules \ Installation$

p31 Automated phosphorus acquisition (M)

Syntax p31<(solvent)>

Description	Prepares parameters for automatically acquiring a standard ^{31}P spectrum. The parameter wexp is set to 'procplot' for standard processing. If p31 is used as the command for automation via the enter command, then the macro au is supplied automatically and should not be entered on the MACRO line of the enter program. However, it is possible to customize the standard p31 macro on the MACRO line by following it with additional commands and parameters. For example, p31 nt=1 will use the standard p31 setup but with only one transient.			
Arguments	solvent is the name of the solvent. The default is CDC13. In automation mode, the solvent is supplied by the enter program.			
Examples	p31 p31('DMSO')			
See also	NMR Spectroscopy User Guide			
Related	auSubmit experiment to acquisition and process data (M)enterEnter sample information for automation run (C)p31pProcess 1D phosphorus spectra (M)proc1dProcessing macro for simple, non-arrayed 1D spectra (M)procplotAutomatically process FIDs (M)wexpWhen experiment completes (P)			

p31p Process 1D phosphorus spectra (M)

Syntax	p31p		
Description	Processes non-arrayed 1D ³¹ P spectra using a set of standard macros. p31p is called by the proc1d macro but can also be used directly. Fully automatic processing (up to a point where a spectrum could be plotted) is provided: Fourier transformation (using preset weighting functions), automatic phasing (aphx macro), automatic integration (integrate macro, if required only), vertical scale adjustment (vsadjc macro), avoiding excessive noise (noislm macro), threshold adjustment (thadj macro), and referencing to the TMS signal, if present (tmsref macro).		
Related	anhy	Perform and check automatic phasing (M)	
nelateu	integrate	Automatically integrate 1D spectrum (M)	
	noislm	Avoids excessive noise (M)	
	p31	Automated phosphorus acquisition (M)	
	proc1d	Automatically process non-arrayed 1D fids (M)	
	thadj	Adjust threshold (M)	
	tmsref	Reference spectrum to TMS line (M)	
	vsadjc	Adjust vertical scale for carbon spectra (M)	

Set phase angle mode in directly detected dimension (C)

Description Selects the phase angle mode by setting the parameter dmg='pa'. In the phase angle display mode, each real point in the displayed spectrum is calculated from the phase angle of the real and imaginary points comprising each respective complex data point. The phase angle also takes into account the phase parameters rp and lp.

> For 2D data, if pmode='partial' or pmode='' (two single quotes with no space in between), pa has an effect on the data prior to the second Fourier transform. If pmode='full', pa acts in concert with the commands pa1, av1, pwr1, or ph1 to yield the resultant contour display for the 2D data.

See also NMR Spectroscopy User Guide

Related Set abs. value mode in directly detected dimension (C) av Data display mode in directly detected dimension (P) ft Fourier transform 1D data (C) ft1d Fourier transform along f_2 dimension (C) ft2d Fourier transform 2D data (C) First-order phase in directly detected dimension (P) Set phase angle mode in 1st indirectly detected dimension (C) pa1 Set phased mode in directly detected dimension (C) ph Processing mode for 2D data (P) Set power mode in directly detected dimension (C) pwr Set power mode in 1st indirectly detected dimension (C) pwr1 Zero-order phase in directly detected dimension (P) rp wft Weight and Fourier transform 1D data (C) wftld Weight and Fourier transform f₂ of 2D data (M) wft2d Weight and Fourier transform 2D data (M)

Set phase angle mode in 1st indirectly detected dimension pa1 **(C)**

Description Selects the phase angle spectra display mode along the first indirectly detected dimension by setting the parameter dmg1 to the string value 'pa1'. If the parameter dmg1 does not exist, pa1 will create it and set it to 'pal'.

> In the phase angle mode, each real point in the displayed trace is calculated from the phase angle of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the phase angle uses the real-real and imaginary-real points from each respective hypercomplex data point. The phase angle also takes into account the phase parameters rp1 and lp1.

> The pal command is only needed if mixed-mode display is desired. If the parameter dmg1 does not exist or is set to the null string, the display mode along the first indirectly detected dimension defaults to

pa

the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of pal is the same as for traces provided that pmode='partial' or pmode=''.

See also NMR Spectroscopy User Guides

Relatedav1Set abs. value mode in 1st indirectly detected dimension (C)dmg1Data display mode in 1st indirectly detected dimension (P)lp1First-order phase in 1st indirectly detected dimension (P)paSet phase angle mode in directly detected dimension (C)ph1Set phased mode in 1st indirectly detected dimension (C)

pmode Processing mode for 2D data (P)

pwr1 Set power mode in 1st indirectly detected dimension (C)

rp1 Zero-order phase in 1st indirectly detected dimension (P)

pacosy Plot automatic COSY analysis (C)

Description Automatically analyzes and plots a COSY data set with fn=fn1 and sw=sw1. Symmetrization of the data with the command foldt is recommended, but not required. First, select a proper threshold and perform a 2D line listing with the command 112d. Next, plot the 2D data with the contour plot command pcon; leaving enough room at the left side of the plot for the connectivity table. Then, pacosy will analyze the data and plot the connectivities on the plotter. pacosy gets its input from the file 112d.out in the current experiment directory. The command acosy performs the same analysis and displays the connectivities on the screen.

See also	NMR	Spectroscopy	User	Guide
----------	-----	--------------	------	-------

Related	acosy	Automatic analysis of COSY data (C)			
	fn	Fourier number in directly detected dimension (P)			
	fn1	Fourier number in 1st indirectly detected dimension (P)			
	foldt	Fold COSY-like spectrum along diagonal axis (C)			
	hcosy Automated proton and COSY acquisition (M)				
	112d	Automatic and interactive 2D peak picking (C)			
	pcon	Plot contours on plotter (C)			
	relayh	Set up parameters for COSY pulse sequence (M)			
	SW	Spectral width in directly detected dimension (P)			
	sw1	Spectral width in 1st indirectly detected dimension (P)			

pad Preacquisition delay (P)

Description Each NMR experiment starts with a single delay time equal to pad over and above the delay d1 that occurs before each transient. Normally, pad is set to a small, nominal time (0.5 seconds) to allow any hardware changes that may be required at the start of the

	acquisition to "settle in." During experiments in which the temperature			
	is changed, the acquisition starts pad seconds after the temperature			
	regulation system comes to regulation. Since the sample temperature			
	does not actually come to equilibrium for some time after that, it is			
	generally desirable to increase pad to perhaps 300 seconds. This is			
	especially true when running experiments involving arrays of			
	temperatures. The pad parameter is most useful for running kinetics			
	experiments. For example, pad=0, 3600, 3600, 3600, 3600 will run an			
	experiment immediately when go is typed (pad=0), then wait an hour			
	(3600 seconds), run the second experiment, etc.			
Values	0.0.1 us to 8190 sec in 12.5 ns steps			
	0.0.2 us to 150.000 sec in 0.1 us steps.			
Soo also	NMR Snactroscony User Guide: Unmr I Waltun			
See also	Mill Specifoscopy Oser Guide, Vitility Walkap			
Related	d1 First delay (P)			
	go Submit experiment to acquisition (C)			

padept Perform adept analysis and plot resulting spectra (C)

Syntax	padept<(<'noll'><,'coef'><,'theory'>)>				
Description	Performs the adept analysis and plots the resulting spectra with a scale and the assigned line listing. Leave enough space at the left end of the display for the line list.				
Arguments	The following	g arguments can be supplied in any order:			
	'noll' is a	keyword that specifies no line listing.			
	'coef' is a keyword that causes the combination coefficients to be printed.				
	'theory' is a keyword that causes the theoretical coefficients rather than optimized coefficients to be used.				
Examples	<pre>padept('noll','coef')</pre>				
See also	NMR Spectroscopy User Guide				
Related	adept	Automatic DEPT analysis and spectrum editing (C)			
	autodept	Automated complete analysis of DEPT data (M)			
	cdept	Automated carbon and DEPT acquisition (C)			
	Dept	Set up parameters for DEPT experiment			
	deptproc	Process DEPT data (M)			
	hcdept	Automated proton, carbon, and DEPT acquisition (C)			
	pldept	Plot DEPT data, edited or unedited (M)			

page

Submit plot and change plotter page (C)

Syntax page<(number_pages<, 'clear' | file>)>

Description	Submits the current plotter file, which has been created by all previous plotter commands, and changes the paper after the plot has been completed. Actual plotting is controlled by the vnmrplot script in the bin subdirectory of the system directory. The page command can also clear the current plotter file or save the data to a specified file name.				
Arguments	number_pages is the number of pages to move the plotter forward. The default is 1. If number_pages is 0, page submits the plot but does not change the paper.				
	'clear' is a keyword to clear the plot made thus far; that is, clear the data in the current plotter file.				
	file is the name of a file to save the plot for import into a document. If the file already exists, it is overwritten.				
Examples	<pre>page page(0) page('clear') page('myplotfile')</pre>				
See also	NMR Spectroscopy User Guide				
Related	vnmrplot Plot files (U)				

page Name of page (P)

Description	Specifies the page of a sample. It is saved with a study.			
Related	cqsavestudy	Macro to save study queue parameters (M)		
	notebook	Notebook name (P)		
	samplename	Sample name (P)		
	studypar	Study parameters (P)		

panellevel Display level for VnmrJ interface pages (P)

Description Determines which VnmrJ interface pages are available under the tabs in the parameter page area. The higher the number, the more pages are available. The only time panellevel is changed is during the login process of an operator in the Walkup interface. For the Walkup interface, the value is set by the VnmrJ Administrator (default is 10).

Values 0-9 – shows the minimum number of pages.

No shim, lock, or processing, and minimal parameter control is available. This may be used for routine automation users.

10-29 – typical for a basic Walkup user.

Shim and lock are available only if there is a sample changer. Basic processing is available. Pages are not fully populated, allowing control of a few basic parameters.

30-100 – typical for the system owner.

All pages are available and fully populated. See also VnmrJ Installation and Administration Related operator Operator name (P) operatorlogin Sets workspace and parameters for the operator (M)

pap Plot out "all" parameters (C)

Syntax pap<(<template><, x><, y><, character_size>)> Description Plots a parameter list containing "all" parameter names and values. Arguments template is the name of a template that controls the display. The default is the string parameter ap, which can be modified using paramvi('ap'). See the manual User Programming for rules on building a template. x is the starting position in the x direction of the plot on the paper, in mm. The default is a preset value. y is the starting position in the y direction of the plot on the paper, in mm. If y is specified, the x position must be also. The default is a preset value. character_size is the character size of the list and is specified as a multiplier. The default is 0.70 (not available on all plotters or printers acting as plotters). Examples pap pap(wcmax-40) $pap(10, wc2max^*.9)$ pap('newpap',wcmax-50,100,1.4) See also NMR Spectroscopy User Guide, User Programming Related Print out "all" parameters (C) ap "All" parameters display control (P) ap Plot parameters on special preprinted chart paper (C) paramvi Edit a variable and its attributes using vi text editor (M) Plot a parameter list in "English" (M) ppa

par2d Create 2D acquisition, processing, and display parameters (M)

Description Creates the acquisition parameters ni, sw1, and phase, which can be used to acquire a 2D data set. par2d also creates any missing processing and display parameters for the ni (or second) dimension, including flcoef, reffrq1, refpos1, and refsource1. The par2d macro is functionally the same as addpar('2d').

See also	NMR Spectroscopy User Guide					
Related	addpar	Add selected parameters to the current experiment (M)				
	flcoef	Coefficient to construct F1 interferogram (P)				
	ni	Number of increments in 1st indirectly detected				
		dimension (P)				
	phase	Phase selection (P)				
	reffrq1	Reference frequency of reference line in 1st indirect				
		dimension (P)				
	refpos1	Position of reference line in 1st indirect dimension (P)				
	refsource1	Center frequency in 1st indirect dimension (P)				
	set2d	General setup for 2D experiments (M)				
	sw1	Spectral width in 1st indirectly detected dimension (P)				

par3d Create 3D acquisition, processing, and display parameters (M)

Description Creates the acquisition parameters ni2, sw2, d3, and phase2 that can be used to acquire a 3D data set. par3d also creates any missing processing or display parameters for the ni2 (or third) dimension, including f2coef, fiddc3d, specdc3d, and ptspec3d. The par3d macro is functionally the same as addpar('3d').

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to the current experiment (M)
	d3	Incremented delay in 2nd indirectly detected dimension
		(P)
	f2coef	Coefficient to construct F2 interferogram (P)
	fiddc3d	3D time-domain dc correction (P)
	ni2	Number of increments in 2nd indirectly detected
		dimension (P)
	phase2	Phase selection for 3D acquisition (P)
	ptspec3d	Region-selective 3D processing (P)
	specdc3d	3D spectral drift correction (P)
	sw2	Spectral width in 2nd indirectly detected dimension (P)

par3rf Get display templates for 3rd rf channel parameters (M)

Applicability Systems with a second decoupler.

Description Retrieves the dg2 and modified ap display templates from the parameter set s2pul3rf in the system parlib directory. These two templates support the display of second decoupler acquisition parameters and 3D acquisition and processing parameters.

See also	User Progr	amming
Related	ap	"All" parameters display control (P)
	dg2	Control dg2 parameter group display (P)

par4d Create 4D acquisition parameters (M)

Applicability	Systems with a third decoupler.				
Description	Creates the acquisition parameters ni3, sw3, d4, and phase3 that can be used to acquire a 4D data set. The par4d macro is functionally the same as addpar('4d').				
See also	NMR Spe	NMR Spectroscopy User Guide			
Related	addpar Add selected parameters to the current experiment (M)				
	d4	Incremented delay for 3rd indirectly detected dimension (P)			
	ni3 Number of increments in 3rd indirectly detected dimension				
	(P)				
	phase3	phase3 Phase selection for 4D acquisition (P)			
	sw3	Spectral width in 3rd indirectly detected dimension (P)			

paramedit Edit a parameter and its attributes with user-selected editor (C)

Syntax	<pre>paramedit(parameter<,tree>)</pre>
Description	Opens a parameter file for editing with a user-selected text editor. The default editor is vi. If vi is used as the editor, paramedit is functionally the same as the paramvi command. To select another editor, set the UNIX environmental variable vnmreditor to the editor name (change .login line setenv vnmreditor old_editor to become setenv vnmreditor new_editor (e.g., setenv vnmreditor emacs) and make sure a script with the prefix vnmr_followed by the name of the editor is placed in the bin subdirectory of the system directory (e.g., vnmr_emacs). The script file makes adjustments for the type of graphic interface in use.
	Scripts in the software release include vnmr_vi and vnmr_textedit. To create other scripts, refer to the vnmr_vi script for non-window editor interfaces and to vnmr_textedit for window-based editor interfaces. The vnmreditor variable must be set before starting VnmrJ.
Arguments	parameter is the name of the parameter file to be edited.
	tree is a keyword for one of the parameter trees 'current', 'global', or 'processed'. The default is 'current'.
Examples	paramedit('ap') paramedit('b','global')

See also	NMR Spectro	scopy User Guide; User Programming
Related	paramvi	Edit a parameter and its attributes with vi editor
		(M)
	vi	Edit text file with the vi text editor (C)

paramgroup Create a set of new parameters in a workspace and optionally add a display string to the dg and ap parameters.

Syntax Syntax1:

paramgroup('all','Mytitle','',''<<<,'pt1','amplitude',2 000><<,'pt2','pulse',4.0>,(..etc)>)

Create a group of parameters pt1, pt2 .. etc with types 'amplitude' 'pulse' ... etc and display them in dg and ap with title Mytitle. If no parameters are present only the title is displayed.

Syntax Syntax2:

paramgroup('all','Mytitle','','suffixH'<<<,'pt1','ampli tude',2000><<,'pt2','pulse',4.0>,(..etc)>)

Create a group of parameters pt1Hsuffix, pt2Hsuffix .. etc as above and display them in dg and ap with title Mytitle (SolidsPack Convention). Syntax Syntax3:

paramgroup('all','Mytitle','(mycon=1)',''<<<,'pt1','amp litude', 2000><<,'pt2','pulse',4.0>,(..etc)>)

Create a group of parameters pt1, pt2 .. etc as above and display them in dg and ap with title Mytitle with the conditional string '(mycon=1)'.

For syntax1 to syntax3, if \$1 = 'dgapstring' only the display strings in dg and ap are created. Creation of the parameters is suppressed. The values 'dgstring' and 'apstring' can be used to create dg or ap strings individually.

Syntax Syntax4:

paramgroup('params','','pt1','amplitude',2000<<,'pt2','
pulse',4.0>,(..etc)>)

Create parameters pt1, pt2...etc only. Do not create a display string in dg or ap. Parameters that require only a dg or an ap string, but not both, should be created with Syntax 4. Then use syntax1 to syntax3 with 'dgstring' or 'apstring' to create the template separately.

Syntax Syntax 5 paramgroup('dgapinit')

Initialize dg='' and ap = '' to remove exisiting dg and ap displays. Also initialize ap = '' in the 'processed' tree. Use 'apinit' and 'dginit' to initialize the displays individually.

Description Create a set of new parameters in a workspace and optionally add a display string to the dg and ap parameters. Parameters are created and given a default value, only if they do not already exist. Bit 14 is set to 1 for use with rtx for Modules and Protocols. Parameters are displayed with a title and an optional conditional string. Parameters

Ρ

can be created directly or as a prefix only (SolidsPack Convention). In the later case, argument 4 must contain a parameter-group name.

Arguments Argument 1 is the function 'all','params','dgapstring','dgstring', 'apstring', 'dgapinit', 'apinit' or 'dginit'.

Argument 2 is a title for the dg/ap display (syntax1 to syntax3 only).

Argument 3 is a conditional for the dg/ap display (syntax1 to syntax3 only).

Argument 4(2) is a a string used to label all parameters in the group. The string must be one or more upper-case "channel identifiers" following a lower case "suffix". the order of the suffix and channel identifiers are reversed an appended to the parameter names (prefixes) in the following arguments (Solidpack Convention).

Argument 5(3) is a string containing a parameter name or prefix (see 4)

Argument 6(4) is the "solids type" of a parameter. The solids type controls the VnmrJ type, the limits and the significant figures in the dg/ap display. See the following table.

Solids Type:	VJ Type:	Max	Min	Step	Figures	Clear
'amplitude'	real	4095.0	0.0	0.0(0.06248)* 3	n	
'delay'	delay	8190(s)	0.0	0.0125e-6	6	n
'frequency'	frequency	1e9	1e9	0.0	1	n
'pulse'	pulse	8192	0.0	0.0125	1	n
'string'	string	na	na	na	na	n
'flag'	flag	na	na	na	na	n
'integer'	real	1e7	1e-7	1	0	n
'idphase'	real	9(12=1,2) 0	1	0	n	
'scaler'	real	63.0	-37****	0.5****	1	n
'phase'	real	360.0	-360.0	0.0(0.00549)*	3	n
'real'	real	none	none	none	6	n
'channel'***	string	na	na	na	na	y**

* The paramgroup macro does not set a step size for 'amplitude' and 'phase' but the step is set by hardware (16 bit for DD2), (12 bit amplitude and 13 bit phase for VnmrS).

**All parameters are created with bit 14 set (for modules and protocols). All 'channel' parameter names are added to the string parameter "clearparams", which can be used to clear bit 14 after loading. Subsequent protocols need to change the values of 'channel' parameters, for example to change from direct to indirect detection.

- *** The paramgroup macro always sets the default values of 'channel' parameters, whether or not the parameter previously exists. Existing values of all other parameters are preserved.
- **** For VnmrS the lower limit and step of 'scaler' parameters are set by hardware to -16 and 1.0.

Argument 7(5) is a default value, which is set only if the parameter is newly created.

Blocks of 3 subsequent arguments (for example, 8(6),9(7) and 10(8)) are additional parameters. You can create any number of parameters within a single paramgroup call.

Programming functions

(These functions are used internally. See the argument descriptions under the individual headings. They are available, but not recommended for use in other macros.)

- 1 paramgroup('dgaptitle'... arguments) creates a title and conditional string in dg and ap (also 'aptitle' and 'dgtitle').
- 2 paramgroup('dgap' arguments) appends a string for a single parameter to dg and ap (also 'ap' and 'dg').
- 3 paramgroup('dgapend') replaces the last character (usually a comma) with a semicolon to conclude the dg and ap parameter group string (also 'apend' and 'dgend').
- 4 paramgroup('dgapnull') appends a semicolon to a string constructed from 'dgaptitle' only to conclude dg and ap strings without parameters (also 'apnull' and 'dgnull').
- 5 paramgroup('setparam' ... arguments) creates a single parameter with type and a default value. Function 5 can be used individually in a macro or on the command line.

Functions 1 to 3 must be used together to create a valid dg and ap entry.

These functions create and use the temporary parameters dgcharindex, dgarrayindex, apcharindex, and aparrayindex to keep track of the length of the dg and ap strings. The dg and ap strings can hold an arbitrary list of parameters as an array of strings of up to 1024 characters.

Function 1 determines the length of the exisiting dg or ap string and creates a new array if the existing number of characters is greater than 768. If an existing group string approaches 768 characters, the string length of a new group should not cause the total to be greater than 1024 characters. If it does, the macros will abort with an error.

Syntax	paramvi(parameter<,tree>)						
• ,•	0		c	1			

Description Opens a parameter file for editing using the UNIX vi text editor. The parameter file contains various attributes of the parameter in a format documented in the manual *User Programming*. Be sure you understand the format before modifying the parameter because if an error in the format is made, the parameter will not load. When the editor is exited, the modified parameter is reloaded into the system. Arguments parameter is the name of the parameter file to be edited.

tree is a keyword for one of the parameter trees 'current', 'global', or 'processed'. The default is 'current'.

Examples paramvi('ap')

paramvi('b','global')

See also NMR Spectroscopy User Guide, User Programming

Related	create	Create new parameter in a parameter tree (C)
	destroy	Destroy a parameter (C)
	destroygroup	Destroy parameters of a group in a tree (C)
	display	Display parameters and their attributes (C)
	fread	Read parameters from file and load them into a tree
		(C)
	fsave	Save parameters from a tree to a file (C)
	groupcopy	Copy parameters of group from one tree to another
		(C)
	paramedit	Edit a parameter and its attributes with
		user-selected editor (C)
	prune	Prune extra parameters from current tree (C)
	setgroup	Set group of a parameter in a tree (C)
	setlimit	Set limits of a parameter in a tree (C)
	setprotect	Set protection mode of a parameter (C)
	vi	Edit text file with the vi text editor (C)

pardiff Report differences between parameter sets (M)

Syntax	pardiff(set1<,set2<,parameter_group>>)
Applicability	VnmrJ 3.1
Description	Reports differences between VNMR parameter sets, based on the output of the listparam command. Calls the UNIX diffparam shell script.
Arguments	set1 and set2 are VNMR directories or parameter sets, like experiments, parameter (*.par) or FID (*.fid) files, or actuall parameter text files, like curexp+'/procpar', or userdir+'/exp4/curpar'. Experiments can also be specified by giving just their number. Unless 'procpar' is specified, for experiments the subfile 'curpar' will be taken, for FID or parameter file the subfile 'procpar' is selected for the comparision. If only one file is specified, this is compared with the current experiment.
The '.fid' or '.par' extension can be omitted if an FID or parameter file (directory) is specified.

parametergroup is an optional argument that permits specifying the parameter type. By default, only acquisition parameters are compared. The following options exist (only the first two characters are relevant):

- acquisition compare acquisition parameters (default)
- processing compare processing parameters only
- display compare display parameters only
- spsim compare spin simulation parameters only
- sample compare sample parameters only
- all compare ALL parameters (output indicates group for for each parameter)
- JCAMP compare acquisition and processing parameters in JCAMP-DX compatible format. Inactive parameters are suppressed.

Examples pardiff(3)

pardiff(1,3,'processing')

pardiff('abc.fid')

pardiff(2,'abc.fid')

pardiff('abc.fid',3)

pardiff('xyz.par','abc.fid','all')

Related listparam list parameters in simple format (UNIX) diffparam report differences between parameter sets (UNIX)

pards Create additional parameters used by downsampling (M)

Description	Creates the parameters downsamp, dscoef, dsfb, dslsfrq, and filtfile necessary for digital filtering and downsampling. The pards macro is functionally the same as addpar('downsamp')	
See also	NMR Spectroscopy User Guide	
Related	addpar	Add selected parameters to current experiment (M)
	downsamp	Downsampling factor applied after digital filtering (P)
	dscoef	Digital filter coefficients for downsampling (P)
	dsfb	Digital filter bandwidth for downsampling (P)
	dslsfrq	Bandpass filter offset for downsampling (P)
	filtfile	File of FIR digital filter coefficients (P)
	movedssw	Set downsampling parameters for selected spectral region (M)

parfidss Create parameters for time-domain solvent subtraction (M)

Description Creates solvent subtraction parameters ssfilter, sslsfrq, ssntaps, and ssorder. Entering addpar('ss') is functionally equivalent to parfidss.

In a 1D transform, subtraction of the zero-frequency component from the time-domain data, usually in the context of solvent subtraction, is selected by setting ssorder and ssfilter to desired values and entering wft:

- The zfs (zero-frequency suppression) option is selected if both ssfilter and ssorder are set to a value other than "Not Used."
- The lfs (low-frequency suppression) option is selected if ssfilter is set to a value other than "Not Used" and ssorder is set to "Not Used."
- The zfs and lfs options are both turned off if ssfilter is set to "Not Used."

The zfs option leads to the following series of processing events: (1) the raw FID is frequency-shifted by sslsfrq Hz, (2) the raw FID is subjected to a low- pass digital filter, (3) the filtered FID is fit to a polynomial of order ssorder, (4) the polynomial function is subtracted from the raw FID, and (5) the resulting FID is frequency-shifted by -sslsfrq Hz.

The lfs option does not include a polynomial fit (step 3 of the zfs option), which leads to the following series of processing events: (1) the raw FID is frequency-shifted by sslsfrq Hz, (2) the raw FID is subjected to a low-pass digital filter, (3) the filtered FID is directly subtracted from the raw FID, (4) the resulting FID is frequency-shifted by -sslsfrq Hz.

The quality of filtering with zfs diminishes rapidly as the solvent peak moves off the exact center of the digital filter. It may be necessary to adjust lsfrq or sslsfrq to move the solvent peak to within \pm 0.2 Hz of the center of the filter to obtain optimal solvent suppression. The lfs option is less sensitive to small offsets, but typically removes or distorts peaks near to the solvent peak.

In a 2D transform, solvent correction to the t_2 FIDs is invoked in the same manner with the ftld, ft2d, wftld, and wft2d commands and with the ft2da, ftlda, wft2da, and wftlda macros.

In a 3D transform, solvent suppression works on t_3 FIDs of 3D spectra just like in the 1D and 2D cases.

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to the current experiment (M)	
	ft	Fourier transform 1D data (C)	
	ft1d	Fourier transform along f ₂ dimension (C)	
	ft2d	Fourier transform 2D data (C)	
	ft3d	Perform a 3D Fourier transform on a 3D FID data set	
		(M,U)	

lsfrq	Frequency shift of the fn spectrum in Hz (P)
ntype3d	N-type peak selection in f_1 or f_2 (P)
ssfilter	Full bandwidth of digital filter to yield a filtered FID (P)
sslsfrq	Center of solvent-suppressed region of spectrum (P)
ssorder	Order of polynomial to fit digitally filtered FID (P)
ssntaps	Number of coefficients to be used in the digital filter (P)
wft	Weight and Fourier transform 1D data (C)

parfix Update parameter sets (M)

Description Corrects upper limits, lower limits, and step sizes of a number of parameters in the current experiment. In addition, the template parameter dgs is updated. This is automatically done via the macro fixpar if the parameter parversion is less than 4.3. parfix is used by the macro updatepars to correct saved data. This macro has been applied to all parameters as of VNMR version 4.3 and should be run on older parameter sets (e.g., rtp('pars') svp('pars') update a parameter set named pars).

See also NMR Spectroscopy User Guide

Related	ap	"All" parameters display control (P)	
	dgs	Control dgs parameter group display (P)	
	fixpar	Correct parameter characteristics in experiment (M)	
	parversion Version of parameter set (P)		
	updatepars	Update all parameter sets saved in a directory (M)	

parlc Create parameters for LC-NMR experiments (M)

Applicability Systems with LC-NMR accessory.

Description Creates the following parameters used for a variety of LC-NMR experiments: curscan, dtrig, inject, ntrig, and savefile. The parlc macro also creates ni and sw1 (if they don't exist) for use in isocratic runs. Finally, it creates a display parameter dglc, so that the dg('dglc') command (or the equivalent macro dglc) can be used to display all the LC-related parameters.

Note that parlc can be used without worrying about losing existing values or attributes; if the parameters already exist, they are left untouched.

See also NMR Spectroscopy User Guide

Related	curscan	Scan currently in progress (P)	
	dglc	Control LC-NMR parameter display (P)	
dtrig Delay to wait for another trigge (P)	Delay to wait for another trigger or acquire a spectrum		
		(P)	
	inject	Trigger the injection of a sample (P)	

ntrig Number of trigger signals to wait before acquisition (P)
savefile Base file name for saving FIDs or data sets (P)

parlist List complete parameters in simple format (M)

Syntax	parlist<(parameter_group)>		
Applicability	VnmrJ 3.1		
Description	Reports differences between VNMR parameter sets, based on the output of the listparam command. Calls the UNIX diffparam shell script		
Arguments	s parametergroup is an optional argument that permits specifying the parameter type. By default, only acquisition parameters are listed. The following options exist (only the first two characters are relevant):		
	• acquisition - list acquisition parameters (default)		
	• processing - list processing parameters only		
	• display - list display parameters only		
	• spsim - list spin simulation parameters only		
	• sample - list sample parameters only		
	• all - list ALL parameters (output indicates group for for each parameter)		
	• JCAMP - list acquisition and processing parameters in JCAMP-DX compatible format. Inactive parameters are suppressed.		
Examples	s parlist		
	parlist('processing')		
	parlist('JCAMP')		
Related	listparamlist parameters in simple format (UNIX)pardiffreport differences between parameter sets (M)diffparamreport differences between parameter sets (UNIX)		

par112d Create parameters for 2D peak picking (M)

Description	Creates additional parameters th2d and xdiag for use with 112d 2D peak picking program. par112d is functionally the same as addpar('112d').	
See also	NMR Spectroscopy User Guide	
Related	Add selected parameters to the current experiment (M) Automatic and interactive 2D peak picking (C) Threshold for integrating peaks in 2D spectra (P) Threshold for excluding diagonal peaks when peak picking (P)	

Ρ

parlp Create parameters for linear prediction (M)

Syntax	parlp<(d	parlp<(dimension)>		
Description	Creates pa experiment if necessa	ates parametrized options for linear prediction (LP) in the current eriment. The display template for the dglp macro is also created eccessary. parlp is functionally the same as addpar('lp').		
Arguments	dimension is the dimension of a multidimensional data set. The default is to create the LP parameters lpalg, lpopt, lpfilt, lpnupts, strtlp, lpext, strtext, lptrace, and lpprint.			
	parlp(1) lpnupts1 addpar(.) creates LP parameters lpalg1, lpopt1, lpfilt1, s1, strtlp1, lpext1, strtext1, lptrace1, and lpprint1. ('lp',1) is functionally equivalent to parlp(1).		
	<pre>parlp(2) lpnupts2 addpar('</pre>) creates LP parameters lpalg2, lpopt2, lpfilt2, 2, strtlp2, lpext2, strtext2, lptrace2, and lpprint2. 'lp',2) is functionally equivalent to parlp(2).		
Examples	parlp parlp(1)			
See also	NMR Spectroscopy User Guide			
Related	lpalg	LP algorithm for np dimension (P)		
	lpext	LP data extension for np dimension (P)		
	lpfilt	LP coefficients to calculate for np dimension (P)		
	lpnupts	Lpnupts LP number of data points for np dimension (P)		
	lpopt	LP algorithm data extension for np dimension (P)		
	lpprint	LP print output for np dimension (P)		
	lptrace	LP output spectrum for np dimension (P)		
	proc	Type of processing on np FID (P)		
	proc1	Type of processing on ni interferogram (P)		
	proc2	Type of processing on ni2 interferogram (P)		
	strtext	Starting point for LP data extension for np dimension (P)		
	strtlp	Starting point for LP calculation for np dimension (P)		

parmax Parameter maximum values (P)

- Description An array that holds the maximum values of other parameters. The maximum value of a parameter is an index into the array, and more than one parameter can have the same index into parmax. Several global parameters set in the Spectrometer Configuration window are part of parmax. To display all parmax values, enter display('parmax', 'systemglobal').
 - See also User Programming
 - RelatedconfigDisplay current configuration and possibly change it (MdisplayDisplay parameters and their attributes (C)parameditEdit a parameter and its attributes with user-selected
editor (C)

paramvi	Edit a parameter and its attributes using vi text editor
	(M)
parmin	Parameter minimum values (P)
parstep	Parameter step size values (P)

parmin Parameter minimum values (P)

Description	An array that holds the minimum values for other parameters. The minimum value of a parameter is the index into the parmin array. More than one parameter may have the same index into the array. To display all the values in parmin, enter display('parmin', 'systemglobal').	
See also	User Programming	
Related	paramvi	Edit a parameter and its attributes using vi text editor (M)
	display	Display parameters and their attributes (C)

· · · 1 · · 1			
paramedit	Edit a parameter and its attributes with		
	user-selected editor (C)		
parmax	Parameter maximum values (P)		
parstep	Parameter step size values (P)		

paros Create additional parameters used by oversampling (M)

Description Creates the parameters def_osfilt, filtfile, oscoef, osfb, osfilt, oslsfrq, and oversamp for oversampling and digital filtering. paros is functionally the same as addpar('oversamp'). See also NMR Spectroscopy User Guide Related addpar Add selected parameters to current experiment (M def_osfilt Default value of osfilt parameter (P) filtfile File of FIR digital filter coefficients (P) oscoef Digital filter coefficients for oversampling (P) osfb Digital filter bandwidth for oversampling (P) osfilt Oversampling filter for real-time DSP (P) Bandpass filter offset for oversampling (P) oslsfrq Oversampling factor for acquisition (P) oversamp

parside Sets Up Parameters for Plotting Reference on Side

Description Sets up plotting parameters for plotting a reference spectrum on top of a 2D data set using pl('side').

Syntax	parside
Related	partop

parstep Parameter step size values (P)

Description An array that holds the step size values for other parameters. The step size value of a parameter is the index into the array. More than one parameter can have the same index into parstep. Several configuration parameters set in the Spectrometer Configuration window are part of parstep. To display all parstep values, enter display('parstep','systemglobal'). User Programming See also Related config Display current configuration and possibly change it (M) display Display parameters and their attributes (C) paramedit Edit a parameter and its attributes with user-selected editor (C) Edit a parameter and its attributes using vi text editor paramvi

	(M)
parmax	Parameter maximum values (P)
parmin	Parameter minimum values (P)

partop Sets Up Parameters for Plotting Reference on Top

Description	Sets up plotting parameters for plotting a reference spectrum on top
	of a 2D data set using pl('top').
Syntax	partop
Related	parside

parversion Version of parameter set (P)

- Description Stores the version of a parameter set. When a parameter set is updated with updatepars or parfix, parversion is set to 4.3 to indicate that fact. When a parameter set is retrieved into an experiment, fixpar checks parversion to determine if other parameters need to be updated using parfix.
 See also NMR Spectroscopy User Guide
 - RelatedfixparCorrect parameter characteristics in experiment (M)parfixUpdate parameter sets (M)updateparsUpdate all parameter sets saved in a directory (M)

patchinstall Install a VnmrJ patch

Syntax patchinstall pathname_of_patch where pathname_of_patch can be either a relative or absolute path name.

Description The patchinstall script installs a VnmrJ patch. VnmrJ patches are made for a variety of reasons. They provide a mechanism to distribute bug fixes or provide support for new computers or computer operating systems (OS). The patch name is used to encode the applicability of a patch to a given VnmrJ installation. The patch name is encoded with the VnmrJ version, OS, Console and patch number. These attributes are separated by underscores in the patch name. For example $3.2_LNX_mmi_101.ptc$ and $3.2A_LNX_ddr_102.ptc$ are potential patch names. The names are case-insensitive. Patches have a .ptc suffix. They are actually zip files, but files with a .zip suffix are often blocked by email systems.

The VnmrJ software versions are of the form VERSION x.y REVISION z and are in the first line of the /vnmr/vnmrrev file. The first field of the patch name can match the VERSION or the VERSION and REVISION. The special key ANY will match any VnmrJ version.

In the examples above, the patches can be installed on VnmrJ 3.2 or VnmrJ 3.2A systems.

The second field of the patch name signifies the computer operating system. Supported OS values are LNX, MAC, and WIN. The special key ANY will match any OS. In the examples above, the patches can be installed only on Linux systems.

The third field of the patch name signifies the spectrometer console. Supported values are VNMRS, MR400, Inova, and Mercury. This third field can also be set to keywords that represent groups of spectrometer consoles. The keyword Mercury applies to both MERCURY-Vx and MERCURYplus. The keyowrd MMI applies to MERCURY-Vx,

MERCURYplus, and UNITY INOVA. The keyword DDR applies to VNMRS, VNMRSDD2, MR400, and MR400DD2. The special key ANY will match any console. The console value is taken from the third line of the /vnmr/vnmrrev file.

The fourth and final field is a patch version. Generally, three ranges of patch versions are made. The 100 series patches are the main patch. These patches are cumulative.

Each subsequent patch in the series contains all the contents of the previous patches. So one can install a 103 patch, for example, without first installing the 101 and 102 patches.

The 300 series patches are "hot-fixes" to solve an urgent problem. The 300 patches are generally single purpose patches. They are not cumulative. The 300 series patches will generally be included in a subsequent 100 series patch.

The 500 series patches are also single purpose patches, often to support new PC or OS versions. They are not cumulative. These patches are made when only a subset of users might be interested. For example, if there is a problem with Japanese fonts, Agilent might make a 500 series patch. The 500 series patches will generally be included in the next VnmrJ release. They may or may not make it into a 100 series patch.

The patchinstall script installs patches such that they can be removed with the patchuninstall script. The patchuninstall script can be provided with a single argument that is the name of the last patch installed. In this case, patchuninstall will remove that patch in a noninteractive way. Without an argument, patchuninstall will interactively remove patches, starting with the last patch that was installed.

A patch.ptc file contains the following files:

patch.zip contains the files that will be installed into the \$vnmrsystem directory

checksum contains the checksum of the patch.zip file. Used for validation.

The patch.zip file has an optional Readme file describing the content of the patch.

The patch.zip file has an optional p_install script to do additional tasks by the patch. See the file /vnmr/bin/p_install for a description of the use of this file.

Older patches were suffixed with a tar.Z extension. If patchinstall is used to try to install one of these older patches, it will call patchinstall_ver1 to do the installation. The patchuninstall utility will not be able to remove these older patches.

Related patchmake Build a custom VnmrJ patch patchuninstall Uninstall a VnmrJ patch

patchmake Build a custom Vnmr patch

Syntax Syntax patchmake pathname_of_patch <name> where pathname of patch can be either a relative or absolute path name. Description Most patches are made and delivered by Agilent. It may be useful for a patch to be constructed from changes made by users of VnmrJ or when Agilent field service engineers make user requested changes. For example, if someone makes a local modification or customization, these can bundled into a patch with the patchmake script. A typical scenario would be for you to make an appdir with the modifications. You would then run patchmake, giving the path name of the appdir as an argument. The patchmake utility will build a patch and give it the name custom <date>. This patch can then be installed using patchinstall and removed with patchuninstall. The patchmake utility can be given a second optional argument. It will be used as the find patch name instead of custom_<date>.

The patchmake script installs patches such that they can be removed with the patchuninstall script. The patchuninstall script can be provided with a single argument that is the name of the last patch installed. In this case, patchuninstall will remove that patch in a noninteractive way. Without an argument, patchuninstall will interactively remove patches, starting with the last patch that was installed.

Description

Examples patchmake myappdir gatchmake myappdir 3.2_lnx_ddr

Related patchuninstall Uninstall a VnmrJ patch

patchuninstall Uninstall a VnmrJ patch

Description The patchinstall script installs patches such that they can be removed with the patchuninstall script. The patchuninstall script can be provided with a single argument that is the name of the last patch installed. In this case, patchuninstall will remove that patch in a noninteractive way. Without an argument, patchuninstall will interactively remove patches, starting with the last patch that was installed.

Related patchinstall Install a VnmrJ patch

path3d Path to currently displayed 2D planes from a 3D data set (P)

Description Stores the absolute path to the current 3D data directory tree. If path3d does not exist, it is created by the macro par3d. The command select, as well as the many macros that make use of select, require path3d in order to know where the 2D planes extracted from a 3D data set can be found.

path3d is set automatically by the macros ft3d and getplane:

• ft3d sets path3d to curexp/datadir3d if ft3d is not supplied with a directory path for the transformed 3D data. If ft3d is supplied with such a directory path (e.g., /home/data/test3D), path3d is set equal to that directory path. In this case, the 3D spectral data would reside in the directory /home/data/test3D/data. • getplane sets path3d to curexp/datadir3d if getplane is not supplied with a directory path to the transformed 3D data. If getplane is supplied with such a directory path (e.g., /home/data/test3D), path3d is set equal to that directory path. In this case, the extracted 3D planes would reside in the directory /home/data/test3D/extr.

See also NMR Spectroscopy User Guide

Related	dplane	Display a 3D plane (M)
	dproj	Display a 3D plane projection (M)
	dsplan	Display a series of 3D planes (M)
	es	
	ft3d	Perform a 3D Fourier transform on a 3D FID data set (M)
	getpla	Extract planes from a 3D spectral set (M)
	ne	
	nextpl	Display the next 3D plane (M)
	par3d	Create 3D acquisition, processing, display parameters (C)
	plane	Currently displayed 3D plane type (P)
	plplan	Plot a series of 3D planes (M)
	es	
	prevpl	Display the previous 3D plane (M)
	select	Select a spectrum or 2D plane without displaying it (C)

paxis Plot horizontal LC axis (M)

Applicability	Systems with the LC-NMR accessory.
Syntax	<pre>paxis(time,major_tic,mino_tic)</pre>
Description	Plots a horizontal LC axis. Horizontal axes are assumed to be used with "LC plots" of an entire LC run are labeled accordingly. It is assumed that relevant parameters (e.g., sc, wc, vo, vp) have not been changed after plotting the data.
Arguments	time is the time scale, in minutes (decimal values are fine), of the axis.
	<pre>major_tic is spacing, in minutes (decimal values are fine), of major tics.</pre>
	<pre>minor_tic is spacing, in minutes (decimal values are fine), of minor tics.</pre>
See also	NMR Spectroscopy User Guide

Pbox Pulse shaping software (U)

Syntax Pbox file options

Description Main Pbox (Pandora's Box) program for the generation of shape files for RF and gradients. (See *NMR Spectroscopy User Guide* manual for description of interactive Pbox usage). options is any of the Pbox parameters initialized by the '-' sign and followed by the parameter value. The following options can be in any order and combinations:

-	-b time	Activates Bloch simulator, sets simtime, in sec.
-	-C	Calibrate only, do not create a shape file.
-	-f file	Set name of the output file.
-	-h wave	Print wave file header.
-	-i wave	Print wave file parameters.
-	-l ref_pw90	Length, in µs, of reference pw90 pulse.
-	-0	List options.
-	-p ref_pwr	Reference power level, in dB.
-	-r file	Reshape Pbox pulse.
-	-s stepsize	Define length, in µs, of a single step in waveform.
-	-t wave	Print wave title.
-	-w wavestr	Set wave data string.
-	-v	Run in verbose mode. Also print Pbox version.
-	-value	Sets reps to value.
Examples	Pbox -i eb Pbox newsh Pbox sel.R Pbox -w 'e Pbox tst - -ref_pwr 4	purp2 ape -wc 'eburp1 450 -1280.0' -1 F -w 'eburp1 420 -800' 'eburp1 420 1200' burp1 200 -1200' -attn e -p1 45 54.2 -b w 'esnob 20p 170p' -sfrq 150.02 -refofs 55p 5 -ref_pw90 54.2
See also	NMR Spectre	oscopy User Guide
Related	l cpx	Create Pbox shape file (M)
	dprofile	Display pulse excitation profile from Pbox software (M)
	dshape	Display pulse shape (M)
	dshapef	Display last generated pulse shape (M)
	dshapeı	Display pulse shape interactively (M)
	opx	Open shape definition file for Pbox (M)
	wd_xodq	Define excitation band (M)
	pbox_bws	pulses (M)
	pbox_dmf	Extract dmf value from Pbox shape file (M)
	pbox_dres	Extract dres value from Pbox shape file (M)
	pbox_name	Extract name of last shape file generated by Pbox (M)
	pbox_pw	Extract pulse length from Pbox shape file (M)
	pbox_pwr	Extract pulse power from Pbox shape file (M)
	pbox_pwrf	Extract pulse fine power from Pbox (M)
	pboxget	Extract all calibration data from a Pbox shape file (M)
	pboxget pboxpar	Extract all calibration data from a Pbox shape file (M) Add parameter definition to the pbox.inp file (M)
	pboxget pboxpar pboxrst	Extract all calibration data from a Pbox shape file (M) Add parameter definition to the pbox.inp file (M) Reset temporary Pbox/VnmrJ variables (M)

pph	Print pulse header (M)
pprofile	Plot pulse excitation profile from Pbox software (M)
pshape	Plot pulse shape (M)
pshapef	Display pulse shape or modulation pattern interactively
	(M)
putwave	Write a wave into Pbox.inp file (M)
pxset	Assign Pbox calibration data to experimental
	parameters (M)
pxshape	Generates a single-band shape file (M)
Pxsim	Simulate Bloch profile for a shaped pulse (M)
Pxspy	Create shape definition using Fourier coefficients (U)
selex	Defines excitation band (M)
setwave	Sets a single excitation band in Pbox.inp file (M)
shdec	Shaped observe excitation sequence (M)

pbox_bw Define excitation band (M)

Syntax	pbox_bw<(shapename)>
Description	Defines the excitation band from the position of cursors in the graphics window and reports them to user. It also sets $r1$ to excitation bandwidth and $r2$ to offset. This macro is used mainly in Pbox menus and macros.
Arguments	shapename is the name of a shape as in wavelib; mainly for use with menus.
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

pbox_bws Define excitation band for solvent suppression (notch) pulses (M)

Syntax	pbox_bws<(shapename)>
Description	Defines the excitation band from the position of cursors in the graphics window and reports them to user. It also sets $r1$ to excitation bandwidth and $r2$ to offset. Note, the left cursor should be placed on the left side of the excitation band and the right cursor on resonance of the solvent signal. This macro is mainly used in Pbox menus and macros.
Arguments	shapename is the name of a shape file as in wavelib, mainly for use with menus.
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

Syntax	pbox_dmf<(shapefile.DEC)>:exp_param	
Description	Extracts the dmf value from the file shapefile.DEC created by Pbox or, if file name is not provided, from the pbox.cal file containing parameters of the last created Pbox shape file.	
Arguments	shapefile.DEC is the name of a shape file.	
	exp_param is a dmf type experiment parameter.	
Examples	<pre>pbox_dmf('myfile.DEC'):mydmf pbox_dmf:dmf2</pre>	
See also	NMR Spectroscopy User Guide	
Related	dmfDecoupler modulation frequency for first decoupler (P)PboxPulse shaping software (U)	

pbox_dres Extract dres value from pbox.cal or Pbox shape file (M)

Syntax	pbox_dres<(shapefile.DEC)>:exp_param	
Description	Extracts the dres value from the file shapefile.DEC created by Pbox or, if file name is not provided, from the Pbox.cal file containing parameters of the last created Pbox shape file.	
Arguments	shapefile.DEC is the name of a shape file.	
	exp_param is a dres type experiment parameter.	
Examples	<pre>pbox_dres('myfile.DEC'):mydres pbox_dres:dres2</pre>	
See also	NMR Spectroscopy User Guide	
Related	dres Tip-angle resolution for first decoupler (P)	
	Pbox Pulse shaping software (U)	

pbox_name Extract name of last shape generated by Pbox from pbox.cal (M)

Syntax	pbox_name:exp_name	
Description	Extracts name of the last shape file generated by Pbox and stored in the Pbox.cal file. Note, that the file name extension is not stored explicitly and is not provided by this macro.	
Arguments	exp_name returns the name of last shape file.	
Examples	pbox_pw:shname pbox_pw:pwpat	
See also	NMR Spectroscopy User Guide	
Related	Pbox Pulse shaping software (U)	

Ρ

pbox_pw Extract pulse length from pbox.cal or Pbox shape file (M)

Syntax	pbox_pw<(shapefile.RF)>:exp_param
Description	Extracts pulse length from the file shapefile.RF generated by Pbox or, if file name is not provided, from pbox.cal file containing parameters of the last created Pbox shape file. Returns the pulse length, in μ s.
Arguments	shapefile.RF is the shape file name, including the extension.
	exp_param is a pw type experiment parameter.
Examples	pbox_pw('myfile.RF'):softpw pbox_pw:selpw
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

pbox_pwr Extract power level from Pbox.cal or Pbox shape file (M)

Syntax pbox_pwr<(shapefile.ext)>:exp_param

Description	Extracts the power lever from the file shapefile.ext generated by Pbox or, if file name is not provided, from the pbox.cal file containing parameters of the last created Pbox shape file. Returns the power level, in dB. The exp_param parameter will not be changed by this macro if the parameter is previously set to 'n' (not used).	
Arguments	shapefile.ext is the name of the shape file.	
	exp_param is a power type experiment parameter.	
Examples	pbox_pwr('myfile.DEC'):mypwr pbox_pwr:dpwr2	
See also	NMR Spectroscopy User Guide	
Related	Pbox Pulse shaping software (U)	

pbox_pwrf Extract fine power level from pbox.cal or Pbox shape file (M)

Syntax	pbox_pwrf<(shapefile.ext)>:exp_param
Description	Extracts the fine power lever from the file shapefile.ext generated by Pbox or, if file name is not provided, from the pbox.cal file containing parameters of the last created Pbox shape file. Returns the value of fine power, in dB. Note that the parameter will not be changed by this macro if it was previously set to 'n' (not used).
Arguments	shapefile.ext is the name of the shape file.
	exp_param is a fine power type experiment parameter.

Examples	<pre>pbox_pwrf('myfile.DEC'):mypwrf</pre>	
	pbox_pwrf:dpwrf	
See also	NMR Spectroscopy User Guide	
Related	Pbox Pulse shaping software (U)	

pbox_rst Reset temporary Pbox/Vnmr variables (M)

Syntax	pbox_rst	
Applicability	VnmrJ 3.1	
Description	pbox_rst resets variables $r1-r4 = 0$, $n2='n'$ and $n3=''$. The macro adds also some standrd comment lines to Pbox.inp file. Used in menues and other Pbox macros.	
Examples	opx selex('isnob3') pbox_rst pboxpar('name','selinv.RF') cpx	
Related	opx selex cpx setwave	

pbox_shapeinfoReturns Pbox Shape Information

Description	Returns values of shape, bandwidth, offset, and pulsewidth for a given Pbox shapefile.
Syntax	<pre>pbox_shapeinfo(shapefile)</pre>
Examples	pbox_shapeinfo('WURST40.DEC'):\$shape,\$bandwidth,\$offset,\$pulsewidth Pbox

pboxget Extract Pbox calibration data (M)

Syntax	<pre>pboxget<(shfile.ext)>:\$name,\$pw,\$pwr,\$pwrf,\$dres,\$dmf</pre>
Description	Extracts calibration data from the file shfile.ext generated by Pbox or, if a file name is not provided, from the pbox.cal file containing parameters of the last created Pbox shape file. Returns shape name and the values of total pulse length (in μ s), power (dB), fine power, dres, and dmf. The parameter will not be changed by this macro if the parameter was previously set to 'n' (not used).
Arguments	shfile.ext is the name of the shape file, including the extension. name is the experiment parameter receiving the shape name (without the extension).
	$p_{\ensuremath{\mathbb{W}}}$ is the experiment parameter receiving the total pulse length, in $\mu s.$
	pwr is the experiment parameter receiving the power level, in dB.

	pwrf is the experiment parameter receiving the fine power level.	
	dres is the experiment parameter receiving the decoupler resolution.	
	dmf is the experiment parameter receiving the decoupler modulation frequency.	
Examples	<pre>pboxget('myfile.DEC'):dseq,r1,dpwr,dpwrf,dres,dmf pboxget('selshape.RF'):pwpat,selpw,selpwr pboxget:dseq2,r1,dpwr2,dpwrf2,dres2,dmf2</pre>	
See also	NMR Spectroscopy User Guide	
Related	Pbox Pulse shaping software (U)	

pboxget Extract Pbox calibration data from pbox.cal or Pbox shapefile (M)

Syntax	<pre>pboxget<(shapefile.EXT)>:\$name,\$par1,\$par2,\$par3,\$par4,\$par5</pre>		
Applicability	VnmrJ 3.1		
Description	pboxget extraction or, if filename is of the last gene is as follows : :	ets calibration data from shapefile.ext generated by Pbox s not provided, from pbox.cal file containing parameters rated Pbox shapefile. Order of the returned parameters name, pw, pwr, pwrf, dres, dmf.	
	Warning : paras (not used)!	meter is not changed by this macro if it was set to 'n'	
Arguments	shapefile.EXT -	shapefile name including extension.	
	name - name v	vithout extension	
	pw - length of the waveform (us)		
	pwr - power level (dB)		
	pwrf - fine pov	ver level	
	dres - decoupler resolution		
	dmf - decouple	r modulation frequency	
Examples	pboxget('myfile.	DEC'):dseq,dres,dpwr,dpwrf,dres,dmf	
	pboxget('selshaj	pe.RF"):pwpat,selpw,selpwr	
	pboxget:dseq2,d	res2,dpwr2,dmf2,dres2,dmf2	
Related	pbox_dmf e	xtract dmf value from Pbox shapefile	
	pbox_dres e	xtract dres value from Pbox shapefile	
	pbox_name e	xtract name of last shapefile generated by Pbox	
	pbox_pw e	xtract pulse length from Pbox shapefile	
	pbox_pwr e	xtract pulse power from Pbox shapefile	
	pbox_pwrf e	xtract pulse fine power from Pbox shapefile	
	Pbox P	andora's box pulse/pattern generator (UNIX)	

Syntax	pboxpar('name'<,value>)	
Applicability	VnmrJ 3.1	
Description	pboxpar adds a parameter definition to Pbox.inp file.	
Arguments	name - parameter name	
	value - value of the parameter	
Examples	pboxpar('name','myfile.DEC')	
	pboxpar('bsim','y')	
	pboxpar('T1', 0.24)	
Related	opx	
	selex	
	срх	
	setwave	

pboxrst Reset temporary Pbox variables (M)

Description	Resets r1=0, r2=0, r3=0, r4=0, n2='n', n3='', and adds some		
	standard comment lines to the Pbox.inp file. This macro is used in		
	menus and other Pbox macros.		
See also	NMR Spectroscopy User Guide		
Related	Pbox Pulse shaping software (U)		

pboxunits Converts to Pbox default units (M)

Syntax	pboxunits	
Description	Used by Pbox menus to scale parameters related to time or frequency down to Pbox default units (Hz or seconds) before the parameter is stored in the Pbox.inp file.	
See also	NMR Spectroscopy User Guide	
Related	Pbox Pulse shaping software (U)	

pcmapapply Apply Phase Correction Map to Data (C)

Syntax	<pre>pcmapapply([<filename>,]<index>)</index></filename></pre>
Applicability	VnmrJ 3.1

Ρ

correction map \$vnmruser/expN/datdir/<filename>.

It assumes the phase correction map file to be opened resides in the user's \$vnmruser/expN/datdir directory where N is the current experiment number. If the "filename" argument is not given, the command defaults to the filename "pcmap". The "index" argument must always be supplied, ranges from 1 to n, and specifies the desired correction map block within the file.

The phase correction values are generated by "pcmapgen". One or more phase correction maps may be generated. In the case of a multislice EPIexperiment there may be one phase correction map for each slice. As mentioned before, the command uses data from the current datafile; which means that a fourier transform must have been performed on the data. For images, a "ftld" should be done on the data before using this command.

"pcmapapply" will open and close the phase map file unless it has been explicitly opened with "pcmapopen". Explicitly opening a phase correction map file may be desired if there are a large number of images or data sets to be processed.

The "pcmap" commands have been developed for processing Echo Planar images, but can also be used for other applications if applicable.

Arguments 'filename' optional argument specifying the phase correction map file name residing in the user's \$vnmruser/expN/datdir directory. The default file is \$vnmruser/expN/datdir/pcmap.

'index' argument specifying which phase correction map to use in the file. This value will usually be 1.

Examples ftld('nf',2)

Description

pcmapapply(1)

ft2d('nf',2)

RelatedpcmapopenPhase Correction Map OpenpcmapgenGenerate Phase Correction Map

pcmapgen Generate Phase Correction Map (C)

Syntax	<pre>pcmapgen([<filename>,]<index>)</index></filename></pre>
Applicability	VnmrJ 3.1
Description	"pcmapgen" generates pixel by pixel complex phase correction values from the current datafile and stores them into the <index> block in the phase correction map file \$vnmruser/expN/datdir/<filename>.</filename></index>
	It assumes the phase correction map file to store the values resides in the user's \$vnmruser/expN/datdir directory where N is the current experiment number. If the "filename" argument is not given, the command defaults to the filename "pcmap". The "index" argument must

always be supplied, ranges from 1 to n, and specifies the desired correction map block within the file.

One or more phase correction maps may be generated, although they can only be generated one at a time. As mentioned before, the command uses data from the current datafile; which means that a fourier transform must have been performed on the data. For images, a "ftld" should be done on the data before using this command.

"pcmapgen" will create, open, and close the phase map file unless it has been explicitly opened with "pcmapopen". Explicitly opening a phase correction map file may be desired if there are a large number of images or data sets to be processed.

The "pcmap" commands have been developed for processing Echo Planar images, but can also be used for other applications if applicable.

Arguments 'filename' optional argument specifying the phase correction map file name residing in the user's \$vnmruser/expN/datdir directory. The default file is \$vnmruser/expN/datdir/pcmap.

'index' argument specifying which phase correction map to use in the file. This value will usually be 1.

Examples ftld('nf',1)

pcmapgen(1)

Related pcmapopen Phase Correction Map Open pcmapapply Apply Phase Correction Map to Data

pcmapclose Phase Correction Map Close (C)

Syntax	<pre>pcmapopen([<filename>,]<max_index>)</max_index></filename></pre>
	pcmapclose

Applicability VnmrJ 3.1

Description "pcmapopen" explicitly opens a phase correction map file using memory mapped I/O. It assumes the phase correction map file to be opened resides in the user's \$vnmruser/expN/datdir directory where N is the current experiment number. If the "filename" argument is not given, the command defaults to the filename "pcmap". The "max_index" argument must always be supplied and be greater than or equal to the maximum number of phase maps stored in the file. Once the phase correction map is opened the phase correction commands "pcmapgen" and "pcmapapply" can be used to generate maps and correct data.

> Explicitly opening a phase correction map file can significantly speed up the data processing. The "pcmap" commands have been developed for processing Echo Planar images, but can also be used for other applications if applicable.

> Once the file has been opened a "pcmapclose" command must be used to close the file when finished. "pcmapclose" closes phase correction map file that has been explicitly opened with a "pcmapopen" command.

Arguments	'filename' optional argument specifying the phase correction map name residing in the user's \$vnmruser/expN/datdir directory.			
	'max_index' argument specifying the maximum number of phase correction maps in the file. This is to ensure the memory mapping extends to or past the end of the file.			
Examples	pcmapopen('pcmap',2) pcmapclose			
Related	pcmapapplyApply Phase Correction Map to DatapcmapgenGenerate pcmap			

pcon Plot contours on a plotter (C)

Syntax	pcon<(<'pos' 'neg'><,'noaxis'><,levels><,spacing>)>				
Description	Plots positive and negative peaks of a contour plot display using different colors. Specifically, if maxpen is set for n pens, positive peaks are plotted using colors 1 through $(n+1)/2$, and negative peaks are plotted using colors $((n+1)/2)+1$ through n (i.e., half the colors for each, plus one extra for positive if an odd number of pens is specified). Pen 1 is always used for the axes, and the lowest contour of the positive peaks is also plotted with pen1. In all cases, the pen colors are cycled if more contours are to be plotted than there are pens available.				
	To plot both negative and positive contours of a phase-sensitive spectrum on a monochrome device such as a LaserJet or a plotter with a single pen, different numbers of contours may be plotted for the different sign. For example, $pcon('pos', 10, 1.4) pcon('neg', 1)$ will plot ten closely spaced positive contours and one negative contour.				
Arguments	'pos' is a keyword specifying that phase-sensitive spectra plot positive peaks only. The default is to plot both positive and negative peaks.				
	'neg' is a keyword specifying that phase-sensitive spectra plot negative peaks only. The default is to plot both positive and negative peaks.				
	'noaxis' is a keyword to omit outlining the plot and omit plotting the horizontal and vertical axes.				
	levels is maximum number of contour levels to plot. The default is 4.				
	spacing is relative intensity of successive contour levels. The default is 2.				
Examples	<pre>pcon pcon(4,1.4) pcon('pos', 'noaxis') pcon('neg',3)</pre>				
See also	NMR Spectroscopy User Guide				
Related	dpconDisplay plotted contours (C)maxpenMaximum number of pens to use (P)				

pcss Calculate and show proton chemical shifts spectrum (M)

Syntax	pcss<(<threshold><,max_cc><,max_width>)></threshold>				
Description	Calculates and shows the proton chemical shifts spectrum. The dsp command is used to display the results. The list of chemical shifts is saved in the file pcss.outpar. The original spectrum can be calculated by the wft command.				
Arguments	threshold sets the level whether a point belongs to a peak or is noise. The default is that pcss automatically calculates the threshold.				
	max_cc is the maximum allowable coupling constant in the spectrum. The default is 20 Hz.				
	max_width is the maximum width of a spin multiplet in the spectrum. The default is 60 Hz.				
Examples	pcss pcss(10) pcss(9,20,80)				
See also	NMR Spectroscopy User Guide				
Related	do_pcssCalculate proton chemical shifts spectrum (C)dspDisplay pulse sequence (C)wftWeight and Fourier transform 1D data (C)				

peak Find tallest peak in specified region (C)

Syntax	<pre>peak<(min_freq,max_freq)><:height,freq></pre>
Description	Returns the height and frequency of the tallest peak in the selected region, including any referencing (i.e., the same frequency that you would measure by placing a cursor on the peak). A spectrum need not actually be displayed for peak to work.
Arguments	With no return arguments, peak displays on the screen information about peak height and frequency. If two cursors are displayed, peak without arguments finds the tallest peak between the cursors.
	<pre>min_freq is minimum frequency limit of the region to be searched. The default value is sp.</pre>
	max_freq is maximum frequency limit, in Hz, of the region to be searched. The default value is $sp + wp$.
	height returns the height, in mm, of the tallest peak in the selected region.
	${\tt freq}$ returns the frequency, in Hz, of the tallest peak in the selected region.
Examples	<pre>peak:\$ht,\$freq peak(0,2000):r3 peak:\$ht,cr</pre>

See also	User	Programming
Related	sp	Start of plot (P)
	wp	Width of plot (P)

peak2d Return information about maximum in 2D data (C)

Syntax	<pre>peak2d:\$maximum_intensity<,\$trace,\$point></pre>				
Description	Searches the area defined by sp, wp, sp1, and wp1 in a 2D data set for a maximum intensity.				
Arguments	\$maximum_i	ntensity returns the maximum intensity value found.			
	$trace$ returns the trace number of the maximum. The parameter trace defines whether f_1 or f_2 traces are counted.				
	\$point returns the data point number of the maximum on that trace.				
See also	NMR Spectroscopy User Guide				
Related	sp	Start of plot (P)			
	sp1 Start of plot in 1st indirectly detected dimension (P)				
	trace Mode for <i>n</i> -dimensional data display (P)				
	wp	Width of plot (P)			
	wpl	Width of plot in 1st indirectly detected dimension (P)			

peakmin Find the minimum point

Syntax	<pre>peakmin<(highfield,lowfield)>:ht,frq,amp</pre>
Applicability	VnmrJ 3.1
Description	 peak finds the height and frequency of the maximum point in the specified region. peakmin finds the height and frequency of the minimum point in the specified region. For both peak and peakmin, height is measured in mm, and frequency is measured in Hz, including any referencing (i.e. the same frequency that you would measure by placing a cursor on that point). Default parameters for highfield and lowfield are "sp" and "sp+wp", respectively. The value of the height and frequency of the point can be returned to the caller if the command is suffixed with a colon and parameter names. An unscaled amplitude may be returned as the third value. This unscaled amplitude is independent of the current value of vs and whether the spectrum is in absolute intensity or normalized mode (ai)
	or nm).

pen Select a pen or color for drawing (C)

Description Selects the pen number for a plotter or the color for the graphics screen. This command is part of a line drawing capability that includes the move and draw commands. move sets the coordinates from which the line starts. draw draws a line from that point to the new coordinates specified by draw. Refer to the description of draw for examples of using the line drawing capability.

Arguments 'graphics' and 'plotter' are keywords selecting the output device. The default is 'plotter'. The output selected is passed to subsequent pen, move, or draw commands and remains active until a different output is specified.

> 'xor' and 'normal' are keywords selecting the drawing mode for the 'graphics' output device. In the 'xor' mode, if a line is drawn such that one or more points of the line are in common with a previously drawn line, the common points are erased. In the 'normal' mode, the common points remain. The mode selected is passed to subsequent pen, draw, or move commands and remains active until a different mode is specified. The default mode is 'normal'.

pen is the plotter pen number: 'pen1', 'pen2', 'pen3', etc.
'pen1', 'pen2', 'pen3', ...

color is the active color for the graphics screen: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', 'white', 'cursor', 'integral', 'threshold', 'scale', 'fid', 'spectrum', 'imaginary', 'parameter'

This list includes eight symbolic color names (cursor, integral, etc). The actual colors associated with the symbolic names may be set with the "Display options..." tool in the Edit menu. The advantage of using the symbolic names is that they are probably adjusted to look good with the chosen background color. For example, using the color white for drawing on the graphics screen may look fine with a dark background, but will be invisible if the background is white. Using the color 'spectrum' will probably look good for both light and dark backgrounds.

	-	-	-	-		-		-	
Examples	pen('pe	n2')							
	pen('gr	aphics',	red')						
See also	NMR Spe	ctroscopy	User Gı	uide					
Related	draw	Draw lin	e from	current]	locatior	n to ano	ther loca	tion (C)
	move	Move to	an abso	olute loca	ation ((C)			

pexp1 Plot exponential or polynomial curves (C)

Syntax pexpl<(<options,><line1,line2, ...)>

Description	Plots exponential curves resulting from T_1 , T_2 , or kinetics analysis. Also
	plots polynomial curves from diffusion or other types of analysis. The
	analyze.out file is the data input file used to make the plot. Refer
	to the expl entry for the format of this file. The parameters sc, wc,
	sc2, and $wc2$ control the size of the plot.

Arguments options are any of the following keywords:

- 'linear', 'square', and 'log' provide for plotting of the data points against the square or log of the data. 'linear' controls x-axis scale, 'square' controls the y-axis. The default is 'linear'.
 - 'link' causes the data points to be connected rather than a plot of the theoretical curve.
- 'nocurve' produces a plot of data points only.
- 'oldbox' plots an additional curve on an existing plot. Only the first data set in analyze.out is plotted. It causes the program to get box and scale description from expfit.out in the current experiment.
- 'file' followed by a file name replaces analyze.out as the input.

line1, line2,... specify curves to be plotted. The default is to plot the first six curves (if that many exist) along with the data points.

Examples pexpl

pexpl(1,3,6)

See also NMR Spectroscopy	User Guide,	User	Programming
---------------------------	-------------	------	-------------

Related	expl	Display exponential or polynomial curves (C)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	WC	Width of chart (P)
	wc2	Width of chart in second direction (P)

pexpladd Add another diffusion analysis to current plot (M)

Applicability	Systems with the diffusion option.		
Syntax	<pre>pexpladd(integral_region)</pre>		
Description	Adds results of another diffusion analysis to the currently plotted results.		
Arguments	integral_region specifies the number of the region whose results are to be added to the existing plot.		
Examples	pexpladd(1)		
See also	NMR Spectroscopy User Guide		
Related	explDisplay exponential or polynomial curves (C)pexplPlot exponential or polynomial curves (C)expladdAdd another diffusion analysis to current display (M)		

pfgon Pulsed field gradient amplifiers on/off control (P)

Applicability Systems with pulsed field gradient (PFG) modules.

- Description A global string parameter controlling the X, Y, and Z gradients for the PFG current amplifiers. Entering su or go sets the amplifiers at the current value of pfgon. For pfgon to take effect, gradtype must equal p, q, 1, t, or u for the corresponding X, Y, or Z gradient, and a su or a go must be issued.
 - Values A three-character string, with the first character controlling the X gradient, the second the Y gradient, and the third the Z gradient. For each gradient, setting the value to y turns on an amplifier and setting the value to n turns it off. For example, pfgon='nny' turns on only the PFG amplifier on the Z channel, and pfgon='nnn' turns off the PFG amplifiers on all channels.
 - See also NMR Spectroscopy User Guide

Related	go	Submit experiment to acquisition (M)
	gradtype	Gradients for X, Y, and Z axes (P)
	setup	Set up parameters for basic experiments (M)
	su	Submit a setup experiment to acquisition (M)

pfww Plot FIDs in whitewash mode (C)

Syntax	pfww<(<sta< th=""><th>rt><,finish><,step><,'all' 'imag'>)></th></sta<>	rt><,finish><,step><,'all' 'imag'>)>	
Description	Plots FIDs in out in region the first FID	whitewash mode (after the first FID, each FID is blanked s in which it is behind an earlier FID). The position of is governed by parameters wc, sc, and vpf.	
Arguments	s start is the index of a particular FID for arrayed 1D or 2D data s For multiple FIDs, start is the index of the first FID.		
	finish is th	e index of the last FID for multiple FIDs.	
	step specifie	es the increment for the FID index. The default is 1.	
	'all' is a l	reyword to plot all of the FIDs. This is the default.	
	'imag' is a default is 'a	keyword to plot only the imaginary FID channel. The ll'.	
Examples	pfww pfww(4,10,	2,'imag')	
See also	NMR Spectro	scopy User Guide	
Related	dfs	Display stacked FIDs (C)	
	dfww	Display FIDs in whitewash mode (C)	
	plfid	Plot FIDs (C)	
	SC	Start of chart (P)	
	vpf	Current vertical position of FID (P)	
	WC	Width of chart (P)	

Applicability Systems with the diffusion option.

Description Adds all necessary parameters to perform the PGE (Pulse Gradient Experiment) pulse sequence, taking those parameters from the file /vnmr/parlib/pge.

See also NMR Spectroscopy User Guide

Related pge_cali Calibrate gradient strengths for PGE pulse sequence (M) Extract data from single element of PGE pulse sequence pge_data (M) Output results from PGE pulse sequence (M) pge_outp ut Automated processing of data from PGE pulse sequence pge_proc (M) ess pge_resu Calculate diffusion constant for integral region (M) lts Set up gradient control parameters for PGE pulse pge_setu sequence (M) σ

pge_calib Calibrate gradient strengths for PGE pulse sequence (M)

- Description Calibrates the parameters grad_cw_coef and grad_p_coef, which relate the DAC values (in DAC units) to the gradient strengths (in gauss/cm). Given a diffusion constant measurement (made with pge_results) for a known diffusion constant, pge_calib then adjusts the calibration parameters to produce the correct diffusion constant.
 - See also NMR Spectroscopy User Guide
 - Related pge Calibrate gradient strengths for PGE pulse sequence (M) pge_resu Calculate diffusion constant for integral region (M) lts

pge_data Extract data from single element of PGE pulse sequence (M)

Applicability Systems with the diffusion option. Syntax pge_data(array_index) Description Extracts integral information from a currently displayed element of a PGE (Pulse Gradient Experiment) and writes the results in the current experiment directory as the file info_#, where # is the value of the array_index argument (e.g., if array_index is 5, the file is info_5)

P

Arguments	array_index is the number of the array element from which the data is extracted.
Examples	pge_data(5)
See also	NMR Spectroscopy User Guide
Related	pge Calibrate gradient strengths for PGE pulse sequence (M)

pge_output Output results from PGE pulse sequence (M)

Applicability Systems with the diffusion option.

- Description Prints the calculated results from the PGE (Pulse Gradient Experiment) pulse sequence on a printer and plots the graphs of calculated decay curves.
 - See also NMR Spectroscopy User Guide
 - Related pge Calibrate gradient strengths for PGE pulse sequence (M)

pge_process Automated processing of data from PGE pulse sequence (M)

Applicability	Systems with the diffusion option.
Syntax	pge_process
Description	Performs full automated processing of data from a PGE (Pulse Gradient
	Experiment) pulse sequence.
See also	NMR Spectroscopy User Guide
Related	pge Calibrate gradient strengths for PGE pulse sequence (M)

pge_results Calculate diffusion constant for integral region (M)

Applicability	Systems with the diffusion option.
Syntax	<pre>pge_results(integral_region<,reference_region>)</pre>
Description	Calculates a diffusion coefficient based on a single integral region in the spectrum (if one input argument) or calculates diffusion coefficient of an integral region consisting of two components (if two input arguments).
Arguments	integral_region is the number of the integral region on which to perform the analysis
	reference_region is the number of the integral region used to get the value of the diffusion coefficient.
Examples	<pre>pge_results(2) pge_results(1,3)</pre>

See also	NMR	Spectroscopy	y User (Huide		
Deleted		Calibrata	and diant	atu an atla a	f	DO

Related pge Calibrate gradient strengths for PGE pulse sequence (M)

pge_setup Set up gradient control parameters for PGE pulse sequence (M)

Applicability	Systems with the diffusion option.
Syntax	pge_setup<('no')>
Description	Prompts the user for the values of the g_max, g_min, g_steps, g_array, nt_first, nt_aray, and other parameters for the PGE (Pulse Gradient Experiment) pulse sequence. These parameters are then used to calculate the grad_p1 and nt arrays.
Arguments	'no' is a keyword to turn off prompting the user and instead use the current values of the parameters to calculate the grad_p1 and nt arrays.
Examples	pge_setup pge_setup('no')
See also	NMR Spectroscopy User Guide
Related	pge Calibrate gradient strengths for PGE pulse sequence (M)

ph

Set phased mode in directly detected dimension (C)

Description Selects the phased mode by setting the parameter dmg='ph'. In the *phased spectra display mode*, each real point in the displayed spectrum is calculated from a linear combination of the real and imaginary points comprising each respective complex data point. The coefficients for this linear combination are derived from the phase parameters rp and lp.

For 2D data, if pmode='partial' or pmode='' (two single quotes with no space in between), ph has an effect on the data prior to the second Fourier transform. If pmode='full', ph acts in concert with the commands ph1, av1, or pwr1 to yield the resultant contour display for the 2D data.

See also NMR Spectroscopy User Guide

Related	av	Set abs. value mode in directly detected dimension (C)
	av1	Set abs. value mode in 1st indirectly detected dimension (C)
	dmg	Data display mode in directly detected dimension (P)
	ft	Fourier transform 1D data (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ft2d	Fourier transform 2D data (C)
	lp	First-order phase in directly detected dimension (P)

pa	Set phase angle mode in directly detected dimension (C)
pa1	Set phase angle mode in 1st indirectly detected dimension
	(C)
ph1	Set phased mode in 1st indirectly detected dimension (C)
ph2	Set phased mode in 2nd indirectly detected dimension (C)
pmode	Processing mode for 2D data (P)
pwr	Set power mode in directly detected dimension (C)
pwr1	Set power mode in 1st indirectly detected dimension (C)
rp	Zero-order phase in directly detected dimension (P)
wft	Weight and Fourier transform 1D data (C)
wft1d	Weight and Fourier transform f2 of 2D data (M)
wft2d	Weight and Fourier transform 2D data (M)

ph1 Set phased mode in 1st indirectly detected dimension (C)

Description Selects the phased spectra display mode along the first indirectly detected dimension by setting the parameter dmg1 to the string value 'ph1'. If the parameter dmg1 does not exist, ph1 will create it and set it to 'ph1'.

In the phased mode, each real point in the displayed trace is calculated from a linear combination of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the linear combination uses the real-real and imaginary-real points from each respective hypercomplex data point. The coefficients for this linear combination are derived from the phase parameters rp1 and lp1.

The ph1 command is only needed if mixed-mode display is desired. If the parameter dmg1 does not exist or is set to the null string, the display mode along the first indirectly detected dimension defaults to the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of ph1 is the same as for traces provided that pmode='partial' or pmode=''.

See also NMR Spectroscopy User Guide

Related	av1	Set abs. value mode in 1st indirectly detected dimension (C)
	dmg1	Data display mode in 1st indirectly detected dimension (P)
	lp1	First-order phase in 1st indirectly detected dimension (P)
	pa	Set phase angle mode in directly detected dimension (C)
	pa1	Set phase angle mode in 1st indirectly detected dimension
		(C)
	ph	Set phased mode in directly detected dimension (C)
	pmode	Processing mode for 2D data (P)
	pwr1	Set power mode in 1st indirectly detected dimension (C)
	rp1	Zero-order phase in 1st indirectly detected dimension (P)

ph2 Set phased mode in 2nd indirectly detected dimension (C)

Description Selects phased spectrum display mode processing along the second indirectly detected dimension by setting the parameter dmg2='ph2'. If dmg2 does not exist or is set to the null string, ph2 creates dmg2 and sets it to 'ph2'.

In the phased mode, each real point in the displayed trace is calculated from a linear combination of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the linear combination uses the real-real and imaginary-real points from each respective hypercomplex data point. The coefficients for this linear combination are derived from the phase parameters rp2 and lp2.

The ph2 command is only needed if mixed-mode display is desired. If the parameter dmg2 does not exist or is set to the null string, the display mode along the second indirectly detected dimension defaults to the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of ph2 is the same as for traces provided that pmode='partial' or pmode=''.

See also NMR Spectroscopy User Guide

Related av2 Set abs. value mode in		Set abs. value mode in 2nd indirectly detected dimension
		(C)
	dmg2	Data display mode in 2nd indirectly detected dimension (P)
	ft1d	Fourier transform along f_2 dimension (C)
ft2d Fourier transform 2D data (C)		Fourier transform 2D data (C)
1p2 First-order phase in 2nd indirectly detected		First-order phase in 2nd indirectly detected dimension (P)
	ph	Set phased mode in directly detected dimension (C)
pmode Processing mode for 2D data (P)		Processing mode for 2D data (P)
	pwr2	Set power mode in 2nd indirectly detected dimension (C)
	rp2	Zero-order phase in 2nd indirectly detected dimension (P)

phase Change frequency-independent phase rp (M)

Syntax	phase(phase_change)	
Description	Changes the phase of all peaks in the spectrum by adding a value to the current rp value. Any excess over 360° is removed.	
Arguments	phase_change is the value to be added to the current rp value (i.e., <i>new</i> rp = <i>old</i> rp + phase_change).	
Examples	phase(45)	
See also	NMR Spectroscopy User Guide	
Related	rp Zero-order phase in directly detected dimension (P)	

Ρ

Description	elects the phase cycling that determines the experiment type. To reate the parameters $phase$, ni , and $sw1$ for acquisition of a 2D data et in the current experiment, enter $addpar('2d')$.		
Values	The following values are generally used in experiments with phase cycling. For more details, see the specific pulse sequence.		
	phase=0 selects an absolute-value 2D experiment.		
	hase=1,2 selects the required two components of a hypercomplex States-Haberkorn) experiment.		
	phase=3 selects TPPI (Time Proportional Phase Incrementation).		
See also	NMR Spectroscopy User Guide		
Related	addparAdd selected parameters to the current experiment (M)cosypsSet up parameters for phase-sensitive COSY (M)DqcosySet up parameters for double quantum filtered COSY (M)HmqcSet up parameters for HMQC pulse sequence (M)hmqcrSet up parameters for HMQCR pulse sequence (M)inadqtSet up parameters for INADEQUATE pulse sequence (M)mqcosySet up parameters for MQCOSY pulse sequence (M)NoesySet up parameters for NOESY pulse sequence (M)RoesySet up parameters for ROESY pulse sequence (M)TocsySet up parameters for TOCSY pulse sequence (M)		

phase1 Phase of first pulse (P)

Applicability	Systems with a solids NMR module.		
Description	Controls the first pulse phase in the cycle, in multipulse experiments		
See also	NMR Spectroscopy User Guide		
Related	br24	Set up BR24 multiple pulse experiment (M)	
	flipflop	Set up sequences for multipulse (M)	

phase2 Phase selection for 3D acquisition (P)

Description	Selects phase cycling type for 3D data acquisitions. Also selects the		
	phase of the second pulse in the sequence set up by flipflop. To		
	create the parameters phase2, d3, ni2, and sw2 for acquisition of a		
	3D data set in the current experiment, enter addpar('3d').		
See also	NMR Spectroscopy User Guide; User Guide: Solid-State NMR		
Related	addpar Add selected parameters to the current experiment (M)		
	(P)		

flipflop	Set up sequences for multipulse (M)		
ni2	Number of increments in 2nd indirectly detected		
	dimension (P)		
par3d	Create 3D acquisition, processing, display parameters (C)		
sw2	Spectral width in 2nd indirectly detected dimension (P)		

phase3 Phase selection for 4D acquisition (P)

Description Selects phase cycling type for 4D data acquisitions. To create the parameters phase3, d4, ni3, and sw3 for acquisition of a 4D data set in the current experiment, enter addpar('4d'). NMR Spectroscopy User Guide See also Related addpar Add selected parameters to the current experiment (M) d4 Incremented delay for 3rd indirectly detected dimension (P) ni3 Number of increments in 3rd indirectly detected dimension (P) Create 4D acquisition parameters (C) par4d Spectral width in 3rd indirectly detected dimension (P) sw3

phasing Control update region during interactive phasing (P)

Description Controls the percentage of the spectrum updated during interactive phasing using the ds command.
Values 10 to 100, in percent, where 100 causes the entire spectrum to be updated, and 20 causes the area between the two vertical cursors to be updated.
See also NMR Spectroscopy User Guide
Related ds Display a spectrum (C)

phfid Zero-order phasing constant for the np FID (P)

Description Specifies the angle of zero-order rotation. This zero-order rotation is executed as a part of retrieving the time-domain data into the active region of the memory and can be used instead of the parameter rp applied to the frequency-domain data. phfid is used only in a complex phase rotation.

phfid (and related parameters lsfid and lsfrq) operate on complex np FID data, referred to as the t_2 dimension in a 2D experiment or as the t_3 dimension in a 3D experiment. phfid is in the processing group and is properly handled through the wti display.

Values -360.0 to +360.0, in degrees; 'n'

see also	NMR Spectroscopy User Guide	
Related	dfid	Display a single FID (C)
	ds	Display a spectrum FID (C)
	ft	Fourier transform 1D data (C)
	ft1d	Fourier transform along f ₂ dimension (C)
	ft2d	Fourier transform 2D data (C)
	lsfid	Number of complex points to left-shift the np FID
		(P)
	lsfrq	Frequency shift of the fn spectrum in Hz (P)
	np	Number of data points (P)
	phfid1	Zero-order phasing constant for ni interferogram
		(P)
	phfid2	Zero-order phasing constant for ni2 interferogram
		(P)
	rp	Zero-order phase in directly detected dimension (P)
	wft	Weight and Fourier transform 1D data (C)
	wft1d	Weight and Fourier transform f ₂ of 2D data (M)
	wft2d	Weight and Fourier transform 2D data (M)
	wti	Interactive weighting (C)

phfid1 Zero-order phasing constant for ni interferogram (P)

Description	Specifies the angle of zero-order rotation. This zero-order rotation is executed as a part of retrieving the time-domain data into the active region of the memory and can be used instead of the parameter rpl applied to the frequency-domain data. phfid1 is used in a complex phase rotation for complex t_1/t_2 interferograms and in a hypercomplex phase rotation for hypercomplex t_1/t_2 interferograms.	
	phfid1 (and related parameters lsfid1 and lsfrq1) operate on ni interferogram data, both hypercomplex and complex. ni interferogram data are referred to as the t_1 dimension in both a 2D and a 3D experiment. phfid1 is in the processing group and is properly handled through the wti display; that is, a wti operation on an ni interferogram applies the parameters phfid1, lsfid1, and lsfrq1, if selected, to the time-domain data prior to the Fourier transformation.	
Values	-360.0 to +360.0, in degrees; 'n'.	
See also	NMR Spectroscopy User Guide	
Related	<pre>lsfid1 Number of complex points to left-shift the ni interferogram (P) lsfrq1 Frequency shift of the fn1 spectrum in Hz (P)</pre>	
	ni Number of increments in 1st indirectly detected dimension (P)	

- Zero-order phasing constant for np FID (P) phfid
- phfid2 Zero-order phasing constant for ni2 interferogram (P)

See

rp1	Zero-order phase in 1st indirectly detected dimension (P)
wti	Interactive weighting (C)	

phfid2 Zero-order phasing constant for ni2 interferogram (P)

Description Specifies the angle of zero-order rotation. This zero-order rotation is executed as a part of retrieving the time-domain data into the active region of the memory and can be used instead of the parameter rp2 applied to the frequency-domain data. phfid2 is used in a complex phase rotation for complex t_1/t_2 interferograms and in a hypercomplex phase rotation for hypercomplex t_1/t_2 interferograms.

phfid2 (and related parameters lsfid2 and lsfrq2) operate on ni2 interferogram data, both hypercomplex and complex. ni2 interferogram data are referred to as the t_2 dimension in a 3D experiment. phfid2 is in the processing group and is properly handled through the wti display.

- Values -360.0 to +360.0, in degrees; 'n'.
- See also NMR Spectroscopy User Guide
 - Related lsfid2 Number of complex points to left-shift ni2 interferogram (P) lsfrq2 Frequency shift of the fn2 spectrum in Hz (P)
 - ni2 Number of increments in 2nd indirectly detected dimension (P)
 - phfid Zero-order phasing constant for np FID (P)
 - phfid1 Zero-order phasing constant for ni interferogram (P)
 - rp2 Zero-order phase in 2nd indirectly detected dimension (P)
 - wti Interactive weighting (C)

Phosphorus Set up parameters for ³¹P experiment (M)

Description Set up parameters for ³¹P experiment.

pi3ssbsq Set up pi/3 shifted sinebell-squared window function (M)

Syntax	pi3ssbsq<(<t1_inc><,t2_inc>)></t1_inc>
Description	Sets up a pi/3 unshifted sinebell-squared window function in 1, 2, or 3 dimensions. The macro checks whether the data is 1D, 2D, and 3D.
Arguments	t1_inc is the number of t1 increments. The default is ni.
	t2_inc is the number of t2 increments. The default is ni2.

See also	NMR Spectroscopy User Guide		
Related	gaussian	Set up unshifted Gaussian window function (M)	
	ni	Number of increments in 1st indirectly detected	
		dimension (P)	
	ni2	Number of increments in 2nd indirectly detected	
		dimension (P)	
	pi4ssbsq	Set up pi/4 shifted sinebell-squared window function	
		(M)	
	sqcosine	Set up unshifted cosine-squared window function (M)	
	sqsinebell	Set up unshifted sinebell-squared window function (M)	

pi4ssbsq Set up pi/4 shifted sinebell-squared window function (M)

Syntax	pi4ssbsq<(<t1_inc><,t2_inc>)></t1_inc>	
Description	Sets up a pi/4 unshifted sinebell-squared window function in 1, 2, or 3 dimensions. The macro checks whether the data is 1D, 2D, and 3D.	
Arguments	t1_inc is the number of t1 increments. The default is ni.	
	t2_inc is the number of t2 increments. The default is ni2.	
See also	NMR Spectroscopy User Guide	
Related	gaussian	Set up unshifted Gaussian window function (M)
	ni	Number of increments in 1st indirectly detected dimension (P)
	ni2	Number of increments in 2nd indirectly detected dimension (P)
	pi3ssbsq	Set up pi/3 shifted sinebell-squared window function (M)
	sqcosine	Set up unshifted cosine-squared window function (M)
	sqsinebell	Set up unshifted sinebell-squared window function (M)

pin Pneumatics Router Interlock ((P)

Description This parameter controls the effect of a Pneumatics Router Fault. The Pneumatic Router can fault in four ways:

- Intake pressure < 20 psi
- Solids narrow bore stack temperature fault
- VT air flow exceeded.
- Power supply fault

When either of these fault occur, and interrupt alerts the console of the problem and this parameter determines how the fault is handled. Once a fault is registered, all subsequent acquisitions will see the error according to 'pin'. The error must be cleared and re-armed with sethw('pneufault','clear')
Values	'n' the fa	ault is ignored
	'w' a wa	rning msg is printed, acquisition continues
	'y' an er	ror msg is printed, acquisition is aborted
Related	tin	Temperature interlock (P)
	vtairflow	VT air flow (P)
	vtairlimit	S VT air flow limits (P)

pintvast Plot VAST Intergral Data in a stacked 1D-NMR matrix format

Applicability VnmrJ 3.1

- Description If an array of 1D spectra have been acquired (in particular if a block of 96 spectra has been acquired using VAST automation, especially in a microtiter-plate format), and if these spectra have been glued into a reconstructed 2D dataset (see vastglue), this macro will arrange and plot the integrals (on the plotter) in a convenient 8 x 12 sample format (as a matrix of 1D spectral integrals).
- Arguments The default is to plot all the intergrals (from 1 through arraydim). An optional argument (plvast(##)) allows one to specify that only intergrals from 1 through ## should be plotted.

See also dsvast dsvast2d plvast plvast2d pintvast

pir Plot integral amplitudes below spectrum (C)

Description	Plots integral amplitudes below the appropriate spectral regions.		
See also	NMR Spectroscopy User Guide		
Related	dpf Display peak frequencies over spectrum (C)		
	dpir Display integral amplitudes below spectrum (C)		
	pirn Display normalized integral amplitudes below spectrum (M)		
	Plot normalized integral amplitudes below spect	rum (M)	
	Plot peak frequencies over spectrum (M)		

pirn Plot normalized integral amplitudes below spectrum (M)

Description Equivalent to the command pir except that the sum of the integrals is normalized to the value of the parameter ins.

See also	NMR Spectroscopy User Guide		
Related	ed dpirn Display normalized integral amplitudes below spect		
	ins	Integral normalization scale (P)	
pir Plot integral amplitudes below spectrum (C		Plot integral amplitudes below spectrum (C)	

piv Plot integral values below spectrum (M)

Syntax piv<(vertical_position)>

Description Labels integrals with a bracket below the spectrum and a vertical number indicating the integral value. See dpiv for description and use.

Related	dpir	Display integral amplitudes below spectrum (C)
	dpiv	Display integral amplitudes below spectrum (M)
	dpirn	Display normalized integral amplitudes below spectrum
		(C)
	dpivn	Display normalized integral amplitudes below spectrum
		(M)
	pirn	Plot normalized integral amplitudes below spectrum (C)
	pir	Plot integral amplitudes below spectrum (C)
	pivn	Plot normalized integral amplitudes below spectrum (M)

pivn Plot normalized integral values below spectrum (M)

Syntax	pivn<(vertical_position)>		
Description	Labels integrals with a bracket below the spectrum and a vertical number indicating the integral value. See $dpiv$ for description and use.		
Related	dpir Display integral amplitudes below spectrum (C)		
	dpiv Display integral amplitudes below spectrum (M)		
	dpirn Display normalized integral amplitudes below spectrum (C		
	dpivn Display normalized integral amplitudes below spectrum (M		
	pirn Plot normalized integral amplitudes below spectrum (C		
	pir Plot integral amplitudes below spectrum (C)		
	piv Plot integral amplitudes below spectrum (M)		

pl Plot spectra (C)

688

entire integral, and intmod='partial' plots every other integral region.

For arrayed 1D spectra or for 2D spectra, a particular trace can be plotted by supplying the index number as an argument. For 2D data sets, spectra can be plotted from either the f_1 or f_2 domain by setting the parameter trace to 'f1' or 'f2', respectively. After the command ft1d, interferogram can be plotted by setting trace='f1' and then typing pl. Multiple spectra can be plotted by supplying the indexes of the first and last spectra.

The position of the first spectrum is governed by the parameters wc, sc, and vp. For 1D data, subsequent spectra are positioned relative to the preceding spectrum by the vertical and horizontal offset parameters vo and ho. For 2D data, ho defines the total horizontal offset between the first and last spectrum. Also for 2D data, vo is inactive while the parameter wc2 defines the total vertical offset between the first and last spectrum.

The parameter cutoff, if it exists and is active, defines the distance above and below the current vertical position vp at which peaks are truncated. By arraying cutoff to have two different values, truncation limits above and below the current vertical position can be controlled. For example, cutoff=50 truncates peaks at vp+50 mm and vp-50 mm. cutoff=50,10 truncates peaks at vp+50 mm and vp-10 mm.

Arguments start is the index of a particular trace for arrayed 1D or 2D spectra. For multiple spectra, start is the index of the first spectrum.

finish is the index of the last spectrum for multiple spectra.

step specifies the increment for the spectral index. The default is 1.

'int' is a keyword that specifies displaying only the integral, independently of the value of intmod.

'all' is a keyword to plot all of the spectra. This value is the default.

options can be any of the following keywords:

- 'top' or 'side' cause the spectrum to be plotted either above or at the left edge of a contour plot. This assumes that the parameters sc, wc, sc2, and wc2 are those used to position the contour plot.
- 'dodc' causes all spectra to be drift corrected independently.
- 'pen1', 'pen2', 'pen3', etc. specify a pen number on a plotter.

Examples pl

pl(1,6,2)

See also NMR Spectroscopy User Guide

Related	cutoff	Data truncation limit (P)
	dssa	Display stacked spectra automatically (C)
	dsww	Display spectra in whitewash mode (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ho	Horizontal offset (P)
	intmod	Integral display mode (P)
	plww	Plot spectra in whitewash mode (C)
	pshr	PostScript High Resolution plotting control (P)

pslw	PostScript Line Width control (P)
SC	Start of chart (P)
sc2	Start of chart in second direction (P)
shownumx	\boldsymbol{x} position counting from bottom left of every
	spectrum (P)
shownumy	y position counting from bottom left of every
	spectrum (P)
trace	Mode for 2D data display (P)
vo	Vertical offset (P)
vp	Vertical position of spectrum (P)
WC	Width of chart (P)
wc2	Width of chart in second direction (P)

pl2d Plot 2D spectra in whitewash mode (C)

pl2d<('nobase' 'fill' 'fillnb')>		
Plots a stacked plot of 2D spectra in whitewash mode (after the first spectra, each spectra is blanked out in regions in which it is behind an earlier spectra). Color does not represent intensity (unlike dcon), since intensity can be seen visually, but instead successive traces are displayed in different colors so that color represents frequency. The horizontal offset parameter ho is not active for this command.		
'nobase' is a keyword to activate th to suppress intensity below th.		
'fill' is a keyword to fill in the peaks. Note that if 'fill' (or 'fillnb') is used, th operates linearly and not logarithmically (w factors of 2) as it does in contour or color intensity displays.		
'fillnb' is a keyword to combine base suppression and peak filling.		
pl2d pl2d('nobase')		
NMR Spectroscopy User Guide		
dconDisplay noninteractive color intensity map (C)ds2dDisplay 2D spectra in whitewash mode (C)dswwDisplay spectra in whitewash mode (C)hoHorizontal offset (P)plwwPlot spectra in whitewash mode (C)thThreshold (P)		

plane Currently displayed 3D plane type (P)

Description Stores the type of 3D plane currently displayed within VnmrJ. If plane does not exist, it is created by the macro par3d. The command select, as well as the many macros that make use of select, requires

the parameter plane to exist for 3D data sets and to contain an appropriate value.

plane is set automatically by the macro getplane; it can also be set by the macro ft3d if automatic plane extraction is requested at the end of the 3D FT. The order of priority for the plane types is 'f1f3', 'f2f3', and then 'f1f2'. In other words, if getplane is requested to extract the f_1f_3 and the f_2f_3 planes, plane will be set to 'f1f3'. plane can also be set manually.

Values 'f1f3', 'f3f1', 'f2f3', 'f3f2', 'f1f2', or 'f2f1'

See also NMR Spectroscopy User Guide

demost Display a 2D plana projection (M)	
aproj Display a 5D plane projection (M)	
dsplanes Display a series of 3D planes (M)	
ft3d Perform a 3D Fourier transform on a 3D FID data	set
(M ,U)	
getplane Extract planes from a 3D spectral set (M)	
nextpl Display the next 3D plane (M)	
par3d Create 3D acquisition, processing, display parameter	ers (C)
path3d Number of complex points to left-shift np FID (P)	
plplanes Plot a series of 3D planes (M)	
prevpl Display the previous 3D plane (M)	
select Select a spectrum or 2D plane without displaying	it (C)

plapt Plot APT-type spectra automatically (M)

Syntax	<pre>plapt<(13Cexp_number)></pre>		
Description	Automatically plots APT spectra. The APT spectrum is plotted on top of a standard carbon spectrum if either an experiment with such data is specified or if a file C13 is found in curexp+'/subexp'. If neither such a subfile is found nor an experiment with standard carbon data is specified, the APT spectrum is plotted alone.		
Arguments	13Cexp_number specifies the number, from 1 to 9, of an experiment with a standard 13 C spectrum.		
Examples	plapt plapt(2)		
See also	NMR Spectroscopy User Guide		
Related	curexp Current experiment directory (P)		

plarray Plotting macro for arrayed 1D spectra (M)

Description A generic macro for plotting arrayed 1D spectra. plarray is called by the plot macro, but can also be used directly. For the plot layout, procarray distinguishes between arrays with few elements (6 or less), which will be stacked vertically (no horizontal offset), and spectra with many (greater than 6) elements. Those are stacked horizontally by default, unless there are too many lines, in which case a diagonally stacked display is chosen. Horizontal stacking is mostly adequate for pulse and power calibrations, where there are usually few lines only; diagonally stacked displays/plots are frequently chosen for T_1 and T_2 experiments on entire spectra, often with many lines.

The automatic stacking mode can be overridden by creating and setting a string parameter stackmode in the startup macro or before calling procplot or procarray. Possible values for stackmode are 'horizontal', 'vertical', or 'diagonal'. DEPT-type spectra can, in principle, also be processed with procarray, but no DEPT editing occurs, of course.

See also NMR Spectroscopy User Guide

Related	aexppl	Automatic expansion plot (M)
	plc	Plot carbon spectrum (M)
	plh	Plot proton spectrum (M)
	plot	Automatically plot spectra (M)
	procarray	Process arrayed 1D spectra (M)
	stackmode	Stack control for processing arrayed 1D spectra (P)

plate_glue Define a glue order for plotting and display (U)

Applicability	Systems with VAST accessory		
Description	In a Unix terminal or shell window type plate_glue. The glue order is determined by clicking on the wells to be displayed. Save the glue order file in the user's vnmrsys/templates/glue directory.		
See also	NMR Spectro	oscopy User Guide	
Related	dsvast2dDisplay VAST data in a pseudo-2D format (M)plvastPlot VAST data in a stacked 1D-NMR matrix (M)plvast2dPlot VAST data in a pseudo-2D format (M)		

plc Plot a carbon spectrum (M)

Syntax	plc<(pltmod)>
Description	Plots a carbon spectrum based on the parameters pltmod (the options 'off', 'full', and 'fixed' are implemented) and intmod ('off', 'full', and 'partial' are implemented). Peak frequency labels, in ppm, are usually plotted.
Arguments	pltmod is an alternate value of pltmod for this macro only. The value of the pltmod parameter is not changed.
Examples	plc plc('full')

See also	NMR Spectro	oscopy User Guide
Related	intmod	Integral display mode (P)
	pltmod	Plotter display mode (P)

plcnmr Plot all forms of LC-NMR data (M)

Applicability	VnmrJ 3.1
Description	This macro is executed with a button on the LC-NMR display pane (labeled spare). Plots on-flow and stopped-flow 1D LC-NMR data. With on-flow data, the NMR data is plotted with the time-aligned LC detector trace(s) along the left side. In the stopped-flow mode, pLCNMR plots the 1D NMR data for each stop code at a position that it is time-aligned with the relevant LC peak.
Examples	pLCNMR(<number contours="" of="">,<contour spacing="">)</contour></number>
See also	dlC
	pLC
	dlcnmr
	plCNMR

plcosy Plot COSY- and NOESY-type spectra automatically (M)

Syntax	plcosy(<'pos' 'neg'><,> <levels<,spacing<,exp1d>>>)</levels<,spacing<,exp1d>
Description	Automatically plots 2D COSY- and NOESY-type spectra (homonuclear correlated spectra). Features include the following:
	• Keeps the orientation (f_1, f_2) of the spectrum on the screen.
	• Plot area is optimized.
	• Number of contour levels and their spacing can be selected.
	• Negative or positive contours can be suppressed.
	• 1D traces can be plotted along both axes; such 1D traces are taken from a full (or reduced) 1D spectrum in an other experiment, or from a subfile from within the current experiment.
	• Works correctly for expansions.
	• 1D traces can be suppressed, allowing a larger area for the 2D spectrum.
	• 1D spectrum can be in any experiment.
	• With phase-sensitive spectra using a plotter with one pen or a printer such as a LaserJet, if 'pos' or 'neg' are not selected, seven positive levels (or the specified number of positive contours) and one negative level are plotted, to distinguish positive and negative signals.
	In multiexperiment mode, for the first plot, the experiment with the 1D spectrum should be specified (at least if it is not in exp1). From then on, the 1D spectrum will be stored <i>within</i> the experiment with

the 2D spectrum, which allows much faster switching between spectra and also frees the other (1D) experiment for other tasks. Because of this internal storage, the exp1D argument is not required for subsequent plots.

Arguments 'pos' is a keyword to plot only positive contours. 'neg' is a keyword to plot only negative contours. levels is the number of contour levels. The default is 7. spacing is the spacing between the contours. The default is 2. explD is the experiment in which the proton 1D spectrum resides. This can be a full 1D spectrum, but the referencing must be the same as for the 2D. A negative number suppresses the proton trace. The default is from a subfile. Examples plcosy plcosy(12, 1.5)plcosy('pos',7,2,3) plcosy(7, 2, -1)plcosy('neg') See also NMR Spectroscopy User Guide

pldept Plot DEPT data, edited or unedited (M)

Description	Plots out DEPT data, either edited or not edited.	
See also	NMR Spectre	pscopy User Guide
Related	adept autodept deptproc padept	Automatic DEPT analysis and spectrum editing (C) Automated complete analysis of DEPT data (M) Process DEPT data (M) Perform adept analysis and plot resulting spectra (C)

plexpinfo Plots Experiment Information

Description	Plots experiment information at a specified position on the page.
Syntax	plexpinfo(x,y)
Examples	plexpinfo(32,210)
Related	pllogo, pltext, pltime, pap, ppa, pll, plexpinfo

plfid Plot FIDs (C)

Description	Plots one or the parameter relative to the parameters v	more FIDs. The position of the first FID is governed by ers wc, sc, and vpf. A subsequent FID is positioned are preceding FID by the vertical and horizontal offset ro and ho.
Arguments	start is the For multiple	index of a particular FID for arrayed 1D or 2D data sets. FIDs, start is the index of the first FID.
	finish is th FIDs, set sta example).	he index of the last FID for multiple FIDs. To include all art to 1 and finish to the parameter arraydim (see
	step specifie	es the increment for the FID index. The default is 1.
	'all' is a k	eyword to plot all of the FIDs. This is the default.
	'imag' is a default is 'a	keyword to plot the imaginary FID channel only. The ll'.
	pen is a key 'pen3', etc.	word with the plotter pen number: 'pen1', 'pen2', The default is 'pen1'.
Examples	<pre>plfid(1,arraydim,3)</pre>	
See also	NMR Spectro	scopy User Guide
Related	arraydim	Dimension of experiment (P)
	dfs	Display stacked FIDs (C)
	dfww	Display FIDs in whitewash mode (C)
	ho	Horizontal offset (P)
	SC	Start of chart (P)
	vo	Vertical offset (P)
	vpf	Current vertical position of FID (P)
	WC	Width of chart (P)

plfit Plot deconvolution analysis (M)

Produces a c the observed component, a	complete output plot of a deconvolution analysis, plotting spectrum, the full calculated spectrum, each individual as well as the numerical results of the analysis.
NMR Spectroscopy User Guide	
fitspec showfit usemark	Perform spectrum deconvolution (C) Display numerical results of deconvolution (M) Use "mark" output as deconvolution starting point (M)
	Produces a of the observed component, a <i>NMR Spectro</i> fitspec showfit usemark

plgrid Plot a grid on a 2D plot (M)

Syntax	<pre>(1) plgrid<(<spacing><,><pen>)></pen></spacing></pre>
	<pre>(2) plgrid<(start_f2,incr_f2,start_f1,incr_f1<,pen>)></pre>
Description	Plots grid lines over a 2D plot.

Arguments	spacing specifies the approximate spacing of the grid lines, in cm. The default is intervals of approximately 1 cm, rounded so that the intervals fall at a multiple of 1, 2, or 5 (in Hz) or 1p, 2p, or 5p (in ppm).
	<pre>pen is a keyword with the plotter pen number: 'pen1', 'pen2', 'pen3', etc. The default is 'pen1'.</pre>
	start_f2, incr_f2, start_f1, incr_f1 define the starting and increment frequencies in both f_2 and f_1 for a grid. Add the p suffix to a value to enter it in ppm (see last example below).
Examples	<pre>plgrid plgrid(2) plgrid('pen5') plgrid(1.5, 'pen2') plgrid(1p, 0.5p, 3p, 0.5p)</pre>
See also	NMR Spectroscopy User Guide
Related	grid Draw a grid on a 2D display (C)

plh Plot proton spectrum (M)

Syntax	plh<(pltmo	d)>
Description	Plots a proton 'off', 'fixe intmod ('of	n spectrum based on the parameters pltmod (the options ed', 'full', and 'variable' are implemented) and f', 'full', and 'partial' are implemented).
Arguments	pltmod is an only. The val	alternate value of the parameter pltmod for this macro ue of the pltmod parameter is not changed.
Examples	plh plh('full')
See also	NMR Spectro	scopy User Guide
Related	intmod	Integral display mode (P)
	pltmod	Plotter display mode (P)
	sp	Start of plot (P)
	wp	Width of plot (P)

plhet2dj Plot heteronuclear J-resolved 2D spectra automatically (M)

Syntox	nlhot2dic(Inogl/Inogle lovelge angginge ovn1D>>>)>
Syntax	pinetzaj<('pos' 'neg'<,ieveis<,spacing<,expiD>>>)>
Description	Automatically plots 2D spectra of type HET2DJ (heteronuclear
	J-resolved 2D spectra) with the following features:
	• Displayed portion of the spectrum is plotted in f2-mode
	• Plot area is optimized

• Number of contour levels and their spacing can be selected

- Negative or positive contours can be suppressed
- A 1D trace can be plotted along the f_2 axis; such a 1D trace is taken from a full (or reduced) 1D spectrum in an other experiment, or from a file from within the current experiment.
- Expansions are handled correctly
- The 1D trace can be suppressed, which allows using a larger area for the 2D spectrum
- The 1D spectrum can be in any experiment
- With phase-sensitive spectra, if 'pos' or 'neg' are not selected and the plotter has only one pen (also for printers like the LaserJet), the specified number of positive contours are plotted (default is 7), but only one negative level, to distinguish positive and negative signals.

In multiexperiment mode, for the first plot the experiment with the 1D spectrum should be specified (at least if it is not in exp1). From then on, the 1D spectrum is stored *within* the experiment with the 2D spectrum, which allows much faster switching between the spectra and also frees the other 1D experiment for other tasks. Because of this internal storage, the exp1D argument is not required for subsequent plots.

Arguments 'pos' is a keyword to only plot positive contours

'neg' is a keyword to only plot negative contours

levels is the number of contour levels. The default is 7.

spacing is the spacing between the contours. The default is 2.

explD is the number from 1 to 9 of the experiment in which the 1D spectrum resides. This can be a full 1D spectrum, but the referencing must be the same as for the 2D. A negative number will suppress the 1D trace. The default is 1 (for expl).

Examples plhet2dj plhet2dj(12,1.5) plhet2dj('pos',7,2,3) plhet2dj(7,2,-1) See also NMR Spectroscopy User Guide

see also Mink Speciroscopy User Guide

plhom2dj Plot homonuclear J-resolved 2D spectra automatically (M)

Syntax (1) plhom2dj<(levels<, spacing<, exp1D>>)>
 (2) plhom2dj('pos'|'neg'<, levels<, spacing<, exp1D>>>)
Description Automatically plots 2D spectra of type HOM2DJ (homonuclear
J-resolved 2D spectra). Features include the following:
 •The displayed portion of the spectrum is plotted in f2-mode
 •The plot area is optimized

- Number of contour levels and their spacing can be selected
- Negative or positive contours can be suppressed

Ρ

	• A 1D trace can be plotted along the f_2 axis; such a 1D trace is taken from a full (or reduced) 1D spectrum in an other experiment, or from a file from within the current experiment.
	• It also works correctly for expansions
	• The 1D trace can be suppressed, which allows using a larger area for the 2D spectrum
	• The 1D spectrum can be in any experiment
	• With phase-sensitive spectra, if 'pos' or 'neg' are not selected and the plotter has only 1 pen (also for printers like the LaserJet) 7 or the specified number of positive contours are plotted, but only one negative level, to distinguish positive and negative signals.
	In multiexperiment mode, for the first plot the experiment with the 1D spectrum should be specified (at least if it is not in exp1). From then on, the 1D spectrum will be stored <i>within</i> the experiment with the 2D spectrum, which allows much faster switching between the spectra and also frees the other (1D) experiment for other tasks. Because of this internal storage, the exp1D argument is not required for subsequent plots.
Arguments	levels is the number of contour levels. The default is 7.
	spacing is the spacing between the contours. The default is 2.
	explD is a number from 1 to 9 for the experiment in which the 1D spectrum resides. The spectrum can be a full 1D spectrum but the referencing must be the same as for the 2D. A negative number will suppress the 1D trace. The default is 1 (for expl).
	'pos' specifies only plot positive contours.
	'neg' specifies only plot negative contours.
Examples	plhom2dj plhom2dj(25,1.2) plhom2dj('pos',7,2,3) plhom2dj(7,2,-1)
See also	NMR Spectroscopy User Guide

plhxcor Plot X,H-correlation 2D spectrum (M)

Syntax	<pre>plhxcor(<'pos' 'neg'><,><levels<,spacing <,exp1d_h<,exp1d_x="">>>)</levels<,spacing></pre>
Description	Automatically plots 2D spectra of type HETCOR, COLOC, HMQC, HMBC (direct and indirect detection). Features include the following:
	• Keeps the orientation (f_1, f_2) of the spectrum on the screen.
	• Plot area is optimized.
	• Number of contour levels and their spacing can be selected.
	• Negative or positive contours can be suppressed.

• 1D proton and X traces can be plotted along both axes; such 1D traces are taken from full (or reduced) 1D spectra in other experiments or subfile within the current experiment. • Works correctly for expansions. • 1D traces can be suppressed, allowing a larger area for the 2D spectrum. • 1D spectra can be in any experiment. Arguments 'pos' is a keyword to plot only positive contours. 'neg' is a keyword to plot only negative contours. levels is the number of contour levels. The default is 7. spacing is the spacing between the contours. The default is 2. explD_H is a number from 1 to 9 of the experiment in which the proton 1D spectrum resides; this can be a full 1D spectrum, but the referencing must be the same as for the 2D. A negative number will suppress the proton trace. The default is a subfile in the current experiment. explD X is a number from 1 to 9 of the experiment in which the X 1D spectrum resides. A negative number suppresses the X trace. the default is a subfile in the current experiment. Examples plhxcor(12,1.5) plhxcor(7,2,3)plhxcor(7,2,1,3) plhxcor('pos',7,2,-1,3) plhxcor(7, 2, -1, -1)plhxcor('neg') See also NMR Spectroscopy User Guide Related hetcor Set up parameters for HETCOR pulse sequence (M)

pll Plot a line list (M)

Syntax	<pre>pll<(x,y,minimum_y)></pre>
Description	Produces a columnar line list on a plotter, similar to what would appear on a printer. pll is quite different from the alternative method of plotting peak frequencies using ppf. The output of pll is automatically formatted into multiple columns, depending on the number of lines.
Arguments	x is the x position of the upper left of the line list.
	$_{\rm Y}$ is the y position of the upper left of the line list.
	minimum_y is the minimum y at which to reset back to top.
Examples	<pre>pll pll(20,150) pll(5,wc2max*.8,wc2max*.5)</pre>

See alsoNMR Spectroscopy User GuideRelatedppfPlot peak frequencies over spectrum (M)

pllogo Plots Logo

Description Plots a logo. Syntax pllogo(x,y) Examples pllogo(32,220) Related pllogo, pltext, pltime, pap, ppa, pll, plexpinfo

pll2d Plot results of 2D peak picking (C)

Syntax	pll2d<(options)>
Description	Plots the results of applying the 112d command to pick 2D peaks in a 2D spectrum or a 2D plane of a 3D spectrum. Refer to the description of 112d for a description of the process and the options available.
See also	NMR Spectroscopy User Guide
Related	Automatic and interactive 2D peak picking (C)

Plock Sets Protection Bit for a Parameter

Description	Sets the protection bit for a parameter given as an argument. This
	causes the specified parameter to be read from the appropriate
	parlib entry upon experiment set up, rather than inherited from the
	current workspace.
Syntax	Plock(parameter)
Examples	Plock('samplename')

plockport Port number to use to lock out multiple ProTune processes (P)

Syntax	plockport= <value></value>
Description	The parameter must be created as a real local parameter before it can
	be used. The parameter is used to override a default port number that

is used internally in ProTune to prevent two Java ProTune process from running simultaneously.

Related	protune	Macro to start ProTune (M)
	create	Create new parameter in a parameter tree
		(C)

plot Automatically plot spectra (M)

Description A universal plotting macro normally called through the procplot macro (which by itself serves as processing and plotting facility for automatic experiments). plot can also be used directly by the user who then doesn't have to remember specific plotting macros. Of course, the specialized macros can still be called directly if the user know their names.

> The main purpose of plot is to automatically call the correct specialized plotting macro, depending on the user definition or otherwise on the type of data in the experiment. A plotting macro is selected automatically as follows:

APT spectra:	plapt
other, non-arrayed 1D data:	plot1d
DEPT type arrayed spectra:	pldept
other arrayed 1D spectra:	plarray
J-resolved 2D spectra:	pl2dj
homonuclear correlation 2D spectra:	plcosy
heteronuclear correlation 2D spectra:	plhxcor

Other types of 2D spectra (mostly multiple-quantum 2D spectra such as 2D-INADEQUATE) are not plotted automatically at this time. For phase-sensitive 2D spectra, automatic plotting is only provided if they were acquired using the method described by States, Haberkorn, and others; TPPI spectra are not covered.

Note that plot macros in general should not adjust the phase, the vertical scale, or change the integral size and reset points; these are assumed to be adjusted either by hand or by a suitable processing macro like procplot and the macros called therein. The plotting macros only make adjustments in order to make spectrum and parameters fit onto the page the desired way.

See also NMR Spectroscopy User Guide

RelatedapptypeApplication type (P)execparsSet up the exec parameters (M)execplotExecute plotting macro (P)plaptPlot APT spectra (M)plarrayPlot arrays (M)plcosyPlot homonuclear 2D correlation spectra (M)

pldept	Plot DEPT type spectra (M)
plhxcor	Plot heteronuclear correlation spectra (M)
plot1d	Plot 1D spectra (M)
plt2Darg	Plot 2D arguments (P)
procplot	Automatically process FIDs (M)

plot_<pslabel>

Description

plot1d Plotting macro for simple (non-arrayed) 1D spectra (M)

- Description A generic macro for plotting non-arrayed 1D spectra using a set of standard macros. plot1d is called by the plot macro, but can also be used directly. plot1d first tries to find a specific macro (e.g., plh, plc, plp) for the current observe nucleus. If such a macro exists, it is called. If a nucleus-specific macro is not found in the command path, a "minimal" 1D plot is produced.
 - See also NMR Spectroscopy User Guide

Related	plc	Plot carbon spectrum (M)
	plh	Plot proton spectrum (M)
	plp	Plot phosphorus spectrum (M)
	plot	Automatically plot spectra (M)

plot2D Plot 2D spectra (M)

- Syntax plot2D('pos'|'neg'|'both',levels,spacing, \ 'top'|'notop'|'proj','side'|'noside'|'proj')
- Description Checks for the presence of appropriate proton or carbon high-resolution spectra in the directory userdir+'/data/'+sample and decides to plot high resolution spectra or a projection depending on whether or not the proton or carbon spectrum exists.

Arguments The plot2D macro accepts the following arguments:

'pos'	keyword to plot positive contours
'neg'	keyword to plot negative contours.
'both'	keyword to plot both positive and negative contours.
levels	number of levels to be plotted.
spacing	spacing between contour levels.
'top'	keyword to plot a high-resolution spectrum on the top.
'notop'	keyword to plot a non-high-resolution spectrum or projection.
'proj'	keyword to plot a projection on top.
'side'	keyword to plot a high-resolution spectrum on the side.

keyword to plot positive contours 'pos' 'neg' keyword to plot negative contours. 'noside' keyword to plot a non-high-resolution spectrum or projection. keyword that plots a projection on the side. 'proj' Examples plot2D('pos',2,5,'top','side') See also NMR Spectroscopy User Guide Related plot Automatically plot spectra (M) Plot spectrum on side (M) plotside plottop Plot spectrum on top (M) plottopside Plot spectrum on top and side (M)

plotfile Plot to a file (M)

Syntax	<pre>plotfile('argument')</pre>
Description	plots automatically to a file. Supported output formats are: ps, pdf,
	jpg, pcl, hpgl. and png.
Arguments	auto - plots automatically.
	manual - plots contents of printer queue to a file.
	Path and file name — plots to specified file in the directory specified. Plots to the data directory using the supplied name if no path is specified.
Examples	<pre>plotfile('xxx.fid/myplotfile.PDF') plots will go into saved data directory.</pre>
	<pre>plotfile('myplotfile.PDF') - plots will go to vnmrsys/plots if FID has not been saved.</pre>

plothiresprepHigh resolution plot output preparation (M)

Description Required for the operation of the "Plot HiRes..." popup window to interactively use plottop/plotside of spectra in work spaces EXPn - creates necessary variables.

plotlcnmr An LC-NMR plotting macro (M)

Syntax

Applicability VnmrJ 3.1

Description The NMR data for a particular peak can be plotted using plotlcnmr with the number of the peak as an argument. While this can also be accomplished with the pl command, plotlcnmr labels the plot with the LC retention time of the peak and the Cascade file name associated with the LC data.

plotmanual Plot manually (M)

Description Makes correct choice of printer (for preview) and correct alignment with respect to parameter output, resets back screen to original size & position based on selections made on the Plot page.

plotlogo Plots a logo (M)

Description Plots a logo Varian logo using image file located in /vnmr/iconlib/varianlogo.gif or a custom logo from location specified in the parameter plotlogo. Reads value for doplotlogo (n/y), plotlogox (x dimension image), and plotlogoy (y dimensions image), and image file in iconlib.

plotpreview Creates temporary plots of the current plot output (M)

<pre>plotpreview<('argument')></pre>
Creates preview of the output from auto-plotting the current spectrum and starts an Acrobat PDF reader. The preview output can be saved in PS, PDF, PCL, HPGL, JPG or PNG formats.
no $\operatorname{argument}$ – creates preview of whatever is ready to send to the plotter.
auto - creates preview of auto-plot based upon plot macro
manual - creates preview of the contents of the print queue.

plotside Plot spectrum on side (M)

Description	Plots projection or high-resolution spectrum on the side of a 2D		
	spectrum. p	lotside is used with $plot2D$ and is not useful by itself.	
See also	NMR Spectr	oscopy User Guide	
Related	plot2D	Plot 2D spectra (M)	

plotter Plotter device (P)

Description	Sets the plotter in use on the system.
Values	A string with entries such as 'DraftPro', 'ThinkJet_96',
	'LaserJet_300', 'jim', 'varian1', and 'Laser1'.

See also NMR Spectroscopy User Guide Related setplotdev Return characteristics of a named plotter (C) showplotter Show list of currently defined plotters and printers (M)

plottop Plot spectrum on top (M)

Description	Plots proje	ection or high resolution spectra on the top of a 2D spectrum.
	plottop	is used with plot2D and is not useful by itself.
See also	NMR Spec	troscopy User Guide
Related	plot2D	Plot 2D spectra (M)

plottopside Plot spectrum on top and side (M)

Description Plots projection or high-resolution spectrum on the top and side of a 2D spectrum. plottopside is used with plot2D and is not useful by itself.
See also NMR Spectroscopy User Guide
Related plot2D Plot 2D spectra (M)

plp Plot phosphorus spectrum (M)

Syntax	plp<(pltmo	d) >
Description	Plots a phosphorus spectrum based on the parameters pltmod (the options 'off', 'full', and 'fixed' are implemented) and intmod ('off', 'full', and 'partial' are implemented). Peak frequency labels, in ppm, are usually plotted.	
Arguments	pltmod is an of the pltmo	alternate value of pltmod for this macro only. The value d parameter is not changed.
Examples	plp plp('full')
See also	NMR Spectro	scopy User Guide
Related	intmod	Integral display mode (P)
	plh	Plot proton spectrum (M)
	pltmod	Plotter display mode (P)

- Description Creates the 2D contour plots for a subset of the 3D planes specified by the parameter plane.
- Arguments start_plot specifies the number, greater than 0, of the 3D plane with which plotting is to begin.

stop_plot specifies the number of the 3D plane with which plotting is to end. If start_plot is greater than stop_plot, only the first plane, whose number is start_plot, is plotted. The range of stop_plot depends on the value of the parameter plane:

- if plane='flf3', stop_plot is between 0 and fn2/2
- if plane='f2f3', stop_plot is between 0 and fn1/2
- if plane='flf2', stop_plot is between 0 and fn/2

'pos' is a keyword specifying that phase-sensitive spectra plot positive peaks only. The default is to plot both positive and negative peaks.

'neg' is a keyword specifying that phase-sensitive spectra plot negative peaks only. The default is to plot both positive and negative peaks.

levels is maximum number of contour levels to plot. The default is 4.

spacing is relative intensity of successive contour levels. The default is 2.

Note that the optional arguments 'pos' | 'neg', number_levels, and spacing are for the VnmrJ plotting command pcon.

Examples plplanes(1,3) plplanes(2,3,'pos',4)

See also NMR Spectroscopy User Guide

Related dplane Display a 3D plane (M) Display a 3D plane projection (M) dsplanes Display a series of 3D planes (M) getplane Extract planes from 3D spectral data set (M) nextpl Display the next 3D plane (M) path3d Path to currently displayed 2D planes from a 3D data set (P) Plot contours on a plotter (C) pcon Currently displayed 3D plane type (P) plane prevpl Display the previous 3D plane (M)

plt2Darg Plot 2D arguments (P)

Applicability Liquids

Description Specifies options for contours and 1D projections on 2D plots, used by the plot2D macro. The plot options are selected on the Defaults page in the Acquire folder for most 2D sequences.

Related plot2D Plot 2D spectra (M)

pltext Plot text file (M)

Syntax	<pre>pltext<(<file><,x<,y<,width>>>)> <:\$x next,\$y next,\$y increment></file></pre>		
Description	Plots a text file.		
Arguments	file is the name of a text file. The default is the current experimen text file.		
	x and y are the location of the page.	coordinates, in mm, of the first line of text. This positions of the output. The default is the upper left-hand corner	
	width is the a word wrap	maximum column text width, in characters. pltext uses to make the text fit into the width specified.	
	\$x_next and line would h plotting.	l y_next are the coordinates where the start of the next ave been plotting. This is useful for subsequent character	
	\$y_increme	ent is the vertical increment between lines.	
Examples	pltext		
	pltext(wcm	ax-70)	
	pltext(use	rdir+'/exp3/text')	
pitext(100,100)		,100)	
	pltext(use	\$v.\$dv	
See also	NMR Spectroscopy User Guide		
Related	dtext	Display a text file in the graphics window (C	
	ptext	Print out a text file (M)	
	text	Display text or set new text for current experiment (C)	
	userdir	User directory (P)	

pltmod Plotter display mode (P)

Description Controls plotting of a proton, carbon, or phosphorus spectrum. Values 'off' sets no plotting. 'fixed' takes sp and wp as is. 'full' adjusts sp and wp to plot the full spectrum. 'variable' adjusts sp and wp to plot only the region of interest.

See also	NMR	Spectroscopy User Guide
Related	plc	Plot carbon spectrum (M)
	plh	Plot proton spectrum (M)
	plp	Plot phosphorus spectrum (M)
	sp	Start plot (P)

plvast Plot VAST Data in a stacked 1D-NMR matrix format

Width of plot (P)

Applicability VnmrJ 3.1

wp

Description If an array of 1D spectra have been acquired (in particular if a block of 96 spectra have been acquired using VAST automation, especially in a microtiter-plate format), and if these spectra have been glued into a reconstructed 2D dataset (see vastglue), this macro will arrange and plot them (on the plotter) in a convenient 8 × 12 sample format (as a matrix of 1D spectra). Uses a file (template) created by plate_glue to display a matrix of data.

The number of spectra displayed, and their order, are controlled by the template file. Each "little spectrum" is labeled with its respective alphanumeric coordinates. The modulo number controls how many spectra appear per row.

Examples plvast(<display order>, <modulo>)

See also dsvast dsvast2d plvast plvast2d intvast pintvast plateglue vastglue

vastget

plvastget Plot VAST spectral data in a vertical stacked plot mode

Applicability Description	VnmrJ 3.1 This macro selects and plots the spectra from any arbitrary well or wells using the label(s) as an argument. The spectra are displayed in a dss stacked plot.
Examples	<pre>vastget("B6","B7","C11","G3") will display four spectra.</pre>
See also	dsvast dsvast2d plvast plvast2d intvast

pintvast plateglue vastglue vastget

Applicability	VnmrJ 3.1		
Description	This macro plots all the spectra in a glued dataset, one spectrum per page of paper. This mimics the plots obtained automatically during data acquisition, but allows the data to be rephased or reprocessed.		
Examples	plvast_replot(96) will replot all 96 spectra		
See also	dsvast dsvast2d plvast plvast2d intvast plateglue vastglue vastglue		

plvast2d Plot VAST data in a stacked pseudo-2D format (M)

Applicability	Systems with the VAST accessory.		
Syntax	plvast2d<(number)>		
Description	If an array of 1D spectra have been acquired (in particular if a block of 96 spectra has been acquired using VAST automation, especially in a microtiter-plate format) and if these spectra have been glued into a reconstructed 2D dataset (see vastglue), plvast2d will arrange and plot them (on the plotter) in a convenient pseudo-2D format (almost like an LC-NMR chromatogram). Well labels are not attached to the spectra and spectra are plotted with 12 spectra per row.		
Arguments	<pre>number specifies that only spectra from 1 through number should be plotted. The default is to plot all the spectra (from 1 through arraydim). An optional argument (plvast(##)) allows one to specify that only spectra from 1 through ## should be plotted.</pre>		
See also	NMR Spectroscopy User Guide plvast2d pintvast		
Related	dsast2d Display VAST data in a pseudo-2D format (M)		

VnmrJ 4 Command and Parameter Reference Guide

dsvastDisplay VAST data in a stacked 1D-NMR matrix (M)plvastPlot VAST data in a stacked 1D-NMR matrix (M)

plww Plot spectra in whitewash mode (C)

Syntax	<pre>plww<(start,finish,step><,'all'>)></pre>		
Description	Plots one or more spectra in whitewash mode (after the first spectra, each spectra is blanked out in regions in which it is behind an earlier spectra).		
Arguments	start — index of the first spectra when plotting multiple spectra. It is also the index number of a particular trace to be plotted when plotting arrayed 1D spectra or 2D spectra. The default is to plot all spectra.		
	finish - index of the last spectra when plotting multiple spectra.		
	step $-$ increment for the spectral index when plotting multiple spectra, default is 1.		
	'all' — (defau	ult) keyword to plot all spectra in the array.	
See also	NMR Spectrosc	opy User Guide	
Related	dss I	Display stacked spectra (C)	
	dsww I	Display spectra in whitewash mode (C)	
	pl F	lot spectra (C)	

pmode Processing mode for 2D data (P)

Description Specifies the type of 2D spectral data that the 2D Fourier transform (FT) will yield. pmode is in the processing group.

Values '' (null string, shown by two single quotes with no space in between) specifies a processing mode in which it is not possible to change either the f_2 or f_1 display mode after the 2D FT. If the f_2 display mode has been set to phased (dmg='ph'), each f_2 spectrum is phase rotated using the phase constants rp and lp prior to the FT along the second dimension. If the f_2 display mode has been set to power (dmg='pwr') or absolute-value (dmg='av'), however, the f_2 spectrum is not processed any further after the first FT. The complex t_1 interferograms are handled in a similar manner. If the f_1 display mode has been set to phased (dmg1='ph1'), each f_1 spectrum is phased using the phase constants rp1 and lp1. If the display mode has been set to power (dmg1='pwr1') or to absolute value (dmg1='av1'), the appropriate magnitude calculation is performed, with the result being placed in the real part of the appropriate complex datum and a 0 being placed in the imaginary part. At the end of the 2D transform, the spectral data file datdir/data is reduced from complex data to real data ("VnmrJ REDUCE" display message).

'partial' specifies a processing mode in which it is not possible to change the f_2 display mode after the 2D FT. It is possible, however, to select between the three f_1 display modes without having to reprocess the 2D data. If the f_2 display mode has been set to phased (dmg='ph'), each f_2 spectrum is phase rotated using the phase constants rp and lp prior to FT along the second dimension. If the f_2 display mode is set to power (dmg='pwr') or absolute value (dmg='av'), the f_2 spectrum is not processed any further after the first FT. Regardless of the requested f_1 display mode, no further processing is performed by ft2d on the f_1 spectra after the second FT. The calculations on 2D spectral data necessary to achieve the requested f_1 display mode are performed by dcon or dconi. If pmode does not exist, it is assigned a value of 'partial' internal to VnmrJ.

'full' specifies a processing mode in which it is possible to select between the three display modes for each dimension without having to reprocess the 2D data. Regardless of any requested display mode, no display mode processing is performed by ft2d on the f_2 spectra after the first or second FT.

The hypercomplex data structure for the 2D time domain data is:

```
{Re(t1)Re(t2), Re(t1)Im(t2), Im(t1)Re(t2),
Im(t1)Im(t2)}
```

and is experimentally composed by the pulse sequence generation arraying mechanism. The hypercomplex data structure for the t_1 interferograms is:

```
{Re(t1)Re(F2), Re(t1)Im(F2), Im(t1)Re(F2),
Im(t1)Im(F2)}
```

where Re represents the real part and Im represents the imaginary part. A hypercomplex FT along t_1 yields a hypercomplex 2D spectrum with the following data structure per hypercomplex point:

```
{Re(F1)Re(F2), Re(F1)Im(F2), Im(F1)Re(F2),
Im(F1)Im(F2)}
```

Note that if pmode='full', the ft2d program will require an array index or coefficients for the construction of the t_1 interferograms.

See also NMR Spectroscopy User Guide

Related	av	Set abs. value mode in directly detected dimension (C)		
	av1	Set abs. value mode in 1st indirectly detected dimension (C)		
	dcon	Display noninteractive color intensity map (C)		
	dconi	ni Interactive 2D data display (C)		
	dmg	Data display mode in directly detected dimension (P)		
	dmg1	Data display mode in 1st indirectly detected dimension (P)		
	ft1d	Fourier transform along f_2 dimension (C)		
	ft2d	Fourier transform 2D data (C)		
	ph	Set phased mode in directly detected dimension (C)		
	ph1	Set phased mode in indirectly detected dimension (C)		
	pwr	Set power mode in directly detected dimension (C)		
	- pwr1	Set power mode in 1st indirectly detected dimension (C)		

wftld Weight and Fourier transform 2D data (C) wft2d Weight and Fourier transform 2D data (C)

poly0 Display mean of the data in regression.inp file (M)

Description	Calculates an	d displays the mean of data in the file regression.inp.
See also	User Program	nming
Related	averag	Calculate average and standard deviation of input (C)
	expl	Display exponential or polynomial curves (C)

pow Find the value of a number raised to a power

Syntax	pow(x,y)<:n>	
Description	Description Finds the value of the first argument (x) raised to the power of the second argument (y) . For example, pow $(2,3)$ is 2 ** 3 or 8. If x is negative and y is not and integer, pow will fail. pow is equivalent to	
	pow(x,y) = exp(y * ln(x))	
Arguments	value is a number.	
	n is the return value giving the value of x $**$ y. The default is to display the value in the status window.	
Examples	pow(2,3)	
	pow(2.1,0.6):val	
See also	See the Agilent VnmrJ User Programming Guide.	

powerfit Fits the diffusional attenuation calcuated by decay_gen to the exponential of a power series in the calibration of the non-uniformity of pulsed field gradients.

Syntax	powerfit()
	powerfit(ncoef)
Applicability	VnmrJ 3.1
Description	Used in the calibration of non-uniform field gradients to fit the diffusional decay calculated by decay_gen to the exponential of a power series.
Arguments	powerfit has one optional argument, the number of coefficients in the power series. The default is 8.

See also decay_gen gradfit nugcalib profile_int

pp Decoupler pulse length (P)

Description	Sets the o DEPT, HE	decoupler pulse length for use by pulse sequences such as T2DJ, and HETCOR.
See also	NMR Spe	ctroscopy User Guide
Related	AC1-AC9	Automatic calibration (M)
	Dept	Set up parameters for DEPT experiment
	dhp	Decoupler high-power control with class C amplifier (P)
	dpwr	Power level for first decoupler with linear amplifier (P)
	hetcor	Set up parameters for HETCOR pulse sequence (M)
	p1	First pulse width (P)
	pw	Pulse width (P)

ppa Plot a parameter list in plain English (M)

Syntax	ppa<(x<,y>) >
Description	Plots parame names and th	ters in plain English (instead of in a table with parameter neir values as plotted by the parameter pap).
Arguments	x controls the x offset, in mm, from the lower left of the plot to the starting position (upper left) of the parameter list. The default is a preset position on the page (upper left corner).	
	y controls th starting posit position on t	e y offset, in mm, from the lower left of the plot to the tion (upper left) of the parameter list. Default is a preset he page (upper left corner).
Examples	ppa ppa(10) ppa(wcmax-	80,wc2max*.9)
See also	NMR Spectro	scopy User Guide
Related	bpa	Plot boxed parameters (M)
	hpa	Plot parameters on special preprinted chart paper (C)
	pap	Plot out "all" parameters (C)
	pltext	Plot a text file (M)

ppcal Proton decoupler pulse calibration (M)

DescriptionProton decoupler pulse calibration for DEPT, HETCOR, INEPT, etc.See alsoNMR Spectroscopy User GuideRelatedAC1S-AC11SAutomatic calibration (M)d2pulSet up parameters for D2PUL pulse sequence (M)DeptSet up parameters for DEPT experimenthetcorSet up parameters for HETCOR pulse sequence (M)ineptSet up parameters for INEPT pulse sequence (M)

ppf Plot peak frequencies over spectrum (C)

Syntax	<pre>(1) ppf<(<'noll'><, 'pos'><, noise_mult><, 'top'>)> (2) ppf<(<'noll'><, 'pos'><, noise_mult><, 'leader'></pre>
Description	Plots peak frequencies, in units specified by the axis parameter, in the plotter device. Only those peaks greater than th high are selected. Two basic modes of label positioning are available: labels placed at the top, with long "leaders" extending down to the tops of the lines (syntax 1 using the 'top' keyword), or labels positioned just above each peak, with short leaders (syntax 2 using the 'leader' keyword). The default is short leaders.
Arguments	<code>'noll'</code> is a keyword to plot frequencies using the last previous line listing.
	'pos' is a keyword to plot positive peaks only ('noneg' is the same as 'pos').
	noise_mult is a numerical value that determines the number of noise peaks plotted for broad, noisy peaks. The default is 3. A smaller value results in more peaks, a larger value results in fewer peaks, and a value of 0.0 results in a line listing containing all peaks above the threshold th. Negative values of noise_mult default to 3. The noise_mult argument is inactive when the 'noll' keyword is specified.
	'top' is a keyword to plot labels at the top with long leaders. In this mode, the height of labels is varied by changing the parameter $wc2$.
	'leader' is a keyword to plot labels positioned just above each peak with short leaders.
	length specifies the leader length, in mm, if labels are positioned just above each peak. The default length is 20 mm.
Examples	<pre>ppf('pos') ppf('leader',30) ppf('top','noll') ppf('pos',0.0,'leader',30)</pre>

See also NMR Spectroscopy User Guide Related axis Axis label for displays and plots (P) dpf Display peak frequencies over spectrum (C) dpir Display integral amplitudes below spectrum (C) dpirn Display normalized integral amplitudes below spectrum (M) Plot integral amplitudes below spectrum (C) pir Plot normalized integral amplitudes below spectrum (M) pirn th Threshold (P)

pph Print pulse header (M)

Syntax	pph(file)		
Description	Prints out the shape file header (i.e., all lines starting with #)		
Arguments	file is the name of the shape file, including the extension.		
Examples	pph('shgrad.GRD')		
See also	NMR Spectroscopy User Guide		
Related	Pbox Pulse shaping software (U)		

ppmm Resolution on printers and plotters (P)

Description An internal software parameter, selected automatically based on the plotter configuration, that contains the resolution in dots/mm on raster graphics printers. On pen plotters, ppmm contains the resolution of points drawn. On PostScript printers, ppmm adjusts linewidths.

pprofile Plot pulse excitation profile (M)

Syntax	pprofile<(axisflag<,profile<,shapefile>>)>
Description	Plots the X, Y and Z excitation (inversion) profile for a pulse shape that has been generated with the Pbox software. If shape names is not provided, the last simulation data stored in the shapelib/pbox.sim file are plotted.
Arguments	The axisflag and profile arguments can be given in any order.
	axisflag is 'y' to display the full spectrum and a frequency scale, or 'n' to suppress the scale and spectrum. The default is 'n'.
	profile is a character string identifying the desired profile. 'xyz' selects X, Y, and Z (inversion) profiles; 'xy' selects only the excitation (transverse) profiles; 'x' selects only the X transverse excitation profile; and 'z' selects only the inversion profile. The default is 'xyz'.

	shapefile i extension.	s the name of a *.RF or *.DEC file, including the
Examples	<pre>pprofile pprofile(' pprofile('</pre>	y','x') xy','n','softpls.RF')
See also	NMR Spectro	scopy User Guide
Related	dprofile	Display pulse excitation profile (M)
	Pbox	Pulse shaping software (U)

pps Plot pulse sequence (C)

Syntax	<pre>pps<(file<,x,y,width,height>)></pre>	
Description	Plots pulse sequences. The plotted picture consists of three to five parts. At the top is the transmitter pulse sequence. Below that is the decoupler pulse sequence. Next is the second decoupler pulse sequence or gradients, depending on the program. At the bottom is the status.	
	The parameter of each pulse is plotted if its length is less than 30 letters. The value of each pulse is also plotted. If its value is less than zero, a question mark "?" is plotted. The time units are displayed as letters (s, m, or u). The height of pulses are plotted according to their power level.	
Arguments	file specifies the pulse sequence to be plotted. The default is \texttt{seqfil}	
	x, y specifies the start of the plotting position with respect to the lower-left corner of the plotter.	
	width, height are in proportion to wcmax and wc2max.	
Examples	pps pps('s2pul')	
~ .	pps (3, 50)	
See also	NMR Spectroscopy User Guide	
Related	dpsDisplay pulse sequence (C)seqfilPulse sequence name (P)wcmaxMaximum width of chart (P)wc2maxMaximum width of chart in second direction (P)	

prealfa Specify a delay for longer ring down (P)

Applicability	Systems with Varian, Inc. Cold Probes
Description	Specify a delay to be used in situations when there is a longer ring down of rf following the last rf pulse.
	This parameter is only active when qcomp='y'. prealfa should be created as a local parameter of type pulse or delay. This parameter

must be created as a local parameter of the type pulse for SpinCad Sequences.

If it is desired to use the software computed value for this delay, destroy the prealfa parameter.

Values User set prealfa value that may be slightly adjusted by the software to better optimize the DSP parameters.

preAmpConfigSet the band of the preamp, high or low, connected to each transmitter channel.

Syntax

Applicability VnmrJ 3.1

Description

Sets the band of the preamp, high or low, connected to each transmitter channel. This global parameter is a string whose entries are the characters "H" "L" and "X" separated by commas. The number of characters must equal the number of channels, numrfch. The characters from left to right refer to the transmitter channels "1", "2", "3"... etc, which for VNMRS correspond to preamps or RF cable outputs from the Front End and fom the transmitters in the RF card cage, right to left. Set: 'H' for a highband preamp, 'L' for a lowband preamp and 'X' for no preamp.

probeConnect and preAmpConfig are required for all experiments that use transmitters "3" and "4" as Obs or Dec. Create probeConnect and preAmpConfig as global parameters on the commandline with:

create('probeConnect','string','global')

create('preAmpConfig','string','global')

If probeConnect is present it will overide the transmitter settings in the 'current' parameter rfchannel.

If probeConnect and preAmpConfig are not created and rfchannel is not present the default transmitters are:

Obs (highband) "1" Dec (lowband) "2"

Obs (lowband) "2" Dec (highband "1"

Dec2 (highband or lowband) "3"

Dec3 (highband or lowband) "4"

If probeConnect and preAmpConfig are not created the default preamps are presumed to be:

- "1" highband
- "2" lowband
- "3" lowband
- "4" lowband

probeConnect and preAmpConfig must both exist or both be absent. If they exist both must have correct values. An empty string or

incorrect string in either parameter will cause errors in channel selection. probeConnect = 'H1 C13 F19 N15', preAmpConfig = 'HLHL', numrfch Examples = 4, tn = 'H1', dn = 'C13', dn2 = 'N15' causes: Obs on channel "1" Dec on channel "2" Dec2 on channel "4" probeConnect = 'H1 N15 F19 C13', preAmpConfig = 'HLHL', numrfch = 4, tn = 'H1', dn = 'C13', dn2 = 'N15' causes: Obs on channel "1" Dec on channel "4" Dec2 on channel "2" probeConnect = 'H1 N15 F19 C13', preAmpConfig = 'HLHL', numrfch = 4, tn = 'C13',dn = 'H1', dn2 = 'N15' causes: Obs on channel "2" Dec on channel "1" Dec2 on channel "4"

prep Run prepare acquisition macro (M)

Applicability	Imaging
Description	Run the prepare acquisition macro specified by the execprep
	parameter. Usually only called from panels.
Related	execprep Execute prepare macro (P)

Presat Set up parameters for presat ¹H experiment (M)

Description Set up parameters for presat ¹H experiment with solvent suppression.

prevp1 Display the previous 3D plane (M)

Description Displays 2D color map of the previous 3D plane in the set of planes defined by the parameters plane and path3d. For example, if dplane (40) has just been executed, prevpl results in the display of 3D plane 39 of that set. (If prevpl immediately follows the command dproj, an error results because there is no 3D plane whose number is -1.) prevpl is more efficient than dplane or dproj because the 3D parameter set (procpar3d) is not loaded into VnmrJ. It is assumed to have already been loaded by, for example, dplane or dproj.

NMR Spectroscopy User Guide		
dplane	Display a 3D plane (M)	
dproj	Display a 3D plane projection (M)	
dsplanes	Display a series of 3D planes (M)	
getplane	Extract planes from a 3D spectral data set (M)	
nextpl	Display the next 3D plane (M)	
path3d	Path to currently displayed 2D planes from a 3D data set	
	(P)	
plane	Currently displayed 3D plane type (P)	
plplanes	Plot a series of 3D planes (M)	
	NMR Spect dplane dproj dsplanes getplane nextpl path3d plane plplanes	

prescan Study queue prescan (P)

Description	This parameter keeps track of the type and status of the prescans in the study queue.		
Related	cqexp	Load experiment from protocol (M)	
	cqrset	Reset study queue parameters (M)	
	sqexp	Load experiment from protocol (M)	
	sqreset	Reset study queue parameters for imaging (M)	

prescan_CoilTableRead or update the CoilTable File (M)

Syntax	prescan_CoilTable(action,rfcoil)
Description	Manages the CoilTable file in ~/vnmrsys. Reads information about rfcoil into the global parameter coil_param; updates/adds information for rfcoil from coil_param; removes the rfcoil entry from CoilTable.
Arguments	actions for the specified rfcoil are:
	read add update remove
Examples	<pre>prescan_CoilTable('read','main')</pre>

prescan_tn Return tn string for a given atomic number (M)

Syntax	<pre>prescan_tn(number):str</pre>
Description	Returns tn string for a given atomic number; for H1, c13, F19, P31, Na23, Xe129 only.
Arguments	Number is the atomic number.

str is a string that can be assigned to tn.

Examples prescan_tn(23):tn

presig Preamp Signal Level Selection Parameter (parameter)

Syntax	
Applicability	VnmrJ 3.1
Description	This parameter is to be used with systems that support large signal handling at the preamp. It allows the user to select high signal handling "presig='h" or low signal handling "presig='l". Currently there are two types of preamps that support this capability.
	UnityPlus Spectrometers with Selectable Large-Signal Mode Preamps support this capability by allowing a current increase the preamp. This allows larger signals, and the overall signal level will be slightly higher.
	UnityPlus SIS Imaging Spectrometers support this capability using attenuation and a current increase. This allows larger signals and results in a lower overall signal level.
	The use of this parameter to control the hardware depends on the Magnet Leg Driver Board Configuration ID being set to 16 for SIS Imaging Systems or 1 for UnityPlus Spectrometers with Selectable Large-Signal Mode Preamp.
Arguments	'h' signifies high signal mode at the preamp.
	'l' signifies low signal mode at the preamp.
	'n' signifies "not used" and will default to low signal mode at the preamp if the hardware is present.
Related	gain

printer Printer device (P)

Description	Selects the printer in use on the system.
Values	A string with entries such as 'ThinkJet_96', 'LaserJet_300', 'jim', 'varian1', and 'Laser1'.
See also	NMR Spectroscopy User Guide
Related	showplotter Show list of currently defined plotters and printers (M)

printfile Path to the print-to-file image (P)

Description Defines the path where an image is saved if it is printed to a file.

printformat Format of saved-to-file image (P)

Description The format of the image to be printed to a file. Values 'jpeg', 'gif', 'tiff', 'bmp'

printlayout Layout of printed image (P)

Description	The layout of the printed image.
Values	'portrait' or 'layout'

printoff Stop sending text to printer and start print operation (C)

Syntax	printoff<('clear' file)>		
Description	Stops redirection of output to printer caused by the printon command and starts the print operation. The command printoff must be entered to obtain output on the printer . Actual printing is controlled by the vnmrprint script in the bin subdirectory of the system directory. printoff can also clear the data in the current print file or save data to a specified file name (i.e., print or plot to a file).		
Arguments	'clear' is a keyword to clear the print file made so far.		
	file specifies the name of a file to save the printout. If the file already exists, it is overwritten.		
Examples	printoff printoff('clear') printoff('vnmrsys/papers/peaks.list')		
See also	NMR Spectroscopy User Guide		
Related	printonDirect text output to printer (C)vnmrprintPrint text files (U)		

printon Direct text output to printer (C)

Description	Sends inform window. Aft window, suc	mation to the printer that is normally displayed in the text er using printon, output from commands that use the text ch as dg and cat, is sent to the printer and does not	
	appear on the screen. The value of the parameter printer is used to select which printer is used.		
See also	NMR Spectroscopy User Guide		
Related	cat	Output one or more files to output text window (C)	
	dg	Display group of acquisition/processing parameters (C)	

printerPrinter device (P)printoffStop sending text to printer and start print operation
(C)

printregion Screen region to be printed (P)

Description The region of the screen to be printed or saved to a file. Values 'vnmrj' -- entire VnmrJ interface. 'graphics' -- the graphics area of the VnmrJ interface. 'frames' -- selected frames from the graphics area.

printsize Size of printed image (P)

Description	The size of the printed image.	
Values	'quarterpage', 'halfpage',	'page'

printsend Defines where image will print (P)

Description Defines whether the selected image will sent to a file or a printer. Values 'file' or 'printer'

probe Probe type (P)

Description Contains a string with the name of the probe currently in the magnet. This parameter is set automatically when the addprobe macro is entered. The getparam and setparams macros use probe to retrieve and write parameters into the current probe file.

See also NMR Spectroscopy User Guide

RelatedaddnucleusAdd new nucleus to existing probe file (M)addprobeCreate new probe directory and probe file (M)getparamReceive parameter from probe file (M)setparamsWrite parameter to current probe file (M)
probeConnectSpecify which nucleus can be acquired on each RF channel (P)

Applicability	VNMRS and 400 MR		
Syntax	<pre>probeConnect = 'nuc1 nuc2 nuc3'</pre>		
Description	Global string parameter that does not exist by default. If present, PSG uses it to determine which RF channel to connect to a given nucleus. The string consists of a series of space-separated nuclei. A nucleus 'X' may be used only once in the string to match any nucleus. The parameter must match the hardware connections. If the parameter does not match the hardware connections or does not exist, default settings are used. Default settings are to use the first channel for tn for high band observe, and the second channel for tn for low band observe.		
Values	Any nucleus name used for tn, or 'X'.		
Examples	create('probeConnect','string','global')		
	probeConnect = 'H1 C13' maps H1 to channel 1, C13 to channel 2		
	probeConnect = 'H1 P31 X' maps H1 to channel 1, P31 to channel 2, any nucleus to channel 3.		
See also	VnmrJ User Programming		
Related	tnNucleus for observe transmitter (P)dmNucleus for first decoupler (P)dm2Nucleus for second decoupler (P)dm3Nucleus for third decoupler (P)		

Probe_edit Edit probe for specific nucleus (U)

Syntax	(UNIX) Probe_edit probe nucleus	
Description	Opens a dialog box showing all the parameters related to a specific nucleus from the probe table.	
Arguments	probe is the name of the probe.	
	nucleus is the specified nucleus from the probe table.	
Examples	Probe_edit 5mmSW H1	
Related	probe_edit Edit probe for specific nucleus (M)	

probe_edit Edit probe for specific nucleus (M)

Syntax	<pre>probe_edit(probe,nucleus)</pre>
Description	Opens a dialog box showing all the parameters related to a specific nucleus from the probe table.
Arguments	probe is the name of the probe.

nucleus is the specified nucleus from the probe table. Examples probe_edit('5mmSW','H1') probe_edit(probe,tn) Related Probe_edit Edit probe for a specific nucleus (U)

probe_protectionProbe protection control (P)

DescriptionControls the power check for probe protection.See alsoNMR Spectroscopy User Guide

proc Type of processing on np FID (P)

Description Specifies the type of data processing to be performed upon the np (t_2) FID. Similarly, parameters proc1 and proc2 specify the type of data processing on the ni (t_1) and ni2 interferograms, respectively.

All Varian data must be processed along np with a complex Fourier transform (FT). Sequentially sampled Bruker data (the usual case) must be processed along this dimension with a real FT, while simultaneously sampled Bruker data must be processed with a complex FT.

Pure absorptive 2D data collected by the States-Haberkorn (hypercomplex) method must be processed along ni or ni2 with a complex FT.

Pure absorptive 2D data collected by the TPPI method on a Varian spectrometer can be processed in one of two ways, depending upon how the data was collected:

phase=3	Complex FT, i.e., proc1='ft' (standard way)
phase=1,4	Real FT, i.e., proc1='rft' (new way)
phase2=3	Complex FT, i.e., proc2='ft'
phase2=1,4	Real FT, i.e., proc2='rft'

Pure absorptive 2D data collected by TPPI method on a Bruker spectrometer must be processed along ni with a real FT (i.e., proc1='rft').

Values 'ft' specifies complex FT data processing.

'rft' specifies real FT data processing.

'lp' specifies linear prediction processing on complex data. If 'lp' is selected, additional parameters must be set to fully define how the time-domain data is to be processed; see the description of the addpar command.

see also	NMR Spectroscopy User Guide	
Related	addpar	Add selected parameters to the current experiment (M)
	ni	Number of increments in 1st indirectly detected dimension
		(P)
	np	Number of data points (P)
	parlp	Create parameters for linear prediction (C)
	phase	Phase selection (P)
	phase2	Phase selection for 3D acquisition (P)
	proc1	Type of processing on ni interferogram (P)
	proc2	Type of processing on ni2 interferogram (P)

See also NMR Spectroscopy User Guide

proc1 Type of processing on ni interferogram (P)

Description	Specifies interfero informat	the type of data processing to be performed upon the ni (t_1) gram (2D). Refer to the description of proc for further ion.
Values	 'ft' specifies complex Fourier transform (FT) data processing. 'rft' specifies real FT data processing. 'lp' specifies linear prediction processing on complex data. If 'lp' is selected, additional parameters must be set to fully define how the time-domain data is to be processed; see the description of the addpar command. 'ht' specifies Hadamard transform processing. If 'ht' is selected, additional parameters must be set with the addpar command. In addition, the data set must be acquired using a Hadamard pulse sequence. 	
See also	NMR Spe	ectroscopy User Guide
Related	addpar ni	Add selected parameters to the current experiment (M) Number of increments in 1st indirectly detected dimension (P)
	proc	Type of processing on np FID (P)

proc1d Processing macro for simple (non-arrayed) 1D spectra (M)

Description A generic macro for processing non-arrayed 1D spectra using a set of standard macros. procld is called by the procplot macro, but can also be used directly. procld first tries to find a macro of the form {tn}p with the name of the observe nucleus in lower case (e.g., h1p, cl3p). If such a macro exists, it is called. If such a nucleus-specific macro is not found in the command path, minimal 1D processing is performed (the intent is to provide a well-processed spectrum in most cases): Fourier transformation (using pre-set weighting functions), automatic phasing (aphx macro), automatic integration (integrate macro), vertical scale adjustment (vsadj macro), avoiding excessive

	noise (noisl proc1d does DEPT-type sp	m macro), and threshold adjustment (thadj macro). a not work with arrayed 1D spectra: use deptproc (for pectra) or procarray (for all other arrayed 1D data).
See also	NMR Spectro	oscopy User Guide
Related	aphx	Perform optimized automatic phasing (M)
	c13p	Process 1D carbon spectra (M)
	deptproc	Process arrayed dept type spectra (M)
	h1p	Process 1D proton spectra (M)
	integrate	Automatically integrate 1D spectrum (M)
	noislm	Avoids excessive noise (M)
	procarray	Process arrayed 1D spectra (M)
	procplot	Automatically process FIDs (M)
	thadj	Adjust threshold (M)
	vsadj	Adjust vertical scale (M)

proc2 Type of processing on ni2 interferogram (P)

- Description Specifies the type of data processing to be performed upon the ni2 interferogram (3D). Refer to the description of proc for further information.
 - Values 'ft' specifies complex Fourier transform (FT) data processing.
 - 'rft' specifies real FT data processing.

'lp' specifies linear prediction processing on complex data. If 'lp' is selected, additional parameters must be set to fully define how the time-domain data is to be processed; see the description of the addpar command.

- See also NMR Spectroscopy User Guide
 - Related addpar Add selected parameters to the current experiment (M) ni2 Number of increments in 2nd indirectly detected dimension (P) proc Type of processing on np FID (P)

proc2d Process 2D spectra (M)

Description A general 2D processing macro that tries to do the appropriate processing for as many types of 2D experiments as possible. It uses wft2da for phase-sensitive spectra, wft2d for absolute-value 2D spectra, wft2d('ptype') for HOM2DJ and COSYPS (absolute value). Symmetric homonuclear correlation spectra (fn=fn1, sw=sw1) in absolute-value mode is symmetrized using foldt. The resulting spectrum is then normalized (adjustment of vs and th) using nm2d and displayed (if not in background mode). proc2d is called as part of the procplot macro, but can also be used directly by the user.

dee also	Man Specifoscopy User Guide	
Related	fn	Fourier number in the directly detected dimension (P)
	fn1	Fourier number in 1st indirectly detected dimension
		(P)
	foldt	Fold COSY-like spectrum along diagonal axis (C)
	nm2d	Normalize intensity of 2D spectrum (M)
	procplot	Automatically process FIDs (M)
	SW	Spectral width in the directly detected dimension (P)
	sw1	Spectral width in the 1st indirectly detected dimension
		(P)
	th	Threshold (P)
	VS	Vertical scale (P)
	wft2d	Weight and Fourier transform 2D data (C)
	wft2da	Weight and Fourier transform for pure absorption 2D
		data (M)

See also NMR Spectroscopy User Guide

procarray Process arrayed 1D spectra (M)

Description A generic macro for processing arrayed 1D data. It is called within the procplot macro, but can also be called directly. It transforms all traces, phase the trace with the largest signal, scale the traces appropriately, and set up the display parameters such that the data can be plotted directly. The plotting is done in a separate macro plarray that is also called in the procplot macro.

For the display setup, procarray distinguishes between arrays with 6 or less elements, which are stacked vertically (no horizontal offset), and spectra with greater than 6 elements, which are stacked horizontally by default, unless there are too many lines, in which case a diagonally stacked display is chosen.

Horizontal stacking is mostly adequate for pulse and power calibrations, where there are usually only a few lines. Diagonally stacked displays and plots are frequently chosen for T_1 and T_2 experiments on entire spectra, often with many lines. The automatic stacking mode can be overridden by creating and setting a string parameter stackmode in the startup macro, or before calling procplot or procarray. Possible values for stackmode are 'horizontal', 'vertical', and 'diagonal'. DEPT-type spectra can, in principle, be also processed with procarray but, of course, no DEPT editing occurs.

See also NMR Spectroscopy User Guide

Related	deptproc	Process arrayed dept type spectra (M)
	plarray	Plot arrayed 1D spectra (M)
	proc1d	Processing macro for simple (non-arrayed) 1D spectra
		(M)
	procplot	Automatically process FIDs (M)

stack Set stacking control parameter (M) Stack control for processing arrayed 1D spectra (P) stackmode

Generic automatic processing (M) process

Description Processes a wide range of data types. If the apptype parameter is set, it runs the execprocess macro if it exists. If the apptype parameter is not set it selects a macro depending on the type of data. For simple 1D spectra, process looks for a macro of form $\{tn\}p$ with the observe nucleus in lower case (e.g., h1p, c13p, f19p). If no such macro is found, process calls proc1d, a generic processing macro for 1D spectra. For DEPT type data, deptproc is called. For other arrays of 1D spectra, procarray is called. For 2D spectra, proc2d is called. process by itself is called within the procplot macro.

See also NMR Spectroscopy User Guide

Related	apptype	Application type (P)
	c13p	Processing of 1D carbon spectra (M)
	deptproc	Process array of DEPT spectra (M)
	execpars	Set up the exec parameters (M)
	execprocess	Execute processing macro (P)
	f19p	Processing of 1D fluorine spectra (M)
	hlp	Processing of 1D proton spectra (M)
	proc1d	Automatically process non-arrayed 1D fids (M)
	proc2d	Process 2D spectra (M)
	procarray	Process arrayed 1D spectra (M)
	procplot	Automatically process FIDs (M)
	tn	Nucleus for observe transmitter (P)

Automatically process FIDs (M) procplot

Syntax procplot<(pltmod_value)>

Description Universal FID processing macro called usually with wexp='procplot' by automatic acquisition macros such as h1, c13, hcapt, and hcosy. The purpose of procplot is not the data processing itself, but rather the selection of the appropriate processing macro for a given data set.

> First, procplot calls a macro process that calculates spectra; that macro by itself then selects an appropriate processing macro, like procld for non-arrayed 1D spectra. Depending whether the parameter pltmod is set to 'none' or not, procplot then calls plot, a universal plotting macro. The setting of the parameter pltmod can be temporarily overridden by specifying an alternative value as argument to procplot.

> One of the concepts behind procplot is that the user should never have to modify any processing macro for customizing the processing or

	the output of happen by se procplot.	f automatic experiments or processing; this outcome can electing a parameter in the calling macro or before calling
Arguments	pltmod_val is only used suppress plo pltmod_val pltmod has if only the ca call the proc	ue is an alternate value for the parameter pltmod that for the current call. The values 'none' and 'off' tting. The range of possible (active) values for ue depends on the plotting macros. Often, the parameter no effect other than turning on or off plotting. Note that alculation of a spectrum is desired, it is usually easier to cess macro.
Examples	procplot procplot('	none')
See also	NMR Spectro	oscopy User Guide
Related	deptproc	Process arrayed dept type spectra (M)
	plot	Determine plot mode (\mathbf{P})
	procld	Processing macro for simple (non-arrayed) 1D spectra (M)
	proc2d	Process 2D spectra (M)
	procarray	Process arrayed 1D spectra (M)

profile Set up pulse sequence for gradient calibration (M)

process Automatically calculate spectra (M)

Applicability	Systems with the pulsed field gradients (PFG) module.
Description	Performs an rf and gradient echo sequence that gives a high quality profile of the sample. This sequence is used with the macro setgeal to provide gradient strength calibration.
See also	Performa I Pulsed Field Gradient Module Installation; Pulsed Field Gradient Modules Installation; User Programming
Related	gcalGradient calibration constant (P)setgcalCalibrate gradient strength from measured data (M)

profile_int Normalise the experimental signal profile during calibration of non-uniform pulsed gradients.

Syntax	<pre>profile_int(lowfrq,highfrq)</pre>
Applicability	VnmrJ 3.1
Description	Integrates the signal in the file Signal_profile, normalises it and writes it to the file Normalised_profile.
Arguments	profile_int takes two arguments: lowfrq is the lower frequency limit of the profile, highfrq is the high frequency limit of the profile.

```
See also decay_gen
gradfit
nugcalib
powerfit
```

proj Project 2D data (C)

Syntax proj(exp_number<, 'sum'><, start<, width>>)

Description Projects 2D data onto the axis parallel to the screen x-axis, which can be f_1 or f_2 , depending upon the parameter trace. Two projections are available:

- Summing projection. The data at each frequency are summed and the result becomes the projection.
- *Skyline projection.* The data are searched and the maximum intensity at any given frequency becomes the intensity in the projection (similar to looking at the skyline of a city where only the largest building along any given line of sight is visible).

Phase-sensitive data can be projected, but the resulting projection can only be displayed in an absolute-value mode

Arguments exp_number is the number of the experiment, from 1 through 9, in which the resulting spectrum is stored.

'sum' is a keyword to use the summing projection. The default is skyline.

start defines the starting trace, in Hz. The default is to project all data.

width defines the width of the traces, in Hz, to be projected. The default is to project all data. If width is supplied as zero, a single trace corresponding to the start frequency will be stored.

- Examples proj(3) proj(5,'sum')
 - proj(4,3*sfrq,6*sfrq)

See also NMR Spectroscopy User Guide

Related trace Select mode for 2D data display (P)

proshimhelp Proshim help (C)

ApplicabilityVnmrJ 3.2DescriptionUse to bring up help for the Proshim window.

Proton Set up parameters for ¹H experiment (M)

Description Set up parameters for ¹H experiment.

protune Macro to start ProTune (M)

Applicability	Liquids, Walkup, Automation		
Syntax	protune(fr	eq1 <, match1 <, freq2 <, match2>>>)	
	protune('argument',<\$nucleus,<\$target>>)		
	protune('e	xec', command1 <, command2,>)	
Description	Tunes to frequency freq1 MHz if the first argument is the frequency in MHz.		
	Executes a sequence of arbitrary tuning commands if the first argument is the keyword exec. Any command that can be typed into the command line box in the ProTune GUI display is allowed.		
Arguments	First case:		
	freq1 MHz	– first tuning frequency in MHz	
	match1 - %	of optimum for the first frequency, 5% is the default	
	freq2 MHz	– optional second tuning frequency in MHz	
match2 - %		of optimum for the second frequency, 5% is the default.	
	Second case:		
	'argument'	may have the following values:	
no or	argument 'popup'	opens Tune Probe dialog for probe tuning. Select the nucleus to tune and how coarse to tune using the	
'calibrate' 'nucleus'		open ProTune calibration interface. tune using specified nucleus – \$nucleus must be specified.	
	\$nucleus —	Nucleus to tune to, 'H1', 'C13'	
\$target — 5 if no valı		Fune target level, 0.1(finest) to 100 (coarsest), defaults to e is specified.	
	Third case:	'hird case:	
	exec - keyv	vord that precedes a command or string of commands.	
Examples	protune('e	<pre>xec', 'setTuneFrequency 0 599.96e6')</pre>	
	Tunes the pr	robe to 599.96 MHz.	
See also	User Guide Liquids and VnmrJ Walkup		
Related	atune	ProTune present (P)	
	protunegui	Macro to start ProTune in graphical user interface (M)	

plockport	Port number to use to lock out multiple ProTune processes (P)
probeConnect	Specify which nucleus can be tuned on each RF channel (P)
settune	set up tune parameters for automation
showprotunegui	show the graphical interface while tuning (P)
tchan	RF channel number used for tuning (P)
tugain	Receiver gain used in tuning (P)
tunehf	Tune both H1 and F19 on an HFX probe (M)
tunesw	Width of the tuning sweep in Hz (P)
tunematch	Default match target, in percent of optimum (P)
tupwr	Transmitter power used in tuning (P)
tuneResult	Message indicating how well the tuning succeeded
	(P)
tunemethod	Method to use for tuning (P)
wtune	Specify when to tune (P)
wtunedone	What to do after tuning is done (P)
xmtune	Check tune parameter during automation (M)

protune Shell script for start ProTune operation (U)

Applicability	Automation	
Description	Starts and stops ProTune. Usually called from Protune macros.	
See also	NMR Spectroscopy User Guide and VnmrJ Walkup	
Related	protune (M Macro to start ProTune (M)	

protunegui Macro to start ProTune in graphical user interface (M)

Applicability	Liquids, VnmrJ Walkup, Automation		
Syntax	<pre>protune('argument',<\$nucleus,<\$target>>)</pre>		
Description	Starts ProTune in graphical mode.		
Arguments	see protune (M)		
See also	NMR Spectroscopy User Guide and VnmrJ Walkup		
Related	protune Macro to start ProTune (M)		

prune Prune extra parameters from current tree (C)

Syntax prune(file)

Description Destroys parameters in the current parameter tree that are not also defined in the supplied parameter file. prune is used to remove

	leftover parameters from previous experimental setups. Recalling a new parameter set into an experiment has a similar effect and, in general, prune is not required.		
Arguments	file is the path of a parameter file.		
Examples	<pre>prune(systemdir+'/parlib/cosyps.par/procpar') prune('/vnmr/par400/stdpar/H1.par/procpar') prune(userdir+'/exp3/curpar')</pre>		
See also	User Programming		
Related	create	Create new parameter in a parameter tree (C)	
	destroy	Destroy a parameter (C)	
	display	Display parameters and their attributes (C)	
	fread	Read parameters from file and load them into a tree (C)	
	fsave	Save parameters from a tree to a file (C)	

pscale Plot scale below spectrum or FID (C)

Syntax	<pre>pscale<(<rev><,axis><,label><,vp0><,sp0><,color><,pen>)></rev></pre>
Description	Plots a scale under a spectrum or FID.
Arguments	rev – reverses the direction of the scale. That is, the smaller numbers will be at the left side of the scale. If used, 'rev' must be the first argument.
	axis - If the letter p, h, k, etc. is supplied, it will be used instead of the current value of the parameter axis. For an FID scale, if the letter s, m, or u is supplied, it will be used instead of the current value of the parameter axisf.
	label - If a string of 2 or more characters is supplied, it will be used as the axis label.
	vp0 – This is supplied as the first real number. It defines the vertical position where the scale is drawn. The default is 5 mm below the current value of the parameter vp .
	sp0 – This is supplied as the second real number. It is a modified start of plot. If, for example, the display is from 347 to 447 hz, but the scale is desired to read 0 to 100 hz., $sp0$ would be input as 0.
	wp0 – This is supplied as the third real number. It is a modified width of plot. If, for example, the display is from 347 to 447 hz, but the scale is desired to read 0 to 550 Units. sp0 would be input as 0, $wp0$ would be 550, and the label would be 'Units'.
	An optional color or pen number can be supplied to dscale or pscale. The available colors and pens are: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', 'white' 'pen1', 'pen2', 'pen3',, 'pen8'
Examples	pscale pscale(20) pscale('h',0,'pen2')

	pscale('fid','m') pscale('h',vp-10,0)	
See also	NMR Spectro	scopy User Guide
Related	axis	Axis label for displays and plots (P)
	axisf	Axis label for FID displays and plots (P)
	dscale	Display scale below spectrum or FID (C)
	vp	Vertical position of spectrum (P)

pseudo Set default parameters for pseudo-echo weighting (M)

Syntax	pseudo<(C1,C2,C3,C4)>		
Description	Generates an initial guess at good weighting parameters for absolute-value 2D experiments. To generate modified guesses, four coefficients are allowed to set the values of the weighting functions.		
Arguments	C1 sets $lb=-0.318/(C1*at)$. The default value of C1 is 0.0625.		
	C2 sets $gf=C2*at$. The default value of C2 is 0.25.		
	C3 sets lb1=-0.318/(C3*(ni/sw1)) but is used with 2D experiments only. The default value of C3 is 0.0625.		
	C4 sets gf1=C4*(ni/sw1) but is used with 2D experiments only. The default value of C4 is 0.25.		
Examples	pseudo pseudo(.1,.4,.2,.5)		
See also	NMR Spectroscopy User Guide		
Related	sinebell Select default parameters for sinebell weighting (M)		

psg Display pulse sequence generation errors (M)

Description Helps identify the problem if, after entering go or su, etc., the message is returned that pulse sequence generation (PSG) aborted abnormally. Any parameters that are not found are listed. This information is stored in the user's directory (vnmrsys) in a text file named psg.error. If the message "Maximum communication retries exceeded, Experiment unable to be sent" is displayed, a program communications problem is indicated. Consult the system operator for assistance. See also User Programming

psggen Compile a user PSG object library (M,U)

Description A user PSG (pulse sequence generation) kit is supplied that allows editing low-level pulse sequence code. psggen compiles these edits so

Ρ

that subsequent pulse sequence generation with the seqgen command uses the customized pulse sequence source.

See also User Programming

psgset Set up parameters for various pulse sequences (M)

Syntax	<pre>psgset(file,par1,par2,,parN)</pre>		
Description	Sets up parameters for various pulse sequences using information in a parlib file. Rather than returning the entire parameter file, psgset returns the parameters listed. psgset, in general, is never entered from the keyboard but is used as part of experiment setup macros.		
Arguments	file is the file from the user or system parlib that provides information on setting up the parameters listed. The parameters seqfil and pslabel are set to the supplied file name.		
	par1,par2,,pN are 1 to 11 parameters to be returned from parlib.		
Examples	psgset('cosy','dg','ap','ss','d1','axis','phase')		
See also	User Programming		
Related	pslabelPulse sequence label (P)seqfilPulse sequence name (P)		

psgupdateon Enable update of acquisition parameters (C)

Description	Permits the interacti	ve updating of acquisition parameters.
See also	SpinCAD	
Related	psgupdateoff	Prevent update of acquisition parameters (C)
	updtparam	Update specified acquisition parameters (C)

psgupdateoffPrevent update of acquisition parameters (C)

Description	Prevents the interact	tive updating of acquisition parameters.
See also	SpinCAD	
Related	psgupdateon	Enable update of acquisition parameters (C)
	updtparam	Update specified acquisition parameters (C)

Syntax	<pre>pshape<(pattern.ext)></pre>		
Description	Plots the real (X) and imaginary (Y) components of a shaped pulse. Any type of waveform (.RF, .DEC or ,GRD) can be plotted.		
Arguments	pattern is the name of a shape or pattern file specified by an absolute file name, relative file name, or a simple pattern file name. ext is a file name extension that specifies the file type. In the case of a simple file name, dshape searches for the file in the local directory, then in the user's shapelib, and finally in the directory /vnmr/shapelib. If pattern.ext is not given, pshape displays the last created waveform stored in the pbox.fid file.		
Examples	pshape pshape('my_shape.DEC')		
See also	NMR Spectroscopy User Guide		
Related	dshapeDisplay the lasPboxPulse shaping state	t created pulse shape (M) software (U)	

pshapef Plot the last created pulse shape (M)

Description	Plots real (X) and imaginary (Y) components of the last created shaped pulse.	
See also	NMR Spectroscopy User Guide	
Related	dshape Pbox	Display the last created pulse shape (M) Pulse shaping software (U)

pshr PostScript High Resolution plotting control (P)

Applicability	ALL	
Syntax	pshr= <value></value>	
Description	Global parameter that controls whether a 1D spectrum is plotted in hi-resolution mode or not. A hi-resolution plot is one in which every data point is represented in the plot. The standard resolution plot determines maximum and minimum values over small regions and plots those. The parameter pshr can have the values 1 for hi-res and 0 for standard plot.	
Values	0 for standard resolution	
	1 for high resolution.	
Related	pl pslw	Plot spectra (C) PostScript Line Width control (P)

pslabel Pulse sequence label (P)

Description Contains the text to be displayed in the Seq: field on the top line of the screen. This string may be different from the pulse sequence name selected with seqfil. However, the string in seqfil is the name of the pulse sequence searched for when an experiment is started. Generally seqfil=pslabel, and when seqfil is set, the system sets pslabel to the same string. See also NMR Spectroscopy User Guide

Related seqfil Pulse sequence name (P)

<pslabel>_setup Experiment-Specific Setup Macro (M)

Syntax	
Applicability	VnmrJ 3.1
Description	Macro is executed to set up sequence-specific parameters.
Examples	User Guide: Automation-User Space Customization
Related	execpslabel('setup') Pulse sequence name (P)

ps1w PostScript Line Width control (P)

Applicability	ALL	
Syntax	pslw= <value></value>	
Description	Global parameter that adjusts the line width of PostScript plots.	
Values	0 (narrowest) to 100 (widest) line width.	
Related	pl Plot spectra (C)	
	pshr PostScript High Resolution plotting control	
	(P)	

psMain Prescan controlling macro

Syntax		
Applicability	VnmrJ 3.1	
Description	Prior to acquiring data, a number of operations may be performed to condition the data acquisition. These may include probe tuning, acquiring a lock, shimming, adjusting receiver gain, and performing an equilibration delay. These operations are collectively referred to as prescan operations. The order of executing the various prescans, and the name of the macro to call for a specific prescan is defined in the	

"templates/vnmrj/choicefiles" application directory by the prescanInfo file.

The psMain macro is the controlling macro that executes each prescan. The individual prescans are controlled by macros, conventionally named psX, where X is Gain, Lock, etc.

Arguments Calling the psMain macro with no arguments will execute all defined prescans, in the order given in the prescanInfo file. Calling psMain with the name of a specific prescan, or a list of specific prescans, will execute those. For example, psMain('psGain') will execute the autogain prescan. psMain('psTune psLock') will tune the probe and then autolock.

> The prescan process can also be executed in steps. psMain('setup') initializes the prescans, but does not start the process. At this point, the setup may be customized. For example, a specific prescan could be removed from the list with the command psCmd('remove','psTune'). The command psMain('start'): \$ret starts the execution. Depending an what specific prescans are requested, a data acquisition may or may not be started. Depending on whether the prescans start an acquisition or not, the \$ret value will be set to 'psAcquiring' or 'psDone', respectively. In the case of 'psAcquiring', you can schedule the post-prescan acquisition with psMain('acquireAfterPs'). See the cpgo macro for an example.

Plot Arrayed Numbers (C) pssl

Syntax pssl(<options>)

Description Plots a label for each element in a set of stacked spectra. The label is an integer value from 1 up to the number of spectra in the display. Arguments options can be any of the following:

- 'all' is a keyword to display all of the spectra.
- 'int' is a keyword to display only the integral, independently of the value of the parameter intmod
- 'top' or 'side' are keywords that cause the spectrum to be displayed either above or at the left edge, respectively, of a contour plot. This assumes that the parameters sc, wc, sc2, and wc2 are those used to position the contour plot.
- 'dodc' is a keyword for all spectra to be drift corrected independently.
- 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'black', and 'white' are keywords that select a color.
- 'pen1', 'pen2', 'pen2' ... are keywords that pens.
- 'nopars' prevents the display commands from drawing the parameters at the bottom of the graphics screen.
- 'custom' uses the parameters shownumx (x position) and shownumy (y position), counting from bottom left of every spectrum.

	• 'reverse' - rotate the text by 90° - useful if the arrayed parameter values are long with respect to the width of the individual sub-spectra.		
• 'value' —The values of up to two simultaneous arrays are dis Diagonal arrays are allowed. The second parameter is shown different color). The name of the arrayed parameter(s) is also If used on a one-dimensional array representation of a 2D sp ni and phase (in case of phase sensitive 2Ds) parameters are			
	• 'list=xxx' produces a display of the values contained in the arrayed parameter xxx.		
	• 'format=yyy' uses the format yyy to control the plot of each label. See the write command for information about formats.		
Examples	pssl pssl('top','left') pssl('value','format=%3.1f')		
See also	NMR Spectroscopy User Guide		
Related	dsslLabel a display of stacked spectra (M)writeWrite formatted text to a device (C)		

ptext Print out a text file (M)

Syntax	<pre>ptext(file)</pre>		
Description	Prints out a text file.		
Arguments	file is the name of the text file.		
Examples	<pre>ptext('/vnmr/maclib/ptext') ptext(curexp+'/dept.out')</pre>		
See also	NMR Spectroscopy User Guide		
Related	curexp Current experiment directory (P)		
	dtext	Display a text file in the graphics window (C)	
	lookup	up Look up words and lines from a text file (C)	
	pltext Plot a text file (C)		
text Display text or set new text for current expe		Display text or set new text for current experiment	
		(C)	
	textvi	Edit text file of current experiment (M)	
	vi	Edit text file with vi text editor (C)	

ptspec3d Region-selective 3D processing (P)

Description Sets whether region-selective 3D processing occurs. If ptspec3d does not exist, it is created by the macro par3d. ptspec3d is functional at this time only for the f_3 dimension. If ptspec3d='ynn', only the currently displayed region of f_3 is retained as non-zero values after the f_3 transform in the 3D FT. A larger f_3 region may be kept to ensure

that the number of hypercomplex f_3 points is a power of 2; but that portion of the f_3 spectrum that is retained outside of the currently displayed region contains only zeroes. This 3D utility can reduce the fully transformed 3D data size by factors of 2 to 4, especially in some

of the triple resonance experiments.

- Values A three-character string such as 'nnn', 'nny', 'nyn', etc. The default is 'nnn'. The first character refers to the f_3 dimension (sw, np, fn); the second character, to the f_1 dimension (sw1, ni, fn1); and the third character, to the f_2 dimension (sw2, ni2, fn2). Each character may take one of two values: 'n' for no region-selective processing in the relevant dimension, or 'y' for region-selective processing in the relevant dimension.
- See also NMR Spectroscopy User Guide

Related	fiddc3d	3D time-domain dc correction (P)
	fn	Fourier number in directly detected dimension (P)
	fn1	Fourier number in 1st indirectly detected dimension (P)
	fn2	Fourier number in 2nd indirectly detected dimension (P)
	ft3d	Perform a 3D Fourier transform (M)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	ni2	Number of increments in 2nd indirectly detected
		dimension (P)
	np	Number of data points (P)
	ntype3d	N-type peak selection in f_1 or f_2 (P)
	par3d	Create 3D acquisition, processing, display parameters
		(C)
	specdc3d	3D spectral drift correction (P)
	SW	Spectral width in directly detected dimension (P)
	swl	Spectral width in 1st indirectly detected dimension (P)
	sw2	Spectral width in 2nd indirectly detected dimension (P)

ptsval PTS frequency synthesizer value (P)

- Description Configuration parameter for the frequency of the PTS synthesizer on each channel. Every broadband system is equipped with a PTS frequency synthesizer as part of broadband frequency generation. The frequency of the unit is marked on its front panel. The value is set for each channel using the Synthesizer label in the Spectrometer Configuration window.
 - Values 0 (Not Present choice in Spectrometer Configuration window); 160, 200, 250, 320, 500, 620, 1000 (PTS 160, PTS 200, PTS 250, PTS 320, PTS 500, PTS 620, PTS 1000 choices in Spectrometer Configuration window, respectively).

See also VnmrJ Installation and Administration. Related config Display current configuration and possibly change it (M) latch Frequency synthesizer latching (P) overrange Frequency synthesizer overrange (P)

pulseinfo Shaped pulse information for calibration (M)

Syntax	<pre>pulseinfo<(shape,pulse_width<,reference_power>)> :width,power</pre>		
Description	Returns or prints a table with the bandwidth and predicted pulse power settings for a given pulse shape. No parameter settings are changed. The necessary data is contained in the file shapeinfo in the system shapelib subdirectory.		
Arguments	shape is the name of the pulse shape. The default is the system interactively prompts the operator for the name of the shape and the duration of the pulse and then prints a table containing the bandwidth of that pulse and the predicted pulse power settings.		
	pulse_width is the duration of the pulse, in μ s. reference_power is a value, in dB, for power calculations. The default is 55. This value replaces the assumption used for power calculation that pw90 is set for a tpwr of 55.		
	width returns the bandwidth of that pulse, in Hz.		
	power returns the predicted 90° pulse power settings.		
Examples	pulseinfo('gauss',1000):bw,pwr		
See also	User Programming		
Related	bandinfo pw90 tpwr	Shaped pulse information for calibration (M) 90° pulse width (P) Observe transmitter power level with linear amplifiers (P)	

pulsetool RF pulse shape analysis (U)

Syntax	pulsetool <-shape filepath>
Description	Enables examination of shaped rf pulses. It is started from a UNIX window.
Arguments	The optional -shape filepath specifies the name of an rf pulse template file that is displayed when pulsetool is started.
Examples	pulsetool pulsetool -shape /vnmr/shapelib/sinc.RF
See also	NMR Spectroscopy User Guide

Description	Removes	one	or	more	macros	from	memory,	freeing	extra	memory
	space.									

Arguments file is the name of a macro file to be removed from memory. The default is to remove all macros that have been loaded into memory.

CAUTION

The purge command with no arguments should never be called from a macro. The purge command with an argument should never be called by the macro being purged.

Examples	purge			
	purge('_sw')			
See also	User Programming			
Related	macrold Load a macro into memory (C)			

puttxt Put text file into a data file (C)

Syntax	<pre>puttxt(file)</pre>				
Description	Copies text from current experiment into a data file.				
Arguments	file is the name of a data file (i.e., a directory with a .fid or .par suffix). Do not include the suffix in the name provided to file.				
Examples	puttxt('mydata')				
See also	NMR Spectroscopy User Guide				
Related	gettxt Get text file from another file (C)				

putwave Write a wave into Pbox.inp file (M)

Syntax	putwave(sh,bw,pw,ofs,st,ph,fla,trev,d1,d2,d0)
Description	Sets up a single excitation band in the Pbox.inp file. An unlimited number of waves can be combined by reapplying putwave.
Arguments	1 to 11 wave parameters in the following predefined order:
	sh is the name of a shape file.
	bw is the bandwidth, in Hz.
	pw is the pulsewidth, in sec.
	ofs is the offset, in Hz.
	st is a number specifying the spin status: 0 for Mz, or 1 for Mxy.

ph is the phase (or phase cycle, see wavelib/supercycles).

fla is the flip angle. Note that fla can override the default flip angle.

trev concerns time reversal. It can be used to cancel time reversal if spin status (st) is set to 1 for Mxy.

 ${\tt d1}$ is the delay, in sec, prior the pulse.

d2 is the delay, in sec, after the pulse.

d0 is a delay or command prior to d1. If d0=a, the wave is appended to the previous wave.

Examples	putwave('e	burp1')				
	putwave('G	putwave('GARP',12000.0)				
	putwave('e	'esnob',600,-1248.2,1,90.0,'n','n',0.001)				
See also	NMR Spectro	scopy User Guide				
Related	Pbox	Pulse shaping software (U)				
	setwave	Write a wave definition string into the Pbox.inp file				
		(M)				

Enter pulse width pw in degrees (C)

pw(flip_an	gle,<90_pulse_width>)	
Calculates the flip tim, in μ s, given a desired flip angle and 90° pulse. The value is entered into the parameter pw.		
flip_angle is the desired flip angle, in degrees.		
90_pulse_w value of para	idth is the 90° pulse length, in μ s. The default is the meter pw90, if it exists.	
pw(30) pw(90,12.8)	
NMR Spectro	scopy User Guide	
ernst pw pw90	Calculate the Ernst angle pulse (C) Pulse width (P) 90° pulse width (P)	
	<pre>pw(flip_ang Calculates the The value is flip_angle 90_pulse_w value of para pw(30) pw(90,12.8 NMR Spectro ernst pw pw90</pre>	

pw Pulse width (P)

Description	Length of the final pulse in the standard two-pulse sequence. In "normal" 1D experiments with a single pulse per transient, this length is the observe pulse width.		
Values	0, 0.1 μ s to 8190 sec, smallest value possible is 0.1 μ s, finest increment possible is 12.5 ns.		
See also	NMR Spectroscopy User Guide		
Related	p1 First pulse width (P)		
	pw Enter pulse width parameter pw in degrees (C)		

pw

pw90 90° pulse width (P)

Descrij	ption	Length of the 90° pulse. $pw90$ is not used by pulse sequences directly, but is used by a number of commands to assist in setting up special experiments $pw90$ is also used by certain output programs to be able				
		to print the value of the pulse width in degrees instead of				
		microseconds. Note that this parameter must be updated by the user and is not automatically determined or magically correct under all circumstances.				
Va	alues	0, 0.1 μs to 8190 sec, smallest value possible is 0.1 $\mu s,$ finest increment possible is 12.5 ns.				
See	also	NMR Spectroscopy User Guide				
Re	elated	AC1S-AC11S Autocalibration macros (M) pw Enter pulse width parameter pw in degrees (C)				

pwd Display current working directory (C)

Syntax	pwd<:direc	tory>	
Description	Displays the path of the current working directory.		
Arguments	directory is a string variable with the path of the current directory.		
Examples	pwd:\$name		
See also	NMR Spectroscopy User Guide		
Related	cd	Change working directory (C)	
	dir	List files in current directory (C)	
	lf	List files in current directory (C)	
	ls	List files in current directory (C)	

pwpat Shape of refocusing pulse (P)

Systems with imaging capabilities.			
Specifies the shape of the refocusing pulse $\ensuremath{\mathtt{pw}}$ in imaging experiments			
'hard', 'sinc', 'gauss', 'sech', 'sine', or any shape resident in the system pulse shape library or libraries.			
VnmrJ Imaging NMR			
p1pat pw	Shape of an excitation pulse (P) Pulse width (P)		
	Systems with Specifies the 'hard', 'sin the system p <i>VnmrJ Imaga</i> plpat pw		

pwr Set power mode in directly detected dimension (C)

Description Selects the power spectra display mode by setting dmg='pwr'. In the *power mode*, each real point in the displayed spectrum is calculated as the sum of the squares of the real and imaginary points comprising each respective complex data point. All information, including noise, is positive and the relationship between signal and noise is non-linear.

For multidimensional data, pwr has no effect on data prior to the second Fourier transform. If pmode='full', pwr acts in concert with the commands ph1, av1 or pwr1 to yield the resultant contour display for the 2D data.

See also NMR Spectroscopy User Guide

Related	av	Set abs. value mode in directly detected dimension (C)
	av1	Set abs. value mode in 1st indirectly detected dimension
		(C)
	dmg	Data display mode in directly detected dimension (P)
	ft	Fourier transform 1D data (C)
	ft1d	Fourier transform along f_2 dimension (C)
	ft2d	Fourier transform 2D data (C)
	pa	Set phase angle mode in directly detected dimension (C)
	pa1	Set phase angle mode in 1st indirectly detected dimension
		(C)
	ph	Set phased mode in directly detected dimension (C)
	ph1	Set phased mode in 1st indirectly detected dimension (C)
	pmode	Processing mode for 2D data (P)
	pwr1	Set power mode in 1st indirectly detected dimension (C)
	pwr2	Set power mode in 2nd indirectly detected dimension (C)
	wft	Weight and Fourier transform 1D data (C)
	wft1d	Weight and Fourier transform f2 of 2D data (M)
	wft2d	Weight and Fourier transform 2D data (M)

pwr1 Set power mode in 1st indirectly detected dimension (C)

Description Selects the power spectra display mode along the first indirectly detected dimension by setting dmg1='pwr1'. If the parameter dmg1 does not exist, pwr1 creates it and sets it to 'pwr1'. In the *power mode*, each real point in the displayed trace is calculated as the sum of the squares of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the real-real and imaginary-real points from each respective hypercomplex data point are used in the summation. In this mode, all information, including noise, is positive and the relationship between signal and noise is non-linear.

The pwrl command is only needed if mixed-mode display is desired. If the parameter dmgl does not exist or is set to the null string, the display mode along the first indirectly detected dimension defaults to

the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of pwr1 is the same as for traces, provided that pmode='partial' or pmode=''. See also NMR Spectroscopy User Guide Related dmg1 Data display mode in 1st indirectly detected dimension (P) Set phase angle mode in directly detected dimension (C) pa pa1 Set phase angle mode in 1st indirectly detected dimension (C) Processing mode for 2D data (P) Set power mode in directly detected dimension (C) pwr

pwr2 Set power mode in 2nd indirectly detected dimension (C)

pwr2 Set power mode in 2nd indirectly detected dimension (C)

Description Selects the power spectra display mode along the second indirectly detected dimension by setting dmg2='pwr2'. If dmg2 does not exist or is set to the null string, pwr2 will create dmg2 and set it equal to 'pwr2'. In the *power mode*, all information, including noise, is positive and the relationship between signal and noise is non-linear. Each real point in the displayed trace is calculated as the sum of the squares of the real and imaginary points comprising each respective complex data point. For hypercomplex data, the real-real and imaginary-real points from each respective hypercomplex data point are used in the summation.

The pwr2 command is only needed if mixed-mode display is desired. If the parameter dmg2 does not exist or is set to the null string, the display mode along the second indirectly detected dimension defaults to the display mode of the directly detected dimension (characterized by the parameter dmg). For the contour display of multidimensional data, the result of pwr2 is the same as for traces, provided that pmode='partial' or pmode=''.

See also NMR Spectroscopy User Guide

Relatedav2Set abs. value mode in 2nd indirectly detected dimension
(C)dmg2Data display mode in 2nd indirectly detected dimension
(P)ft1dFourier transform along f_2 dimension (C)ft2dFourier transform 2D data (C)ph2Set phased mode in 2nd indirectly detected dimension (C)pmodeProcessing mode for 2D data (P)pwrSet power mode in directly detected dimension (C)

pwsadj Adjust pulse interval time (M)

Applicability Systems with waveform generators. Syntax pwsadj(shape_file,pulse_parameter) Adjusts the pulse interval time so that the pulse interval for the Description specified shape is an integral multiple of 100 ns. This ensures there is no time truncation error in executing the shaped pulse by waveform generators. Arguments shape_file is a file name of a shaped pulse file. The name can be specified with or without the .RF file extension. pwsadj first looks for the file name specified by shape_file in the user's shapelib directory. If the file specified is not found there, pwsadj then looks in the system shapelib directory. pulse parameter is a string containing the adjusted pulse interval time. Examples pwsadj('pulse12','pulseparam') See also User Programming Related dmfadj Adjust decoupler tip-angle resolution time (M) Adjust second decoupler tip-angle resolution time dmf2adj (M)

pwxcal Decoupler pulse calibration (M)

Description Provides an interactive method of selecting the decoupler (first, second, or third) and the nucleus $({}^{13}C, {}^{15}N, \text{ or }{}^{31}P)$ to calibrate. The pwxcal pulse sequence determines the pulse width characteristics of the probe's decoupler channel(s) in indirect detection or triple resonance experiments. pwxcal can also be used to determine the rf field homogeneity of the decoupler.

The parameter pwx1 is arrayed to calibrate the 90° pulse width on the first decoupler. If a second decoupler is present, the parameter pwx2 is arrayed to calibrate the 90° pulse width on that decoupler. If a third decoupler is present, the parameter pwx3 is arrayed to calibrate the 90° pulse width on that decoupler. If a third decoupler is present, the parameter pwx3 is arrayed to calibrate the 90° pulse width on that decoupler. Other parameters include: jC13 is the ¹³C-¹H coupling, constant, jN15 is the ¹⁵N-¹H coupling constant, jP31 is the ³¹P-¹H coupling constant, and jname is a selected calibration nucleus.

See also System Administration

Description	A flag to enable or disable Bloch-Siegert shift correction during the creation of Pbox pulses.		
Values	'y' enable Bloch-Siegert shift correction 'n' disable Bloch-Siegert shift correction Default value is 'y'.		
See also	NMR Spectroscopy User Guide		
Related	htfrq1 Hadamard frequency list in ni (P)		

pxrep Flag to set the level of Pbox reports (P)

Description	A flag to set the level of Pbox debug messages displayed at the start of acquisition.	
Values	'y' shows all Pbox reports. 'h' shows the Hadamard matrix. 'n' shows no reports. Default value is 'nnn'.	
See also	NMR Spectroscopy User Guide	
Related	htfrq1 Hadamard frequency list in ni (P)	

pxsetAssign Pbox calibration data to experimental parameters(M)

Syntax	pxset<(fil	e.ext)>
Description	Retrieves experimental settings from a file and assigns them to corresponding experimental parameters using a dialog form. If no file name is provided, pxset extracts data from the Pbox.cal file that contains the output data of the last created waveform	
Arguments	file.ext is	the name of a shape or pattern file.
Examples	pxset pxset('Pbox.RF')	
See also	NMR Spectroscopy User Guide	
Related	Pbox pboxget	Pulse shaping software (U) Extract Pbox calibration data (M)

pxshape Generates a single-band shape file (M)

- Syntax pxshape('sh bw/pw ofs st ph fla trev \ d1 d2 d0',name,disp)
- Description Generates a single-band waveform based on wave definition provided as a single string of wave parameters.
- Arguments A single string of 1 to 12 wave parameters in predefined order. Note that a single quote is required at the start and the end of the entire string, but no single quotes are required surrounding characters and strings inside the entire string.

sh is the name of a shape file.

bw/pw is either the bandwidth, in Hz, or the pulsewidth, in sec.

ofs is the offset, in Hz.

st is a number specifying the spin status: 0 for Mz, or 1 for Mxy.

ph is the phase (or phase cycle, see wavelib/supercycles).

fla is the flip angle. Note that fla can override the default flip angle.

trev is a time reversal. This can be used to cancel time reversal if spin status (st) is set to 1 for Mxy.

d1 is the delay, in sec, prior the pulse.

d2 is the delay, in sec, after the pulse.

d0 is a delay or command prior to d1. If d0=a, the wave is appended to the previous wave.

name is the output file name. An extension is optional and can be used to override an internally defined shape type.

disp is the shape is displayed by default in the graphics window. If disp is set to 'n', the shape is not displayed.

Examples pxshape('eburp1', 'myshape.RF') pxshape('GARP 12000.0', 'shape2', 'y') pxshape('esnob 600.0 -1248.2 n 180.0 n n 0.001', 'xxx') See also NMR Spectroscopy User Guide

Related Pbox Pulse shaping software (U)

Pxsim Simulate Bloch profile for a shaped pulse (U)

Syntax	Pxsim file <simtime <add="" <num_steps="" sub="">>></simtime>
Description	Used by the dprofile macro to simulate a Bloch profile for a shaped pulse. Pxsim extracts the information necessary for simulation from the shape header. Only shape files containing this information can be processed.
Arguments	file is the name of a shape or pattern file including an .RF or .DEC extension. Pxsim searches for the file in the user's shapelib

	(~/vnmrsys/shapelib), and if not found there, it searches in the system shapelib (vnmr/shapelib).		
simtime is the maximum simulation time (in sec) that can provided.			
	num_steps is the number of steps in the profile.		
	add/sub is add (a) or subtract (s) from the previous simulation.		
Examples	Pxsim myshape.RF		
See also	NMR Spectroscopy User Guide		
Related	Pbox Pulse shaping software (U)		

Pxspy Create shape definition using Fourier coefficients (U)

Syntax	Pxspy file
Description	An interactive program that converts shaped pulse files into a Fourier series and produces an output file pbox.cf in the user's shapelib (~/vnmrsys/shapelib), which can be used to create a wave definition file in the wavelib directory. Pxspy can also be used to convert hard pulse decoupling sequences into soft ("cool") decoupling waveforms. The resulting Fourier coefficients can depend on the number of points in the waveform.
Arguments	file is the name of a shape or pattern file, including an .RF, .DEC, or .GRD extension. The name can be given as a relative name, absolute name, or as a simple name (i.e., with a path). If given as a simple name, Pxspy searches for the file in the user's shapelib (~/vnmrsys/shapelib), and then if not found there, it searches in the system shapelib (vnmr/shapelib).
Examples	Pxspy myshape.RF Pxspy /vnmr/shapelib/myshape.RF Pxspy ~vnmrsys/shapelib/myshape.RF
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

<pslabel>_plotExperiment-Specific Plot Macro

Description The <pslabel>_plot macro, if it exists, is executed at set-up and is used to configure plotting and display features on a pslabel-specific basis.

Ρ

<pslabel>_processExperiment-Specific Processing Macro

Description The <pslabel>_process macro, if it exists, is executed at set-up and is used to configure processing parameters on a pslabel-specific basis.

<pslabel>_setupExperiment-Specific Setup Macro

Description The <pslabel>_setup macro is executed to set up sequence-specific parameters.

Related cpsetup



Agilent VnmrJ 4 Command and Parameter Reference Guide

qcomp	Longer dead time for longer ring down (P)
QKexp	Set up quick experiment (M)
qtune	Tune probe using swept-tune graphical tool (C)
?	Display the value of an individual parameter)
quadtt	Prints differences in wideline receiver channels

gcomp Longer dead time for longer ring down (P)

Applicability Systems with Varian, Inc. Cold Probes

- Description Global parameter to handle longer ring down times following the rf pulse. This is only active if dsp='i' or if dsp='r' and fsq='y'. The dead time is calculated by the software and the DSP parameters are appropriately adjusted for flat baseline and good phase properties. If it is necessary to use a user specified delay, create the prealfa parameter. qcomp is not effective in explicit acquisition experiments. Not compatible with srof2.
 - Values qcomp='y' triggers a longer dead time before the receiver is gated on for the acquisition.

Related prealfa Specify a delay for longer ring down (P) dsp Type of DSP for data acquisition (P)

QKexp Set up quick experiment (M)

Syntax	QKexp(arguments)
Description	Set up parameters for quick experiment for a chained acquisition. Multiple arguments can be given to define the chain. Default parameter values are used by the macro and or the probe file is used.
Examples	QKexp('PROTON','COSY','HMQC') QKexp('PROTON','CARBON','HETCOR','gCOSY')



0

gtune Tune probe using swept-tune graphical tool (C)

qtune<(gai	n<,power>)>
Displays a real-time graph showing reflected power versus frequency for tuning probes. If the acquisition system has been recently rebooted, enter su before running qtune. Refer to the manual <i>NMR Spectroscopy</i> <i>User Guide</i> for a detailed description of this tool.	
gain specifies the gain value, typically 20 to 50. The default is 50. power specifies the power value, typically 60 to 70. The default is 60.	
qtune qtune(20) qtune(38,6)	5)
NMR Spectroscopy User Guide	
tugain su tune	Amount of receiver gain used by qtune (P) Submit a setup experiment to acquisition (M) Assign frequencies (C)
	<pre>qtune<(gain Displays a re for tuning pro enter su befo User Guide ff gain specific qtune qtune(20) qtune(38,65 NMR Spectroo tugain Su tune</pre>

?

Display the value of an individual parameter (C)

Syntax	parameter_name<[index]>?	
Description	The question mark displays the current numerical or string value of a parameter when the parameter name is followed by a question mark. No change is made to the value of the parameter. To display an individual element of an parameter array, provide the index in square brackets (e.g., nt[3]? might display "nt[3]=2")	
	Certain parameters can be "turned off" by setting the parameter to 'n'. The display of a parameter that is turned off will be the phrase "Not Used" followed by the actual value in parentheses. For example, if lb is set to 1.5 and then set to 'n', entering lb? will display lb= Not Used (1.5). Such a parameter can be "turned on" by setting it to 'y'. It will then have its prior value.	
	To show a parameter's array of values or learn about its attributes, use the display command.	
Arguments	index is the integer for a selected member of an arrayed parameter.	
Examples	1b? sw? pw[2]?	
See also	NMR Spectroscopy User Guide	
Related	displayDisplay parameters and their attributes (C)getvalueGet value of a parameter in a tree (C)	

quadtt Prints differences in wideline receiver channels

Syntax	quadtt
Applicability	VnmrJ 3.1
Description	Prints differences in wideline receiver channels.
See also	See Wideline Accessory Installation Manual Pub. No. 87-178257-00 Rev B788 or later. Used with pulse sequence s2pulg.



Agilent VnmrJ 4 Command and Parameter Reference Guide

r	Recall display parameter set (M)
r(n)	Recall some display parameters (C)
r1-r7	Real-value storage for macros (P)
ra	Resume acquisition stopped with sa command (C)
random	Return a random number
rcvrwt	Weighting for different receivers (P)
react	Recover from error conditions during werr processing (M)
readallshims	Read all shims from hardware (M)
readbrutape	Read Bruker data files from 9-track tape (U)
readfile	Read the contents of a text file into two parameters (C)
readhw	Read current values of acquisition hardware (C)
readlk	Read current lock level (C)
readparam	Read one of more parameters from a file (C)
readultra	Read shim coil setting for Ultra•nmr shim system (M)
real	Create a real variable without a value (C)
recon_all	Reconstruct images from 2D MRI fid data (C)
record	Record keyboard entries as a macro (M)
redor1	Set up parameters for REDOR1 pulse sequence (M)
redosy	Restore 2D DOSY display from sub experiment (M)
reff1	Reference f2 Indirect Dimension from Observe Dimension (M)
reff2	Reference f2 Indirect Dimension from Observe Dimension (M)
reffrq	Reference frequency of reference line (P)
reffrq1	Reference freq. of reference line in 1st indirect dimension (P)



reffrq2	Reference freq. of reference line in 2nd indirect dimension (P)
refpos	Position of reference frequency (P)
refpos1	Position of reference frequency in 1st indirect dimension (P)
refpos2	Position of reference frequency in 2nd indirect dimension (P)
refsourcel	Center frequency in 1st indirect dimension (P)
refsource2	Center frequency in 2nd indirect dimension (P)
region	Divide spectrum into regions (C)
relayh	Set up parameters for RELAYH pulse sequence (M)
rename	Move and/or rename a file (C)
reorder3D	Reorders array elements in arrayed phase sensitive 2D experiment (M)
reqparcheck	Flag which enables/disables required parameters (P)
reqparclear	Clears the parameters in required parameter list (M)
reqparlist	List of required parameters (P)
reqpartest	Tests whether required parameters are set (M)
resetf3	Reset parameters after a partial 3D Fourier transform (M)
resetplotter	Reset plotter to system plotter (M)
resetsampglob al	Clears sample global parameters
resolv	Set resolution enhancement parameters (M)
restorenuctab le	Calculate and (Re-)store accurate nuctable (M)
resume	Resume paused acquisition queue (C)
return	Terminate execution of a macro (C)
rev	System software revision level (P)
revdate	System software preparation date (P)
rfband	RF band in use (P)
rfblk	Reverse FID block (C)
rfchannel	Independent control of rf channel selection (P)
rfchtype	Type of rf channel (P)
rfdata	Reverse FID data (C)
rfl	Reference peak position in directly detected dimension (P)
------------	--
rfl1	Reference peak position in 1st indirectly detected dimension (P)
rfl2	Reference peak position in 2nd indirectly detected dimension (P)
rfp	Reference peak frequency in directly detected dimension (P)
rfp1	Reference peak freq. in 1st indirectly detected dimension (P)
rfp2	Reference peak freq. in 2nd indirectly detected dimension (P)
rftempcomp	RF Transmitter Board Temperature Compensation (P)
rftrace	Reverse FID trace (C)
rftype	Type of rf generation (P)
rfwg	RF waveform generator (P)
right	Set display limits to right half of screen (C)
rights	Determine an operator's specified right (C)
rinput	Input data for a regression analysis (M)
rl	Set reference line in directly detected dimension (M)
rl1	Set reference line in 1st indirectly detected dimension (M)
r12	Set reference line in 2nd indirectly detected dimension (M)
rm	Delete file (C)
rmdir	Remove directory (C)
rmsAddData	Add transformed data files with weighting (U)
Roesy	Convert the parameter to a ROESY experiment (M)
Roesyld	Convert the parameter set to a Roesy1d experiment (M)
rofl	Receiver gating time preceding pulse (P)
rof2	Receiver gating time following pulse (P)
rof3	Receiver gating time following T/R switch (P)
rotate	Rotate 2D data (C)
rotorsync	Rotor synchronization (P)
rp	Zero-order phase in directly detected dimension (P)

rp1	Zero-order phase in 1st indirectly detected dimension (P)
rp2	Zero-order phase in 2nd indirectly detected dimension (P)
rt	Retrieve FIDs (M)
rtcmx	Return Spinsight data into current experiment (C)
rtp	Retrieve parameters (M)
rts	Retrieve shim coil settings (C)
rttmp	Retrieve experiment data from experiment subfile (M)
rtv	Retrieve individual parameters (C)
rtx	Retrieve parameters based on rtx rules (C)

Recall display parameter set (M)

Syntax (1) rset_number

(2) r(set_number)

Description Recalls the parameters sp, wp, sp1, wp1, sp2, wp2, sc, wc, sc2, wc2, ho, vo, vs, and ai/nm of a selected display parameter set. Not recalled are phase parameters, drift correction parameters, integral reset parameters, and reference parameters. This allows, for example, saving a set of display parameters, adjusting the phase or drift correction, and later recalling the display parameters without und0oing the new phase or drift correction.

Arguments set_number is the number, from 1 to 9, of a display parameter set. Examples r2

r(3)

See also NMR Spectroscopy User Guide

Related	ai	Select absolute intensity mode (C)
	fr	Full recall of a display parameter set (M)
	ho	Horizontal offset (P)
	nm	Select normalized intensity mode (C)
	S	Save display parameters as a set (M)
	SC	Start of chart (P)
	sc2	Start of chart in second direction (P)
	sp	Start of plot in directly detected dimension (P)
	sp1	Start of plot in 1st indirectly detected dimension
		(P)
	sp2	Start of plot in 2nd indirectly detected dimension
		(P)
	vo	Vertical offset (P)

r

vs	Vertical scale (P)
WC	Width of chart (P)
wc2	Width of chart in second direction (P)
wp	Width of plot in directly detected dimension (P)
wp1	Width of plot in 1st indirectly detected dimension
	(P)
wp2	Width of plot in 2nd indirectly detected dimension
	(P)

r(n) Recall some display parameters (C)

Applicability	All		
Syntax	r(n<,noupdate>)		
Description	r(n) recalls only the following parameters: sp, wp, sp1, wp1, sp2, wp2, sc, wc, sc2, wc2, ho, vo, vs, and ai/nm.		
	noupdate – as a second argument prevents the automatic update of interactive programs.		
Arguments	n=1 to 9		
See also	User Programming		
Related	<pre>fr(n) Recall all the parameters of the specified display parameter set (C)</pre>		
	s (n) Save a copy of the current values of all display parameters (C)		

r1-r7 Real-value storage for macros (P)

Description	The seven parameters r1, r2, r3, r4, r5, r6, and r7 are available in each experiment for macros to store a real value.		
See also	User Programming		
Related	dgs	Display group of special/automation parameters (M)	
	n1,n2,n3	Name storage for macros (P)	

ra Resume acquisition stopped with sa command (C)

Description Resumes an experiment acquisition that was stopped with the sa command. ra is not permitted after any parameters have been brought into the stopped experiment with the rt or rtp macros. The parameters dp and np may not be altered. ra applies to the experiment that you are joined to at the time the

command is entered. If experiment 1 has been previously stopped with sa, you must be joined to experiment 1 for ra to resume that

acquisition. If you are in experiment 2, entering ra has no effect on experiment 1.

If an experiment has been stopped with sa, you can increase the number of transients nt and resume the acquisition with ra. You cannot, however, increase nt and enter ra if the experiment had completed in a normal fashion (i.e., it was not stopped with sa).

Note that the completion time and remaining time shown in the Acquisition Status window are not accurate after ra is executed.

See also NMR Spectroscopy User Guide

Related	dp	Double precision (P)
	np	Number of data points (P)
	nt	Number of transients (P)
	rt	Retrieve FID (M)
	rtp	Retrieve parameters (M)
	sa	Stop acquisition (C)

random Return a random number

Syntax random<(max <, 'real'>)>:val

Applicability VnmrJ 3.1

Description Return a random number. By default, it returns a random integer between 0 and 2^31-1. (2^31-1 is 2147483647 or, in hexadecimal, 0x7fffffff). If an optional number is supplied, the returned value will be between 0 and that value. If an optional keyword 'real' is supplied, the random number will be returned as a real number.

Arguments The difference between random(10) and random(10,'real') is that the first will only return whole numbers between 0 to 10. The second call, with the 'real' option, will return fractional numbers such as 2.342, 7.324, etc.

If a max value is supplied, the conversions are slightly different if a real number or integer is returned. This is to avoid truncation problems with integer math.

For real numbers:

double val = random();

val = val / 2147483647.0; /* results in value from 0.0 to 1.0 */

val = val * max; /* scales from 0.0 to max (max may be negative) */

For integers:

long val = random();

long range = 2147483647 / (abs(max) + 1); /* determine size of max + 1 ranges of integers */

val = val / range;

if (max < 0)
val = -val;</pre>

Arguments

Examples To return a random real number between 0.0 and 1.0, use random(1,'real'):\$val

rcvrwt Weighting for different receivers (P)

Applicability Systems with multiple receivers.

Description An array of real numbers giving weighting factors to use when combining multiple receiver data. The i'th array element is used to weight data from the i'th receiver. Applying a weight factor is like increasing the gain of the receiver by the same factor (but the weights are specified as numerical factors rather than in dB).
 Examples rcvrwt=10, 12, 8

react Recover from error conditions during werr processing (M)

Syntax react<('wait')>

- Description When an acquisition error occurs, any action specified by the werr parameter is executed. The react macro is a prototype for handling these errors. This macro can be invoked for error handling by setting werr='react'. The acqstatus parameter is provided so that react can determine which specific error has occurred.
- Arguments 'wait' is a keyword for a special type of error handling during an automation run. The react macro always uses the 'next' option when it calls the command au. Under certain conditions, it is also appropriate to use the 'wait' option. react checks to see if an argument was passed to it; that is, werr='werr(\'wait\')' to determine whether to use the 'wait' option of au.
 - See also NMR Spectroscopy User Guide

Related	acqstatus	Acquisition status (P)
	au	Submit experiment to acquisition and process data (C)
	werr	Specify action when error occurs (C)
	werr	When error (P)

readallshimsRead all shims from hardware (M)

Description Reads all shims from the hardware and sets the values into the shim parameters in the current parameter tree. The shims used depend on the shimset configuration. For the shim set on the Ultra•nmr shim

system, readallshims is active only if hardware-to-software shim communication is enabled. See also NMR Spectroscopy User Guide Related load Load status of displayed shims (P) Read current values of acquisition hardware (C) readhw setallshims Set all shims into hardware (M) sethw Set values for hardware in acquisition system (C) shimset Type of shim set (P) su Submit a setup experiment to acquisition (M)

readbrutape Read Bruker data files from 9-track tape (U)

Syntax	(From UNIX) readbrutape file <number_skipped></number_skipped>	
Description	A shell script that reads one file from a Bruker tape into a UNIX file with the name specified. Bruker tapes are likely to be made at 1600 bpi, although 1600 bpi is not a requirement.	
Arguments	file is the name of the file read into UNIX. For identification, the .bru extension is added to the file name.	
	number_skipped is the number of files skipped and <i>includes</i> the header file (which is assumed to be the first file on the tape). The default is the script reads the first file after the header file. If number_skipped equals 0, there is no rewinding and the first file (or the next file) on the tape is read.	
See also	NMR Spectroscopy User Guide	
Related	convertbru Convert Bruker data (M,U)	

readfile Read the contents of a text file into two parameters (C)

Examples readfile (path, par1, par2, <, cmpstr <, tree> >):num

Description readfile reads the contents of a file and puts the contents into two supplied parameters. The first word on each line in the file is placed in the first parameter. The remainder of the line is placed in the second parameter. An optional fourth argument specifies a string which is used to match the first word of the line. For example, if the file contained:

H1pw 10 H1pwr 55 C13pw 14 C13pwr 50 and the comparison string was set to H1, only the lines starting with H1 would be put into the parameters. Namely, H1pw and H1pwr. Arguments path is the path name of the file to read. par1 is the name of the parameter to hold the first word of the line. par2 is the name of the parameter to hold the remainder of each line.

cmpstr is the optional comparison string for matching the first word.

tree is an optional parameter to select the tree for par1 and par2. The possibilities are current, global, and local. Current is the default. Local is used if the parameters are \$macro parameters. If tree is used, the cmpstr must also be supplied. If cmpstr is '', then it is ignored.

The par1 and par2 parameters must already exist. If par1 or par2 are defined as a real parameter, as opposed to a string parameter, then if the value does not have a number as the first word, a zero will be assigned.

num will be set to the number of items in the arrayed parameters par1 and par2.

Lines that only contain white space are not added to the parameters. Lines that start with a # are not added to the parameters. Lines which start with a # can be used as comment lines. If a line only contains a single word, that word is put into the first parameter. The corresponding array element of the second parameter will be set to an empty string. The readfile will return the number of lines added to the parameters.

Examples Examples using a prototype file containing the following:

A readfile test case # Proton values H1pw 10 H1pwr 55 # Carbon values C13pw 14 C13pwr 50 H1macro ft f full aph vsadj End

readfile(systemdir+'/probes/testcase','attr','vals')

This sets the attr and vals parameters to arrays of six strings.

attr='H1pw','H1pwr','C13pw','C13pwr','H1macro','End' vals='10','55','14','50','ft f full aph vsadj',''

readfile(systemdir+'/probes/testcase','attr','vals','H1
')

This sets the attr and vals parameters to arrays of three strings.

attr='H1pw','H1pwr','H1macro'
vals='10','55','ft f full aph vsadj'

The readfile command might be used in conjunction with the teststr command. The teststr command can be used to search an arrayed parameter to determine the index of a specified element.

For example,

teststr(attr, 'H1pwr'):\$e
vals[\$e] will be the value of 'H1Pwr'

Syntax readhw("param1", "param2", ...)<:r1, r2, ...>

readhw("keword"):\$res1,...

Description Returns or displays the current values of the lock system parameters lockpower, lockgain, lockphase, lock, temp, loc, and z0.

The values of the shims can also be obtained. The particular shims that can be read depends upon the type of shim hardware present in the system. See the description of shimset for a list of the shim names for each type of shim hardware.

Shim DACs read by readhw:

- •Axial shim: z1, z2, z3, z4, z1c, z2c
- •Non-axial shims: x1, y1, xz, yz, xy, x2y2, x3, y3
- Special Oxford magnets shims: z5, xz2, yz2, zx2y2, zxy
- Arguments param1, param2, ... parameter to read maximum of 10 parameters.

 $r1, r2, \ldots$ Vnmr variables hold the returned results

no variables supplied – results are displayed in the text panel Keywords:

loc -sample changer location.

temp – returns the sample temperature, controller status, and set point. Results are displayed in the text panel if no variables are supplied

Returned value	Status
0	Regulation off
1	Regulated
2	Not regulated
3	No controller

status — returns the systems status as an integer. The returned values are:

Returned value	Status
10	IDLE
15	PARSE
16	PREP
17	SYNCED
20	ACQUIRE
25	PAD
30	VTWAIT
40	SPINWAIT
50	AGAIN
60	ALOCK
61	AFINDRES
62	APOWER
63	APHASE
70	SHIMMING
80	SMPCHANGE
81	RETRIEVSMP
82	LOADSMP
90	INTERACTIVE
100	TUNING

Returned value S		us CTIVE
Erro	r messages	
-1 -2	Avai acqu	lable on spectrometer only (i.e. system = 'datastation') isition not active (acquisition communication programs
-7	cons	sole powered down or not connected
	Results are disp	played in the text panel if no variables are supplied.
	readhw cannot be used when an acquisition is in progress or when acqi is connected to the acquisition system.	
Arguments	param1, param2, are the names of the parameters to be read. value1,value2, are return variables to store the settings of the parameters specified. The default is to display the setting in the status window.	
Examples	readhw('z1c','z2c','z1','z2') readhw('z1c','z2c','z1','z2'):r1,r2,r3,r4 readhw('temp'):\$t sets \$t	
See also	NMR Spectroscopy User Guide	
Related	lockgain	Lock gain (P)
	lockphase	Lock phase (P)
	lockpower	Lock power (P)
	readallshims	Read all shims from hardware (M)
	sethw	Set values for hardware in the acquisition system (C)
	shimset	Type of shim set (P)

readlk Read current lock level (C)

Syntax	readlk<:lock_level>		
Description	Returns the same information as would be displayed on the digital lock display using the manual shimming window. readlk can be used in developing automatic shimming methods such as shimming via grid searching. It <i>cannot</i> be used during acquisition or manual shimming.		
Arguments	lock_level returns the current lock level.		
Examples	readlk readlk:\$levell		
See also	User Programming		
Related	alock Automatic lock status (P)		

readparam Read one or more parameters from a file (C)

Syntax	readparam(file,parlist[,tree[,type]])-
Description	The readparam command will read one or more parameters from a
	specified file. The first argument is the name of the file. The second

argument is either a list of the names of the parameters to be read or it is the name of an arrayed temporary \$ variable. If it is a list, it is a string parameter and the names can be separated either by a space or a comma.

If it is an arrayed temporary \$ varaible, each array element is a single parameter name. If a parameter in the list or array is not present in the file being read, no error is generated. The optional third argument is the tree into which the parameters are read.

The variable trees are 'current', 'global'. 'processed' and 'systemglobal'.

The optional fourth argument controls the behavior of the readparam command.

The options are 'read', 'replace', 'add', 'list', and 'alist \$par'.

The default type is 'read'. In order to specify a type other than 'list' or 'alist', the tree must also be specified.

The list and alist types is somewhat special, since they only return the list of parameter names in the file. They does not actually set any parameters in any tree. The tree argument can be included or not. It is not used. The second argument also is not used by the list or alist types. It can be passed as a empty string. The typical invocations will be readparam(file, '', 'list'):\$parlist or

readparam(file,'','alist \$parlist'):\$num

In the 'list' case, the \$parlist return variable will be set to a space separated list of parameter names. If the specified file does not contain any parameters, or the file does not exist, \$parlist will be returned as an empty string (\$parlist='').

In the 'alist \$parlist' case, the \$num return variable will be set to the number of returned values. The actual values will be returned as an array in the \$parlist variable.

If the specified file does not contain any parameters, or the file does not exist, \$parlist will be returned as an empty string (\$parlist='').

Examples In order to specify the type, the tree must also be specified. The behaviors are best illustrated with specific examples. Lets say that there is a temporary file containing only the parameters a and b. We are going to use the readparam command to read parameters into a current tree which contains the parameters a and c but does not contain the parameters b and d. This can be summarized as:

Parameters in mypar: a=1 b=2

Initial parameters in current tree: a=4 c=8 (b and d do not exist) Example 1.

readparam(curexp+'/mypar','a b c d','current','read')

Parameter in a current tree is replaced with parameter from mypar. Parameter b in current tree is read in from mypar Parameter c in current tree is unaltered Parameter d in current tree still does not exist. Final parameters in current tree: a=1 b=2 c=8 (d does not exist). Example 2.

readparam(curexp+'/mypar','a b c
d','current','replace')

Parameter in a current tree is replaced with parameter from mypar. Parameter b in current tree still does not exist. Parameter c in current tree is deleted. Parameter d in current tree still does not exist. Final parameters in current tree: a=1 (b c and d do not exist).

Example 3.

readparam(curexp+'/mypar','a b c d','current','add')

Parameter in a current tree is unaltered. Parameter b in current tree is read in from myper Parameter c in current tree is unaltered. Parameter d in current tree still does not exist. Final parameters in current tree: a=4 b=2 c=8 (d does not exist).

Example 4.

```
$list='a b c d'
readparam(curexp+'/mypar',$list,'current','add')
This is the same as Example 4.
$arraylist='a','b','c','d'
```

readparam(curexp+'/mypar','\$arraylist','current','add')

This is also the same as Example 3, however the variable names are passed as an arrayed temporary \$ variable \$arraylist. Note the single quotes around the second argument to readparam. The name of the local temporary \$variable is passed to the command, not its value. This format is useful if the list of parameters to read becomes large.

This command may be used to read temporary values which have been saved with the writeparam command.

More Examples:

readparam(curexp+'/mypar', 'in') reads the parameter in from the file mypar in the current experiment directory.

readparam(curexp+'/mypar','sw ct np','processed') reads the parameters sw, ct, and np into the processed tree from the file mypar in the current experiment directory.

readparam(curexp+'/mypar','','list'):\$parlist sets
\$parlist='a b in sw ct np'

readparam(curexp+'/mypar','','alist \$parlist'):\$num sets
\$num=6 sets \$parlist='a','b','in','sw','ct','np'

Applicability	Systems with the Ultra • nmr shim system.		
Syntax	readultra<(file_number)>		
Description	Reads shim set files for a Ultra•nmr shim system from a Sun floppy disk into VnmrJ. The floppy disk for Ultra•nmr contains up to 63 shim sets named file1.dac to file63.dac.		
Arguments	file_number is the number of the shim set file, from 1 to 63. The default is to read all of the shim set files.		
Examples	readultra readultra(6)		
See also	NMR Spectroscopy User Guide		
Related	shimsetType of shim set (P)svsSave shim coil settings (C)		

real Create a real variable without a value (C)

Syntax	real(variable)		
Description	Creates a real variable without a value.		
Arguments	variable is the name of the variable to be created.		
Examples	real('realval1')		
See also	User Programming		
Related	create Create a new parameter in a parameter tree (C)		
	string Create a string variable (C)		

recon_all Reconstruct images from 2D MRI fid data (C)

Applicability	Imaging Systems				
Syntax	recon_all(acqstring, <pc option="">)</pc>				
	or				
	<pre>recon_all(acqstring,<image directory=""/>,<pc option=""></pc></pre>				
	or				
	recon_all				

- Description Produces 2D images (in fdf format) from FID data acquired with most 2D imaging sequence, including sems, gems, fsems, and epi. Supported features:
 - Compressed/Standard/Arrayed experiments supported (relevant VNMR parameter: seqcon)
 - Capable of running concurrently with acquisition (set acqstring to acq after first wnt; empty or dummy string initially).
 - Disable image display (relevant parameter: recondisplay. Create in processed tree as a real variable and set it to 0)
 - Display every N images (relevant parameter: recondisplay. Create in processed tree as a real variable and set it to N)
 - DC removal (relevant parameter: dcrmv)
 - Image shifting (relevant VNMR parameter: lsfrq, lsfrq1)
 - Multi-shot/sorting (relevant parameters: petable, etl, and/or nseg)
 - Multi-slice (interleaved) acquisitions (relevant VNMR parameter: ns)
 - Separate output from multiple receivers (relevant VNMR parameter: rcvrout, a string. Set to i, will yield either raw- (if VNMR parameter raw is set) or image-domain magnitude and phase images for separate coils)
 - Multi-echo imaging support (sems, epi) (relevant VNMR parameter: ne)
 - Multiple receiver data (magnitude sum) (relevant parameter: rcvrs)
 - Weighting (through VnmrJ panel selections) (relevant parameter: ftproc)
 - Zero filling (through VnmrJ panel selections) (relevant parameters: fn and/or fn1)
 - Output magnitude and/or phase raw data components. (relevant (optional) parameter: raw. Create in processed tree as a string which can be set to 'm' (magnitude), 'p' (phase), or 'b' (both))
 - Partial k-space conjugation. Relevant parameters are fract_kx and fract_ky, which denote the number of points/echoes acquired beyond the intended N/2. Example: nv=80, fract_ky=16 results in the central 32 echoes used as a correction map prior to conjugate

synthesis. Resulting image has $128 (2^{*}(80-16))$ lines in the phase encoded direction.

• Phase correction (relevant parameters: image, epi_pc). Implemented for epi sequences. Phase of transformed imaging data (image=1) is corrected by phase of transformed reference data (image=0). Accepted values for pc option in command string or for the optional parameter epi_pc are:

POINTWISE (the default; direct use of the phase of profile)

LINEAR (1st order fit of phase of profile)

QUADRATIC (2nd order fit of phase of profile)

CENTER_PAIR (even/odd pair at center of echo train used for all even/odd echoes)

PAIRWISE (even/odd pair phase differences along echo train used)

 $6.\ensuremath{\mathsf{FIRST}}\xspace$ PAIR (1st and 2nd echoes used for even/odd correction)

- Navigator Echo correction. Requires acquisition of *echo train* data (fsems, epi), some of which are not phase encoded. Adjusts phase of encoded echoes according to the phase of navigator echoes of the same echo train, relative to the first such navigator echo. Relevant parameters are:
- navigator (can be string set to 'y' or 'n', or array of integers giving navigator echo positions within the echo train (i.e., navigator=1,2).)
- nav_type (optional; string, set to 'off' to disable correction or 'POINTWISE' (default)).

Order of operation per echo in block:

- 1 DC removal
- 2 echo reversal if necessary
- **3** raw data output if requested
- 4 windowing if necessary
- 5 read direction Fourier transform
- **6** phase correction if necessary
- **7** sorting if necessary

Order of operation per slice:

- 1 navigator correction if necessary
- 2 windowing in phase direction if necessary
- **3** partial Fourier correction if necessary
- **4** phase direction Fourier transform
- 5 accumulation of multi-receiver data
- 6 write fdf output file

Arguments

acqstring	Set to ' perform retrosp	acq' to indicate concurrent reconstruction; is no initialization. Any other value can be used for ective reconstruction or the first pass through
pc option	Optiona description	rent reconstruction (initialization is performed). Al argument to specify phase correction method (see
image directory	Optiona	al argument to specify the directory which will contain
NB	recon for cont variable	a_all accesses parameters in the PROCESSED tree trol of some features. It is in the PROCESSED tree that es should be created and/or modified for
Input/Output	of the c written when ru in study files are the curr generat recon director	a_all reads the FID file in the acqfil subdirectory surrent experiment, and creates fdf files that are to the recon subdirectory of the current experiment un in standalone mode, or to the study tree when run y mode. If raw data output is selected, the resulting fdf written to the rawmag or rawphs subdirectory of rent experiment. If phase images are optionally ted, the resulting fdf files are written to the uphs subdirectory of the current experiment's ry.
Exar	nples	recon_all(",'/usr/home/myimages') recon_all(",'/usr/home/myimages','CENTERPAIR') recon_all('ignorethis','LINEAR') recon_all('acq')
See	e also	VnmrJ Imaging User's Guide

record Record keyboard entries as a macro (M)

Syntax record<(file|'off')>

Description Records keyboard entries and stores the entries as a MAGICAL macro in the user's maclib directory. To start recording keyboard entries, enter record. You are prompted for a macro name (you can also give the name as an argument to record). The command line prompt then becomes "Command?" to indicate that the record macro is active. Type the MAGICAL commands to be recorded on the keyboard. Function keys can be included by entering F1 to F8 for function keys 1 to 8, respectively. Enter off or record('off') to finish the recording.
Arguments file is the name of the macro file in which the entries are saved. The default is that the user is prompted for a file name. If the macro file name already exists, the user is asked if the file should be overwritten. 'off' is a keyword to stop recording the entries.
Examples record record('mymacro') record('off')
See also User Programming

redor1 Set up parameters for REDOR1 pulse sequence (M)

Applicability	Three-channel systems with a triple-tuned MAS solids probe.		
Description	Sets up a parameter set, obtained withXPOLAR1, for REDOR (rotational echo double-resonance) experiment.		
See also	User Guide: Solid-State NMR		
Related	xpolar1 Set up parameters for XPOLAR1 pulse sequence (M)		

redosy Restore 2D DOSY display from sub experiment (M)

- Description Restores the previous 2D DOSY display (if one exists) by recalling the data stored by the dosy macro in the file subexp/dosy2Ddisplay in the current experiment. undosy and redosy enable easy switching between the 1D DOSY data (spectra as a function of gzlvl) and the 2D DOSY display (signal as a function of frequency and diffusion coefficient).
 - See also NMR Spectroscopy User Guide
 - Related dosy Process DOSY experiments (M) undosy Restore original 1D NMR data from subexperiment (M)

reff1 Reference f1 Indirect Dimension from Observe Dimension (M)

Syntax	reff1<(refsource1)>
Description	Macros uses the ratio of the Ξ values for the relevant nuclei from
	refsource1 or the reference source specified to determine the

reference frequency in the f1 indirect dimension directly from the reference frequency in the observe dimension using the formula:

reffrq1 = (reffrq / Ξ[tn]) * Ξ[nucf1]
rfp1=0
rfl1 = sw1/2 - (frq[f1] - reffrq1)*1e6

 Ξ is the normalized frequency such that the $^1\mathrm{H}$ signal from TMS is 100.00 MHz.

Referencing in the observe dimension using setref and this method is same as using setref1 (apart from minor round-off errors).

Referencing the observe dimension to an internal reference standard as proposed by IUPAC references all dimensions to that single reference signal and not the lock as with setref, setref1, and setref2.

Limitations: the macro works with data recalled from an archive or acquired on an other system provided the data was acquired using VNMR6.1C or newer.

Referencing is based on nuctables/nuctabrefBio if bioref='y' (global or local). Setting bioref='n' (global or local) or if the flag does not exist the standard IUPAC / organic chemistry referencing (nuctables/nuctabref) is used.

See /vnmr/nuctables/nuctabref.

Arguments No argument – reference source is determined from refsource1. If the relevant parameter is missing, the macro tries to determine the (indirect) reference source from the axis parameter.

'sfrq', 'dfrq2', 'dfrq3', or 'dfrq4' as a reference source

Examples reff1 reff1('sfrq')

Related reff2 Reference f2 Indirect Dimension from Observe Dimension (M)

- setref Set Frequency Referencing for Proton Spectra (M)
- setref1 Set Frequency Referencing for f1 Evolution Dimension (M)
- setref2 Set Frequency Referencing for f2 Evolution Dimension (M)

mref Set Referencing Based on Spectrum from the same sample (M)

bioref Flag for Bio-NMR Referencing (P)

reff2 Reference f2 Indirect Dimension from Observe Dimension (M)

Syntaxreff2<(refsource2)>DescriptionMacros uses the ratio of the E values for the relevant nuclei from
refsource1 or the reference source specified to determine the
reference frequency in the f1 indirect dimension directly from the
reference frequency in the observe dimension using the formula:

R

reffrq1 = (reffrq / Ξ [tn]) * Ξ [nucf1]
rfp1=0
rfl1 = sw1/2 - (frq[f1] - reffrq1)*1e6

 Ξ is the normalized frequency such that the $^1\mathrm{H}$ signal from TMS is 100.00 MHz.

Referencing in the observe dimension using setref and this method is same as using setref1 (apart from minor round-off errors).

Referencing the observe dimension to an internal reference standard as proposed by IUPAC references all dimensions to that single reference signal and not the lock as with setref, setref1, and setref2.

Limitations: the macro works with data recalled from an archive or acquired on an other system provided the data was acquired using VNMR6.1C or newer.

Referencing is based on nuctables/nuctabrefBio if bioref='y' (global or local). Setting bioref='n' (global or local) or if the flag does not exist the standard IUPAC / organic chemistry referencing (nuctables/nuctabref) is used.

See /vnmr/nuctables/nuctabref.

Arguments No argument – reference source is determined from refsource2. If the relevant parameter is missing, the macro tries to determine the (indirect) reference source from the axis parameter.

'sfrq', 'dfrq', 'dfrq2', 'dfrq3', or 'dfrq4' as a reference source

Examples reff2('dfrq3')

Relatedreff1Reference f2 Indirect Dimension from Observe Dimension
(M)setrefSet Frequency Referencing for Proton Spectra (M)setref1Set Frequency Referencing for f1 Evolution Dimension (M)setref2Set Frequency Referencing for f2 Evolution Dimension (M)mrefSet Referencing Based on Spectrum from the same sample
(M)biorefFlag for Bio-NMR Referencing (P)

reffrq Reference frequency of reference line (P)

Description Reference frequency, in MHz, of the reference line. This parameter is set by the rl macro. By defining reffrq as the conversion factor between Hz and ppm using the unit command, ppm calculations can be made.

> If referencing is on (i.e., refpos is not set to 'n'), the go, ga, and au macros calculate values of rfl and rfp based on reffrq and refpos. If referencing is off, go, ga, and au set reffreq to sfrq.

See also	NMR Spectroscopy User Guide		
Related	au	Submit experiment to acquisition and process data (M)	
	crl	Clear reference line in directly detected dimension (M)	
	ga	Submit experiment to acquisition and FT the result (M)	
	go	Submit experiment to acquisition (M)	
	reffrq1	Ref. frequency of reference line in 1st indirect dimension	
		(P)	
	reffrq2	Ref. frequency of reference line in 2nd indirect dimension	
		(P)	
	refpos	Position of reference frequency (P)	
	rfl	Reference peak position in directly detected dimension (P)	
	rfp	Reference peak frequency in directly detected dimension	
		(P)	
	rl	Set reference line in directly detected dimension (M)	
	sfrq	Transmitter frequency of observe nucleus (P)	
	unit	Define conversion units (C)	

reffrq1 Reference freq. of reference line in 1st indirect dimension (P)

Description	Reference frequency, in MHz, of the reference line in the first indirect dimension of a nD experiment. This parameter should be used as the		
	conversion factor between hertz and ppm in the first indirect dimension.		
See also	NMR Spectroscopy User Guide		
Related	crl1	Clear reference line in 1st indirectly detected dimension (M)	
	reffrq	Reference frequency of reference line (P)	
	refpos1	Position of reference frequency in 1st indirect dimension (P)	

reffrq2 Reference freq. of reference line in 2nd indirect dimension (P)

Description Reference frequency, in MHz, of the reference line in the second indirect dimension of a 2D experiment. This parameter should be used as the conversion factor between hertz and ppm in the second indirect dimension.
 See also NMR Spectroscopy User Guide
 Related cr12 Clear reference line in 2nd indirectly detected dimension (M)

refpos Position of reference frequency (P)

Description	Position of reference frequency, set by the setref and rl macros. Setting refpos='n' indicates that referencing has been turned off. The crl macro turns referencing off.		
Values	Because all spectra are (by definition) referenced to a frequency at 0 ppm, refpos is either 0 or "not used".		
See also	NMR Spectroscopy User Guide		
Related	crl	Clear reference line in directly detected dimension (M)	
	reiirq	Reference frequency of reference line (P)	
	refpos1	Position of reference frequency in 1st indirect dimension (P)	
	refpos2	Position of reference frequency in 2nd indirect dimension	
		(P)	
	rl	Set reference line indirectly detected dimension (M)	
	setref	Set frequency referencing (M)	

refpos1 Position of reference frequency in 1st indirect dimension (P)

Description	Position of reference frequency in the first indirect dimension of a nD experiment, set by setref1 and rl1 macros. Setting refpos1='n' indicates that f1 referencing has been turned off. The crl1 macro turns f1 referencing off.		
Values	Because all spectra are (by definition) referenced to a frequency at 0 ppm, refpos1 is either 0 or "not used".		
See also	NMR Spectroscopy User Guide		
Related	crl1	Clear reference line in 1st indirectly detected dimension (M)	
	reffrq1	Ref. frequency of reference line in 1st indirect dimension (P)	
	refpos	Position of reference frequency (P)	
	rl1	Set reference line in 1st indirect dimension (M)	
	setref1	Set frequency referencing for 1st indirectly detected dimension (M)	

refpos2 Position of reference frequency in 2nd indirect dimension (P)

- Description Position of reference frequency in the second indirect dimension of a 3D experiment, set by setref2 and rl2 macros. Setting refpos2='n' indicates that f2 referencing has been turned off in 3D spectra. The crl2 macro turns f2 referencing off.
 - Values Because all spectra are (by definition) referenced to a frequency at 0 ppm, refpos2 is either 0 or "not used".
 - See also NMR Spectroscopy User Guide
 - Related crl2 Clear reference line in 2nd indirectly detected dimension (M) reffrq2 Ref. frequency of reference line in 2nd indirect dimension
 - (P) refpos Position of reference frequency (P)
 - r12 Set reference line in 2nd indirect dimension (M)
 - setref2 Set frequency referencing for 2nd indirectly detected
 dimension (M)

refsource1 Center frequency in 1st indirect dimension (P)

Description Holds a parameter name to be used as the center frequency in the first indirect dimension of 2D experiments. If refsource1 does not exist, the default is 'sfrq'.

For 2D experiments, the second dimension may be related to sfrq if it is a homonuclear experiment. The second dimension may also be related to dfrq if it is a heteronuclear experiment. refsource1 would then be set as refsource1='sfrq' and refsource1='dfrq', respectively.

See also NMR Spectroscopy User Guide

RelateddfrqTransmitter frequency of first decoupler (P)refsource2Center frequency in 2nd indirect frequency (P)sfrqTransmitter frequency of observe nucleus (P)

refsource2 Center frequency in 2nd indirect dimension (P)

Description Holds a parameter name to be used as the center frequency in the second indirect dimension. refsource2 is analogous to refsource1
See also NMR Spectroscopy User Guide
Related refsource1 Center frequency in 1st indirect dimension (P)

R

region Divide spectrum into regions (C)

Description Breaks a spectrum up into regions containing peaks.

Arguments tail_length is the length from 0.0 to sw, in Hz, that is added to the start and end of each calculated peak region; default value is sw/10. The default value is used if a negative number is entered for this argument. If the addition of these wings would cause overlap between adjacent regions, the wings are reduced until the regions no longer overlap.

relative_number is a number that, in combination with other factors, governs the relative number of regions to be found. The default is 12, which is used if 0 is entered for this argument.

relative_number is used as part of a test to determine whether two spectral areas containing peaks are close enough together to be represented as a single region. There are no strict rules that associate the value of relative_number to the total number of regions that will be found. In general, increasing this number decreases the number of regions that will be found and increases the size of an individual region. A value of 1 would give more regions; a value of 100 would give fewer regions.

threshold is a sensitivity factor used to decide if a data point is large enough, relative to the noise level, to qualify it as part of a peak. The default value is 0.6, which is used if 0 is entered for this argument. Smaller values of threshold make peak selection more sensitive; larger values make peak selection less sensitive.

number_points governs the number of successive data points, normally from 7 to 40, that must qualify as part of a peak (see the description of threshold above) in order for that spectral area to be considered a real peak. The default value is a function of fn, sw, weighting functions, and other values. The default is used if 0 is entered for this argument. For carbon spectra with large spectral windows, experimental peaks often contain only one or two data points. Adjust number_points to 1 or 2 in those cases.

tail_size is a number that, in combination with relative_number and other factors, governs whether two spectral areas that contain peaks are close enough together to be represented as a single region. The default value is used if 0 is entered for this argument.

number_regions is the total number of regions determined by region.

 Examples
 region

 region:\$1
 region(50,0,1)

 region(-1,0,0,2):r1
 See also

 See also
 NMR Spectroscopy User Guide

 Related
 fn

 Sw
 Spectral width in directly detected dimension (P)

R

relayh Set up parameters for RELAYH pulse sequence (M)

Description	Sets up parameters for absolute-value COSY, or a single or double RELAY-COSY pulse sequence.		
See also	NMR Spect	roscopy User Guide	
Related	Cosy	Set up parameters for COSY pulse sequence (M)	
	cosyps	Set up parameters for phase-sensitive COSY (M)	
	Dqcosy	Set up parameters for double quantum filtered COSY (M)	

rename Move and/or rename a file (C)

Syntax	<pre>rename(from_file,to_file)</pre>		
Description	Renames and/or moves a file or directory. rename is identical in function to the command mv .		
Arguments	from_file	is the name of the file to be moved to renamed.	
	to_file is t from_file a sure the to_	the name of the file after moving or renaming it. If the argument has an extension such as .fid or .par, be file argument has the same extension.	
Examples	rename('/home/vnmr1/vnmrsys/seqlib/d2pul', '/vnmr/seqlib/d2pul')		
See also	NMR Spectroscopy User Guide		
Related	сору	Copy a file (C) Copy a file (C)	
	delete	Delete a file, parameter directory, or FID directory (C)	
	mv	Move and/or rename a file (C)	
	rm	Delete file (C)	

reorder3D Reorders array elements in arrayed phase sensitive 2D experiment

Syntax	reorder3D
Applicability	VnmrJ 3.1
Description	Exchanges the order of the two arrayed parameters in an arrayed phase sensitive 2D experiment. Useful if 3D DOSY data are acquired with array='phase,gzlvll' instead of array='gzlvll,phase'.
See also	dosy

reqparcheck Flag which enables/disables required parameters (P)

Syntax	reqparcheck=	'Y' or 'n'	
	Description:		
Description	The parameter r 'n'. Only if it is If it is set to 'n'	reqparcheck is a flag with the possible values of 'y' or set to 'y' are actual parameters compared to the file. ', reqpartest will always return 0.	
Values	'y' or 'n', indicating whether required parameters are to be checked.		
Related	callacq	Utility macro to call Acq command (M)	
	reqparlist	List of required parameters (P)	
	reqparclear	Clears the parameters in required parameter list	
		(M)	
	reqpartest	Tests whether required parameters are set (M)	

regparclear Clears the parameters in required parameter list (M)

Syntax reqparclear

Description Clears the parameters listed in reqparlist. If for some reason reqparlist has been destroyed, then this macro exits without a message. The parameter is cleared on the current tree, if it exists there, or on the global tree, if it exists there. If it exists in neither place, a message is printed and the routine moves on to the next parameter in reqparlist.

The definition of "clear" is that real parameters are turned "off" and string parameters are set to the empty string ".

There is a known issue with this macro, which due to its obscurity will remain as "user beware." The issue is that if a parameter of the same name exists in both the 'global' and 'current' trees, and if that parameter is part of reqparlist, then it will be cleared in the 'current' tree but not in the global tree. Users should just not be doing this.

Also note that while this macro checks for reqparlist=", if it is an array and any element in the array is " then it assumes " is a parameter and reports a "does not exist" message.

 Related
 callacq
 Utility macro to call Acq command (M)

 reqparcheck
 Flag which enables/disables required parameters (P)

 reqparlist
 List of required parameters (P)

 reqpartest
 Tests whether required parameters are set (M)

R

regparlist List of required parameters (P)

Description The parameter reqparlist holds the parameter names. It is an array of strings. It will not array the experiment.

RelatedcallacqUtility macro to call Acq command (M)gettokenUtility macro to separate a string into tokens (M)reqparcheckFlag which enables/disables required parameters (P)reqparclearClears the parameters in required parameter list (M)reqpartestTests whether required parameters are set (M)

regpartest Tests whether required parameters are set (M)

Syntax reqpartest<('showtext'|'showgui'<,callback_string>)>

Description If the parameter reqparcheck='y', then this macro examines the list of parameter names in reqparlist and if all of them exist and are properly set, returns 0. Properly set is defined as a non- empty string for string parameters, or the active bit set (parameter is 'on') for real parameters.

This macro also checks the string which is the concatenation of autoname + globalauto + sqname for any parameters in that string. Parameters in this string are delimited by \$.

For convenience, this macro will return different values depending on the specific non-true condition, as defined in the following table (X is "don't care").

All parameters exist	Т	Х	F	Т	F
All parameters set	Т	Х	Т	F	F
regparcheck='y	'T	F	Т	Т	Т
return value	0	-1	1	2	3

Also note that the non-existence of either reqparcheck or reqparlist is equivalent to reqparcheck not set to 'y'.

Parameters are checked in the current tree first for existence, and if that parameter exists there, then that tree is checked for whether it is set. If it does not exist in the current tree, then the global tree is checked. If and only if it exists in neither tree is it considered to not exist.

If the argument to this macro is 'showtext' then if one or more parameters do not exist or are not properly set, then they are listed on the alphanumeric (text) screen.

If the argument to this macro is 'showgui', then an entry popup is displayed for both creation (of non-existing parameters) and value entry. The return value is not affected by the fact that the values are now being entered - in other words, the return value is to be interpreted as 'did not exist' or 'was not set' prior to running the macro.

	The comprehen reqparlist+a occur, and this	sive list to check is utoname+globalauto+sqname. Some duplicates may macro checks and eliminates duplicates.	
	The argument of passed onto Vn required param otherwise synch	callback_string is an optional argument that gets mrJ, and then gets passed back to vnmrbg when the eters entry popup closes. VnmrJ and vnmrbg are not pronized, so this allows for re-entrance.	
Arguments	'showgui' showtext'		
	'showgui' dis set;	plays an entry popup in the required parameter is not	
	'show text' displays information about the required parameters in the text window		
	callback_str (ignored in 'sh	ing - optional callback to vnmrbg from VnmrJ nowtext' option)	
See also	VnmrJ User Programming		
Related	callacq	Utility macro to call Acq command (M)	
	reqparcheck	Flag which enables/disables required parameters (P)	
	reqparclear	Clears the parameters in required parameter list (M)	

resetf3 Reset parameters after a partial 3D Fourier transform (M)

Description Restores the acquisition parameter sw, the processing parameter fn, and the display parameters sp, wp, rfl, and rfp in the 3D parameter set, which are read into VnmrJ by either the select command or the dplane or dproj macros. These parameters were modified due to the selection of regional f_3 processing (ptspec3d ='ynn'). The original value for each of these parameters is stored in the parameter \$sv, where \$ represents sw, fn, sp, wp, rfl, or rfp (e.g., swsv).

If a 2D plane into VnmrJ is retrieved from a 3D transformed data set that was processed with regional f_3 processing, resetf3 must be run before executing ft3d in that particular VnmrJ environment.

See also NMR Spectroscopy User Guide

Related	dplane	Display a 3D plane (M)
	dproj	Display a 3D plane projection (M)
	fn	Fourier number in directly detected dimension (P)
	ft3d	Perform a 3D Fourier transform (M)
	ptspec3d	Region-selective 3D processing (P)
	rfl	Ref. peak position in directly detected dimension (P)
	rfp	Ref. peak frequency in directly detected dimension (P)
	select	Select a spectrum or 2D plane without displaying it (C)
	sp	Start of plot (P)
	SW	Spectral width in directly detected dimension (P)

R

vnmrjcmd() Commands to invoke the GUI popup (C)
wp Width of plot (P)

resetplotterReset plotter to system plotter (M)

Description Command to reset a (temporarily chosen) plotter back to the system plotter sysplotter. Command is called by all plotfile/plotpreview and plot/autoplot buttons on plot panels.

resetsampglobalClears sample global parameters

Description	Clears sample global parameter values in the current workspace.
Examples	resetsampglobal
Related	getsampglobal, resetsampglobal, savesampglobal, mvsampglobal, showsampglobal

resolv Set resolution enhancement parameters (M)

Syntax	resolv<(a,b)>		
Description	Calculates a default resolution enhancement function, setting up 1b and gf based on the acquisition time at. "Zero-filling" is also accomplished, if possible, by making fn $\geq \geq 2 \cdot np$.		
Arguments	a sets a value of 1b using $lb=-0.318/(a^*sw)$. The default for a is 0.1.		
	b sets a valu	e of gf using gf=b*sw. The default for b is 0.3 .	
Examples	resolv resolv(.2,.4)		
See also	NMR Spectroscopy User Guide		
Related	at	Acquisition time (P)	
	fn	Fourier number in directly detected dimension (P)	
	gf	Gaussian function in directly detected dimension (P)	
	lb	Line broadening in directly detected dimension (P)	
	np	Number of data points (P)	
	SW	Spectral width in directly detected dimension (P)	

restorenuctableCalculate & store accurate nuctable for current system (M)

Syntax restorenuctable

Description The setref contribution is a generic nucleus table, /vnmr/nuctables/nuctable, based on a standard proton frequency of 1000.0 MHz. All standard nucleus tables in the /vnmr/nuctables are symbolic links pointing to a generic table.

> The restorenuctable is used to replace the standard links with specific links that to files containing proper and accurately calculated nucleus tables. Problems arising with custom macros and third party software that are not aware of the symbolic links pointing to a generic table can be fixed using this macro.

> Commands and utilities that do not scale nuctable entries to the actual proton frequency (as they should) will work better than with the standard tables.

Limitations: restorenuctable is not compatible with quune and certain commands in current software.

Examples restorenuctable

Related nuctable Display nucleus table for a given H1 frequency (M)

resume Resume paused acquisition queue (C)

- Description Enables continuing submitting experiments to the acquisition system. For experiments initiated with the command au('wait'), the acquisition is paused during the time of data processing in order to prevent the acquisition from submitting new experiments that might be queued. resume then allows the data processing macro to initiate another acquisition with au('next'), which is then performed immediately instead of at the end of the queue.
 - See also NMR Spectroscopy User Guide
 - Related au Submit experiment to acquisition and process data (C)

return Terminate execution of a macro (C)

Syntax	return<(expression1,expression2,)>
Description	Terminates the execution of a macro and optionally returns values to another calling macro. This is usually used after testing some condition. return is used only in macros and not entered from the keyboard.
Arguments	expression1, expression2, are return values to another calling macro

See also User Programming

Related abort Terminate action of calling macro and all higher macros (C)

rev System software revision level (P)

- DescriptionStores a string identifying the VnmrJ software version for the system.
This parameter is not be entered by the user, but can be examined by
entering rev?.See alsoVnmrJ Installation and Administration
 - Related revdate System software preparation date (P)

revdate System software preparation date (P)

Description Stores a string identifying the date the current VnmrJ software version was prepared. This parameter is not be entered by the user, but can be examined by entering revdate?.
See also VnmrJ Installation and Administration
Related rev System software revision level (P)

rfband RF band in use (P)

- Description Indicates which rf band of the amplifier is in use for each channel.
 Values A string, such as 'hlc', in which the first channel is determined by the first character, the second channel is determined by the second character, and so forth. The following values are available for each channel:

 'h' indicates the high rf band is in use on the channel.
 'l' indicates the low rf band is in use on the channel.
 'c' indicates the system software will calculate whether to use the high band or the low band for the channel.
 - See also NMR Spectroscopy User Guide

rfblk Reverse FID block (C)

Syntax	rfblk(<src_expno>,src_blk_no,dest_expno,dest_blk_no)</src_expno>
Description	Reverses and copies data from a source FID block specified by
	src_blk_no to a destination FID block specified by dest_expno and

dest_blk_no, using memory-mapped input and output. The file header determines the size and type of data to reverse.

rfblk searches for the source and destination FID file in the directory \$vnmruser/expN/acqfil; N is the requested experiment number or the current experiment number. If the FID file is not open, rfblk opens the file, copies the data, and closes the file. If a number of blocks need to be copied, explicitly opening and closing the files with the commands mfopen and mfclose can significantly speed up the data reformatting process.

rfblk can also be used to append blocks of data to a FID file by specifying that the dest_blk_no is greater than the number of blocks in a file.

Be aware that rfblk can modify data returned to an experiment with the rt command. To avoid modification, enter the following sequence of commands before running rfblk:

```
cp(curexp+'/acqfil/fid', curexp+'/acqfil/fidtmp')
rm(curexp+'/acqfil/fid')
mv(curexp+'/acqfil/fidtmp', curexp+'/acqfil/fid')
```

Arguments src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.

src_blk_no specifies the source block of data to be copied. Block
numbers run from 1 to the number of blocks in a file.

dest_expno specifies the experiment number of the destination FID file.

dest_blk_no specifies the destination block to send the copied data.

Examples rfblk(1,2,1) reverses and copies block 1 from the current experiment to block 1 of experiment 2.

See also User Programming

Related	mfblk	Move FID block (C)
	mfclose	Memory map close FID file (C)
	mfdata	Move FID data (C)
	mfopen	Memory map open FID file (C)
	mftrace	Move FID trace (C)
	rfdata	Reverse FID data (C)
	rftrace	Reverse FID trace (C)

rfchannel Independent control of rf channel selection (P)

Description Gives override capability over the selection of rf channels. rfchannel does not normally exist but can be created by a user with the command create('rfchannel','flag').

The control of each rf channel is built around a collection of parameters and pulse sequence statements. The frequency of channel 1 is set by sfrq and tof, its power by tpwr and tpwrf. The first decoupler uses the corresponding parameters dfrq, dof, dpwr, and dpwrf, respectively. Furthermore, the decoupler can have modulation modes specified with the parameters dmf, dm, dmm, dres, and dseq,. The second decoupler has the same set of parameters as the first decoupler and they are distinguished by appending a 2 to each name. That is, the names aer dfrq2, dof2, dpwr2, dpwrf2, dmf2, dm2, dmm2, dres2, and dseq2. The third decoupler would use parameters with a 3 appended: dfrq3, dof3, dpwr3, dpwrf3, dmf3, dm3, dmm3, dres3, and dseq3. The rfchannel parameter provides a mechanism to override the default parameter usage.

- Values A string of one to four characters in which the position of each character identifies the rf channel controlled.
 - The first character selects which rf channel (1 to 4) the parameters sfrq, tof, tpwr, etc. control. The first character also identifies the rf channel used as the receiver.
 - The second character selects which rf channel (1 to 4) the parameters dfrq, dof, dpwr, etc. control.
 - The third character maps the parameter set dfrq2, dof2, dpwr2, etc. to an rf channel (1 to 4).
 - The fourth character maps tdfrq3, dof3, dpwr3, etc. to an rf channel (1 to 4).

For example, rfchannel='132' would exchange control of the second and third rf channels from the default parameter usage.

The number of characters in the rfchannel parameter must match the number of real rf channels (defined by the parameter numrfch) and each rf channel must be selected by the parameter.

Besides remapping the parameters to different rf channels, pulse sequence statements are also remapped. For example, if rfchannel='132', then statements decpulse, decshaped_pulse, decoffset, decpower, decspinlock, and so on are applied on rf channel 3 and dec2pulse, dec2shaped_pulse, and so on are applied on rf channel 2.

An obvious use for this remapping is on systems with the decoupler set to U+ H1 Only in the Spectrometer Configuration window. On these systems, if multinuclear pulses are needed and ¹H needs to be observed, the parameter sets that assume a dual-broadband system can be used and the parameters remapped by setting rfchannel='21'. However, internal logic checks if the first decoupler is set to U+ H1 Only, tn is set to 'H1', and dn is not set to 'H1'. If these settings are the case, the parameter mapping for rf channels 1 and 2 is exchanged automatically.

See also NMR Spectroscopy User Guide; User Programming

create	Create new parameter in parameter tree (C)
dfrq	Transmitter frequency for first decoupler (P)
dm	Decoupler mode for first decoupler (P)
dmf	Decoupler modulation frequency for first decoupler (P)
dmm	Decoupler modulation mode for first decoupler (P)
dn	Nucleus for first decoupler (P)
dof	Frequency offset for first decoupler (P)
	create dfrq dm dmf dmm dn dof

dpwr	Power level for first decoupler with linear amplifier (P)
dpwrf	First decoupler fine power (P)
dres	Tip-angle resolution for first decoupler (P)
dseq	Decoupler sequence for first decoupler (P)
numrfch	Number of rf channels (P)
sfrq	Transmitter frequency for observe nucleus (P)
tn	Nucleus for observe transmitter (P)
tof	Frequency offset for observe transmitter (P)
tpwr	Observe transmitter power level with linear amplifiers (P)
tpwrf	Observe transmitter fine power (P)

rfchtype Type of rf channel (P)

Description	Configuration parameter for type of rf on each channel. The value for
	a channel is set using the Type of RF label in the Spectrometer
	Configuration window. Pulse sequence programs check rfchtype to
	determine if indirect detection should be used for some experiments.
	Indirect detection occurs automatically if the decoupler is set to U+ H1
	Only in the Spectrometer Configuration window, tn is set to 'H1', and
	dn is not set to 'H1'.

Values The values of rfchtype parallel the rftype values. The only distinction is that the setting for rftype is 'd' on the U+ Direct Synthesis and U+ H1 Only entries.

'U+ Direct Synthesis' is the setting for a system with direct synthesis (U+ Direct Synthesis in the Spectrometer Configuration window).

'U+ H1 Only' is a fixed-frequency proton system (U+ H1 Only in Spectrometer Configuration window).

'Deuterium Decoupler' is the setting for a system deuterium decoupler channel.

'Direct Synthesis' is the setting for direct synthesis (Direct Synthesis in the Spectrometer Configuration window).

'Broadband' is the setting for broadband (Broadband in the Spectrometer Configuration window).

'Fixed Frequency' is the setting for fixed frequency (Fixed Frequency in the Spectrometer Configuration window).

'SIS Modulator' is the setting for imaging modulator (SIS Modulator in the Spectrometer Configuration window).

See also VnmrJ Installation and Administration

Related config Display current configuration and possibly change it (M) dn Nucleus for first decoupler (P) rftype Type of rf generation (P) tn Nucleus for observe transmitter (P)

rfdata Reverse FID data (C)

Description Reverses and copies data specified by src_start_loc from a FID block specified by src_blk_no to a destination location specified by dest_expno, dest_blk_no, and dest_start_loc, using memory-mapped input and output. The data point locations and the num_points to be reversed are specified by data points corresponding to the np parameter, not bytes or complex points; however, when reversing the data, rfdata looks at the file header to determine the size and type of data to reverse.

> rfdata searches for the source and destination FID file in the directory \$vnmruser/expN/acqfil; N is the requested experiment number or the current experiment number. If the FID file is not open, rfdata opens the file, copies the data, and closes the file. If a number of blocks need to be copied, explicitly opening and closing the files with the commands mfopen and mfclose can significantly speed up the data reformatting process.

> Be aware that rfdata can modify data returned to an experiment with the rt command. To avoid modification, enter the following sequence of commands before running rfdata:

cp(curexp+'/acqfil/fid', curexp+'/acqfil/fidtmp')
rm(curexp+'/acqfil/fid')
mv(curexp+'/acqfil/fidtmp', curexp+'/acqfil/fid')

Arguments src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.

src_blk_no specifies the source block of data to be copied. Block
numbers run from 1 to the number of blocks in a file.

src_start_loc specifies the starting data location within the specified block to copy the data. Data locations start from 0 and are specified as data points corresponding to the np parameter.

dest_expno specifies the experiment number of the destination FID file.

dest_blk_no specifies the destination block to send the copied data.

dest_start_loc specifies the starting data destination location within the specified block to send the copied data.

Examples rfdata(1,0,2,1,(nv-1)*np,np) copies and reverses np points of data from the starting location 0 of block 1 of the current experiment to the data location (nv-1)*np of block 1 of experiment 2.

See also User Programming

Related	mfblk	Move FID block (C)		
	mfclose	Memory map close FID file (C)		
	mfdata	Move FID data (C)		
	mfopen	Memory map open FID file (C)		
	mftrace	Move FID trace (C)		

rfblk	Reverse	FID	block	(C)
rftrace	Reverse	FID	trace	(C)

rfl Reference peak position in directly detected dimension (P)

Description Actual position of the reference line in the spectrum (i.e., the distance from the right edge of the spectrum to the reference line). If there is no reference line in the spectrum, rfl can be used to enter the frequency where the reference line would appear if the line were present in the spectrum.

Values Number, in Hz.

See also NMR Spectroscopy User Guide

- Related rfl1 Reference peak position in 1st indirectly detected dimension (P)
 - rf12 Reference peak position in 2nd indirectly detected dimension (P)
 - rfp Reference peak frequency in directly detected dimension (P)

rfl1 Reference peak position in 1st indirectly detected dimension (P)

- Description Analogous to the rfl parameter except that rfll applies to the first indirectly detected dimension of a multidimensional data set. rfll can either be set manually or be adjusted automatically when the macro rll is used to assign a reference line.
 - Values Number, in Hz.
 - See also NMR Spectroscopy User Guide
 - Related rfl Reference peak position in directly detected dimension (P)
 - rfl2 Reference peak position in 2nd indirectly detected dimension (P)
 - rfp1 Reference peak frequency in 1st indirectly detected dimension (P)

rf12 Reference peak position in 2nd indirectly detected dimension (P)

Description Analogous to the rfl parameter except that rfl2 applies to the second indirectly detected dimension of a multidimensional data set. rfl2 can either be set manually or be adjusted automatically when the macro rl2 is used to assign a reference line.

Values Number, in Hz.
See also NMR Spectroscopy User Guide
Related rfl Reference peak position in directly detected position (P)
rfl1 Reference peak position in 1st indirectly detected dimension (P)
rfp2 Reference peak frequency in 2nd indirectly detected dimension (P)

rfpReference peak frequency in directly detected dimension
(P)

Description	Sets the frequency to be assigned to the reference line in the spectrum. rfp is always stored in Hz, but can be entered in ppm by using the p suffix (e.g., rfp=2.1p).		
Values	Number, in Hz.		
See also	NMR Spectroscopy User Guide		
Related	rfl	Reference peak position in directly detected dimension (P)	
	rfp1	Ref. peak frequency in 1st indirectly detected dimension (P)	
	rfp2	Ref. peak frequency in 2nd indirectly detected dimension (P)	
	rl	Set reference line in directly detected dimension (M)	

rfp1Reference peak freq. in 1st indirectly detected dimension
(P)

Description Analogous to the rfp parameter except that rfp1 applies to the first indirectly detected dimension of a multidimensional data set. rfp1 can either be set manually or be assigned a value when rl1 is called with an argument (e.g., rl1(7.2p) assigns the value of 7.2 ppm to rfp1). Values Number, in Hz.

See also NMR Spectroscopy User Guide

Related rfl1 Ref. peak position in 1st indirectly detected dimension (P) rfp Ref. peak frequency in directly detected dimension (P) rfp2 Ref. peak frequency in 2nd indirectly detected dimension (P) rl1 Set reference line in 1st indirectly detected dimension (M)

rfp2 Reference peak freq. in 2nd indirectly detected dimension (P)

Description Analogous to the rfp parameter except that rfp2 applies to the second indirectly detected dimension of a multidimensional data set. rfp2 can be set manually or be assigned a value when rl2 is called with an argument. For example, entering rl2(7.2p) assigns the value of 7.2 ppm to rfp2.

Values Number, in Hz.

- See also NMR Spectroscopy User Guide
 - Related rfl2 Reference peak position in 2nd indirectly detected dimension (P)
 - rfp Reference peak frequency in directly detected dimension (P)
 - rfp1 Reference peak frequency in 1st indirectly detected dimension (P)
 - r12 Set reference line in 2nd indirectly detected dimension (C)

rftempcomp RF Transmitter Board Temperature Compensation (P)

C-		
2)	/nta	ıΧ

Applicability VnmrJ 3.1

Arguments If rftempcomp='n' temperature compensation on the RF transmitter board is turned off.

If rftempcomp='c' temperature compensation on the RF transmitter board is turned on continuously and will continuously update until it is turned off.

To create the rftempcomp parameter, enter:

create('rftempcomp','string','global')

rftrace Reverse FID trace (C)

Syntax	<pre>rftrace(<src_expno,src_blk_no,src_trace_no,< pre=""></src_expno,src_blk_no,src_trace_no,<></pre>	\
	<pre>dest_expno,<dest_blk_no,dest_trace_no)< pre=""></dest_blk_no,dest_trace_no)<></pre>	
orintion	Powerses and copies FID treases specified by and trease	ma fi

Description Reverses and copies FID traces specified by src_trace_no from a FID block specified by src_blk_no to a destination location specified by dest_expno, dest_blk_no, and dest_trace_no, using memory-mapped input and output. The file header determines the size and type of data to be reversed.

> rftrace searches for the source and destination FID file in the directory \$vnmruser/expN/acqfil; N is the requested experiment number or the current experiment number. If the FID file is not open,
rftrace opens the file, copies the data, and closes the file. If a number of blocks need to be copied, explicitly opening and closing the files with the commands mfopen and mfclose can significantly speed up the data reformatting process.

You cannot use rftrace to append data to a FID file. Its purpose is for moving around data.

Be aware that rftrace can modify data returned to an experiment with the rt command. To avoid modification, enter the following sequence of commands before running rftrace:

cp(curexp+'/acqfil/fid', curexp+'/acqfil/fidtmp')
rm(curexp+'/acqfil/fid')
mv(curexp+'/acqfil/fidtmp', curexp+'/acqfil/fid')

Arguments src_expno specifies the experiment number of the source FID file. The default is the FID file of the current experiment.

src_blk_no specifies the source block of data to be copied. Block
numbers run from 1 to the number of blocks in a file.

src_trace_no specifies the source trace of data within the specified block to be copied. Trace numbers run from 1 to number of traces in a file.

dest_expno specifies the experiment number of the destination FID file.

dest_blk_no specifies the destination block to send the copied data.

src_trace_no specifies the destination trace of data within the specified block to be copied. Trace numbers run from 1 to the number of traces in a file.

- Examples rftrace(1,1,2,1,nv) copies and reverses trace 1 from block 1 of the current experiment to trace nv of block 1 of experiment 2.
- See also User Programming

Related	mfblk	Move FID block (C)
	mfclose	Memory map close FID file (C)
	mfdata	Move FID data (C)
	mfopen	Memory map open FID file (C)
	mftrace	Move FID trace (C)
	rfblk	Reverse FID block (C)
	rfdata	Reverse FID data (C)

rftype Type of rf generation (P)

Description Configuration parameter for type of rf generation on each rf channel. On other systems, the value is set using the Type of RF label in the Spectrometer Configuration window.

Values The values of rftype parallel the rfchtype values. The setting for rftype is 'd' on the entries U+ Direct Synthesis and U+ H1 Only.

	'd' is the setting for a system with direct synthesis (U+ Direct Synthesis in the Spectrometer Configuration window) or a fixed-frequency proton system (U+ H1 Only in Spectrometer Configuration window).		
	'l' is the setting for a deuterium decoupler channel.		
'c' is the setting for direct synthesis Spectrometer Configuration window).		etting for direct synthesis (Direct Synthesis in the Configuration window).	
	'b' is the setting for broadband (Broadband in the Spect Configuration window).		
	'a' is the setting for fixed frequency (Fixed Frequency in the Spectrometer Configuration window).		
	'm' is the se Spectrometer	etting for imaging modulator (SIS Modulator in the Configuration window).	
See also	VnmrJ Instal	llation and Administration	
Related	config	Display current configuration and possibly change it (M)	
	rfchtype	Type of rf channel (P)	

rfwg RF waveform generator (P)

Description	Configuration parameter for whether a waveform generator board is present or not on the current rf channel. The value for each channel is set using the Waveform Generator label in the Spectrometer Configuration window.	
Values	'n' is setting for no waveform generator board on the channel (Not Present choice in Spectrometer Configuration window).	
	${}^{\prime}{}_{\rm Y}{}^{\prime}$ is setting for a waveform generation board on the channel (Present choice in Spectrometer Configuration window).	
See also	VnmrJ Installation and Administration	
Related	config Display current configuration and possibly change it (M)	

right Set display limits to right half of screen (C)

Description	Sets the horizontal control parameters, sc and wc, to produce a display (and subsequent plot) in the right portion of the screen (and page). For 2D data, space is left for the scales.		
See also	NMR Spectroscopy User Guide		
Related	center	Set display limits for center of screen (C)	
	full	Set display limits for a full screen (C)	
	fullt	Set display limits for full screen with room for traces (C)	
	left	Set display limits for left half of screen (C)	

R

SC	Start of chart (P)
WC	Width of chart (P)

rights Determine an operator's specified right (C)

Applicability	Walkup		
Syntax	rights('right'<,'errval'>)<:\$ret>		
Description	The rights program queries the rights database to determine if the current operator has the specified right. This command is used by the interface designer to determine if and how certain options are presented. An operator does not typically use this command. The system administrator sets (restricts) the rights for an operator using VnmrJ administrator interface. By default, the rights command grants any requested right. Rights requested that are not in the rights database are granted. Granting a right means that the rights program returns a 1 to the calling macro.		
Arguments	right $-a$ specific operator right, not case sensitive.		
	• 1 is returned by the command if the specified right is granted or the right is not in the rights data base		
	• 0 is default value returned by the command if the right is both in the database and the operator does not have the specified right.		
	errval – optional argument specifying return value if a right is both in the database and the operator does not have the specified right.		
	\$ret - variable holding the return value from the right command.		
Examples	rights('prioritySample',-1):\$ok		
	Sets \$ok to -1 if the prioritySample right is not granted. A value of 1 is returned if the prioritySample is granted. Returning either a 0 or -1 if a right is not granted lets the interface designer choose to show or gray out a control.		
See also	VnmrJ Installation and Administration and VnmrJ Walkup manuals.		

rinput Input data for a regression analysis (M)

Description Formats data for regression analysis and places the data into the file regression.inp. The program is interactive. If a regression.inp already exists, rinput starts by asking if you want to overwrite the file. Type y and press the Return key. It then asks for an x-axis title and a y-axis title. Enter the titles as asked (for no title, simply press Return). Next, rinput asks you to input the data in pairs. Separate each pair of values with a blank and press Return after the second value. At the end of the data set, press Return in response to the request for data. If you have another data when it is asked for.

See also	NMR Spectro	scopy User Guide; User Programming
Related	expl	Display exponential or polynomial curves (C)
	poly0	Find mean of data in the file regression.inp (C)

r1 Set reference line in directly detected dimension (M)

Syntax	<pre>rl<(frequency)></pre>	
Description	Sets the direct dimension reference line, taking into account any frequency scaling with the scalesw parameter.	
Arguments	frequency is a value, in Hz, to assign to the reference line. The default is the cursor position cr. To enter the value in ppm, add a p suffix.	
Examples	rl	
	rl(0)	
	rl(7.2p)	
See also	NMR Spectroscopy User Guide	
Related	cr	Current cursor position in directly detected dimension (P)
	crl	Clear ref. line in directly detected dimension (C)
	reffrq	Reference frequency of the reference line (P)
	rl1	Set ref. line in 1st indirectly detected dimension (M)
	r12	Set ref. line in 2nd indirectly detected dimension (M)
	scalesw	Scale spectral width in directly detected dimension (P)

rll Set reference line in 1st indirectly detected dimension (M)

Syntax	<pre>rl1<(frequency)></pre>	
Description	Sets the first indirect dimension reference line, taking into account any frequency scaling with the scalesw1 parameter.	
Arguments	frequency is a value, in Hz, to assign to the reference line. The default is the cursor position cr1. You can enter the suffixes p, d, or k to mean ppm, decoupler ppm, and kilo, respectively. These suffixes are exactly equivalent to using $*sfrq$, $*dfrq$, and $*1000$. Thus, if you are doing a 2D experiment in which the indirect axis is determined by the decoupler channel, you might enter, for example, rl1(10d), which is equivalent to rl1(10*dfrq).	
Examples	rl1 rl1(0) rl1(7.2p)	
See also	NMR Spectroscopy User Guide	
Related	cr1Cursor position in 1st indirectly detected dimension (P)cr11Clear ref. line in 1st indirectly detected dimension (M)dfrqTransmitter frequency of first decoupler (P)	

refpos2	Position of reference frequency in 2nd indirect dimension		
	(P)		
rl	Set ref. line in directly detected dimension (M)		
r12	Set ref. line in 2nd indirectly detected dimension (M)		
scalesw1	Scale spectral width in 1st indirectly detected dimension		
	(P)		
sfrq	Transmitter frequency of observe nucleus (P)		

r12 Set reference line in 2nd indirectly detected dimension (M)

Syntax	rl2<(frequency)>	
Description	Sets the second indirect dimension reference line, taking into account any frequency scaling with the scalesw2 parameter.	
Arguments	frequency is a value, in Hz, to assign to the reference line. The default is the cursor position $cr2$. You can enter the suffixes p, d, or k to mean ppm, decoupler ppm, and kilo, respectively. These suffixes are exactly equivalent to using *sfrq, *dfrq, and *1000. Because there is no suffix for the second decoupler (i.e., the third channel), to reference the third axis using r12 you might enter (e.g., r12(45*dfrq2)).	
Examples	rl2 rl2(0) rl2(7.2p)	
See also	NMR Spectroscopy User Guide	
bee albo	Ninh Speciroscopy User Guide	
Related	cr2Cursor position in 2nd indirectly detected dimension (P)cr1Clear ref. line in directly detected dimension (C)cr11Clear ref. line in 1st indirectly detected dimension (C)cr12Clear ref. line in 2nd indirectly detected dimension (C)dfrqTransmitter frequency of first decoupler (P)dfrq2Transmitter frequency of second decoupler (P)r11Set ref. line in 1st indirectly detected dimension (M)scalesw2Scale spectral width in 2nd indirectly detected dimension (P)sfrqTransmitter frequency of observe nucleus (P)	

 \mathtt{rm}

Delete file (C)

Syntax rm(file1<, file2, ...>)

Description Removes one or more files from the file system, functioning like the UNIX command of the same name. Because it allows wildcard characters (* and ?) in the command argument and recursive file deletion with the -r option, rm is very powerful. But it can be quite dangerous—without warning important files can be inadvertently

deleted, even by experienced users. Using rm to delete files in VnmrJ is not recommended. The delete command is provided as a safer alternative.

file1, file2, are names of files to delete.		
NMR Spectroscopy User Guide		
delete	Delete a file, parameter directory, or FID directory (C)	
delexp	Delete an experiment (C)	
exists	Determine if a parameter, file, or macro exists (C)	
mv	Move and/or rename a file (C)	
rename	Move and/or rename a file (C)	
	file1, file3 NMR Spectro delete delexp exists mv rename	

rmdir Remove directory (C)

Syntax	rmdir(directory)		
Description	Removes one or more empty directories (i.e., directories without files).		
Arguments	directory is the name of the directory to be removed.		
Examples	rmdir('/home/dan/temp')		
See also	NMR Spectroscopy User Guide		
Related	delete Delete a file, parameter directory, or FID directory (C)		
	dir	List files in current directory (C)	
	lf	List files in current directory (C)	
	ls	List files in current directory (C)	
	mkdir	Create new directory (C)	

rmsAddData Add transformed data files with weighting (U)

- Applicability Systems with multiple receivers.
- Description This command is not normally executed directly by the user.

Roesy Convert the parameter to a ROESY experiment (M)

Description Convert the parameter to a rotating frame Overhauser effect spectroscopy (ROESY) experiment.

Description	Convert the parameter set to a 1D rotating frame Overhauser effect spectroscopy (Roesy1D) experiment.		
See also	NMR Spectroscopy User Guide		
Related	Proton	Set up parameters for ¹ H experiment (M).	
	sel1d	Selective 1D protocols to set up (M).	

rof1 Receiver gating time preceding pulse (P)

Description	Sets the period of time in most pulse sequences when the receiver is gated off before each pulse. This allows the amplifier to fully turn on before the start of the pulse. Systems are configured with linear
	amplifiers that are normally "blanked" to give the best possible
	signal-to-noise (i.e., the amplifiers are turned off when the receiver is turned on). The ${}^{1}\text{H}/{}^{19}\text{F}$ amplifiers have a short turn-on time, usually
	1 to 5 μs following the removal of blanking by turning the receiver off.
	The low-frequency amplifier modules have a longer turn-on time, about 40 to 60 $\ensuremath{\mu s}$.
Values	Typically 2-5 microseconds.
See also	NMR Spectroscopy User Guide

Related rof2 Receiver gating time following pulse (P)

rof2 Receiver gating time following pulse (P)

Description Sets the time after the final pulse in each pulse sequence that the receiver is gated off before acquisition begins. If "pulse breakthrough" effects are seen (a spike in the beginning of the FID), increasing rof2 can reduce or eliminate the problem, particularly for low-frequency nuclei.

values	Typically 10 microseconds.
See also	NMR Spectroscopy User Guide

Related	rof1	Receiver gating time preceding pulse (P)
	setlp0	Set parameters for zero linear phase (M	()

rof3 Receiver gating time following T/R switch (P)

Description Sets the time when the receiver is gated on following the T/R switch during the pulse. This allows for the elimination of pulse artifacts during the acquisition period.

rotate Rotate 2D data (C)

Syntax	rotate<(number_degrees)>		
Description	Rotates a 2D spectrum. Both complex and hypercomplex 2D data will work.		
Arguments	number_degrees is the amount of counter-clockwise rotation, in degrees. The default is 45.		
See also	NMR Spectroscopy User Guide		
Related	foldcc Fold INADEQUATE data about 2-quantum axis (C)		
	foldj Fold J-resolved 2D spectrum about $f1=0$ axis (C)		
	foldt Fold COSY-like spectrum along diagonal axis (C)		

rotorsync Rotor synchronization (P)

Applicability	Systems with the solids rotor synchronization module.		
Description	Configuration parameter that identifies if the system has the optional solids rotor synchronization module. The value of rotorsync is set using the Rotor Synchronization label in the Spectrometer Configuration window. Rotor synchronization requires either the Acquisition Controller board (Part No. 969204) or the Pulse Sequence Controller board (Part No. 992560) in the system.		
Values	1 is setting the choice in the	hat system has solids rotor synchronization (Present Spectrometer Configuration window).	
	0 is setting that system does not have solid rotor synchronization (Not Present choice in the Spectrometer Configuration window).		
See also	VnmrJ Installation and Administration		
Related	config	Display current configuration and possibly change it (M)	

\mathbf{rp}

Zero-order phase in directly detected dimension (P)

Description	Specifies the right phase-correction angles along the directly detected dimension according to	
	absorption spectrum(ω) = real channel(ω) * cos θ + imaginary channel(ω) *sin θ	
	where the phase angle θ is a function of frequency:	
	$\theta = rp + (\omega - \omega_0)/sw * lp$	
	ω_0 is defined as the right end of the spectrum. This dimension is referred to as the f ₂ dimension in 2D data sets, f ₃ dimension in 3D data sets, and so on.	
Values	-360 to +360, in degrees.	

NMR S ₁	pectroscopy User Guide
aph	Automatic phase adjustment of spectra (C)
aph0	Automatic phase of zero-order term (C)
lp	First-order phase in directly detected dimension (P)
rp1	Zero-order phase in 1st indirectly detected dimension (P)
rp2	Zero-order phase in 2nd indirectly detected dimension (P)
setlp0	Set parameters for zero linear phase (M)
	NMR Sp aph aph0 lp rp1 rp2 setlp0

rp1 Zero-order phase in 1st indirectly detected dimension (P)

Description	Specifies the right phase parameter along the first indirectly detected dimension, in degrees, for the f_1 dimension of a multidimensional data set during the process of phase-sensitive 2D transformation.		
See also	NMR Spectroscopy User Guide		
Related	1p1 First-order phase in 1st indirectly detected dimension (P)		
	rp Zero-order phase in directly detected dimension (P)		
	rp2 Zero-order phase in 2nd indirectly detected dimension (P)		

rp2 Zero-order phase in 2nd indirectly detected dimension (P)

Description	Controls the zero-order phase constant along the second indirectly detected dimension during a ds, dconi, or equivalent display operation on the 2D data or a 1D trace therein. This dimension is often referred to as the f_2 dimension.	
See also	NMR Spectroscopy User Guide	
Related	dconi	Interactive 2D contour display (C)
	ds	Display a spectrum (C)
	lp2	First-order phase in 2nd indirectly detected dimension (P)
	rp	Zero order phase in directly detected dimension (P)

rt Retrieve FIDs (M)

Syntax rt<(file<, 'nolog'>)>
Description Retrieves FIDs from a file into the current experiment.
The rt macro does not copy the FID into the experiment. Instead, it
links access to the original FID from the experiment. Most of the time,
this behavior is desired, because the FID file is seldom changed. By
making a link, disk space is also conserved. However, if the FID file in
the experiment is written to, the data in the original file is also written
to. It is best to make a copy of a FID file before altering it. The

	makefid cor gives details	nmand alters the FID file. The manual entry for makefid on how to make a copy of the FID.	
	As another s is a link to a the experime to be there, e using rt, you	omewhat subtle point, because the FID in the experiment another .fid file, if that .fid file is removed, the link from nt may be gone. If you expect the FID in the experiment even if you delete the .fid file from where it was retrieved a should explicitly copy the file into the experiment.	
Arguments	file is the r the FIDs to b the name (in If file.fid parameters f	name of the file that, with the suffix .fid added, contains be retrieved. The default is that the system prompts for that case, the name can be given without single quotes). does not exist and file.par does, rt retrieves the rom file.par.	
	'nolog' is a	a keyword specifying that the log file is not to be retrieved	
Examples	rt		
	rt('/vnmr/	fidlib/fid1d')	
See also	NMR Spectroscopy User Guide		
Related	fixpar	Correct parameter characteristics in experiment (M)	
	makefid	Make a FID element using numeric text input (C)	
	rtp	Retrieve parameters (M)	
	rtv	Retrieve individual parameters (C)	
	svf	Save FIDs in current experiment (M)	

rtcmx Return Spinsight data into current experiment (C)

Syntax	<pre>rtcmx<(file)></pre>
Description	Retrieves Spinsight data into the current experiment.
Arguments	file is the name of the file. The default is that the macro prompts for the file name.
	Alternate: Load button in the files program.
Examples	rtcmx
	rtcmx('redor.data')
See also	NMR Spectroscopy User Guide
Related	files Interactively handle files (C)

rtp Retrieve parameters (M)

Syntax	<pre>rtp<(file)></pre>
Description	Retrieves parameters from a file into the current experiment.
Arguments	file is the name of the file that, with the suffix .par added, contains
	the parameters to be retrieved;. The default is that the system prompts
	for the name (in that case, the name can be given without single

quotes). If file.par does not exist and file.fid does, rtp retrieves the parameters only from file.fid.		
rtp		
rtp('/vnmr/stdpar/P31')		
NMR Spectroscopy User Guide		
fixpar	Correct parameter characteristics in experiment (M)	
rt	Retrieve FIDs (M)	
rtv	Retrieve individual parameters (C)	
svp	Save parameters from current experiment (M)	
	<pre>quotes). If fi the paramete rtp rtp('/vnmr, NMR Spectro fixpar rt rtv svp</pre>	

rts Retrieve shim coil settings (C)

Syntax	<pre>rts(file)<:status></pre>		
Description	Locates a preexisting file of shim settings and copies the settings into the current parameter set of the current experiment and sets load='y' to facilitate subsequent loading of shims with su (or related commands or macros). If the shim file is not found, rts displays the file names it tried.		
	The rts command returns shims from a .fid file or a .par file, selecting the shim parameters from the parameters stored there.		
Arguments	file — the name of a file containing the shim coil settings to be retrieved. If the file name is an absolute path, rts uses it with no modifications. Otherwise, rts searches the applications directories.		
	status – the return variable with one of the following values after rts finishes searching for the shim coil settings file:		
	•0 indicates that rts failed to find requested file.		
	•1 indicates that rts found the requested file, either as an absolute path or in the shims directory of the first application directory.		
	•>=2 indicates that rts found the requested file in shims subdirectory of the second, third, or later application directory.		
Examples	rts('acetone') rts('bb10mm'):r1		
See also	NMR Spectroscopy User Guide		
Related	loadLoad status of displayed shims (P)suSubmit a setup experiment to acquisition (M)svsSave shim coil settings (C)		

rttmp Retrieve experiment data from experiment subfile (M)

Syntax rttmp(file)

Description	Retrieves experiment data-parameters, FID, and transformed spectrum-from the file specified in a subdirectory inside curexp+'/subexp'.		
Arguments	file is the name of the subfile from which to retrieve the experiment data.		
Examples	rttmp('H1') rttmp('cosy')		
See also	NMR Spectroscopy User Guide		
Related	cptmp	Copy experiment data into experiment subfile (M)	
	curexp	Current experiment directory (P)	
	svtmp	Move experiment data into experiment subfile (M)	

rtv Retrieve individual parameters (C)

Syntax	<pre>rtv<(file,par1<,index1<,par2,index2>>)><:val></pre>
	rtv('parmaster','noabort','parameter'):\$pm
Description	Retrieves one or more parameters from a parameter file. The file might have been made with svf or svp or sd commands, or it might be from another experiment. If no return argument is added, the parameters are copied into the experiment's current tree. If the parameter does not already exist in the current tree, it is created. If the returned parameter is an array, the entire array is returned.
	rtv returns values into the macro if a return argument is added. This form of rtv command, in which values are passed only to macro variables, avoids the creation of additional parameters in the experiment's current tree.
Arguments	file — name of the directory or a parameter file. If the supplied value for file is a directory (with or without the .fid or .par extension), the parameters are retrieved from the procpar file in that directory. If the supplied value does not correspond to a directory but rather is a parameter file, that file is used. The default is that rtv prompts for a file name. In that case, the file name can be given without single quotes.
	par1, index1, par2, index2, name and array index of one or more parameters to be retrieved. The default for each array index argument is the first index. Including the array index for a parameter is only useful when returning values to the macro through a return argument.
	val — return argument for values to return to the macro. If the requested parameter do not exist in the parameter file, rtv will abort.
	noabort - keyword option must follow the 'parmaster' keyword and precede the parameter argument. This option applies to a single parameter. Command does not abort if the requested parameter does not exist.

parmaster - filename of the parameter set.

806

parameter - the parameter name.

Executing rtv without macro return values causes the fixpar macro run. The macro fixpar is not executed if return values are requested. rtv will prompt for a file name if the command is executed without an argument. The filename given in response to the prompt does not require single quotes.

In LC-NMR, rt will retrieve the lcdata (and drunlog) files if these files were saved along with the NMR data by using svf.

Examples rtv

rtv('/vnmr/parlib/cosy.par','phase')
rtv('/vnmr/parlib/cosy.par','noabort','phase')

See also NMR Spectroscopy User Guide and User Programming manuals

Related	rt	Retrieve FIDs (M)
	rtp	Retrieve parameters (M)
	sd	Set first decoupler frequency to cursor position (M)
	svf	Save FIDs in current experiment (M)
	svp	Save parameters from current experiment (M)

rtx Retrieve parameters based on rtx rules (C)

Syntax	rtx(filename <,tree <, keyword1 <, keyword2 >>>)	
Description	The rtx command retrieves parameters from filename, based on the setting of the P_LOCK protection bit and using the rules below.	
Arguments	tree is 'current', 'processed', 'global', or 'systemglobal'.	
	keyword1 may be 'keep' or 'rt'. The default is 'keep'.	
	keyword2 may be 'clear' or 'noclear'. The default is 'clear'.	
	keyword2 determines if the P_LOCK bit is cleared after rtx is executed.	
	Truth table for rtx.	

Status of P_LOCK	Status of P_LOCK	keyword1	result
bit in current exp	bit in filename		
on	on	keep or rt	do not rt
on	off	keep or rt	do not rt
off	on	keep or rt	do rt
off	off	keep	do not rt
off	off	rt	do rt
<no parameter=""></no>	on	keep or rt	do rt
<no parameter=""></no>	off	keep	do not rt
<no parameter=""></no>	off	rt	do rt

See also NMR Spectroscopy User Guide

Related execpars rtp

Set up the exec parameters (M) Retrieve parameters (M)



Agilent VnmrJ 4 Command and Parameter Reference Guide

S	Save display parameters as a set (M)
s(n)	Save display parameters (C)
s2pul	Set up parameters for standard two-pulse sequence (M)
sa	Stop acquisition (C)
sample	Submit change sample, Autoshim experiment to acquisition (M)
samplechange	Automation utility
samplename	Sample name (P)
sampling	Parameter to control elliptical k-space sampling
save	Save data (M)
savefdfspec	Save 1D or arrayed 1D spectra as fdf file (C)
savefid	Save fid
savefile	Base file name for saving files (P)
saveglobal	Save selected parameters from global tree (P)
savemodule	Save a module
savesampglobal	Saves Sample Global Parameters
sb	Sinebell constant in directly detected dimension (P)
sb1	Sinebell constant in 1st indirectly detected dimension (P)
sb2	Sinebell constant in 2nd indirectly detected dimension (P)
sbs	Sinebell shift in directly detected dimension (P)
sbs1	Sinebell shift in 1st indirectly detected dimension (P)
sbs2	Sinebell shift in 2nd indirectly detected dimension (P)
SC	Start of chart (P)
sc2	Start of chart in second direction (P)
scalelimits	Set limits for scales in regression (M)



scalesw	Set scaling factor for multipulse experiments (M)
scalesw	Scale spectral width in directly detected dimension (P)
scalesw1	Set f ₁ scaling factor for 2D multipulse experiments (M)
scalesw1	Scale spectral width in 1st indirectly detected dimension (P)
scalesw2	Scale spectral width in 2nd indirectly detected dimension (P)
schedulerhelp	Proshim Maintenance Scheduler help(C)
sd	Set first decoupler frequency to cursor position (M)
sd2	Set second decoupler frequency to cursor position (M)
sd3	Set third decoupler frequency to cursor position (M)
sda	Set first decoupler frequency array (M)
sd2a	Set second decoupler frequency array (M)
sd3a	Set third decoupler frequency array (M)
sdp	Show diffusion projection (M)
sel1d	Apptype macro for Selective 1D experiments
select	Select spectrum, FID, trace, or 2D plane without display (C)
selex	Defines excitation band (M)
selexcit	Set up PFG selective excitation pulse sequence (M)
selexHT	Set up a selective Hadamard experiment (M)
send2vnmr	Send a command to VnmrJ (U)
seqfil	Pulse sequence name (P)
seqgen	Initiate compilation of user's pulse sequence (M,U)
seqgenupdate	Update compilation of user's pulse sequence
serverport	Returns the VnmrJ network listening port value (C)
set2D	General setup for 2D experiments (M)
set2d	General setup for 2D experiments (M)
set3dproc	Set 3D processing (C)
setallshims	Set all shims into hardware (M)
setcolor	Set colors for graphics window and for plotters (C)
setdecpars	Set decoupler parameter values from probe file (M)
setDECpars	Sets Decoupler Parameters
setdec2pars	Set decoupler 2 parameter values from probe file (M)

setdgroup	Set the Dgroup of a parameter in a tree (C)
setenumeral	Set values of a string parameter in a tree (C)
setether	Connect or reconnect host computer to Ethernet (U)
setexport	Set parameter bits for use with protocols (M)
setfrq	Set frequency of rf channels (C)
setgauss	Set a Gaussian fraction for lineshape (M)
setgcal	Set the gradient calibration constant (M)
setgcoil	Assign sysgcoil configuration parameter (M)
setgrid	Divide graphics window into rows and columns (C)
setgroup	Set group of a parameter in a tree (C)
sethtfrq1	Set a Hadamard frequency list from a line list ((M)
sethw	Set values for hardware in acquisition system (C)
sethwshim	Set values for hardware in acquisition system (C)
setint	Set value of an integral (M)
setlimit	Set limits of a parameter in a tree (C)
setlk	Set up lock parameters (M)
setlockfreq	Set lock frequency (M)
setLP	Set up linear prediction in the direct dimension (M)
setLP1	Set F1 linear prediction parameters (M)
setlp0	Set parameters for zero linear phase (M)
setnoether	Disconnect host computer from Ethernet (U)
setobspars	Sets Observe Parameters
setoffset	Calculate offset frequency for given nucleus and ppm (M)
setparams	Write parameter to current probe file (M)
setpen	Set maximum number of HP plotter pens (M)
setplotdev	Return characteristics of a named plotter (C)
setpower	Set power and pulsewidth for a given $\gamma B1$ value (M)
setprotect	Set the protection bits of a variable in a tree (C)
setpw180ad	Creates and sets observe adiabatic pulse shapes (M)
setpwx180ad	Creates and sets decoupler adiabatic pulse shapes (M)
setrc	Set receiver constants (M)
setref	Set frequency referencing (M)

setref1	Set freq. referencing for 1st indirectly detected dimension (M)
setref2	Set freq. referencing for 2nd indirect detected dimension (M)
setscout	Set up a scout run (M)
setssfilter	Set sslsfrq to the frequencies of each suppressed solvents (M)
setsw	Set spectral width (M)
setsw1	Set spectral width in evolution dimension (M)
setsw2	Set spectral width in 2nd evolution dimension (M)
setselfrqc	Set selective frequency and width (M)
setselinv	Set up selective inversion (M)
settcldefault	Select default display templates for pulse sequence (M)
settune	Opens the Auto Tune Setup dialog (M)
settype	Set the type of a parameter (C)
setup	Set up parameters for basic experiments (M)
setup_dosy	Set up gradient levels for DOSY experiments (M)
setuserpsg	Creates/initializes user PSG directory
setvalue	Set value of any parameter in a tree (C)
setwave	Write a wave definition string into Pbox.inp file (M)
setwell	Adjust the label of the "t1" axis for VAST contour maps (M)
setwin	Activate selected window (C)
sf	Start of FID (P)
sf1	Start of interferogram in 1st indirectly detected dimension (P)
sf2	Start of interferogram in 2nd indirectly detected dimension (P)
sfrq	Transmitter frequency of observe nucleus (P)
sh2pul	Set up for a shaped observe excitation sequence (M)
shdec	Set up for shaped observe excitation sequence (M)
shell	Start a UNIX shell (C)
shelli	Start an interactive UNIX shell (C)
shim	Submit an Autoshim experiment to acquisition (C)
shimmult	Multiple the shim dacs of the current shimset

shimnames	Returns shim names
shimset	Type of shim set (P)
showconfig	Show system configuration settings (M)
showconsole	Show console configuration parameters (U)
showdosy	Show DOSY Plot (M)
showdosyfit	Plots the experimental signal attenuation, fitted attenuation and residual for one peak from a 2D or 3D DOSY experiment (M)
showdosyresidu al	Plots the residual for one peak from a 2D or 3D DOSY experiment
showfit	Display numerical results of deconvolution (M)
showgradfit	Plots the experimental gradient variation with position and the power series fit in non-uniform gradient calibration (M)
showloginbox	Shows operator login dialog (M)
shownugfit	Plots the logarithm of the calculated diffusional attenuation and of the power series fit in non-uniform gradient calibration (M)
shownumx	x position counting from bottom left of every spectrum (P)
shownumy	y position counting from bottom left of every spectrum (P)
showoriginal	Restore first 2D spectrum in 3D DOSY experiment (M)
showplotter	Show list of currently defined plotters and printers (M)
showplotq	Display plot jobs in plot queue (M)
showprintq	Display print jobs in print queue (M)
showprotunegui	Show the graphical interface while tuning (P)
showrfmon	Show RF Monitor Button in Hardware Bar (P)
showsampglobal	Shows sample global parameters
showstat	Display information about status of acquisition (M,U)
sim	Sample in magnet (For systems equipped with a robot)
sin	Find sine value of an angle (C)
sine	Find values for a sine window function (M)
sinebell	Select default parameters for sinebell weighting (M)
sinesq	Find values for a sine-squared window function (M)

size	Returns the number of elements in an arrayed parameter (O)
slfreq	Measured line frequencies (P)
slw	Spin simulation linewidth (P)
smaxf	Maximum frequency of any transition (P)
sminf	Minimum frequency of any transition (P)
smsport	Sample Management System serial port connection (P)
sn	Signal-to-noise ratio (P)
solppm	Return ppm and peak width of solvent resonances (M)
solvent	Lock solvent (P)
solvinfo	Retrieve information from solvent table (C)
sort	Sort real values of a parameter (M)
sp	Start of plot in directly detected dimension (P)
spl	Start of plot in 1st indirectly detected dimension (P)
sp2	Start of plot in 2nd indirectly detected dimension (P)
spadd	Add current spectrum to add/subtract experiment (C)
spcfrq	Display frequencies of rf channels (M)
specdc3d	3D spectral drift correction (P)
spin	Submit a spin setup experiment to acquisition (C)
spin	Sample spin rate (P)
spinll	Set up a slfreq array (M)
spinner	Open the Spinner Control window (C)
spins	Perform spin simulation calculation (C)
split	Split difference between two cursors (M)
spintype	Spinner Type ((P)
splmodprepare	Used by the dosy macro to prepare data for the program SPLMOD (C)
splmodread	Used by the dosy macro to convert the output of the SPLMOD program into a form suitable for ddif (C)
spmax	Take the maximum of two spectra (C)
spmin	Take minimum of two spectra in add/subtract experiment (C)
spsm	Enter spin system (M)

spsub	Subtract current spectrum from add/subtract experiment (C)
sqcosine	Set up unshifted cosine-squared window function (M)
sqdir	Study queue directory (P)
sqend	End a study queue (M)
sqexp	Load experiment from protocol (M)
sqfilemenu	Study queue file menu commands (M)
sqLog	Records specific events from a study queue
sqmode	Study queue mode (P)
sqname	Study queue parameter template (P)
sqpars	Create study queue parameters for imaging (M)
sqprotocol	Macro to create protocols (M)
sqreset	Reset study queue parameters for imaging (M)
sqrt	Return square root of a real number (0)
sqsavestudy	Macro to save study parameters for imaging (M)
sqsinebell	Set up unshifted sinebell-squared window function (M)
srate	Spinning rate for magic angle spinning (P)
sread	Read converted data into VnmrJ (C)
srof2	Calculate exact rof2 value for Cold Probes (M)
SS	Steady-state transients (P)
ssecho	Set up solid-state echo pulse sequence (M)
ssecho1	Set up parameters for SSECH01 pulse sequence (M)
ssfilter	Full bandwidth of digital filter to yield a filtered FID (P)
sslsfrq	Center of solvent-suppressed region of spectrum (P)
ssntaps	Number of coefficients in digital filter (P)
ssorder	Order of polynomial to fit digitally filtered FID (P)
stack	Stacking mode for processing and plotting arrayed spectra (M)
stackmode	Stacking control for processing arrayed 1D spectra (P)
startq	Start a chained study queue (M)
status	Display status of sample changer (C,U)
stdld	Apptype macro for Standard 1D experiments (M)
stdshm	Interactively create a method string for autoshimming (M)

sth	Minimum intensity threshold (P)
string	Create a string variable (C)
string2array	Formats a String Variable into an Array
strstr	Find position of one string in another
strsv2array	Formats a String Separated Variable into an Array
strtext	Starting point for LP data extension in np dimension (P)
strtext1	Starting point for LP data extension in ni dimension (P)
strtext2	Starting point for LP data extension in ni2 dimension (P)
strtlp	Starting point for LP calculation in np dimension (P)
strtlp1	Starting point for LP calculation in ni dimension (P)
strtlp2	Starting point for LP calculation in ni2 dimension (P)
studyid	Study identification (P)
studypar	Study parameters (P)
studystatus	Study status (P)
studytime	Determine start and end times for studies (P)
su	Submit a setup experiment to acquisition (M)
sub	Subtract current FID from add/subtract experiment (C)
substr	Select a substring from a string (C)
suselfrq	Select peak, continue selective excitation experiment (M)
svdat	Save data (C)
svf	Save FIDs in current experiment (M)
svfdf	Save FID data in FDF format (M)
svfdir	Directory for non-study data (P)
svfj	Save FID in JCAMP-DX format (M)
svfname	Filename parameter template for non-study data ((P)
svfname	Create path for data storage (C)
svimg	Generate and Save images as FDF files (macro)
svllj	Save peak listing in JCAMP-DX X,Y or X,Y,M format (M)
svlsj	Save large dynamic range spectrum in JCAMP-DX format (M)
svp	Save parameters from current experiment (M)
svpdp	Compare workspace parameters to parameter file.

svr	Save secured REC data for VnmrJ SE
SVS	Save shim coil settings (C)
svs	Spin simulation vertical scale (P)
svsis	Generate and Save images as FDF files (macro)
svsj	Save spectrum in JCAMP-DX format (M)
svtmp	Move experiment data into experiment subfile (M)
svxyj	Save spectrum in JCAMP-DX X,Y format (M)
SW	Spectral width in directly detected dimension (P)
swl	Spectral width in 1st indirectly detected dimension (P)
sw2	Spectral width in 2nd indirectly detected dimension (P)
sw3	Spectral width in 3rd indirectly detected dimension (P)
sysgcoil	System gradient coil (P)
system	System type (P)
systemdir	VnmrJ system directory (P)

Save display parameters as a set (M)

Syntax	<pre>(1) sset_number (2) s(set_number)</pre>	
Description	Saves a copy of the current values of all display parameters. The set is data-independent because the parameters that govern a display (sp, wp, vs, etc.) are saved but no data is saved.	
Arguments	set_number is number of the display parameter set to be saved.	
Examples	s2 s(3)	
See also	NMR Spectroscopy User Guide	
Related	frFull recall of display parameter set (M)rRecall display parameter set (M)	

s (n) Save display parameters (C)

Applicability	All
Syntax	s(n<,noupdate>)
Description	Saves a copy of the current values of all display parameters as display
	parameter set n in the current experiment

S

noupdate as second argument prevents the automatic update of interactive programs.

Arguments n=1 to 9

- Related fr(n) Recall all the parameters of the specified display parameter set (C) $% \left(C\right) =0$
 - r (n) Recalls limited number of display parameters)

s2pul Set up parameters for standard two-pulse sequence (M)

DescriptionConverts the current experiment to an experiment suitable for the
standard two-pulse sequence (S2PUL).See alsoNMR Spectroscopy User Guide

sa Stop acquisition (C)

Syntax sa<(option | number) >

Description Stops an experiment that has been submitted to acquisition. If experiment is active, it is stopped. Data is retained. sa applies to the experiment that you are joined to at the time the sa command is entered. Thus, if experiment 1 is active, you must be joined to experiment 1 for sa to stop that acquisition. If you are in experiment 2, entering sa has no effect on experiment 1.

> When experiments are queued, the behavior of sa is more complex. If an experiment is active in exp1 and queued in exp2, entering sa from exp1 stops that experiment and immediately begins acquisition on exp2. Entering sa from exp2, on the other hand, removes exp2 from the queue, without affecting the active experiment 1.

> Entering sa from an experiment that is not active or queued has no effect.

Arguments option is one of the following:

- 'eos', 'ct', 'scan' are keywords to stop at the next ct.
- 'eob', 'bs' are keywords to stop at the next block size.
- 'eof', 'nt', 'fid' are keywords to stop at the next complete FID.
- 'eoc', 'il' are keywords to stop at next complete il cycle (i.e., the latest block size that has been completed for all FIDs in interleave cycle.

number is an integer number to stop at the next ct, where the value of ct is a multiple of number. This is useful when you want to complete a phasecycle before stopping.

Examples

```
sa('ct')
sa(4)
```

sa

Related	bs	Block size (P)
	ct	Completed transients (P)
	il	Interleave arrayed and 2D experiments (P)
	nt	Number of transients (P)
	ra	Resume acquisition stopped with sa command (C)

sample Submit change sample, Autoshim experiment to acquisition (M)

Applicability	Systems with a sample changer.	
Description	Performs the combined operations change, spin, lock, and shim, making it a convenient setup command for a new sample.	
See also	NMR Spectroscopy User Guide	
Related	au	Submit experiment to acquisition and process data (C)
	change	Submit a change sample experiment to acquisition (M)
	ga	Submit experiment to acquisition and FT the result (C)
	go	Submit experiment to acquisition (C)
	lock	Submit an Autolock experiment to acquisition (C)
	shim	Submit an Autoshim experiment to acquisition (C)
	spin	Submit a spin setup experiment to acquisition (C)
	su	Submit a setup experiment to acquisition (M)

sample ChangeAutomation utility

ApplicabilityVnmrJ 3.1DescriptionThis is a utility macro to remove the sample from the magnet after an
automation queue finishes. It is only available with systems with the
7600-AS or 7510-AS robot systems. The choice to either put a
reference sample into the magnet, leave the current sample in the
magnet, or remove the current sample from the magnet, is made from
the Preferences pop-up window.

samplename Sample name (P)

Description	Specifies the name	of the sample. It is saved with a liquids study,
See also	NMR Spectroscopy	User Guide, VnmrJ Walkup
Related	cqsavestudy	Macro to save study queue parameters (M)
	notebook	Notebook name (P)

pageName of page (P)studyparStudy parameters (P)

sampling Parameter to control elliptical k-space sampling

Syntax sampling='e' selects the elliptical sampling schedule for data acquisition.

Description A 3D data set can be viewed as a rectangle where the lengths of the sides are set by the ni and ni2 parameters. If an ellipse is inscribed inside this ni x ni2 rectangle, the elliptical sampling schedule selects those traces inside the ellipse. The data in traces outside the ellipse will be set to zero.

If the sampling parameter does not exist or is not set to 'e', the standard sampling schedule of acquiring every point within the rectangle will be used.

If sampling='e', four additional parameters control the size and position of the ellipse.

These parameters are:

samplingEScale - multiplier for the size of the ellipse. Default is 1.01.

samplingEAngle - rotation angle of the ellipse, in degrees. Default is 0.

samplingETransX - translation of the ellipse in the "X" direction. Default is 0.0.

<code>samplingETransY</code> — translation of the ellipse in the "Y" direction. Default is 0.0.

save Save data (M)

- Description Macro to save data. In a study, it uses sqdir and autoname to construct the data filename. If not in a study, it uses svfdir and svfname to construct the data filename.
 - See also NMR Spectroscopy User Guide, VnmrJ Walkup

acquire	Acquire data (M)
autoname	Create path for data storage (C)
autoname	Prefix for automation data file (P)
sqdir	Study queue directory (P)
svfdir	Directory for non-study data (P)
Svfname	Create path for data storage (C)
svfname	Filename parameter template for non-study data ((P) $% \left(P\right) =\left(P\right) \left(P\right) \left$
	acquire autoname sqdir svfdir Svfname svfname

savefdfspec Save 1D or arrayed 1D spectra as fdf file (C)

Syntax savefdfspec<(fullpath)>

Description This command saves 1D or arrayed 1D spectra in vnmrbg phasefile buffer to a fdf file. All traces of arrayed 1D data are saved to a single fdf file. Default path is curexp/datdir/spec.fdf.

savefdfspec is implemented to save CSI spectral data, which is an arrayed 1D data. CSI spectral data is generated by 1D FT of spatially reconstructed CSI data. Zerofilling and/or spatial cropping may be applied during spatial reconstruction, resulting in a final array size of fnv*fnv2*fnv3. It is required that fid (after spatial reconstruction) is in uncompressed format (one trace per block). If fid file is in compressed format, "ft" or "wft" only transforms nf traces. In this case, only nf traces will be saved.

fdf spectral data is stored in the order of $\{np, fnv, fnv2\}$ or $\{np, fnv, fnv2, fnv3\}$, with np being the inner loop and fnv2 or fnv3 the outer loop.

fdf header contains the following required fields (values are examples):

```
#!/usr/local/fdf/startup
float rank = 3;
char *storage = "float";
float bits = 32;
int bigendian = 0;
float matrix[] = {512,64,64};
float spec_matrix[] = {512,128};
float spec_data_rank = 2;
float spec_display_rank = 1;
```

The header may contain optional fields (not required for loading data), such as

```
char *type = "phased";
char *apptype = "im2Dcsi";
char *sequence = "csi2d";
char *studyid = "s_2012011801";
char *fidpath =
"/home/imaging/vnmrsys/exp4/test.csi/spatial.fid";
float location[] = {0.043000,0.933000,0.142000};
float roi[] = {6.000000,7.000000,0.000000};
int slices = 1;
float gap = 0.000000;
float thk = 0.400000;
```

S

```
float psi = 90.000000;
float phi = 0.000000;
float theta = 90.000000;
float orientation[] =
\{-0.000001, -1.000000, -0.000000, 0.000001, -0.000000, 1.000
000,-1.000000,0.000001,0.000001};
char *position1 = "";
char *position2 = "";
char *dataType = "spectrum";
char *nucleus[] = {"H1"};
float sfreq[] = {499.721973};
float sw[] = \{4006.410256\};
float upfield[] = {-580.622558};
float wp[] = {3998.585236};
float sp[] = \{-572.797538\};
float rp[] = {-51.335980};
float lp[] = {-21.599991};
```

savefid Save fid

Description	This utility saves the data in the current workspace according to the templates in the Preferences/Templates panel.
Syntax	savefid
Related	svf

savefile Base file name for saving files (P)

Applicability	Systems with LC-NMR accessory.		
Description	Contains the base file name using the format savefile.001,		
	savefile.002, etc., to which a series of FIDs or data sets are saved.		
	If savefile does not exist, the parlc macro can create it.		
See also	NMR Spectroscopy User Guide		
Related	parlc Create LC-NMR parameters (M)		

saveglobal Save selected parameters from global tree (P)

Description Saves an array of parameter names from the global or systemglobal tree. Whenever go is executed, the parameters listed are saved in the

	current tree	with an underscore (_) appended. These parameters are	
	copied back into the global tree (without the underscore) whenever		
	processing by wbs, wnt, wexp, or werr occurs.		
See also	NMR Spectro	scopy User Guide	
Related	go	Submit experiment to acquisition (C)	
	loc	Location of sample in tray (P)	

savemodule Save module

Syntax	<pre>savemodule('modulename'<,dirpath<,tree<,parameter>>>)</pre>
	arg1 - modulename
	<pre>arg2 - (optional) pathname where the module should be saved default is studydir/dirinfo/modules if arg2 is an empty string, it is set to default arg2='cp' is a keyword for curexp (auto='n') or autodir (auto='y')</pre>
	arg3 - (optional) (save from which) tree (default is current)
	arg4 - (optional) specific parameter

savesampglobalSaves Sample Global Parameters

Description	Updates sample global parameters in the study directory from th current workspace.		
See also	savesampglobal		
Related	getsampglobal, resetsampglobal, savesampglobal, mvsampglobal, showsampglobal		

sb Sinebell constant in directly detected dimension (P)

Description Applies a sinebell constant along the directly detected dimension. This dimension is often referred to as the f₂ dimension in 2D data sets, the f₃ dimension in 3D data sets, etc. Values A positive value applies a sinebell of the form $sin\left(\frac{t \cdot \pi}{2 \cdot sb}\right)$ A negative value applies a squared sinebell function of form $sin^2\left(\frac{t \cdot \pi}{2 \cdot sb}\right)$

sb is given in seconds. Typical value is sb='n'.

See also	NMR Spectr	oscopy User Guide
Related	sb1	Sinebell constant in 1st indirectly detected dimension
		(P
	sb2	Sinebell constant in 2nd indirectly detected dimension
		(P)
	sbs	Sinebell shift constant in directly detected dimension
		(P)
	sine	Find values for a sine window function (M)
	sinebell	Select default parameters for sinebell weighting (M)
	sinesq	Find values for a sine squared window function (M)

sb1 Sinebell constant in 1st indirectly detected dimension (P)

Description	Applies a sinebell constant along the first indirectly detected
	dimension. This dimension is often referred to as the f_1 dimension in
	multidimensional data sets. sb1 works analogously to the parameter
	sb. The "conventional" parameters, such as 1b and gf, operate on the
	detected FIDs, while this "2D" parameter is used during processing of
	the interferograms.
Values	A positive value applies a sinebell of the form $\sin\left(\frac{t \cdot \pi}{2 - \tau t}\right)$

A negative value applies a squared sinebell function of form $\sin^2\left(\frac{t \cdot \pi}{2 \cdot sbl}\right)$ sb1 is given in seconds. Typical value is sb1='n'.

See also NMR Spectroscopy User Guide

sb2 Sinebell constant in 2nd indirectly detected dimension (P)

Description Applies a sinebell constant along the second indirectly detected dimension. This dimension is often referred to as the f_2 dimension in multidimensional data sets. sb2 works analogously to the parameter sb. The value of sb2 can be set with wti on the 2D interferogram data.

Values A positive value applies a sinebell of the form $\sin\left(\frac{t \cdot \pi}{2 \cdot sb^2}\right)$ A negative value applies a squared sinebell function of form $\sin^2\left(\frac{t \cdot \pi}{2 \cdot sb^2}\right)$

sb2 is given in seconds. Typical value is sb2='n'.

- See also NMR Spectroscopy User Guide
 - Related sb Sinebell constant in directly detected dimension (P)

sb1	Sinebell constant in 1st indirectly detected dimension
	(P)
wti	Interactive weighting (C)

sbs Sinebell shift in directly detected dimension (P)

Description	Working in combination with the parameter sb, sbs allows shifting the origin of the sinebell function along the directly detected dimension. This dimension is often referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, etc.		
varues	The square of this function is applied if sb is negative. sbs is given in seconds. The typical value is $sbs='n'$		
See also	NMR Spectroscopy User Guide		
Related	sb sbs1	Sinebell constant in directly detected dimension (P) Sinebell shift in 1st indirectly detected dimension (P)	
	sbs2	Sinebell shift in 2nd indirectly detected dimension (P)	
	sine	Find values for a sine window function (M)	
	sinesq	Find values for a sine squared window function (M)	

sbs1 Sinebell shift in 1st indirectly detected dimension (P)

- Description Working in combination with the parameter sb1, sbs1 allows shifting the origin of the sinebell function along the first indirectly detected dimension. This dimension is often referred to as the f₁ dimension in multidimensional data sets. sbs1 works analogously to parameter sbs. The "conventional" parameters, such as 1b and gf, operate on the detected FIDs, while this "2D" parameter is used during processing of the interferograms. Values The origin is shifted according to the form $sin\left(\frac{(t-sbs1)\cdot\pi}{t}\right)$
 - Values The origin is shifted according to the form $\sin\left(\frac{(t-sbs1)\cdot \pi}{2\cdot sb1}\right)$ The square of this function is applied if sb1 is negative. sbs1 is given in seconds. The typical value is sbs1='n'. See also NMR Spectroscopy User Guide
 - Related sb1Sinebell constant in 1st indirectly detected dimension (P)sbsSinebell shift constant in directly detected dimension (P)sb2Sinebell constant in 2nd indirectly detected dimension (P)

sbs2 Sinebell shift in 2nd indirectly detected dimension (P)

Description Working in combination with the parameter sb2, sbs2 allows shifting the origin of the sinebell function along the second indirectly detected dimension. This dimension is often referred to as the f₂ dimension in multidimensional data sets. sbs2 works analogously to parameter sbs. sbs2 can be set with wti on the 2D interferogram data. The origin is shifted according to the formula $\sin\left(\frac{(t-sbs2)\cdot\pi}{2sbc2}\right)$ Values $2 \cdot sb2$ The square of this function is applied if sb2 is negative. sbs2 is given in seconds. The typical value is sbs2='n'. NMR Spectroscopy User Guide See also Related sbs Sinebell shift constant in directly detected dimension (P) Sinebell constant in 2nd indirectly detected dimension (P) wti Interactive weighting (C)

sc Start of chart (P)

Description	Positions of the start of the plotting position (the "chart") with respect to the right edge of the plotter.		
Values	0 to wemax, in mm		
See also	NMR Spectroscopy User Guide		
Related	sc2	Start of chart in second direction (P)	
	WC	Width of chart (P)	
	wcmax	Maximum width of chart (P)	

sc2 Start of chart in second direction (P)

Description	Controls the a 2D contour	start of plotting position of the second axis (or y axis) of plot. The parameter wc2 controls the width of the chart.	
Values	0 to wc2max, in mm.		
See also	NMR Spectroscopy User Guide		
Related	sc wc2	Start of chart (P Width of chart in second direction (P) Maximum width of chart in second direction (P)	
	WCZIIICA	maximum which of chart in second uncerton (1)	

scalelimits Set limits for scales in regression (M)

Syntax scalelimits(x_start, x_end, y_start, y_end)

Description	Causes the command $expl$, which is used by regression to display data, to use typed-in scale limits. The limits are retained as long as an $expl$ display is retained.		
Arguments	x_start,x_e ending limits	end,y_start,y_end are x -axis and y -axis starting and . The default is that scalelimits prompts for the limits.	
See also	NMR Spectro	scopy User Guide, User Programming	
Related	autoscale	Resume autoscaling after limits set by scalelimits (M)	
	expl	Display exponential or polynomial curves (C)	

scalesw Set scaling factor for multipulse experiments (M)

Description	Sets the spectral width scaling factor for the multipulse sequences se up by macros br24 and mrev8. The value of the scaling factor is stored in the parameter scalesw.		
See also	User Guide	: solid-State NMR	
Related	br24	Set up BR24 multiple pulse experiment (M)	
	mrev8	Set up MREV8 multiple pulse experiment (M)	

	1	-	1	1		
scalesw	Scale spectral	width in	directly	detected	dimension (I	P)

scalesw1 Set f_1 scaling factor for 2D multipulse experiments (M)

scalesw Scale spectral width in directly detected dimension (P)

Description	Adjusts the frequency scale dimension used with the parameter sets in
	the sequences set up by the br24, mrev8, ssecho, and xpolar1
	macros. If scalesw is active, the labels for the frequency scales
	includes the letters sc in parentheses. A scaled frequency can be
	referenced using the 11 macro.
Values	'n'. number greater than 0.0

See also User Guide: Solid-State NMR

Related	br24	Set up BR24 multiple pulse experiment (M)
	mrev8	Set up MREV8 multiple pulse experiment (M)
	rl	Set reference line (M)
	scalesw	Set scaling factor for multipulse experiments (M)
	scalesw1	Scale spectral width in 1st indirectly detected
		dimension (P)
	scalesw2	Scale spectral width in 2nd indirectly detected
		dimension (P)
	ssecho	Set up solid-state echo pulse sequence (M)
	xpolar1	Set up parameters for XPOLAR1 pulse sequence
		(M)

Description	Sets the f_1 spectral width scaling factor for the multipulse sequences set up by the br24 and mrev8 macros. The value of the scaling factor is stored in the parameter scalesw1.	
See also	User Guide:	Solid-State NMR
Related	br24 mrev8 scalesw1	Set up BR-24 multiple pulse experiment (M) Set up MREV8 multiple pulse experiment (M) Scale spectral width in 1st indirectly detected dimension (P)

scalesw1 Scale spectral width in 1st indirectly detected dimension (P)

Description Analogous to the scalesw parameter except that scalesw1 applies to first indirectly detected dimension of a multidimensional data set. A scaled frequency along this dimension can be referenced using the rl1 macro.

Values 'n', number greater than 0.0

- See also User Guide: Solid-State NMR

scalesw2 Scale spectral width in 2nd indirectly detected dimension (P)

Description	Analogous to the scalesw parameter except scalesw2 applies to second indirectly detected dimension of a multidimensional data set. A scaled frequency along this dimension can be referenced using the rl2 macro.		
Values	'n', number greater than 0.0		
See also	User Guide: Solid-State NMR		
Related	r12	Set reference line in 2nd indirectly detected dimension (M)	
	scalesw	Set scaling factor for multipulse experiments (M)	
	scalesw1	Set f ₁ scaling factor for 2D multipulse experiments	

(M)

schedulerhelp Proshim Maintenance Scheduler help(C)

Applicability VnmrJ 3.2 Description Brings up help for the Proshim Maintenance Scheduler.

sd Set first decoupler frequency to cursor position (M)

Description Sets the first decoupler frequency offset parameter dof to place the first decoupler at the cursor position in the spectrum. This works only if the transmitter nucleus and first decoupler nucleus are the same (tn=dn).

See also	NMR Spectro	scopy User Guide
Related	dof	Frequency offset for first decoupler (P)
	dn	Nucleus of first decoupler (P)
	sd2	Set second decoupler frequency to cursor position
		(M)
	sd3	Set third decoupler frequency to cursor position
		(M)
	sda	Set first decoupler frequency array (M)
	tn	Nucleus for observe transmitter (P)

sd2 Set second decoupler frequency to cursor position (M)

Applicability	Systems with a second decoupler.		
Description	Sets the second decouple frequency offset parameter $dof2$ to place the second decoupler at the cursor position in the spectrum. This works only if the transmitter nucleus and second decoupler nucleus are the same (tn=dn2).		
See also	NMR Spectroscopy User Guide		
Related	dn2	Nucleus for second decoupler (P)	
	dof2	Frequency offset for second decoupler (P)	
	sd	Set first decoupler frequency to cursor position (M)	
	sd2a	Set second decoupler frequency array (M)	
	tn	Nucleus for observe transmitter (P)	

sd2a Set second decoupler frequency array (M)

ApplicabilityVnmrJ 3.1DescriptionWith the cursor set to some position in the spectrum, "sd2" sets the
decoupler offset parameter "dof2" to place the second decoupler at

that position in the spectrum. To set up an array of offset values for the second decoupler, use "sd2" for the first position and "sd2a" for all subsequent positions. Either command will only work if the parameter "tn" is the same as the parameter "dn2".

sd3 Set third decoupler frequency to cursor position (M)

Applicability Systems with a third decoupler.

Description Sets the third decoupler frequency offset parameter dof3 to place the third decoupler at the cursor position in the spectrum. This works only if the transmitter nucleus and third decoupler nucleus are the same (tn=dn3).

See also	NMR Spectro	scopy User Guide
Related	dn3	Nucleus for third decoupler (P)
	dof3	Frequency offset for third decoupler (P)
	sd	Set first decoupler frequency to cursor position (M)
	sd3a	Set third decoupler frequency array (M)
	tn	Nucleus for observe transmitter (P)

sda Set first decoupler frequency array (M)

- Description Sets up an array of offset values for the first decoupler, using sd for the first decoupler position and sda for subsequent positions. This works only if the transmitter nucleus and first decoupler nucleus are the same (tn=dn).
 - See also NMR Spectroscopy User Guide

Related	dn	Nucleus for first decoupler (P)
	sd	Set first decoupler frequency to cursor position (M)
	sd2a	Set frequency array for second decoupler (M)
	sd3a	Set frequency array for third decoupler (M)
	tn	Nucleus for observe transmitter (P)

sd3a Set third decoupler frequency array (M)

ApplicabilitySystems with a third decoupler.DescriptionSets up an array of offset values for the third decoupler, using sd3 for
the first position and sd3a for subsequent positions. This works only
if the transmitter nucleus and third decoupler nucleus are the same
(tn=dn3).
See also	o NMR Spectroscopy User Guide	
Related	dn2	Nucleus for third decoupler (P)
	sd3	Set third decoupler frequency to cursor position
		(M)
	sda	Set first decoupler frequency array (M)
	tn	Nucleus for observe transmitter (P)

Show diffusion projection (M) sdp

- Displays projection onto diffusion axis using the dsp facility. Use with Description 2D or 3D DOSY data after DOSY analysis. The unit of the resulting axis is D (10^{-10} m²/sec). Because sdp overwrites the parameters in the current experiment, use it in only an experiment in which it is okay for existing data to be overwritten.
 - See also NMR Spectroscopy User Guide
 - Related dosy Process DOSY experiments (M)

sel1d Apptype macro for Selective 1D experiments (M)

Description	Perform the actions f plot experiments.	for Selective 1D protocols to set up, process, and
Examples	<pre>sel1d('setup') - sel1d('process') sel1d('plot') - end</pre>	execute sel1d experimental setup – execute sel1d processing xecute sel1d plotting
Related	apptype execpars	Application type (p) Set up the exec parameters (M)

Select spectrum, FID, trace, or 2D plane without display (C) select

Syntax	<pre>(1) select<('next' 'prev' selection)><:index> (2) select<(<'f1f3' 'f2f3' 'f1f2'><,'proj'></pre>
Description	Directs future actions to apply to a particular spectrum or FID in a 1D array, to a trace in 2D (syntax 1), or to a particular 2D plane from a 3D data set (syntax 2). If select is called with no arguments, it returns the current index. When VnmrJ is first booted up, select is in 1D mode. select enters the 2D mode if any of the keywords 'f1f3', 'f2f3', 'f1f2', or 'proj' are present in the argument list. Entering the ds and jexp commands set select back in the 1D mode.

Arguments For 1D operations (syntax 1):

- 'next' is keyword to increment by 1 the 1D spectrum or trace index.
- 'prev' is keyword to decrement by 1 the 1D spectrum or trace index.
- •selection is a number selecting a 1D spectrum, FID, or trace.
- index returns the number of the current 1D spectrum, FID, or trace. For selecting various 2D planes of a 3D data set (syntax 2):
- 'f1f3', 'f2f3', and 'f1f2' are types of 2D planes. The parameters plane and index2 serve to indicate the exact 2D plane that is currently viewable by VnmrJ. Note that index2 cannot be entered from the keyboard (i.e., you cannot select a new 2D plane by changing the value of index2); you must use the select command instead.
- 'proj' is keyword to use the 2D projection whose plane type is determined by the parameter plane.
- 'next' is keyword to increment the parameter index2 to its next value and sets up VnmrJ to be ready to display the 2D plane whose number is the new index2 value.
- 'prev' performs analogously except that index2 is decremented.
- •plane is a number selecting the plane.
- index returns the number of the current plane.

Examples select('next')
 select(2):r1
 select('f1f3')
See also NMR Spectroscopy User Guide, User Programming
Related arraydim Dimension of experiment (P)

Display a spectrum (C)
Projection or 3D plane index selected (P)
Join existing experiment (C)
Currently displayed 3D plane type (P)

selex Defines excitation band (M)

Syntax	<pre>selex<(sh<,pw<,st<,ph<,fla<,trev>>>>)></pre>
Description	Defines the excitation band from the position of cursors in the graphics window and reports them to user. It also sets r1 to excitation bandwidth and r2 to offset. selex is part of the Pbox software environment and uses the Pbox macros pbox_bw and putwave.
Arguments	sh is the name of a shape file.
	pw is the pulsewidth, in sec.
	st is the spin status: 0 for excitation, 0.5 for refocusing, or 1 for de-excitation.
	ph is the phase (or phase cycle, see wavelib/supercycles).
	fla is the flip angle.

	trev is the time reversal. This argument can be used to cancel time reversal introduced by setting the spin status (st) to 1 for de-excitation.
Examples	selex('esnob',0.0,1,90.0)
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

selexcit Set up PFG selective excitation pulse sequence (M)

Applicability	Systems with a pulsed field gradient module.
Description	Prepares an experiment for PFG (pulsed field gradient) selective
	excitation, with presaturation option.
See also	NMR Spectroscopy User Guide

selexHT Set up a selective Hadamard experiment (M)

Description	Sets up parameters for a selective shaped pulse Hadamard-encoded test experiment.	
See also	NMR Spectroscopy User Guide	
Related	htofs1	Hadamard offset in ni (P)
	fn1	Fourier number in 1st indirectly detected dimension (P)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	ft2d	Fourier transform 2D data (C)
	sethtfrq1	Set Hadamard frequency list from a line list (M)

send2vnmr Send a command to VnmrJ (U)

Syntax	send2Vnmr \$vnmruser/.talk command		
Description	Sends a command from UNIX to VnmrJ using the port number stored in the <pre>\$vnmruser/.talk</pre> file. This file is created when the macro listenon is entered on the VnmrJ command line.		
Arguments	command is any character string (commands, macros, or if statements) normally typed into the VnmrJ command line.		
Examples	send2Vnmr \$vnmruser/.talk dg		
See also	User Programming		
Related	Macro executed automatically when VnmrJ activated (Enable receipt of messages from send2Vnmr (M) Disable receipt of messages from send2Vnmr (M)	(M)	

S

seqfil Pulse sequence name (P)

Description Identifies the name of the pulse sequence to be used. The value of seqfil is displayed on the top line of the screen after the "Seq:" label. Macros used to set up new pulse sequences, such as Dept and Apt, automatically change the seqfil parameter.

See also NMR Spectroscopy User Guide

Related pslabel Pulse sequence label (P)

seggen Initiate compilation of user's pulse sequence (M,U)

Syntax (From VnmrJ) seqgen(<-static,>file<.c>)
 (From VnmrJ) seqgen(file<.c>)
 (From VnmrJ) seqgen
 (From VnmrJ) seqgen('file<.c> file2 file3 ...')
 (From UNIX) seqgen <-static> file<.c> <file1,...>

Description Begins compilation of a user pulse sequence. When used from VnmrJ, the macro seqgen calls the UNIX shellscript seqgen, which can also be called directly from UNIX, as shown above. The seqgen shellscript then calls the compilation makefile seqgenmake, located in the directory /vnmr/acqbin.

> The specified pulse sequence can be located in ~/vnmrsys/psglib or in /vnmr/psglib. If two files with the same name exist in these two directories, the local directory (~/vnmrsys/psglib) takes precedence. For sequences in /vnmr/psglib, seqgen first copies the file into the local directory ~/vnmrsys/psglib and then compiles it there; the resulting executable is then placed in ~/vnmrsys/seqlib. A copy of the pulse sequence is also copied into the seqlib directory along with the executable. As it is running, seqgen reports where it found the specified sequence(s).

> seqgen uses library files (object modules) found in /vnmr/lib. If setuserpsg and psggen has been run, the library files in the local directory ~/vnmrsys/psg take precedence of those in /vnmr/lib.

> Error messages are written into the file file.errors, where file is the name of the pulse sequence in psglib in which compilation is performed.

> Note that seqgen not only accepts file names with and without extensions, but also accepts files specified with wildcards and complex paths (seqgen strips the directory part, and seqgen /vnmr/psglib/apt will compile ~/vnmrsys/psglib/atp.c if it exists).

Arguments -static is a keyword for seqgen to use static rather than dynamic binding. Static binding results in larger executables in seqlib (several hundred Kbytes), but these sequences execute slightly faster (i.e., the go command). While insignificant generally, faster execution is helpful in some special applications such as the Scout Scan[™] mode of LC-NMR, where the time spent on the go command becomes critical. Static binding results in a fixed-size time gain, regardless of the number of increments; for large multidimensional experiments, the speed difference is not noticeable. file is the file name of a standard two-pulse sequence. .c is the extension on the file name. file1,file2,... are the names of files containing more sequences. Examples (From VnmrJ) seqgen('/vnmr/psglib/*.c') (From UNIX) seqgen /vnmr/psglib/*.c (From UNIX) seqgen apt dept noesy (From UNIX) seqgen -static lcld See also User Programming

seggenupdateUpdate compilation of user's pulse sequence

Applicability	VnmrJ 3.1		
Description	seqgenupdate has the same syntax as seqgen. Just like seqgen, one or more pulse sequence names can be supplied. seqgenupdate proceeds in two steps. In the first step, if any arguments are given, it passes them to seqgen for compilation. In the second step, it looks at the results of a preceeding seqgen. If permissions allow, it will move the compiled sequences back to the application directories or absolute		
	paths they were copied from.		
See also	VNMR User Programming, Chapter 2, "Pulse Sequence Programming".		
Related	psggen compile a user PSG object library (M.U)		

serverport Returns the VnmrJ network listening port value (C)

Applicability	VnmrJ
Syntax	serverport
Description	The serverport command returns the port number when VnmrJ opens a network port (socket) for other programs to send it network messages. See the write('net',) command for an example on how to use this port number.
Related	write Write formatted text to a device (C)

set2D General setup for 2D experiments (M)

Syntax set2D<(F2_dig_res<,F1_dig_res>)>

Description	Similar to set2d but does not execute par2d and does not make sw1, rfl1, and rfp1 decisions based on tn=dn condition.		
Arguments	s F2_dig_res is the f_2 digital resolution desired, in Hz/pt. Default i		
	F1_dig	<code>g_res</code> is the $\rm f_1$ digital resolution desired, in Hz/pt. Default is 12.	
Related	rfl1	Reference peak position in 1st indirectly detected dimension (P)	
	rfp1	Reference peak frequency in 1st indirectly detected dimension (P)	
	set2d	General setup for 2D experiments (M)	
	sw1	Spectral width in 1st indirectly detected dimension (P)	

set2d General setup for 2D experiments (M)

Syntax	<pre>set2d(experiment<,F2_dig_res<,F1_dig_res>>)</pre>	
Description	Runs the macro par2d to create new parameters needed for 2D experiments, then selects starting values for a number of parameters. The set2d macro is "internal" and not normally typed directly by the user.	
Arguments	experiment is the name of a 2D experiment (e.g., 'noesy').	
	<code>F2_dig_res</code> is the f_2 digital resolution desired, in Hz/pt.	
	<code>F1_dig_res</code> is the f_1 digital resolution desired, in Hz/pt.	
Examples	set2d('cosyps') set2d('hetcor',16) set2d('het2dj',16,(2*sw1)/fn1)	
See also	NMR Spectroscopy User Guide	
Related	par2d Create 2D acquisition parameters (M)	

set3dproc Set 3D processing (C)

Syntax set3dproc<(<'nocoef'><,directory>)>

Description Creates the file procdat that contains binary 3D information used by ft3d in processing the 3D FID data. It also creates the 3D parameter set procpar3d that is used by the select command to display the 2D planes from the 3D transformed data. set3dproc can only create the proper 3D coefficient file if the parameters phase and phase2 are used to generate States-Haberkorn (hypercomplex) or TPPI data along the t_1 and t_2 dimensions.

set3dproc creates the coefficient file for the following five values of array (where SH is States-Haberkorn):

- if array='' (null string), type of 3D data is $TPPI(t_1) TPPI(t_2)$
- if array='phase', type of 3D data is $SH(t_1) TPPI(t_2)$
- if array='phase2', type of 3D data is $SH(t_2) TPPI(t_1)$

• if array	='phase2,phase', type of 3D data is $SH(t_1)$ - $SH(t_2)$		
If <mark>array</mark> coefficier	is set to some other value, set3dproc cannot create the 3D at file and an error is reported within VnmrJ.		
'nocoef created.	' is a keyword that the 3D coefficient file $\verb"coef"$ is not to be		
directo The defa	ry is the name of the directory for procdat and procpar3d. ult is the subdirectory info in the directory curexp.		
set3dpr	set3dproc		
set3dpr	oc('nocoef','curexp/info3d')		
NMR Spe	NMR Spectroscopy User Guide		
array	Parameter order and precedence (P)		
ft3d	Perform a 3D Fourier transform (M,U)		
phase	Phase selection (P)		
phase2	Phase selection for 3D acquisition (P)		
select	Select a spectrum or 2D plane without displaying it (C)		
wftt3	Process f_3 dimension during 3D acquisition (M)		
	• if array If array coefficier 'nocoef created. directo The defa set3dpr set3dpr set3dpr <i>NMR Spe</i> array ft3d phase phase2 select wftt3		

setallshims Set all shims into hardware (M)

Description	Sets shims from the current parameter tree into hardware.		
	setallshims is equivalent to entering load='y'su but without		
	setting all the hardware parameters normally set by su (temperature,		
	decoupling, transmitter initialization, etc.). The shims used depend on		
	the shimset configuration. For the shim set on the Ultra•nmr shim		
	system, setallshims is active only if hardware-to-software shim		
	communication is enabled.		
See also	NMR Spectroscopy User Guide		
Related	load Load status of displayed shims (P)		

readallshims	Read all shims from hardware (M)
readhw	Read current values of acquisition hardware (C)
sethw	Set values for hardware in acquisition system (C)
shimset	Type of shim set (P)
su	Submit a setup experiment to acquisition (M)

setcolor Set colors for graphics window and for plotters (C)

Syntax (1) setcolor('pcl',item_index,'color')
(2) setcolor('hpgl',item_index,'color')
(3) setcolor('pen',pen_number,'color')
(4) setcolor('graphics',item_index,red,green,blue)
(5) setcolor('ps',item_index,red,green,blue)
(6) setcolor('plotter',black_plane,color_planes)

Description Sets colors used on the graphics window and on plotters. This command is a utility program used by the color macro and other macros. It is not expected that setcolor would be entered directly from the input window.

Arguments 'pcl' is a keyword to set colors on a plotter device that uses the PCL language. PCL plotters are the laser type of plotter.

'hpgl' is a keyword to set colors on a plotter device that uses the HPGL language. HPGL plotters are the pen type of plotter.

'pen' is a keyword that next two arguments set the color for a physical pen on a plotter device that uses the HPGL language.

'graphics' is a keyword to set colors on the graphics window.

'ps' is a keyword to set colors on a plotter using the PostScript language.

red, green, blue are three integers between 0 and 255 that set the amount of red, green, and blue color on the graphics window or PostScript plotter.

'plotter' is a keyword that the next two arguments set the black mode and number of colors available for a plotter device.

item_index is an index number from the following list that represents a specific drawing item.

- 8 background of images
- 9 real channel of an FID
- 10 imaginary channel of an FID
- 11 spectrum
- 12 integral
- 13 parameters
- 14 scale
- 15 threshold line (graphics device only)
- 16 second spectrum or FID in addi (graphics device only)
- 17 result spectrum or FID in addi (graphics device only)
- 18 cursors (graphics device only)
- 19 foreground of images
- 20 background color of graphics window (graphics device only)
- 20-35 contour 0 to contour 15 of absolute value 2D display
- 36-42 contours -7 to -1 of phased 2D display
- 44-50 contours 1 to 7 of phased 2D display

pen_number is an integer from 1 to 8 that specifies the physical pen used.

color is a string for the color set for the device: 'red', 'green', 'blue', 'cyan', 'magenta', 'yellow', 'white', or 'black'.

black_plane is 1 or 0, specifying whether the plotter has a separate black mode. Because all currently supported plotters have this feature, the value is usually 1.

color_planes specifies how many colors are available. Use 3 for color plotters and 0 for black and white plotters.

Examples setcolor('pcl',11,'green')
setcolor('hpgl',11,'red')
setcolor('pen',2,'red')
setcolor('graphics',11,255,0,0)

	setcolor('ps',11,255,255,0)		
	<pre>setcolor('plotter',1,0)</pre>		
See also	NMR Spectroscopy User Guide		
Related	addi	Start interactive add/subtract mode (C)	
	color	Select plotting colors from a graphical interface (M)	

setDECpars Sets Decoupler Parameters

Description	Called to set decoupler parameters when dn is changed during
	customizations.
Syntax	setDECpars
Related	setDECpars, setobspars

setdec2pars Set decoupler 2 parameter values from probe file (M)

Syntax	setdec2pars
Description	Reads from the probe file pwx2lvl, pwx2, dpwr2, dmf2, dmm2, dres2, and dseq2 values, if they exist, and updates the current experiment parameters.
Related	setdecpars Set decoupler parameter values from probe file (M)

setdgroup Set the Dgroup of a parameter in a tree (C)

Syntax	<pre>setdgroup(parameter,dgroup<,tree>)</pre>	
Description	Sets the Dgroup of a parameter in a tree. The application determines the usage of setdgroup. Only Tcl-dg currently uses this feature.	
Arguments	parameter is the name of the parameter.	
	dgroup is an integer.	
	tree is 'current', 'global', 'processed', or 'systemglobal'. The default is 'current'. Refer to the description of the create command for more information on types of trees.	
Examples	<pre>setdgroup('a',1) setdgroup('b',3,'global')</pre>	
See also	User Programming	
Related	create Create new parameter in a parameter tree (C)	

S

setenumeral Set values of a string parameter in a tree (C)

Syntax	<pre>setenumeral(parameter, N, enum1, enum2,, enumN<, tree>)</pre>	
Description	Sets the possible values of a string parameter in a parameter tree. To remove enumerated values from a parameter, set argument N to 0 (see example below).	
Arguments	parameter is the name of the parameter.	
	N is the number of enumeral values to be assigned to parameter (or removed from parameter if N is set to 0).	
	enum1 to enumN are the possible string values of the parameter.	
	tree is 'current', 'global', 'processed', or 'systemglobal'. The default is 'current'. Refer to the description of the create command for more information on types of trees.	
Examples	<pre>setenumeral('size',0) setenumeral('size',2,'large','small') setenumeral('user',3,'user','superuser','master',</pre>	
See also	User Programming	
Related	create Create new parameter in a parameter tree (C)	

setether Connect or reconnect host computer to Ethernet (U)

- Description Connects or reconnects the host computer to the Ethernet network. Only root can execute this shellscript properly. If the system is already connected to the Ethernet network, setether does nothing. On systems running Solaris, setether undoes the work of setnoether. You cannot use setether unless you previously entered the setnoether command. setether restores the files hostname.le0, defaultdomain, and defaultrouter so that Ethernet is activated on the host computer when UNIX is rebooted. See also VnmrJ Installation and Administration
 - Related setnoether Disconnect host computer from Ethernet (U)

setexport Set parameter bits for use with protocols (M)

Description Set the parameter protection bits for use with the rtx command. Usually called by other macros, and not used from the command line.

Related rtx

cqprotocol Create study queue parameters for liquids (M)

setfrq Set frequency of rf channels (C)

Syntax setfrq<(channel)><('nucleus')>

Description Calculates frequencies based on the nucleus (tn, dn, dn2, etc.), referencing (lockfreq), solvent, and the offset parameter (tof, dof, etc.). The result of the calculation is stored in parameters sfrq, dfrq, dfrq2, etc. The parameters are rounded to the resolution of the channel—either 0.1 or 100 Hz.

The setfrq command should never need to be entered from the keyboard. It is called automatically when the appropriate parameters are changed or a parameter set is returned. If a parameter is entered that affects a single frequency, setfrq is called from an internal underscore macro (e.g., _tn, _tof, _dn, _dof) to recalculate the frequency for that channel. Likewise, if a parameter is entered that affects all frequencies, setfrq is called from an internal underscore macro (e.g., _tn, _lockfreq) to recalculate the frequencies.

Arguments channel is a single integer specifying the rf channel to be set. The default is to calculate the frequencies for all rf channels.

nucleus displays or returns the frequency of the supplied nucleus. Channel 1 is assumed for rounding information and an offset (e.g., tof or dof) is not added to the result.

Examples	setfrq
	setfrq(2)
	<pre>setfrq('P31'):freq</pre>
See also	NMR Spectroscopy User Guide
Related	spcfrq Display frequencies of rf channels (M)

setgauss Set a Gaussian fraction for lineshape (M)

Syntax	 setgauss(fraction) setgauss(fraction*) 	
Description	Modifies the output of a deconvolution using pure Lorentzian lineshape (fitspec.outpar) and makes it the input for a subsequent analysis (fitspec.inpar), after first modifying the Gaussian fraction. To allow this fraction to vary, use syntax 1; to fix the fraction, use syntax 2.	
Arguments	fraction is the Gaussian fraction of the lineshape, a number from 0 to 1. To fix the fraction (syntax 2), suffix the value with an asterisk (*) and enclose the value in single quotes (see the second example below).	
Examples	<pre>setgauss(0.4) setgauss('1.0*')</pre>	
See also	NMR Spectroscopy User Guide	
Related	fitspec Perform spectrum deconvolution (C)	

setgcal Set the gradient calibration constant (M)

Applicability Systems with pulsed field gradients (PFG) or imaging capabilities. Description Determines the gradient calibration constant gcal by using a proton phantom of known dimensions. setgcal requests the linear dimension of the phantom in the readout direction. It uses the value entered, together with cursor separation of this dimension from the image profile and the strength of the readout gradient gzlvl1 if pulsed field gradients, to calculate gcal in units of gauss/cm-DAC units. You are then prompted whether this value should be entered. If you answer yes, it is stored as a system constant in the your global file. Note that a particular value of gcal is closely related to the current eddy current compensation settings. If these settings are changed (e.g., reading in a new curecc file), a different value of gcal should be expected. Before running setgcal, use the pulse sequence set up by profile to acquire a signal from a known sized object while the gradient is on.

See also Pulsed Field Gradient Modules Installation; VnmrJ Imaging NMR

RelatedgcalGradient calibration constant (P)profileSet up pulse sequence for gradient calibration (M)

setgcoil Assign sysgcoil configuration parameter (M)

Syntax	<pre>setgcoil<(file)></pre>		
Description	Allows user	rs to change the configured gcoil for the system.	
	setgcoil updates the systemglobal parameter sysgcoil to the		
	named table and updates the assignment value of the parameter gcoil		
	in the named table. The directory		
	Svnmrsystem/imaging/gradtables must have write permission for		
	all users for the macro to be effective. This table now exists in the		
	system local /var/vnmr/gradtables directory, with a soft link from		
	<pre>\$vnmrsystem/imaging/gradtables to that directory.</pre>		
Arguments	file is the any legal file name defined for the parameter gcoil.		
See also	VnmrJ Imaging NMR		
Related	config	Display current configuration and possible change it (M)	
	gcoil	Read data from gradient calibration tables (P)	
	sysgcoil	System value for gcoil parameter (P)	

setgrid Divide graphics window into rows and columns (C)

Syntax setgrid(row<, column>)

Description	Divides graphics window panes). can be activated by entering sets	window into an array of rows and columns (or Only one pane is active at a time. An individual pane by double-clicking in it with the left mouse button or win in the input window.
Arguments	ents row is the number of rows (maximum is 3) in the graphic 0 is entered, the number of rows remains the same; e.g., setgrid(0,2), the number of rows is unchanged and t are created in each row.	
	column is the n window.	umber of columns (maximum is 3) in the graphics
Examples	<pre>setgrid(3) setgrid(3,3) setgrid(0,2)</pre>	
See also	NMR Spectrosco	py User Guide
Related	curwin	Current window (P)
	fontselect	Open FontSelect window (C)
	jwin	Activate current window (M)
	mapwin	List of experiment numbers (P)
	setwin	Activate selected window (C)

setgroup Set group of a parameter in a tree (C)

Syntax	<pre>setgroup(parameter,group<,tree>)</pre>		
Description	Sets the group of a parameter in a tree.		
Arguments	parameter is the	he name of the parameter.	
	group is one of 'acquisition'	the following keywords: 'all', 'sample', , 'processing', 'display', or 'spin'.	
	tree is one of the tree is one of the default is 'on the types of	he keywords 'current', 'global', or 'processed'. current'. See the create command for information trees.	
Examples	<pre>setgroup('a', setgroup('b',</pre>	'sample') 'all','global')	
See also	User Programming		
Related	create	Create new parameter in a parameter tree (C)	
	destroy	Destroy a parameter (C)	
	destroygroup	Destroy parameters of a group in a tree (C)	
	display	Display parameters and their attributes (C)	
	groupcopy	Copy parameters of group from one tree to another (C)	
	paramvi	Edit a parameter and its attributes using vi text editor (M)	
	setlimit	Set limits of a parameter in a tree (C)	
	setprotect	Set protection mode of a parameter (C)	

sethtfrq1 Set a Hadamard frequency list from a line list ((M)

- Description A macro to set the Hadamard frequency list htfrq1 from a line list curexp+'/dll.out'. It assumes that the line list has already been created. The macro also sets ni to the Hadamard matrix size, creates htofs1, and sets fn1 from the minimum frequency difference in htfrq1.
 - See also NMR Spectroscopy User Guide

Related	htfrq1	Hadamard frequency list in ni (P)
	dll	Display listed line frequencies and intensities (C)
	htofs1	Hadamard offset in ni (P)
	fn1	Fourier number in the 1st indirectly detected dimension (P)
	ni	Number of increments in the 1st indirectly detected
		dimension (P)

sethw Set values for hardware in acquisition system (C)

Applicability Syntax 1 through 5 apply to all systems. Syntax 6 applies only to systems with a sample changer. Syntax 7 and 8 apply only to systems with a variable temperature (VT) controller.

Syntax The following syntax is used with the sethw command:

- 1 sethw(<'wait'|'nowait',>par1,val1<,par2,val2,...)</pre>
- 2 sethw('lock','on'|'off')
- 3 sethw('spin',speed)
- 4 sethw('spinner','bump')
- 5 sethw('eject','on'|'off')
- 6 sethw('loc',location)
- 7 sethw('vt','reset'|'off')
- 8 sethw('temp',temperature)
- 9 sethw('lockfreq',lockfreq_value)

Description sethw allows the VNMR program to set values for selected parameters in the acquisition hardware. sethw cannot be used when an acquisition is in progress or when the acqi program is active.

> Syntax 1 can be used to set the lock system parameters lockpower, lockgain, lockphase, and z0. This syntax can also be used to set the values of the shims. The particular shim that can be set depends upon the type of shim hardware present in the system. See the description of shimset for a list of the shim names for each type of shim hardware.

Syntax 2 turns the hardware lock on or off.

Syntax 3 controls spinning speed.

Syntax 4 carries the sample to bump by giving it a short burst of eject air. This is sometimes useful to reseat the sample if it is failing to spin.

Syntax 5 ejects and inserts samples into the probe. Entering the command sethw('eject','on') is equivalent in function to macros eject and e; and sethw('eject','off') is equivalent to macros insert and i.

Syntax 6 sets a location for the sample currently in the magnet on a system with a sample changer. The parameter loc is updated.

Syntax 7 resets the VT controller, useful when changing the probe in a system with VT regulation. By entering sethw('vt', 'reset') after installing a new probe in the magnet and attaching the VT controller interface to the probe, the VT controller is ready to regulate the temperature. No other parameters can be modified by the command. As an alternate, you can manually turn the VT controller unit off and then back on. Syntax 7 also turns the VT controller off by entering sethw('vt','off').

Syntax 8 sets the temperature in degrees celsius. The host computer does not wait for the temperature to regulate.

Syntax 9 sets the lock frequency, in MHz.

Arguments 'wait' or 'nowait' keyword must be either the first or last argument.

- 'wait' sends the new values to the acquisition console, verifies these values, and updates the corresponding parameters. This is the default.
- 'nowait' sends the new values to the console without verifying them or changing parameters.

parameter1, value1, parameter2, value2, ... are paris of parameter names and their values (see the first two examples below). At least one parameter name and its value must be specified. A maximum of ten parameters can be set.

'lock', 'on' is a keyword pair to turn the hardware lock on.

'lock', 'off' is a keyword pair to turn the hardware lock off.

'liqbear' sets the bearing air on level; see liqbear parameter.

'pneufault' second argument is 'clear', 'n', 'w', or 'y' to clear or set the pneumatics fault code.

'spin' is a keyword that identifies the next argument, speed, as the sample spinning speed, in Hz.

'spinner', 'bump' is a keyword pair to bump the sample.

'eject', 'on' is a keyword pair to eject the sample from the probe.

'eject','off' is a keyword pair to insert the sample into the probe.

'loc' is a keyword to identify that the next argument, location, is a number for the sample currently in the magnet ('loc' is unrelated to the loc parameter).

'vt','reset' is a keyword pair to reset the VT controller after the controller has been disconnected from the probe. This is equivalent to turning the VT controller power off and on.

'vt','off' is a keyword pair to turn the VT controller off.

'temp' is a keyword that identifies the next argument, temperature, as the requested sample temperature, in degrees celsius. 'lockfreq' is a keyword that the next argument is the lock frequency. lockfreq_value is the lockfreq value, in MHz, for the lock frequency. 'lockrate' is a number <5000 used internally; usually 20 or 2000. Examples sethw('z1c',30,'z2c',-50) sethw('wait','z1',150,'z2',-400) sethw('lock','on') sethw('spin',20) sethw('spinner','bump') sethw('eject','on') sethw('loc',5) sethw('vt','reset') sethw('lockfreq',46.042) See also NMR Spectroscopy User Guide Related loc Location of sample in tray (P) lockpower Lock power (P) Lock frequency (P) lockfreq lockgain Lock gain (P) lockphase Lock phase (P) Read current values of acquisition hardware (C) readhw sethwshim Set values for hardware in acquisition system (C) Sample spin rate (P) spin

z0 Z0 field position (P)

sethwshim Special case of sethw for setting shims (C)

Applicability	VnmrJ 3.2		
Description	sethwshim sethwshim command is a special case of sethw. It takes two arguments, the shim name and shim value, as in sethwshim('z1',1000)		
Arguments	sethwshim('z1',1000)		
See also	NMR Spectroscopy User Guide		
Related	sethw Set values for hardware in acquisition system (C)		

setint Set value of an integral (M)

Syntax	<pre>setint(int_number<,value>)</pre>
Description	Sets the value of an integral.

Arguments	<pre>int_number is the integral number. It corresponds to the index number displayed by dli if all integrals are shown (i.e., intmod='full') or the region if alternating integrals are shown (i.e. intmod='partial').</pre>	
Francis	value sets th	ne actual value of the selected integral. The default is ins.
Examples	<pre>setint(2) setint(1,3</pre>)
See also	NMR Spectro	scopy User Guide
Related	dli	Display list of integrals (C)
	ins	Integral normalization scale (P)
	intmod	Integral display mode (P)

setlimit Set limits of a parameter in a tree (C)

Applicabili	ty All			
Synta	ix set	<pre>setlimit(name, max,min,step [,tree])</pre>		
	set	<pre>climit(name, index[,tree])</pre>		
Descriptio	on set	limit sets the limits of a variable in a tree.		
	The an size sets lool tab	The limits are max value, min. value and step size. A variable, such a an index into the table, can look up maximum, minimum, and step sizes in a table. Supplying all three (max, min., and step) argument sets the parameter's protection bits (see setprotect) so that the tabl lookup is turned off. The parameter's protection bits are set so that table lookup is turned on if only a single index argument is supplie		
	The	e step value is only used if the parameter is a real number.		
S < > <u>></u>	tep Value - 1 → 1 and < → 0 and <1 2 1	 Parameter setting The parameter is set to the nearest larger value that is a power of 2. The fn parameter uses a step of -2 to select this case. The inverse of the parameter is set to the nearest multiple of the absolute value of the step. The sw parameter uses a step of negative of the minimum dwell time to select this mode. The parameter is set to the nearest multiple of the step value. As an equation, value = n * step where n is a positive or negative integer. The parameter is set to nearest value that is a multiple of step relative to the minimum value. For example, setlimit('var', 3, -3, 2) allows only the following values -3, -1, 1, and 3. As an equation, value = min + n*step where n is an integer >= 0. In this example, the equation is: value = (-3) + (n * 2).		
	Up the	to four optional return arguments can be used. The first will return maximum, the second will return the minimum, and the third will		

the maximum, the second will return the minimum, and the third will return the step size. The fourth argument will return a 0 if the parameter is not using an indexed table lookup for the maximum, minimum, and step size. If the parameter is using the table lookup mechanism, the fourth argument will be set to the index for that table.

	The variable 'systemglo]	trees are 'current', 'global', 'processed' and bal'. The default tree is 'current'.
Arguments	name — the n	name of the variable.
	tree - the v or systemgl	ariable tree: current (the default), global, processed, obal.
Examples	setlimit('a	a',10000,0,.3)
	setlimit('I	b',1e5,-3e2,1,'global')
	setlimit('o	dpwr',9)
See also	User Program	nming
Related	create	Create new parameter in a parameter tree (C
	destroy	Destroy a parameter (C)
	display	Display parameters and their attributes (C)
	fread	Read parameters from file and load them into a tree (C)
	fsave	Save parameters from a tree to a file (C)
	getlimit	Get the limits of a variable in a tree (C)
	paramvi	Edit a parameter and its attributes using vi text editor
		(M)
	parmax	Parameter maximum values (P)
	parmin	Parameter minimum values (P)
	parstep	Parameter step size values (P)
	prune	Prune extra parameters from current tree (C)
	setgroup	Set group of a parameter in a tree (C)
	setprotect	Set protection mode of a parameter (C)
	settype	Change type of a parameter (C)
	setvalue	Set value of any parameter in a tree (C)

setlk Set up lock parameters (M)

Syntax	setlk(solvent)
Description	Called from other macros to provide adjustment of locking and shimming as a function of solvent. Removing quotation marks from around different parts of the text file of the macro places that particular section into effect. If the macro is left unchanged, setting <pre>alock='s'</pre> is required in the parameter sets where used.
Arguments	solvent is the solvent to be used.
See also	NMR Spectroscopy User Guide
Related	alock Automatic lock status (P)

setlockfreq Setlock frequency (M)

Description	Calculates and sets the lock frequency parameter lockfreq. Before using setlockfreq, you must acquire a signal using ¹ H as the transmitter nucleus ($tn='H1'$). To avoid errors in calculating frequencies set lockfreq='n' before starting the acquisition	
See also	VnmrJ Installation and Administration	
Related	lockfreqLock frequency (P)tnNucleus for observe transmitter (P)	

setLP Set up linear prediction in the direct dimension (M)

Applicability	ALL	
Syntax	setLP(n)	
Description	Sets up lir coefficient	near prediction in the direct dimension using the number of s specified.
Examples	setLP(3)	
See also	NMR Spec	troscopy User Guide
Related	lpext	LP data extension in np dimension (P)
	lpfilt	LP coefficients to calculate in np dimension (P)
	lpnupts	LP number of data points in np dimension (P)
	lpopt	LP algorithm data extension in np dimension (P)
	proc	Type of processing on np FID (P)
	setrc	Set frequency referencing based upon lock signal shift (M)
	strtext	Starting point for LP data extension in np dimension (P)
	strtlp	Starting point for LP calculation in np dimension (P)

setLP1 Set F1 linear prediction parameters (M)

Syntax	setLP1<(extended_length<,current_length>)>		
Description	Sets F1 linear prediction parameters. If no arguments are specified, the interferograms are quadrupled in length.		
Arguments	extended_length is the number of complex points now existing (ni).		
	current_length is the number of points desired after the (forward) linear prediction.		
See also	NMR Spectroscopy User Guide		
Related	ni Number of increments in 1st indirectly detected dimension (P)		

setlp0 Set parameters for zero linear phase (M)

Syntax setlp0

Description	A new value of ddrtc is calculated by setlp0 using the current values of alfa, rof2, and lp to achieve a zero linear phase condition (lp=0). A trial experiment must first be acquired and phased for pure absorption before running setlp0. A value of lp near zero is required for flat base line.	
See also	NMR Spectroscopy User Guide	
Related	alfa	Set alfa delay before acquisition (P)
	ddrtc	Set ddr time constant (P)
	lp	First-order phase in directly detected dimension (P)
	rp	Zero-order phase in directly detected dimension (P)
	SW	Spectral width in directly detected dimension (P)

rof2 Receiver gating time following pulse (P)

setnoether Disconnect host computer from Ethernet (U)

Description Disconnects the host computer from the Ethernet network. Only root can execute this shellscript properly. setnoether does nothing if the system is already disconnected from the Ethernet network. On systems running Solaris, setnoether renames the hostname.le0, defaultdomain, and defaultrouter files so that Ethernet is not activated when the system is rebooted.
 See also VnmrJ Installation and Administration
 Related setether Connect or reconnect host computer to Ethernet (U)

setobspars Sets Observe Parameters

Description Called to set observe parameters when the is changed during customizations. Syntax setobspars Related setDECpars, setobspars

setoffset Calculate offset frequency for given nucleus and ppm (M)

Syntaxsetoffset(nucleus,ppm):offsetfreqDescriptionUsing the setref macro, setoffset calculates the offset frequency
for a given chemical shift and returns the value.

Arguments	nucleus is the given nucleus.	
	ppm is the chemical shift.	
	offsetfreq returns the offset frequency for the given chemical shift.	
Examples	setoffset(tn,5):tof setoffset('C13',85):dof	
See also	NMR Spectroscopy User Guide	
Related	setref Set frequency referencing for proton spectra (M)	

setparams Write parameter to current probe file (M)

Syntax	<pre>setparams(param,value<,nucleus>)</pre>		
Description	Writes the value of a parameter to the current probe file. The name of the probe file is referenced from the parameter probe.		
Arguments	param is the n	ame of the parameter to write.	
	value is a str	ing with the value to be written for the parameter.	
	nucleus is the nucleus to write in the probe file. The default is the current value of the parameter tn .		
Examples	setparams('pw90','10') setparams('pplvl','60') setparams('dpwr',\$strdpwr,'H1')		
See also	NMR Spectroscopy User Guide		
Related	addnucleus	Add new nucleus to existing probe file (M)	
	addparams	Add parameter to current probe file (M)	
	addprobe	Create new probe directory and probe file (M)	
	getparam	Retrieve parameter from probe file (M)	
	probe	Probe type (P)	
	tn	Nucleus for the observe transmitter (P)	
	updateprobe	Update probe file (M)	

setpen Set maximum number of HP plotter pens (M)

Syntax	<pre>setpen<(maxpen,max_number_pens)></pre>
Description	Allows the user to interactively define the maximum number of pens when changing to a Hewlett-Packard plotter.
Arguments	maxpen is the current value of the parameter maxpen.
	maximum_number_pens is the maximum number of pens to be used. If the value of max_number_pens is less than or equal to the current value of the parameter maxpen, this value becomes the new value of maxpen.

See also	NMR Spectroscopy User Guide	
Related	color	Select plotting colors from a graphical interface (M)
	maxpen	Maximum number of pens to use (P)

setplotdev Return characteristics of a named plotter (C)

Syntax	<pre>setplotdev<:plotter_type,plotter_host,ppmm,raster></pre>		
Description	Returns information from the devicenames and devicetable files to identify the characteristics of a plotter. This command need never be entered directly by a user because it is automatically called whenever the plotter parameter is set. Note that different "types" of plotters (and printers) are characterized in devicetable. The devicenames file associates different "names" to a given "type."		
Arguments	plotter_type returns the type of the named plotter.		
	plotter_host returns the host associated with the plotter.		
	ppmm returns the plotter resolution in points per millimeter.		
	raster returns the value from the devicetable file.		
See also	VnmrJ Installation and Administration		
Related	plotter Plotter device (P)		

setpower Set power and pulsewidth for a given γ B1 value (M)

Syntax	setpower(γ B1,nucleus)	
Description	Sets power level and pw90 values. For tn, setpower uses ref_pwr and ref_pw90 from the parameter set or from the probe table. For dn, it uses ref_pwx1vl and ref_pwx90 from the parameter set or from the probe table. For dn2, it uses ref_pwx21vl and ref_pwx290 from the parameter set or from the probe table. If the reference power levels and pulse width do not exist, setpower uses tpwr (pw90), dpwr (1/dmf) or dpwr2 (1/dmf2) (if the nucleus is tn, setpower uses tpwr; if the nucleus is dn, it uses dpwr; if the nucleus is dn2, it uses dpwr2).	
Arguments	γB1 is a given γB1 value.	
	nucleu	is a given nucleus.
Examples	<pre>setpower(sw,tn) setpower(5000,H1)</pre>	
Related	dn dn 2	Nucleus for first decoupler (P)
	dnur	Power level for first decoupler (F)
	dowr?	Power level for second decoupler (P)
	pw90	90° pulse width (P)

swSpectral width in directly detected dimension (P)tpwrObserve transmitter power level with linear amplifiers (P)

setprotect Set the protection bits of a variable in a tree (C)

Arguments Every parameter has a set of "protection bits" associated with it. The command "setprotect" allows one to change, list, or test these protection bits. The meaning of each "bit" is shown in the list below. The variable trees are 'current', 'global', 'processed', 'systemglobal', and 'usertree'. The default tree is 'current'. 'names' is either a list of the names of the parameters to be read or it is the name of an arrayed temporary \$ variable. If it is a list, it is a string parameter and the names can be separated either by a space or a comma. If it is an arrayed temporary \$ variable, each array element is a single parameter name.

Either 'set', 'on', 'off', 'list', 'clear', 'getval', or 'ison' must be the second argument. 'Set' causes the current protection bits to be completely replaced with the set specified by the third parameter 'bit_vals'. 'On' causes the specified bits to be turned on. It does not affect other protection bits. 'Off' causes the specified bits to be turned off. It does not affect other protection bits.

'list' and 'alist' cause all parameter with the specified bit_vals turned on to be returned. This list may be returned to the calling macro. The typical invocations will be

setprotect('', 'list',8):\$parlist or setprotect('', 'alist \$parlist',8):\$num In the 'list' case, the \$parlist return variable will be set to a space separated list of parameter names. In the 'alist \$parlist' case, the \$num return variable will be set to the number of returned values. The actual values will be returned as an array in the \$parlist variable.

The 'clear' option clears the specified bit_vals from all parameters. For the list, alist, and clear options, the names argument must be ". The return value when setprotect is called with the list or alist options can be used as the 'names' argument for other forms of setprotect. It can also be names for other commands which use lists of parameter names, such as writeparam and readparam.

'getval' returns the value of the protection bits for the single specified parameter.

'ison' tests the parameter specified by the 'names' argument to see if the bit_vals are on. In this case, the 'names' argument must be a single parameter name. The return value is 1 if the specified bits are all on. The return value is 0 if the specified bits are not all on. The return value is -1 if the specified parameter does not exist in the specified tree.

'Bit_vals' is the sum of the values of the selected bits.

S

Arguments

Examples

<pre>setprotect('syn','on',2)</pre>	cannot set 'syn' to 'y' or 'n'.
<pre>setprotect('pslabel','on',8)</pre>	causes a macro to be executed when this parameter is changed. The name of the macro must, in general, be calledname. In this case, it must be calledpslabel.
<pre>setprotect('','list',8):\$mac</pre>	Puts into the \$mac parameter the list of parameters which have bit 8 on.
<pre>setprotect('','alist \$mac',8)</pre>):\$num Puts into \$num the number of parameters that have bit 8 on. The names of the parameters that have bit 8 on will be returned as an array in the \$mac variable.
<pre>setprotect('sw','ison',8):\$s</pre>	wmac Set \$swmac to 1 if sw has bit 8 on.
\$list='a b c d'	
<pre>setprotect(\$list,'on',2)</pre>	

\$arraylist='a','b','c','d'

setprotect('\$arraylist','on',2) This is the same as the above example, however the variable names are passed as an arrayed temporary \$ variable \$arraylist. Note the single quotes around the first argument to setprotect. The name of the local temporary \$variable is passed to the command, not its value. This format is useful if the list of parameters is large.

parameter protection definitions using a bit field if the bit is set, the comment is true

Bit	Value	Description
0	1	Cannot array the parameter
1	2	Cannot change active/not active status
2	4	Cannot change the parameter value
3	8	Causes macro to be executed
4	16	Avoids automatic redisplay
5	32	Cannot delete parameter
6	64	System ID for spectrometer - datastation
7	128	Cannot copy parameter from tree to tree
8	256	Will not set array parameter
9	512	Cannot set parameter enumeral values
10	1024	Cannot change the parameter's group
11	2048	Cannot change protection bits
12	4096	May cause ipa macro to be executed
13	8192	Look up min, max, step values in table
14	16384	Parameter marked for locking (P LOCK; see rtx)
15	32768	Global parameter not shared in multiple VJ viewports
16	65536	Force automatic redisplay in VJ templates

See also	User Programming

Related	array	Parameter order and precedence (P)
	create	Create new parameter in a parameter tree (C)
	destroy	Destroy a parameter (C)
	display	Display parameters and their attributes (C)
	fread	Read parameters from file and load them into a tree (C)
	fsave	Save parameters from a tree to a file (C)
	getlimit	Get the limits of a variable in a tree (C)
	paramvi	Edit a parameter and its attributes using vi text editor
		(M)
	prune	Prune extra parameters from current tree (C)
	setlimit	Set limits of a parameter in a tree (C)

setpw180ad Creates and sets observe adiabatic pulse shapes (M)

Syntax	setpw180ad(tn,<'make' or 'create'>,<'base shape'>,<'bandwidth in ppm''>)	
Applicability	VnmrJ 3.1	
Description	Based upon probe calibrations, this will create adiabatic pulse shap for a given nucleus as defined in the tn parameter.	
	Based upon the second argument, which defaults to "make", it will set the adiabatic pulse parameter values.	
Examples	setpw180ad(tn)	
	<pre>setpw180ad(tn,'make')</pre>	
	<pre>setpw180ad(tn,'make',wurst2i)</pre>	
	<pre>setpw180ad(tn,'create',wurst2i)</pre>	
	<pre>setpw180ad(tn,'make','wurst2i',115)</pre>	
Related	<pre>setpwx180ad Och_adiabtic_module</pre>	

setpwx180ad Creates and sets decoupler adiabatic pulse shapes (M)

Syntax	setpwx180ad(dn,<'make' or 'create'>,<'base shape'>,<'bandwidth in ppm''>)
Applicability	VnmrJ 3.1
Description	Based upon probe calibrations, this will create adiabatic pulse shapes for a given nucleus as defined in the dn parameter.
	Based upon the second argument, which defaults to "make", it will set the adiabatic pulse parameter values.
Examples	setpwx180ad(dn)
	<pre>setpwx180ad(dn,'make')</pre>

```
setpwx180ad(dn, 'make', wurst2i)
setpwx180ad(dn, 'create', wurst2i)
setpwx180ad(dn, 'make', 'wurst2i', 115)
Related setpw180ad Dch_adiabtic_module
```

setrc Set receiver constants (M)

Applicability Syntax	VNMRS and setrc	400 - MR systems
Description	Sets receiver minimum val a minimum val zero. ddrtc i is set based Linear predic value is more for setting up receiver cons	time constants to optimal values. alfa is set to a ue from the probe file (default is 10 μ s). rof2 is set to value from the probe file (default is 25 μ s). lp is set to is set to a value based upon the ddrpm parameter, which upon pulse sequence type (default value ddrpm = 'p'). etion is turned on in the direct dimension if the ddrtc e than a dwell time. setrc is used in the apptype macros o pulse sequences or from the command line to optimize tants.
Description	sets receiver	time constants to optimal values.
See also	NMR Spectro	scopy User Guide
Related	alfa	Set alfa delay before acquisition (P)
	rof2	Receiver gating time following pulse (P)
	pw	Pulse width (P)
	probe	Probe type (P)
	ddrtc	Set ddr precession mode (P)
	ddrpm	Set ddr precession mode (P)
	SW	Spectral width in directly detected dimension (P)
	setLP	Set F1 linear prediction parameters (M)

setref Set frequency referencing (M)

Syntax	setref<(nucleus)>:\$rfl,\$rfp,\$reffrq,\$refpos
Description	Calculates the referencing for a given parameter or FID data set, for samples locked on deuterium, and based on the chemical shift of the lock solvent line. <code>setref</code> uses information in /vnmr/solvents (² H chemical shift for current solvent) and /vnmr/nuctables/nuctabref (absolute reference frequencies for NMR nuclei) to predict the position of the reference frequency with the current solvent, spectral window, and spectrometer frequency. <code>setref</code> assumes a locked sample.
	The macro calculates the (auxiliary) 2H reference frequency (TMS-d1) from the lock frequency (lockf = lockfreq + $lkof/le6$) as follows:
	H2_TMSfreq = lockf / (1 + solppm/le6)

then takes the Ξ values for ²H and tn and calculates the auxiliary reference frequency (reffrq) for the observe nucleus at the given field strength:

reffrq = $(H2_TMSfreq / \Xi(H2)) * \Xi(tn)$

from this, rfl and rfp are set:

rfp=0 rfl = sw/2 - (sfrq - reffrq)*1e6.

Setting the global (or local) flag bioref ='y' uses Bio-NMR referencing (based on nuctables/nuctabrefBio) rather than standard IUPAC / organic chemistry referencing (based on nuctables/nuctabref)

 Ξ is the normalized frequency such that the $^1\mathrm{H}$ signal from TMS is 100.00 MHz.

This estimate of the frequency based upon the chemical shift value of the lock signal and does not account for temperature, pH, or other factors affecting the chemical shift of the lock solvent.

The default tree is 'current'.

Arguments An argument and return values are beneficial for the use of setref within other macros such as setref1 and setref2. By default (i.e., without an argument), setref calculates the referencing for 1D spectra or for the directly detected dimension in nD spectra (f2 in 2D, f3 in 3D).

When only nucleus is used as an argument, setref returns values without setting parameters.

\$rfl,\$rfp,\$reffrq,\$refpos are return values for reference peak position, reference peak frequency, reference line frequency, and reference line position, respectively.

Examples setref

setref('C13'):\$rfl,\$rfp

See also NMR Spectroscopy User Guide

Related	reffrq	Reference frequency of reference line (P)
	refpos	Position of reference frequency (P)
	rfl	Reference peak position (P)
	rfp	Reference peak frequency (P)
	rl	Set reference line in directly detected dimension
		(M)
	setref1	Set frequency referencing for 1st indirectly detected
		dimension (M)
	setref2	Set frequency referencing for 2nd indirectly
		detected dimension (M)
	setup	Set up parameters for basic experiments (M)
	tmsref	Reference 1D proton or carbon spectrum to TMS
		(M)
	bioref	Use nuctables/nuctabrefBio) rather than
		standard IUPAC / organic chemistry

setref1 Set freq. referencing for 1st indirectly detected dimension (M)

Syntax	setref1(nu	cleus)
Description	Calculates th parameters a for the solver setref mace chemical shif /vnmr/nuct. NMR nuclei) with the curr nucleus.	e referencing for the first indirect dimension (f1) in nD and FID data sets, for samples locked on deuterium, and at specified by the solvent parameter. setref1 uses the ro to calculate the reference frequency and based on the ft of the lock solvent line and ables/nuctabref (absolute reference frequencies for to predict the referencing in f1 (reffrq1, rf11, rfp1) rent solvent, sw1, and for the frequency of the specified
This estimate of the frequency based upon the chemical the lock signal, as in setref, and does not account for pH, or other factors affecting the chemical shift of the l Using setref, setref1, and setref2, maintains a con- reference for all dimensions		e of the frequency based upon the chemical shift value of al, as in setref, and does not account for temperature, factors affecting the chemical shift of the lock solvent. f, setref1, and setref2, maintains a consistent all dimensions.
	Ξ is the norm 100.00 MHz.	nalized frequency such that the ¹ H signal from TMS is
	Setting the gl (based on nu organic chem	obal (or local) flag bioref ='y' uses bio-NMR referencing ctables/nuctabrefBio) rather than standard IUPAC / istry referencing (based on nuctables/nuctabref)
	See /vnmr/nuctables/nuctabref.	
Arguments	nucleus is t	the frequency-relevant nucleus in f1.
Examples	setref1(tn setref1('C) 13')
See also	NMR Spectroscopy User Guide	
Related	reffrq1	Reference frequency of reference line in 1st indirect dimension (P)
	refpos1	Position of reference frequency in 1st indirect dimension (P)
	rfl	Reference peak position (P)
	rfl1	Reference peak position in 1st indirectly detected
	rfp1	dimension (P) Reference peak frequency in 1st indirectly detected dimension (P)
	setref	Set frequency referencing (M)
	bioref	Use nuctables/nuctabrefBio

setref2 Set freq. referencing for 2nd indirect detected dimension (M)

Syntax setref2(nucleus)

Description Calculates the referencing for the second indirect dimension (f2) in nD parameters and FID data sets, for samples locked on deuterium, and for the solvent specified by the solvent parameter. setref2 uses setref to calculate the reference frequency and based on the chemical shift of the lock solvent line and /vnmr/nuctables/nuctabref (absolute reference frequencies for NMR nuclei) to predict the referencing in f2 (reffrq2, rfl2, rfp2) with the current solvent, sw2, and for the frequency of the specified nucleus.

This estimate of the frequency based upon the chemical shift value of the lock signal, as in <u>setref</u>, and does not account for temperature, pH, or other factors affecting the chemical shift of the lock solvent. Using <u>setref</u>, <u>setref1</u>, and <u>setref2</u>, maintains a consistent reference for all dimensions.

Setting the global (or local) flag bioref ='y' uses bio-NMR referencing (based on nuctables/nuctabrefBio) rather than standard IUPAC / organic chemistry referencing (based on nuctables/nuctabref)

See /vnmr/nuctables/nuctabref.

Arguments nucleus is the frequency-relevant nucleus in f2.

Examples setref2(tn) setref2('C13')

See also NMR Spectroscopy User Guide

Related	reffrq2	Reference frequency of reference line in 2nd indirect dimension (P)
	refpos2	Position of reference frequency in 2nd indirect dimension (P)
	rfl2	Reference peak position in 2nd indirectly detected dimension (P)
	rfp2	Reference peak frequency in 2nd indirectly detected dimension (P)
	r12	Set reference line in 2nd indirectly detected dimension (M)
	setref	Set frequency referencing (M)
	bioref	Use nuctables/nuctabrefBio

setscout Set up a scout run (M)

ApplicabilitySystems with LC-NMR accessory.DescriptionDesigned to help run simple experiments during the setup phase of
LC-NMR or to be the first of two experiments run on peaks in a
stopped-flow or loop-flushing mode. In the latter application, you can
set wexp='setwet au' so that the scout run is analyzed, parameters
adjusted, and an appropriate solvent-suppressed experiment run.If parameters already exist in the current experiment for performing
the lcld pulse sequence, setscout turns off the solvent suppression
portion of the sequence; if they do not exist, they are created and set

to default values using lc1d.

See also	NMR Spectro	scopy User Guide
Related	lc1d	Pulse sequence for LC-NMR (M)
	setwet	Set up a solvent-suppressed experiment (M)

ApplicabilitySystems with LC-NMR accessory.DescriptionSets sslsfrq to the frequencies of each of the suppressed solvents.See alsoNMR Spectroscopy User Guide

setsw Set spectral width (M)

Syntax	<pre>setsw(downfieldppm,upfieldppm)</pre>	
Description	Sets sw and referencing.	tof for the given spectral window and also does
Arguments	downfieldppm is the downfield frequency, in ppm.	
	upfieldppm	is the upfield frequency, in ppm.
Examples	setsw(12,0)	
	setsw(235,-15)	
See also	NMR Spectroscopy User Guide	
Related	setsw1	Set spectral width in evolution dimension (M)
	setsw2	Set spectral width in 2nd evolution dimension (M)
	SW	Spectral width in directly detected dimension (P)
	tof	Frequency offset for observe transmitter (P)

setsw1 Set spectral width in evolution dimension (M)

Syntax	<pre>setsw1(nucleus,downfieldppm,upfieldppm):offset</pre>		
Description	Sets sw1 for the given spectral window and also does referencing.		
Arguments	nucleus returns the nucleus.		
	downfieldppm is the downfield frequency, in ppm.		
	upfieldppm is the upfield frequency, in ppm.		
	offset returns the appropriate offset.		
Examples	setsw1(tn,12,0) setsw1(dn,235,-15):dof		

See also	NMR Spectro	scopy User Guide
Related	setsw	Set spectral width (M)
	sw1	Spectral width in 1st indirectly detected dimension (P)

setsw2 Set spectral width in 2nd evolution dimension (M)

Syntax	<pre>setsw2(nucleus,downfieldppm,upfieldppm):offset</pre>		
Description	Sets sw^2 for the given spectral window and also does referencing.		
Arguments	nucleus returns the nucleus.		
	downfieldppm is the downfield frequency, in ppm.		
	upfieldppm is the upfield frequency, in ppm.		
	offset returns the appropriate offset.		
Examples	setsw2(tn,12,0)		
	setsw2(dn,235,-15):dof		
See also	NMR Spectroscopy User Guide		
Related	setsw Set spectral width (M)		
	sw2 Spectral width in 2nd indirectly detected dimension (P)		

setselfrgc Set selective frequency and width (M)

- Description Sets selective frequency and width of the excitation bandwidth for selective excitation. Used after TOCSY1D and Noesy1d selection. Selected frequencies and widths of the excitation bandwidth are used by suselfrg.
 - RelatedNoesy1dChange parameters for NOESY1D experiment (M)suselfrqSelect peak, continue selective excitation experiment (M)TOCSY1DChange parameters for TOCSY1D experiment (M)

setselinv Set up selective inversion (M)

- Description Sets power, pulsewidth, and shape for selective inversion; used by suselfrg. By default, setselinv selects a q3 gaussian cascade pulse if a waveform generator or linear modulator is present. Otherwise, setselinv selects a "rectangular" pulse.
 - Related setselfrqc Select selective frequency and width (M) suselfrq Select peak, continue selective excitation experiment (M)

settcldefaultSelect default display templates for pulse sequence (M)

Syntax	<pre>settcldefault<(<default><,sequence>)></default></pre>		
Description	Selects the display templates to use as the default for a pulse sequence.		
Arguments	default is the name of the set of display templates to use for the default display of the current pulse sequence (defined by the parameter seqfil). If no arguments are given, the user is prompted for the name of the display templates.		
	sequence defines which pulse sequence will use the default displays of the pulse sequence given as the first argument. The default is the pulse sequence defined by the parameter seqfil.		
Examples	settcldefault settcldefault('cosy') settcldefault('default2d','HMQC8')		
See also	User Programming		
Related	seqfil Pulse sequence name (P)		

settune Opens the Auto Tune Setup dialog (M)

Applicability	Automation, VnmrJ Walkup		
Syntax	settune		
Description	Opens a dialog for setting when to tune in automation using ProTune.		
See also	NMR Spectroscopy User Guide and VnmrJ Walkup		
Related	protune Macro to start ProTune (M)		
	wtune Specify when to tune (P)		

settype Set the type of a parameter (C)

Syntax settype(name,type[,tree])

Description The settype command can change the type of an existing variable. A 'string' variable can have it's type set to 'string' or 'flag'. A 'real' variable can have it's type set to 'real', 'delay', 'frequency', 'pulse', or 'integer'.

The settype command can not be used to change a string variable into a 'real' variable, or a 'real' variable into a 'string' variable. The variable trees are 'current', 'global'. 'processed', 'usertree', and 'systemglobal'. The default is to search for the parameter in the 'current', 'global', and 'systemglobal' trees, in that order.

Arguments The first "name" argument is either a list of the names of the parameters or it is the name of an arrayed temporary \$ variable. If it is a list, it is a string parameter and the names can be separated either by a space or a comma. If it is an arrayed temporary \$ variable, each array element is a single parameter name. If the variable does not exist, a warning message will be given.

Appending the settype command with a return value will suppress warnings if the variable does not exist.

Examples

<pre>settype('in','flag')</pre>	Change 'in' into a 'flag' variable.
<pre>settype('p12','pulse')</pre>	Change 'p12' into a 'pulse' variable.
settype('tpwr dpwr dpwr2	dpwr3','real'):\$e Change the
	power parameters to type 'real' and
	suppress messages if any do not exist

Examples

See also User Programming

Related	create	Create new parameter in a parameter tree (C)
	display	Display parameters and their attributes (C)
	setgroup	Set group of a parameter in a tree (C)
	setlimit	Set limits of a parameter in a tree (C)
	setprotect	Set protection mode of a parameter (C)
	setvalue	Set value of any parameter in a tree (C)

setup Set up parameters for basic experiments (M)

Syntax setup<(nucleus<,solvent>)>

- Description Returns a parameter set to do the experiment requested, complete with positioning of the transmitter and decoupler. Parameters set by setup are recalled from the /vnmr/stdpar directory or from the user's stdpar directory if the appropriate file exists there. Any changes made to the files in these directories are reflected in setup. The default parameters for carbon and proton survey spectra are in files /vnmr/stdpar/C13.par and /vnmr/stdpar/H1.par, respectively. These files should be modified as desired to produce spectra under desirable conditions.
- Arguments nucleus is a nucleus chosen from the files in /vnmr/stdpar or in the user's stdpar directory (e.g., 'H1', 'C13', 'P31'). solvent is a solvent chosen from the file /vnmr/solvents (e.g.,

'CDC13', 'C6D6', 'D2O'). The default is 'CDC13'.

Examples setup

setup('H1') setup('C13','DMSO')

See also NMR Spectroscopy User Guide

setup_dosy Set up gradient levels for DOSY experiments (M)

- Description Initiates a dialogue to set up an array of gzlvl1 values for DOSY experiments. setup_dosy requests the number of array increments and an initial and a final gzlvl1 value and sets up an array that gives increments in gzlvl1 squared between these limits. setup_dosy retrieves the gradient strength from the probe calibration file if probe<>'' and stores it in the local experimental parameter DAC_to_G. If probe='' (i.e., the probe is not defined), then DAC_to_G is set to the current value of the global parameter gcal.
 - See also NMR Spectroscopy User Guide

 Related dosy
 Process DOSY experiments (M)

 DAC_to_G
 Parameter to store gradient calibration value in DOSY sequences (P)

 setgcal
 Set the gradient calibration constant (M)

setuserpsg Creates/initializes user PSG directory

Syntax setuserpsg

Applicability VnmrJ 3.1

Description SETUSERPSG is a UNIX shellscript which performs the following functions:

- creates the user PSG directory if one does not already exist;
- and initializes the user PSG directory with the appropriate PSG object libraries from the system PSG directory, if necessary.

For reference, the user PSG object library in the system PSG directory is LIBPSGLIB.A; and the Agilent PSG object library in the same directory is LIBPARAM.A. SETUSERPSG is automatically invoked by the shellscript PSGGEN.

setvalue Set value of any parameter in a tree (C)

Syntax	<pre>setvalue(parameter,value<,index><,tree>)</pre>
Description	Sets the value of any parameter in a tree. This command bypasses the normal range checking for parameter entry, as well as bypassing any action that would be invoked by the parameter's protection mode (see the setprotect command). If the parameter entry normally causes aparameter macro to be executed, this action also is bypassed.
Arguments	parameter - name of the parameter.
	value -set value for the parameter.
	index — number of a single element in an arrayed parameter. The default is 1. A value of 0 for the index resets an arrayed (or

non-arrayed) parameter to the one element supplied as the second argument to setvalue.

tree - keyword 'global', 'current', 'processed', or 'systemglobal'. The default is 'current'. Refer to the create command for more information on the types of parameter trees.

Examples setvalue('arraydim',128,'processed')

See also User Programming

Related create Create new parameter in a parameter tree (C) setprotect Set protection mode of a parameter (C)

setwave Write a wave definition string into Pbox.inp file (M)

Syntax	setwave('sh bw/pw ofs st ph fla trev d1 d2 d0')
Description	Sets up a single excitation band in the Pbox.inp file. An unlimited number of waves can be combined by reapplying setwave.
Arguments	A single string of 1 to 10 wave parameters in predefined order. Note that a single quote is required at the start and the end of the entire string, but no single quotes are required surrounding characters and strings inside the entire string.
sh bw/ ofs st	name of a shape file. 'pw either the bandwidth, in Hz, or the pulsewidth, in sec. offset, in Hz. number specifying the spin status: 0 for excitation 1 for de-excitation 0.5 for refocusing.
ph fla	phase (or phase cycle, see wavelib/supercycles).
tre	fla can override the default flip angle. ev time reversal. This can be used to cancel time reversal if spin status
d1 d2 d0	(st) is set to 1 for Mxy. delay, in sec, prior the pulse. delay, in sec, after the pulse. delay or command prior to d1. If d0=a, the wave is appended to the previous wave.
Examples	setwave('eburp1') setwave('GARP 12000.0') setwave('esnob 600 -1248.2 1 90.0 n n 0.001')
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

setwell Adjust the label of the "t1" axis for VAST contour maps

Applicability VnmrJ 3.1

setwin Activate selected window (C)

Syntax setwin(row<,column>)

- Description Activates a specific pane in the graphics window. Panes are numbered sequentially from left to right and top to bottom.
- Arguments row is the number of the row containing the pane to be activated. column is the number of the column containing the pane to be activated.
 Examples setwin(3)
 - setwin(1,2) See also NMR Spectroscopy User Guide Related curwin Current window (P) fontselect Open FontSelect window (C) jwin Activate current window (M) mapwin List of experiment numbers (P) setgrid Activate selected window (M)

sf Start of FID (P)

- Description Sets the start of the FID display. This parameter can be entered in the usual way or interactively controlled by the sf wf button during a FID display.
 - Values 0 to the value of at, in seconds.

See also NMR Spectroscopy User Guide

Related at Acquisition time (P) Display noninteractive color intensities map (C) dconi Interactive 2D data display (C) df Display a single FID (C) sf1 Start of interferogram in 1st indirectly detected dimension (P) Start of interferogram in 2nd indirectly detected dimension sf2 (P) vf Vertical scale of FID (P) wf Width of FID (P)
Description	Sets the start of the interferogram display in the first indirectly detected dimension.	
Values	0 to $(2 \times ni)/sw1$, in seconds.	
See also	NMR Spectroscopy User Guide	
Related	ni Number of increments in 1st indirectly detected dimension (P)	
	sul Spectral width in 1st indirectly detected dimension (P)	
	wf1 Width of interferogram in 1st indirectly detected dimension (P)	

sf2 Start of interferogram in 2nd indirectly detected dimension (P)

Description	Sets the start of the interferogram display in the second indirectly detected dimension.	
Values	0 to $(2 \times ni2)/sw^2$, in seconds.	
See also	NMR Spectroscopy User Guide	
Related	 ni2 Number of increments in 2nd indirectly detected dimension (P sf Start of FID (P) sw2 Spectral width in 2nd indirectly detected dimension (P) wf2 Width of interferogram in 2nd indirectly detected dimension (P) 	

sfrq Transmitter frequency of observe nucleus (P)

Description	Contains the frequency for the observe transmitter. $sfrq$ is automatically set when tn is changed, and it should not be necessary for the user to manually set this parameter.	
Values	Number, in M	IHz.
See also	NMR Spectroscopy User Guide	
Related	dfrq	Transmitter frequency of first decoupler (P)
	dfrq2	Transmitter frequency of second decoupler (P)
	dfrq3	Transmitter frequency of third decoupler (P)
	tn	Nucleus for observe transmitter (P)
	tof	Frequency offset for observe transmitter (P)
	spcfrq	Display frequencies of rf channels (M)

sh2pul Set up for a shaped observe excitation sequence (M)

Applicability	Systems with	waveform generators.
Syntax	sh2pul	
Description	Behaves like hard pulses of The name of plpat. Infor- bandwidths i	standard two-pulse sequence S2PUL but with the normal changed into shaped pulses from the waveform generator. the shaped pulse associated with pw is pwpat and p1 is mation about the specifics of power settings and s available from the macros bandinfo and pulseinfo.
See also	User Program	nming
Related	bandinfo	Shaped pulse information for calibration (M)
	plpat	Shape of an excitation pulse (P)
	pwpat	Shape of refocusing pulse (P)
	pulseinfo	Shaped pulse information for calibration (M)

shdec Set up for shaped observe excitation sequence (M)

Applicability	Systems with waveform generators.
Description	Sets up the SHDEC pulse sequence that generates a shaped pulse on the observe channel using the waveform generator. It also allows for programmed (e.g.: multiselective) homodecoupling or solvent presaturation using the observe transmitter, and an optional gradient pulse following the excitation pulse.
See also	NMR Spectroscopy User Guide
Related	Pbox Pulse shaping software (U)

shell Start a UNIX shell (C)

Syntax shell<(command)>:\$var1,\$var2,...

- Description Brings up a normal UNIX shell for the user. On the Sun, a pop-up window is created. On the GraphOn terminal, the entire terminal is used.
- Arguments command is a UNIX command line to be executed by shell. The default is to bring up a UNIX shell. If the last character in the command line is the symbol &, the command is executed in background, which allows commands to be entered and executed while the shell command is still running. Note that if this background feature is used, any printed output should be redirected to a file. Otherwise, the output may pop up in the text window at random times.

shell calls involving pipes or input redirection (<) require either an extra pair of parentheses or the addition of; cat to the shell command string.

\$var1, \$var2,... are names of variables to hold text lines that are generated as a result of the UNIX command. The default is to display the text lines. Each variable receives a single display line. shell always returns a text line; in many cases, it is a simple carriage return. To prevent this carriage return from being shown, capture it in a dummy variable, such as

shell('command'):\$dum
Examples shell
shell('ps')
shell('ls -lt'):\$filelist
shell(systemdir+'/acqbin/Acqstat '+hostname+' &')
shell('ls -t|grep May; cat')
or
shell('(ls -t|grep May)')
See also NMR Spectroscopy User Guide, User Programming
Related shelli Start an interactive UNIX shell(C)

shelli Start an interactive UNIX shell (C)

Syntax	shelli(command)	
Description	On a terminal, runs interactively the UNIX command line given as the argument. No return or output variables are allowed.	
Arguments	command is a UNIX command line to be executed.	
Examples	shelli('vi myfile')	
See also	NMR Spectroscopy User Guide, User Programming	
Related	shell Start a UNIX shell (C)	

shim Submit an Autoshim experiment to acquisition (C)

Description	Performs validity checks on the acquisition parameters and then submits an Autoshim experiment to acquisition.	
See also	NMR Spectroscopy User Guide	
Related	au	Submit experiment to acquisition and process data (C)
	change	Submit a change sample experiment to acquisition (M)
	ga	Submit experiment to acquisition and FT the result (C)
	go	Submit experiment to acquisition (C)
	lock	Submit an Autolock experiment to acquisition (C)
	sample	Submit change sample, autoshim experiment to acquisition
		(M)
	spin	Submit a spin setup experiment to acquisition (C)
	su	Submit a setup experiment to acquisition (M)

shimmult Multiple the shim dacs of the current shimset

Syntax shimmult<(multiplier)>

Applicability VnmrJ 3.1

Description The shimmult macro will multiply the value of each dac in the current shimset by a multiplier. The multiplier may be supplied as an argument. The default value is 1.0/1.5. One might use this macro if the current output by the shim power supply has changed. This macro does not load the new values into the hardware. Follow the shimmult macro with a call to "su" to set the hardware. Note also that shim dac values are integer values. Therefore, shimmult(1/3) followed by shimmult(3) may not give the original values, do to truncation effects.

shimnames Returns shim names

Syntax	shimnames<:\$names,\$num>
Applicability	VnmrJ 3.1
Description	This command returns a list of the names of the active shims. These are returned in a single string parameter. A second argument will return the number of active shims. The substr command can be used to extract individual shim names from the returned list.
Arguments	
Examples	shimnames:\$names,\$num

shimset Type of shim set (P)

Description Configuration parameter for the type of shims on the system. The value of shimset is set using the Shimset label in the Spectrometer Configuration window.

Values 1 to 14, where the value identifies one of the following shim sets:

1 is a shim set in a Agilent 13-shim supply with computer-controlled axial shims z1, z1c, z2, z2c, z3, z4, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3. Shims can be adjusted from -2047 to +2047. This value is used with the Ultra•nmr shim system when operated from the HIM box (Agilent 13 Shims choice in Spectrometer Configuration window).

2 is a shim set in a Oxford 18-shim supply with computer-controlled axial shims z1, z1c, z2, z2c, z3, z4, z5, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2. Shims can be adjusted from -2047 to +2047 (Oxford 18 Shims choice in Spectrometer Configuration window).

3 is a shim set in a Agilent 23-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, z6, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy. Shims can be

adjusted from -32767 to +32767 (Agilent 23 Shims choice in Spectrometer Configuration window).

4 is a shim set in a Agilent 28-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, z6, z7, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, z4x, z4y. Shims can be adjusted from -32767 to +32767 (Agilent 28 Shims choice in Spectrometer Configuration window).

5 is a shim set in an Ultra•nmr shim system (39 shim channels) with computer-controlled axial shims z1, z1c, z2, z2c, z3, z3c, z4, z4c, z5, z6, z7, z8, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, z4x, z4y, z3x2y2, z3xy, z2x3, z2y3, z3x3, z3y3, z4x2y2, z4xy, z5x, z5y. Shims can be adjusted from -32767 to +32767 (Ultra Shims choice in Spectrometer Configuration window).

6 is a shim set in a Agilent 18-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2. Shims can be adjusted from -32767 to +32767 (Agilent 18 Shims choice in Spectrometer Configuration window).

7 is a shim set in a Agilent 20-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y. Shims can be adjusted from -32767 to +32767 (Agilent 20 Shims choice in Spectrometer Configuration window).

8 is a shim set in a Oxford 15-shim supply with computer-controlled axial shims z1, z2, z3, z4, and radial shims x1, y1, xz, yz, xy, x2y2, zx2y2, xz2, yz2, zxy. Shims can be adjusted from -2047 to +2047 (Oxford 15 Shims choice in Spectrometer Configuration window).

9 is a shim set in a Agilent Ultra•nmr shim system II (40 shim channels) with computer-controlled axial shims z1, z1c, z2, z2c, z3, z3c, z4, z4c, z5, z6, z7, z8, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, x4, y4, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, z4x, z4y, z3x2y2, z3xy, z2x3, z2y3, z3x3, z3y3, z4x2y2, z4xy, z5x, z5y. Shims can be adjusted from

-32767 to +32767 (Agilent 40 Shims choice in Spectrometer Configuration window).

10 is a shim set in a Agilent 14-shim supply with computer-controlled axial shims z1, z1c, z2, z2c, z3, z4, z5, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3. Shims can be adjusted from -2047 to +2047 (Agilent 14 Shims choice in Spectrometer Configuration window).

11 is a shim set in a Agilent 8-shim supply with computer-controlled axial shims z1, z2, and radial shims x1, y1, xz, yz, xy, x2y2. Shims can be adjusted from -32767 to +32767 (Whole Body Shims choice in Spectrometer Configuration window).

12 is a shim set in a Agilent 26-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, x4, y4.

Shims can be adjusted from -32767 to +32767 (Agilent 26 Shims choice in Spectrometer Configuration window).

13 is a shim set in an Agilent 29-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, z6, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, z4x, z4y, z5x, z5y. Shims can be adjusted from -32767 to +32767 (Agilent 29 Shims choice in Spectrometer Configuration window).

14 is a shim set in a Agilent 35-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, z6, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, x4, y4, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, z4x, z4y, z3x2y2, z3xy, z4x2y2, z4xy, z5x, z5y. Shims can be adjusted from -32767 to +32767 (Agilent 35 Shims choice in Spectrometer Configuration window).

15 is the Agilent 15 Shim.

16 is the Ultra 18 Shims.

17 is a shim set in an Agilent 15-shim supply with computer-controlled axial shims z1, z2, z3, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zx2y2, zxy. Shims can be adjusted from -32767 to +32767 (Agilent Combo Shims choice in Spectrometer Configuration window).

18 is a shim set in an Agilent 28-shim supply with computer-controlled axial shims z1, z2, z3, z4, z5, z6, z7, and radial shims x1, y1, xz, yz, xy, x2y2, x3, y3, xz2, yz2, zxy, zx2y2, z3x, z3y, z2x2y2, z2xy, zx3, zy3, z4x, z4y. Shims can be adjusted from - 32767 to +32767 (Agilent 28 Thin Shims choice in Spectrometer Configuration window).

See also VnmrJ Installation and Administration

Related configDisplay current configuration and possibly change it
(M)readhwRead current values of acquisition hardware (C)

showconfig Show system configuration settings (M)

- See also Displays the system configuration settings in the text window. To print the settings, enter the following in the VnmrJ command line: **printon showconfig printoff**.
 - See also VnmrJ Installation and Administration

Related config Display current configuration and possibly change it (M

showconsole Show system configuration settings (U)

Description Displays console hardware configuration parameters and system versions. This information is recorded during console bootup and represents the system hardware options recognized by the acquisition

computer. The command is used mainly when troubleshooting or performing diagnostics.

See also NMR Spectroscopy User Guide

Related ihwinfo Hardware status of console (C)

showdosy Show DOSY Plot (M)

Syntax	showdosy(<expno>)</expno>
Applicability	VnmrJ 3.1
Description	The macro 'showdosy' is a convenient way of displaying the pseudo 2D DOSY spectrum. Typing showdosy(N) after the completion of the "dosy" macro joins experiment N and displays the DOSY spectrum automatically. It sets fn1=256 and fn=8k, which can be adjusted to achieve better resolution.
Arguments	'expno' experiment number to display the DOSY plot.
Related	dosy ddif

showdosyfit Plots the experimental signal attenuation, fitted attenuation and residual for one peak from a 2D or 3D DOSY experiment

Syntax	showdosyfit(peaknr)
	showdosyfit(peaknr,expFac)
Applicability	VnmrJ 3.1
Description	Displays using expl the result of fitting peak peaknr using dosy. Experimental data points are in red, fitted points in blue, and residuals in magenta.
Arguments	The macro takes one or two arguments (peaknr, expFac), which are the peak number and the expansion factor of the residual respectively. When expansion factor is not given it defaults to 1.
See also	dosy

showdosyresidual Plots the residual for one peak from a 2D or 3D DOSY experiment

Syntax	showdosyresidual(peaknr)
	showdosyresidual(peaknr,expFac)
Applicability	VnmrJ 3.1
Description	Displays using expl the residuals of fitting peak peaknr using dosy.

Arguments	The macro takes one or two arguments (peaknr, expFac), which are
	the peak number and the expansion factor of the residual respectively.
	When expansion factor is not given it defaults to 1.

See also dosy

showgradfit Plots the experimental gradient variation with position and the power series fit in non-uniform gradient calibration.

Syntax	showgradfit
Applicability	VnmrJ 3.1
Description	Displays (using expl) the result of fitting the experimental variation of gradient strength with position, measured during non-uniform gradient calibration, and the result of fitting with a power series.Experimental data points are in red and fitted points in blue.
Arguments	
Examples	
See also	gradfit
	nugcalib
	powerfit
	shownugfit

showfit Display numerical results of deconvolution (M)

Description	After a deconvolution, the results are written into file fitspec.outpar in an abbreviated format. showfit converts these data to an output format more suitable for examination and printing	
See also	NMR Spectroscopy User Guide	
Related	fitspecPerform spectrum deconvolution (C)plfitPlot deconvolution analysis (M)usemarkUse "mark" output as deconvolution starting point (M)	

showloginboxShows operator login dialog (M)

Description Shows the login dialog for operators.

shownugfit Plots the logarithm of the calculated diffusional attenuation and of the power series fit in non-uniform gradient calibration

Syntax	shownugfit
Applicability	VnmrJ 3.1
Description	Displays (using expl) the result of fitting the calculated signal attenuation as a function of gradient squared to the exponential of a power series. Calculated data points are in red and fitted points in blue.
See also	gradfit
	nugcalib
	powerfit
	shownugfit

shownumx Show x position of number (P)

Description	Show the X position of the number. The bottom left of every spectrum is defined as 0.		
See also	User Program	nming	
Related	shownumy	y position counting from bottom left of every spectrum (P)	

shownumy Show y position of number (P)

Description	Show the \mathbf{Y} position of the number. The bottom left of every spectrum is defined as 0.	
See also	User Programming	
Related	shownumx x position counting from bottom left of every spectrum (P)	

showoriginalRestore first 2D spectrum in 3D DOSY experiment (M)

Description	Restores the first 2D spectrum in a 3D DOSY experiment (if it ha		
	been saved b	y the dosy macro).	
See also	NMR Spectroscopy User Guide		
Related	dosy	Process DOSY experiments (M)	

showplotter Show list of currently defined plotters and printers (M)

DescriptionShows a list of currently defined plotters and printers.See alsoNMR Spectroscopy User GuideRelatedplotterPlotterPlotter device (P)printerPrinter device (P)

showplotg Display plot jobs in plot queue (M)

Description	Displays current plot jobs in the plot queue for the active plotter		
See also	NMR Spectroscopy User Guide		
Related	killplot	Stop plot jobs and remove from plot queue (C	
	showprintq	Display print jobs in print queue (C)	

showprintg Display print jobs in print queue (M)

Description	Displays current print jobs in the print queue for the active printer		
See also	NMR Spectroscopy User Guide		
Related	killprint	Stop print jobs and remove from print queue (C)	
	showplotq	Display plot jobs in plot queue (M)	

showprotuneguishow the graphical interface while tuning (P)

Syntax	showprotunegui='argument'		
Description	This is a global string parameter that does not exist by default. The user can create it to force the ProTune GUI to be shown during norma tuning operation.		
Arguments	'n' – Do not force	e the GUI to be shown.	
	'y' - Show the GU	JI, except in automation.	
	'a' – Always show the GUI, even in automation.		
	Set showprotunegui='a' will cause ProTune to fail in automation unless the proper display permission has been set. Set the display permissions on Linux systems by executing "xhost local:" on the Linux command line.		
See also	NMR Spectroscopy	User Guide	
Related	protune	Macro to start ProTune (M)	

showrfmon Show RF Monitor Button in Hardware Bar (P)

Applicability	Imaging
Syntax	showrfmon= <value></value>
Description	Show RF Monitor Button in Hardware Bar.
Values	1 show RF Monitor button.
	-1 hide RF Monitor button.
See also	VnmrJ Imaging User Guide

showsampglobalShows sample global parameters

Description	Shows sample global parameter values in current workspace		
Syntax	showsampglobal		
Related	getsampglobal, resetsampglobal, savesampglobal, mvsampglobal		

showstat Display information about status of acquisition (M,U)

Syntax	(From VnmrJ) showstat<(remote_system)> (From UNIX) showstat <remote_system></remote_system>	
Description	Displays information in the text screen about the status of acquisition on a spectrometer. The command is similar to Acqstat, but displays the information in a non-graphical manner and only once.	
Arguments	remote_system is the host name of a remote spectrometer. The default is to display information about acquisition on the local system	
See also	NMR Spectroscopy User Guide	
Related	Acqstat Bring up the acquisition status display (U)	

sim Sample in magnet (For systems equipped with a robot)

Syntax	sim
Applicability	VnmrJ 3.1
Description	The sim macro generates a pop-up window to set the number of the sample currently in the magnet. The sim macro is only available for systems with a robot. This macro would typically be used only after a manual insert of a sample. In this case, the NMR console is unaware of the proper location of the inserted sample. The sim macro allows a location to be assigned to the inserted sample. In addition to assigning the number of the sample in the magnet, the "Sample in Magnet" popup

can also be used to remove the current sample or to replace the current sample.

sin Find sine value of an angle (C)

Syntax	<pre>sin(angle)<:n></pre>		
Description	Finds the sine value of an angle.		
Arguments	angle is the angle given in radians.		
	n is a return val the sine value ir	ue giving the sine of angle. The default is to display the status window.	
Examples	<pre>sin(.5) sin(val):sin_</pre>	val	
See also	User Programming		
Related	asin Fin	nd arc sine of number (C)	
	atan Fin	nd arc tangent of a number (C)	
	cos Fin	nd cosine value of an angle (C)	
	exp Fin	nd exponential value (C)	
	ln Fi	nd natural logarithm of a number (C)	
	tan Fin	nd tangent value of an angle (C)	

sine Find values for a sine window function (M)

Syntax	sine<(shift<,number_points<,domain>)>		
Description	Calculates appropriate values for parameters sb and sbs (if the domain argument is 'f2') or for parameters sb1 and sbs1 (if the domain argument is 'f1') in order to achieve a sine window function. The value of the parameter trace is used if the domain argument is not entered.		
Arguments	If shift is greater than 1, the sbs parameter is calculated as 2*sb/shift (sbs1 is calculated as 2*sb1/shift). sine(2) gives a "PI/2-shifted" sine window, i.e., cosine weighting. sine(3) gives a "PI/3" shifted sine window, etc. If shift is less than or equal to 1, an unshifted sine window is used (sbs='n' or sbs1='n').		
	number_points specifies the number of real points that the window function spans. The value of the window function for subsequent points is 0. number_points must be greater than 0 and a multiple of 2. The default is ni*2 if trace='f1', or np if trace='f2'.		
	domain is 'f1' or 'f2'. The default is the current setting of trace.		
See also	NMR Spectroscopy User Guide		
Related	npNumber of data points (P)sbSinebell const. in directly detected dimension (P)sb1Sinebell const. in 1st indirectly detected dimension (P)		

sbs	Sinebell shift const. in directly detected dimension (P)
sbs1	Sinebell shift const. in 1st indirectly detected dimension
	(P)
sinesq	Find values for a sine squared window function (M)
trace	Mode for n -dimensional data display (P)

sinebel1 Select default parameters for sinebell weighting (M)

DescriptionGenerates initial guess at good sinebell weighting parameters by setting
the sb and sb1 parameters to one-half the acquisition time and
turning off all other weighting. Use sinebell in absolute-value 2D
experiments only.See alsoNMR Spectroscopy User GuideRelatedpseudoSet default parameters for pseudo-echo weighting (M)
sbSinebell const. in directly detected dimension (P)sb1Sinebell const. in 1st indirectly detected dimension (P)

sinesq Find values for a sine-squared window function (M)

Syntax	sinesq<(shift<,number_points<,domain>)>		
Description	Calculates appropriate values for parameters sb and sbs (if the domain argument is 'f2') or for parameters sb1 and sbs1 (if the domain argument is 'f1') in order to achieve a sine-squared window function. The value of parameter trace is used if the domain argument is not entered.		
Arguments	shift sets the starting value for the window function. If shift is greater than 0, the starting value is given by sin p/shift; otherwise, if shift is less than or equal to 0, the starting value is 0. The default value is 0.		
	number_points specifies the number of real points that the window function spans. The value of the window function for subsequent point is 0. The number_points argument must be greater than 0 and a multiple of 2. The default is ni*2 if trace='f1', or np if trace='f2'.		
	domain is 'f1' or 'f2'. The default is the current setting of trace.		
See also	NMR Spectroscopy User Guide		
Related	ni Number of increments in 1st indirectly detected dimension (P)		
	np Number of data points (P)		
	sb Sinebell const. in directly detected dimension (P)		
	sb1 Sinebell const. in 1st indirectly detected dimension (P)		
	Sinebell shift const. In directly detected dimension (P)		

sineFind values for a sine window function (M)traceMode for *n*-dimensional data display (P)

size Returns the number of elements in an arrayed parameter (0)

Description	In MAGICAL programming, an operator that returns the number of elements in an arrayed parameter.		
Examples	r1 = size('d2')		
See also	User Programming		
Related	arraydim	Dimension of experiment (P)	
	typeof	Return identifier for argument type (0)	
	length	Determine length of a string (C)	

slfreq Measured line frequencies (P)

- Description Contains a list of measured line frequencies. In iterative spin simulation, a calculated spectrum is matched to the lines in the list. The spinll macro fills in slfreq from the last line listing or a mark operation. Use assign to make assignments between the measured lines and the calculated transitions. slfreq is a global parameter and is displayed by dla.
 - See also NMR Spectroscopy User Guide .

assign	Assign transitions to experimental lines (M)
cla	Clear all line assignments (M)
dla	Display spin simulation parameter arrays (M)
fitspec	Perform spectrum deconvolution (C)
mark	Determine intensity of a spectrum at a point (C)
spinll	Set up an slfreq array (M)
	assign cla dla fitspec mark spinll

slw Spin simulation linewidth (P)

Description Sets linewidth for individual transitions in the displayed spectrum. Only one linewidth is provided, so all transitions must be given the same linewidth. If the Set Params button is used in setting up spin simulation parameters, slw is automatically set to the measured linewidth of the tallest line displayed.

> slw is also the starting default linewidth for deconvolution calculations. This linewidth will be set automatically when deconvolution is operated using the menu mode and is bypassed if the

usemark command has been used in conjunction with two cursor input.

Values 0.01 to 1e6. The typical value is 1.

See also NMR Spectroscopy User Guide

Related usemark Use "mark" output as deconvolution starting point (M)

smaxf Maximum frequency of any transition (P)

Description	Sets the maximum frequency limit for the calculation of the final simulated spectrum. It should be set before the calculation is		
	performed. If the Set Params button is used in setting up spin simulation parameters, smaxf is initialized to $sp+wp$; which assumes that you have already expanded the region of the spectrum that you		
	wish to simulate before beginning the spin simulation process.		
Values	-1e10 to $1e10$, in Hz. The typical value is the maximum chemical shift + 50.		
See also	NMR Spectroscopy User Guide		
Related	sminf	Minimum frequency of any transition (P)	
	sp	Start of plot (P)	
	qw	Width of plot (P)	

sminf Minimum frequency of any transition (P)

Description	Sets the minimum frequency limit for the calculation of the final simulated spectrum. It should be set before the calculation is performed. If the Set Params button is used in setting up spin simulation parameters, sminf is initialized to sp, which assumes tha you have already expanded the region of the spectrum that you wish to simulate before beginning the spin simulation process.		
Values	-1e10 to 1e10, in Hz. The typical value is 0.		
See also	NMR Spectroscopy User Guide		
Related	smaxf sp wp	Maximum frequency of any transition (P) Start of plot (P) Width of plot (P)	

smsport Sample Management System serial port connection (P)

Description Sets which serial port on the host computer is connected to a Sample Management System (i.e., a sample changer). The value of smsport is

set using the Sample Changer Serial Port label in the Spectrometer Configuration window.

Values	'a' sets the	connection for serial port A. This value is the default.
	'b' sets the	connection for serial port B.
See also	VnmrJ Insta Guide	llation and Administration; NMR Spectroscopy User
Related	config	Display current configuration and possibly change it (M)

sn Signal-to-noise ratio (P)

Description	Sets a ratio for testing signal-to-noise. The testsn macro checks whether a signal-to-noise ratio equal to sn has been achieved.		
Values	Typical value is 35.		
See also	NMR Spectroscopy User Guide		
Related	dsn	Measure signal-to-noise (C)	
	getsn	Get signal-to-noise estimate of a spectrum (M)	
	testsn	Test signal-to-noise of a spectrum (M)	
	testct	Check ct for resuming signal-to-noise testing (M)	

solppm Return ppm and peak width of solvent resonances (M)

solppm:chemical_shift,peak_width		
Returns to the calling macro information about the chemical shift and peak spread of solvent resonances in various solvents for either ¹ H or 13 C, depending on the observe nucleus tn and the parameter solvent. This macro is used "internally" by other macros only.		
chemical_s	hift returns the chemical shift of the solvent in ppm.	
peak_width returns the approximate peak spread of solvent resonances.		
User Programming		
solvent tn	Lock solvent (P) Nucleus for observe transmitter (P)	
	solppm:chea Returns to the peak spread of ¹³ C, dependin This macro is chemical_si peak_width resonances. User Program solvent tn	

solvent Lock solvent (P)

 are not case- sensitive (e.g., solvent='C6D6' and solvent='c6d6' are identical)

The auto_dir macro now controls most of the automation features,
including setting the value of solvent.

Values Standard values in /vnmr/solvents include:

	Deuterium O D2O Acetone CD3COCD3 Benzene C6D6 DMSO	xide CDCl3 Cyclohexane C6Dl2 Toluene C6D5CH3 Acetic_Acid CD3C00D	MethyleneChloride MethylAlcohol-d4 CD2Cl2 CD3OD Chloroform
See also	NMR Spectro	oscopy User Guide	
Related	lastlk	Last lock solvent u	used (P)
	solvinfo	Retrieve information	on from solvent table (C)
	tof	Frequency offset f	or observe transmitter (P)

solvinfo Retrieve information from solvent table (C)

Syntax	<pre>solvinfo(solvent):\$chemical_shift,\$name</pre>		
Description	Retrieves solvent shift and solvent name from the solvent table.		
Arguments	solvent is the name of a solvent from the /vnmr/solvents file. This argument is not case-sensitive (e.g., 'c6d6' is the same as 'C6D6')		
	chemical_shift returns the chemical shift of the solvent, in ppm.		
	name returns the name of the solvent. The name returned will match the case of the letters (upper or lower) in /vnmr/solvents.		
Examples	solvinfo('acetone'):\$shift solvinfo('d2o'):\$shift,solvent		
See also	NMR Spectroscopy User Guide		
Related	lookupLook up words and lines from a text file (C)solventLock solvent (P)		

sort Sort real values of a parameter (M)

Syntax	<pre>sort(parametername<,sortType>:order,val</pre>
Description	Sorts the real values of a parameter. The sort macro is not used for parameters holding string values. The default behavior is to the array into values of increasing value. A sortType can be given to sort into descending order ('r').
	If only unique values are wanted, the 'u' sortType can be used. The 'ru' sortType given unique values in descending order.

The name of a parameter is the first argument to sort. Two return values hold the results of the sort. The first return value is an array containing the original indexes of the sorted array. The second return value gives the sorted array.

Examples With par=10,8,6,4,2 the display('par') command will show:
[1] = 10
[2] = 8
[3] = 6
[4] = 4
[5] = 2
The command sort('par'):\$order,\$val will set:
\$order=5,4,3,2,1
\$val =2,4,6,8,10

sp Start of plot in directly detected dimension (P)

Description	Low-frequency limit of the display or plotted region of the spectrum. sp is always stored in Hz, but can be entered in ppm by using the p suffix (e.g., sp=2p sets the start of plot to 2 ppm).		
See also	NMR Spectroscopy User Guide		
Related	sp1	Start of plot in 1st indirectly detected dimension (P)	
	sp2	Start of plot in 2nd indirectly detected dimension (P)	

sp1 Start of plot in 1st indirectly detected dimension (P)

Description	Analogous to the sp parameter except that sp1 applies to the first indirectly detected dimension of a multidimensional data set.		
See also	NMR Spectroscopy User Guide		
Related	sp sp2	Start of plot in directly detected dimension (P) Start of plot in 2nd indirectly detected dimension (P)	

sp2 Start of plot in 2nd indirectly detected dimension (P)

Description	Analogous to the sp parameter except that sp2 applies to the second indirectly detected dimension of a multidimensional data set.
See also	NMR Spectroscopy User Guide
Related	sp Start of plot in directly detected dimension (P)

spadd -add the current spectrum to the add/subtract experiment
spsub -subtract the current spectrum from the add/subtract
experiment

spmin - take the minimum of two spectra
spmax - take the maximum of two spectra
spadd<('new')> - add the current spectrum to the add/subtract
experiment
spsub<('new')> - subtract the current spectrum from the
add/subtract experiment
spadd<('trace',index)> - add the current spectrum to the "index"
element in the add/subtract experiment
spsub<('trace',index)> - subtract the current spectrum from the

"index" element in the add/subtract

spadd<('range'<,highfield,lowfield>)> - add a spectral range
to the add/subtract experiment

Description Non-interactive spectral addition and subtraction uses the ``spadd" and ``spsub'' commands. The last displayed or selected spectrum is added to ("spadd") or subtracted from ("spsub") the current contents of the add/subtract experiment. The spmin and spmax, instead of adding or subtracting, take the minimum and maximum, respectively, of the two spectra.

> The commands have two numeric arguments, both of which are optional. The first is the multiplier; the second is the shift. Thus the spectrum can be multiplied and/or shifted. (To shift a spectrum without multiplying it, use a multiplier of 1.0) A positive shift values moves the current spectrum to higher frequency, or to the right. A negative shift value moves the spectrum to lower frequency, or to the left.

> The commands have an optional 'range' argument, which, in turn has optional highfield and lowfield values. The range option adds only the specified range of data points to the add / subtract experiment. Points outside the range are treated as zeros. If the highfield and lowfield arguments are not given, they default to sp and sp+wp. That is, the currently displayed portion of the spectrum will be combined with the add / subtract experiment. If the range option is used, the normal limit that the Fourier numbers of the two data sets being combined is not enforced. The shift option may be used to position a section of a spectrum with a smaller fn value into the addsub spectrum. If used, the multiplier and shift arguments must precede the 'range' argument and its options highfield / lowfield arguments. If used, the shift argument is relative to the left edge of the addsub data set. The range option works for spadd, spsub, spmax, and spmin.

A multi-element add/subtract experiment may be created with the spadd or spsub command. The optional argument 'new' will create a new spectrum in the add/subtract experiment. For example, the commands clradd select(1) spadd from some experiment will create the add/subtract experiment with a single spectrum in it. If the next commands typed are select(2) spadd, then a single spectrum which is the sum of the original spectra one and two will be made in the add/subtract experiment. If, on the other hand, the commands select(2) spadd('new') were typed, then the add/subtract experiment will contain an array of two spectra corresponding to the original spectra one and two, respectively.

Individual spectra in a multi-elemnet add/subtract experiment may subsequently be added to and subtracted from. The spadd and spsub command without a 'trace' argument will add or subtract from the first spectrum in the add/subtract experiment. Adding the 'trace' argument followed by a required index number will select another spectrum to be the target of the add/subtract. For example, select(4) spadd('trace',6) will take the fourth spectrum from the current experiment and add it to the sixth spectrum in the add/subtract experiment. When using the 'trace' argument, that spectrum must already exist in the add/subtract experiment by using an appropriate number of spadd('new') or spsub('new') commands.

The results can be examined by joining the add/subtract experiment with the jaddsub macro and using the normal spectral display and plotting commands.

spmin takes the minimum of two spectra, considered point-by-point in an absolute-value sense. For example, if the two corresponding values are -2 and +3, the spmin spectrum will have -2; if the two values are +2 and -3, the spmin spectrum will have +2 at that point.

The function of spmin is to essentially select for common features within two spectra while eliminating features that are not common between them. In particular, if two CP/MAS spectra are obtained at different spin rates, the peaks stay in the same place (and hence the spmin spectrum also contains the same peaks), but the sidebands move. If spectrum 1 has baseline where spectrum 2 has sideband, and spectrum 2 has baseline where spectrum 1 has sideband, then the spmin spectrum will contain only baseline in these regions, eliminating the spinning sidebands...6101

spmax takes the maximum of two spectra, considered point-by-point in an absolute-value sense. For example, if the two corresponding values are -2 and +3, the spmax spectrum will have +3; if the two values are +2 and -3, the spmax spectrum will have -3 at that point.

Examples

spadd
spsub(0.75)
spadd('new')

spadd('trace',2) spadd('range') // And the spectral range between sp and sp+wp spadd('range', cr-delta, cr) // And the spectral range between the cursors See also NMR Spectroscopy User Guide Related add Add current FID to add/subtract experiment (C) addi Start interactive add/subtract mode (C) clradd Clear add/subtract experiment (C) Display a spectrum (C) jaddsub Join the add/subtract experiment (C) jexp Join existing experiment (C) select Select a spectrum without displaying it (C) Take minimum of two spectra in add/subtract experiment (C) Subtract current spectrum from add/subtract experiment spsub (C)

spcfrq Display frequencies of rf channels (M)

Description	Displays the parameters sfrq, dfrq, dfrq2, and dfrq3 with seven decimal points (to nearest 0.1) to provide the exact frequencies of each rf channel. The number of values displayed depends on numrfch.		
	Prior to VNM channel. The spcfrq.	IR version 4.3, spcfrq set the frequency of the observe parameter sfrq now sets the frequency instead of	
See also	NMR Spectroscopy User Guide		
Related	dfrq	Transmitter frequency of first decoupler (P)	
	dfrq2	Transmitter frequency of second decoupler (P)	
	dfrq3	Transmitter frequency of third decoupler (P)	
	numrfch	Number of rf channels (P)	
	setfrq	Set frequency of rf channels	
	sfrq	Transmitter frequency of observe nucleus (P)	

specdc3d 3D spectral drift correction (P)

Description Sets whether a 3D spectral dc correction occurs. The spectral dc correction is the last operation to be performed upon the data prior to forming linear combinations of the data, using the coefficients in the 3D coefficient file (coef), and then writing the data to disk. If specdc3d does not exist, it is created by the macro par3d.

Values A three-character string selected from 'nnn', 'nny', 'nyn', etc. Each character may take one of two values: n for no spectral dc correction along the relevant dimension, and y for spectral dc correction along

the relevant dimension. The first character refers to the f_3 dimension (sw, np, fn), the second character refers to the f_1 dimension (sw1, ni, fn1), and the third character refers to the f_2 dimension (sw2, ni2, fn2). The default is 'nnn'.

See also NMR Spectroscopy User Guide

Related	dc	Calculate spectral drift correction (C)
	fiddc3d	3D time-domain drift correction (P)
	fn	Fourier number in directly detected dimension (P)
	fn1	Fourier number in 1st indirectly detected dimension (P)
	fn2	Fourier number in 2nd indirectly detected dimension (P)
	ft3d	Perform a 3D Fourier transform (M)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	ni2	Number of increments in 2nd indirectly detected
		dimension (P)
	np	Number of data points (P)
	par3d	Create 3D acquisition, processing, display parameters (C)
	ptspec3d	Region-selective 3D processing (P)
	SW	Spectral width in directly detected dimension (P)
	sw1	Spectral width in 1st indirectly detected dimension (P)
	sw2	Spectral width in 2nd indirectly detected dimension (P)

spin Submit a spin setup experiment to acquisition (C)

Description	Regulates sample spinning according to the <i>parameter</i> spin, using the acquisition computer. It also sets rf frequency, decoupler status, and temperature.	
See also	NMR Spectroscopy User Guide	
Related	au Submit experiment to acquisition and process data (C)	
	change	Submit a change sample experiment to acquisition (M)
	ga	Submit experiment to acquisition and FT the result (C)
	go	Submit experiment to acquisition (C)
	lock	Submit an Autolock experiment to acquisition (C)
	sample	Submit change sample, autoshim experiment to acquisition
		(M)
	shim	Submit an Autoshim experiment to acquisition (C)
	spin	Sample spin rate (P)
	su	Submit a setup experiment to acquisition (M)

spin Sample spin rate (P)

Description Selects a regulated spin rate. The rate is changed when a sample is inserted or spin, go, ga, au, or sample are entered.

Values	0 indicates non-spinning operation.			
	5 to 39 a	5 to 39 are spinning rates.		
	'n' leave for regula	' leaves the spin rate at the currently used value and does not wait regulated spinning before performing acquisition.		
See also	NMR Spectroscopy User Guide			
Related	au	Submit experiment to acquisition and process data (C)		
	ga	Submit experiment to acquisition and FT the result (C)		
	go	Submit experiment to acquisition (C)		
	sample Submit change sample, Autoshim experiment to acquisition			
		(M)		
	sethw	Set values for hardware in acquisition system (C)		
	spin	Submit a spin setup experiment to acquisition (C)		

spinll Set up a slfreq array (M)

Syntax	spinll<('m	ark')>	
Description	Copies a list of frequencies to the slfreq parameter in iterative spin simulation and runs dla. This macro also clears previous line assignments.		
Arguments	'mark' is a keyword to copy the list of frequencies from the mark1d.out file to slfreq. The default is to copy the frequencies from the last line listing by nll or dll to the slfreq. Use the cursor and the mark button to place the lines to be assigned in mark1d.out. Enter mark('reset') to clear the file, and use nl to move the cursor to the center of a selected line.		
See also	NMR Spectroscopy User Guide		
Related	dla	Display line assignments (M)	
	d11	Display listed line frequencies and intensities (C)	
	mark	Determine intensity of the spectrum at a point (C)	
	nl	Position the cursor at the nearest line (C)	
	nll	Find line frequencies and intensities (C)	
	slfreq	Measured line frequencies (P)	

spinner Open the Spinner Control window (C)

Description Opens the Spinner Control window. This window has the following capabilities:

- Turn the sample spinner off.
- Turn the sample spinner on at a specified speed, in Hz.
- Enable spinner control from within an experiment using the spin parameter and the spin, go, ga, or au commands. This mode is the default.

- Alternatively, turn off experiment control of the sample spinner and allow only the Spinner Control window (and acqi and sethw) to set the spinning speed. This mode has the advantage that, often times, the spin parameter is different between experiments. Joining a different experiment and entering go can unexpectedly change the spinning speed. This alternate mode prevents this problem. In this mode, when a go, su, ga, or au is entered, the spin parameter is first set to the speed selected in the Spinner Control window and then the spin parameter is set to "Not Used."
- Select the style of spinner: low-speed style or a high-speed style. If the high-speed style of spinner (used for solids) is selected, the choice of setting the spinning speed or the air flow rate is provided. Setting the air flow rate is useful when setting up the solids spinning apparatus.

If the spinning speed is controlled only through the Spinner Control window, the action to be taken after a spinner error can be selected:

- Display a warning but continue acquisition.
- Stop acquisition and display a warning.

If experiment control of spinning speed is selected, these selections are faded because they are inoperative, and the selection of the action to be taken after a spinning speed error is provided by the parameter in.

See also NMR Spectroscopy User Guide

Related	acqi	Interactive acquisition display process (C)
	au	Submit experiment to acquisition and process data (C)
	change	Submit a change sample experiment to acquisition (M)
	ga	Submit experiment to acquisition and FT the result (C)
	go	Submit experiment to acquisition (C)
	in	Lock and spin interlock (P)
	lock	Submit an Autolock experiment to acquisition (C)
	sample	Submit change sample, autoshim experiment to acquisition
		(M)
	sethw	Set values for hardware in acquisition system (C)
	shim	Submit an Autoshim experiment to acquisition (C)
	spin	Sample spin rate (P)
	su	Submit a setup experiment to acquisition (M)

spins Perform spin simulation calculation (C)

Syntax spins<(options)>

Description Performs a spin simulation, using the current spin system parameters. Refer to the description of spsm for setting up the parameters. Use dsp to display the spectrum resulting from the simulation. The output file is spins.list in the current experiment. This file includes the calculated transitions ordered by frequency. Line assignments are required for the iteration. These consist of a list of observed frequencies, which is stored in the arrayed parameter slfreq, and the line assignments stored in the array clindex. spinll copies the frequencies from the last line listing by nll or dll into the parameter slfreq. The line listing can be from an observed spectrum or from the results of deconvolution. After spinll, line assignments are most easily made by entering assign. dla displays the assignments. Single assignments can also be made by assign(transition_number,line_number), where transition_number is the index of a transition and line_number is the index of the measured line. Setting the line_number argument to 0 deletes assignments. dla('long') produces an expanded display of assignments.

Be aware that spin simulation line numbers and line list line numbers are *not* the same. Conventional line lists produced by dll number the lines from left to right (low- to high-field). The spin simulation software numbers lines according to a more complicated scheme, and these numbers are rarely if ever in frequency order.

The parameters to be iterated are chosen by setting the string parameter iterate (e.g, iterate='A, B, JAB'). If several parameters have the same value due to symmetry, use

iterate='A, B, C, JAB, JAC=JAB'. This string sets the iterated parameter JAC to JAB during the iteration. JAB must be defined as an iterated parameter in the string before it can be used at the right side of the equal sign. Sets of parameters with up to six members may be set up in this way. The member in the set that is used on the right side of the equal sign must always come first in the parameter display (e.g., JAB=JAC would be wrong). A parameter is held constant during iteration if it is not included in the iterate string.

The command initialize_iterate sets iterate to iterate all spins not named X, Y, or Z and the associated coupling constants.

Following an iterative spin simulation, dga displays the new values of the coupling constants and chemical shifts. undospins restores a spin system as it was before the last iterative run. It returns the chemical shifts, coupling constants, and line assignments, making it possible to continue from this state with modified line assignments.

Note that major changes in the starting values of parameters may change the numbering of the energy levels and hence the line numbers. The line assignments would then be incorrect and would have to be reentered.

For a successful iteration, it is often necessary to keep some parameters fixed. For example, it is sometimes useful to alternately iterate couplings and shifts, keeping one group fixed while the other is iterated independently.

- Arguments The following variations of spins are available:
 - spins('calculate', 'energy') puts an energy-level table in the output file.
 - spins('calculate','transitions') puts a second table of transitions ordered by transition number in the output file.

S

- spins('display') and dsp are equivalent.
- spins('system','spinsystemname') and spsm('spinsystemname') are equivalent.
- spins('iterate') runs interactively to match experimental and calculated lines.
- spins('iterate','iteration') lists parameters after each iteration in the output file.
- spins('iterate'<, options>) provides for determining the chemical shifts and coupling constants to produce a spectrum that matches a table of observed lines. spins iterates until the rms (root-mean-square) error of the line matching meets a built-in test, unless it first reaches the value given by number_iterations. Iteration also stops if the rms error increases.
- Put multiple list options into the second argument, separated by a blank (e.g., spins('calculate','transitions energy')).

Examples spins

```
spins('calculate','energy')
spins('iterate')
```

See also NMR Spectroscopy User Guide

Related	assign	Assign transitions to experimental lines (M)
	clindex	Index of experimental frequency of a
		transition (P)
	dga	Display parameter groups (spin simulation)
		(C)
	dla	Display line assignments (M)
	d11	Display listed line frequencies and
		intensities (C)
	dsp	Display calculated spectrum (C)
	initialize_iterate	Set iterate to contain relevant parameters
		(M)
	iterate	Parameters to be iterated (P)
	niter	Number of iterations (P)
	nll	Find line frequencies and intensities (C)
	slfreq	Measured line frequencies (P)
	spinll	Set up slfreq array (M)
	spsm	Enter spin system (M)
	undospins	Restore spin system as before last iterative
		run (M)

split Split difference between two cursors (M)

Description Repositions the left-hand cursor halfway between its original position and the position of the other cursor. This macro is very useful for finding the center of a powder pattern: place the two cursors on the horns of the pattern and then enter split to give the center. See alsoNMR Spectroscopy User GuideRelateddeltaDifference of two frequency cursors (P)

spintype Spinner Type ((P)

Description This global parameter determines which spinner hardware is used. Values 'liquids' for low speed spinning of 5 and 10 mm liquids samples 'tach' for high speed spinning of 5 and 7 mm Jacobsen probes 'mas' for high speed spinning using standalone spinner 'nano' for spinning of nano probes 'none' for no spinner controller is present, e.g. imaging

splmodprepareUsed by the dosy macro to prepare data for the program SPLMOD

Syntax	splmodprepare
Applicability	VnmrJ 3.1
Description	splmodprepare takes a dosy_in file as created by dosy and creates the file dosy_splmod.in in a format suitable for the SPLMOD program (http://s-provencher.com/index.shtml).
See also	splmodread
	continread
	continprepare
	dosy

splmodread Used by the dosy macro to convert the output of the SPLMOD program into a form suitable for ddif

Syntax	splmodread
Applicability	VnmrJ 3.1
Description	splmodread takes the file dosy_splmod.out, created by SPLMOD (run by the splmodrun shell script from the dosy macro) and creates the files diffusion_display.inp and diffusion_spectrum in a suitable format for the ddif and sdp commands respectively.
See also	splmodread
	continread
	continprepare
	dosy

Description Takes the maximum of two spectra, considered point-by-point in an absolute-value sense. For example, if the two corresponding values are -2 and +3, the spmax spectrum will have +3; if the two values are +2 and -3, the spmax spectrum will have -3 at that point.

spmin Take minimum of two spectra in add/subtract experiment (C)

Description Takes the minimum of two spectra, considered point-by-point in an absolute-value sense. For example, if the two corresponding values are -2 and +3, the spmin spectrum will have -2; if the two values are +2 and -3, the spmin spectrum will have +2 at that point.

The function of spmin is to essentially select for common features within two spectra while eliminating features that are not common between them. In particular, if two CP/MAS spectra are obtained at different spin rates, the peaks stay in the same place (and hence the spmin spectrum also contains the same peaks), but the sidebands move. If spectrum 1 has baseline where spectrum 2 has sideband, and spectrum 2 has baseline where spectrum 1 has sideband, then the spmin spectrum will contain only baseline in these regions, eliminating the spinning sidebands.

- See also NMR Spectroscopy User Guide
 - RelatedaddiStart interactive add/subtract mode (C)spaddAdd current spectrum to add/subtract experiment (C)spsubSubtract current spectrum from add/subtract experiment (C)

spsm Enter spin system (M)

Syntax spsm(spin_system)

- Description Enables entry of the spin system for spin simulation and creates and initializes the appropriate parameters to describe the various chemical shifts and coupling constants. Chemical shifts can be entered for the X-nucleus, and the spectrum is calculated if that shift is in the window. Generally, however, it is not necessary to enter the X-nucleus chemical shift, and its value has no effect on the spectrum of the remainder of the spin system.
- Arguments spin_system is an alphanumeric string of upper-case letters for chemical shift and coupling constant parameters. Chemical shifts are stored in parameters A through Z, and the coupling constants are stored in the parameters starting with JAB and ending with JYZ. Different nucleus types are handled by using letters starting with A for

the first type, X for the second, and M for the third. Once created, these parameters are entered and modified in the usual way (e.g., A=78.5 JAC=5.6). Entry of chemical shifts in ppm is entered by using sfrq (e.g., B=7.5*sfrq).

Examples	spsm('AB')		
	spsm('A3B2')		
	spsm('AB2CMXY')		
See also	NMR Spectro	scopy User Guide	
Related	sfrq	Transmitter frequency of observe nucleus (P)	
	spins	Perform spin simulation calculation (C)	

spsub Subtract current spectrum from add/subtract experiment (C)

Syntax	 (1) spsub<(multiplier<,shift>)> (2) spsub('new') (3) spsub('trace',index) 		
Description	Performs non-interactive spectral subtraction. The last displayed or selected spectrum is subtracted from the current contents of the add/subtract experiment (exp5). A multi-element add/subtract experiment can be created using the 'new' keyword. Individual spectra in a multi-element add/subtract experiment can be subsequently subtracted from using the 'trace' keyword followed by an index number of the spectrum.		
Arguments	multiplier is a value to multiply each spectrum being subtracted from the add/subtract experiment (exp5). The normal range of multiplier would be +1 to -1 but is actually unlimited. The default is 1.0.		
	shift is the number of data points to shift each spectrum. A positive value shifts the spectrum being added to a higher frequency, or to the left. A negative value shifts the spectrum to a lower frequency, or to the right. The default is 0.		
	'new' is a keyword to create a new spectrum in the add/subtract experiment.		
	'trace' is a keyword to select the spectrum given by the index number argument (index) and subtract it from the add/subtract experiment. The default is to subtract from the first spectrum in the add/subtract experiment.		
	index is the index number of the spectrum to be used as a target in a multi-element add/subtract experiment.		
Examples	spsub spsub(.5,25) spsub('new') spsub('trace',2)		

See also	NMR Spectroscopy User Guide		
Related	clradd	clradd Clear add/subtract experiment (C)	
	ds	Display a spectrum (C)	
	jexp	Join existing experiment (C)	
	spadd	Add current spectrum to add/subtract experiment (C)	
	select	Select a spectrum without displaying it (C) Take minimum of two spectra in add/subtract experiment	
	spmin		
		(C)	
	sub	Subtract current FID from add/subtract experiment (C)	

sqcosine Set up unshifted cosine-squared window function (M)

Syntax	<pre>sqcosine<(<t1_inc><,t2_inc>)></t1_inc></pre>		
Description	Sets up an unshifted cosine-squared window function in 1, 2, or 3 dimensions. The macro checks whether the data is 1D, 2D, and 3D.		
Arguments	t1_inc is the number of t1 increments. The default is ni.		
	t2_inc is th	e number of t2 increments. The default is ni2.	
See also	NMR Spectroscopy User Guide		
Related	gaussian	Set up unshifted Gaussian window function (M)	
	ni Number of increments in 1st indirectly detected dimension (P)		
	ni2 Number of increments in 2nd indirectly detected dimension (P)		
	pi3ssbsq Set up pi/3 shifted sinebell-squared window functio (M)		
	pi4ssbsq Set up pi/4 shifted sinebell-squared window function		
	sqsinebell Set up unshifted sinebell-squared window function (M)		

sqdir Study queue directory (P)

Description	Specifies the full path directory where a study is stored. It is set when a new study is created.		
See also	NMR Spectroscopy User Guide, VnmrJ Automation, VnmrJ Walkup, VnmrJ Imaging User Guide		
Related	autodir	Automation directory absolute path (P)	
	globalauto	Automation directory name (P)	
	save	Save data (M)	
	sqname	Study queue parameter template (P)	
	startq	Start a chained study queue (M)	
	studyid	Study identification (P)	

sqname	Study queue parameter template (P)
xminit	Initialize an imaging study queue (M)

sgend End a study queue (M)

Description End a study queue. Usually called by other macros, and not used from the command line.

Related sqfilemenu Study queue file menu commands (M)

sgexp Load experiment from protocol (M)

Applicability	Imaging		
Description	Macro to load an experiment from a protocol.		
Syntax	<pre>sqexp(experiment <, 'save'>)</pre>		
	The first argument is the name of the experiment, and is required. The second argument is an optional keyword 'save'. If specified, it first saves parameter changes to the current experiment in the study queue before loading the parameters for the new experiment.		
Examples	sqexp('epidw')		
	<pre>sqexp('spuls','sa</pre>	ve')	
See also	VnmrJ Imaging User Guide		
Related	apptype	Application type (P)	
	execpars	Set up the exec parameters (M)	

sqfilemenu Study queue file menu commands (M)

Description	A macro to perform commands for the study queue operation. Usually the macro is called from the <i>study queue file menu</i> located below the study queue area, and not from the command line.		
See also	VnmrJ Imaging User Guide		
Related	cqinit	Initialize liquids study queue (M)	
	cqreset	Reset study queue parameters (M)	
	sqend	End a study queue (M)	
	sqreset	Reset study queue parameters for imaging (M)	
	xminit	Initialize an imaging study queue (M)	

S

Syntax sqLog(event<, arg>) - log automation events

sqLog:\$path - return log file path

Applicability VnmrJ 3.1, VnmrJ 3.2

Description The sqLog macro records specific events from a study queue. The messages and details of the logging are customizable with the editLog utility.

The sqLog facility will record the following events.

- SampleStart
- SampleEnd
- ExpStart
- ExpEnd
- ExpError

Each event recorded in the logfile may may be preceded by header information. This may include things like the date, time, user, etc. This header information is also customizable. The sqLog macro is very generic. It gets all of its details from a file written be the editLog utility. This file has the same name as the macro and is in the <appdir>/templates/vnmrj/loginfo directory. For example, the current sqLog file is:

Formatting statements for automation log files.

#

1Header Date: %MOC% %DAY% %YR% at %HR%:%MIN%:%SEC%, User: \$operator\$, Sample: \$samplename\$, 1SampleStart Start new sample at location \$loc\$. 1SampleEnd Finish sample at location \$loc\$\\#### 1ExpStart Experiment \$pslabel\$ started.

1ExpEnd Experiment \$pslabel\$ complete.

1ExpError Experiment error:

1ExpPrescan Prescan:

1File \$autodir\$/logfile

1Ifcondition (auto='y')

Lines starting with a hash mark (#) are comments. The first character of each non-comment line is a 1 or 0, indicating enabled or disabled. The rest of the first word, following the 1 or 0, is a keyword that is passed to the sqLog macro. The remainder of a line is the template for writing the log file. The template is passed to the chkname command for translation.

The File keyword defines where the log file will be saved. If this keyword is disabled, all of the sqLog event logging will be disabled. Disabling other keywords only disables that specific event or feature. The Ifcondition keyword allows the logging mechanism to make decisions as to whether to log the event. For example, in the case of

sqLog, we only log events during an automation run. Logging will occur only if the Ifcondition is true. A special keyword of "None" for the Ifcondition specifies no special conditions. That is, events are always logged. The sqLog macro is called from appropriate places in the software. It is called with the keyword as the first argument. A second, optional argument can also be passed. It will be appended to the log message generated by the keyword. For example, when called with ExpError, we pass the actual error message as the second argument.

geterror:\$err

sqLog('ExpError',\$err)

During an automation run, messages written to 'line3', which puts them into the "acqlog". If sqLog is called with no arguments but one return value, the pathname of the log file, defined by the File keyword, is returned.

As defined above, sqLog saves logging information only for automation runs. By changing the File attribute to your userdir directory, and setting the Ifcondition to None, all study queue activities will be logged, both automation and foreground. The log editor can handle menus of choices. Files in templates/vnmrj/loginfo with the same name as the keyword will be used to make menus of choices to select from within the editLog editor. Files prefixed with the name of the logging macro, for example sqLog will make a File menu specific for editLog('sqLog').

The logging macro, along with the editLog editor are very general and can be used to log other events. As an example, suppose one wants to monitor access to the VnmrJ program. A "loginLog" could be made as follows. Make a copy of the sqLog macro called loginLog. Add a loginLog file describing the events to logged to the

<appdir>/templates/vnmrj/loginfo. An example of such a file may be:

Formatting statements for login log files.

#

1Header Date: %MOC% %DAY% %YR% at %HR%:%MIN%:%SEC%, User: \$operator\$

1Login Login

1Logout Logout

1File \$systemdir\$/acqqueue/loginLog

1Ifcondition ((auto='n') and (jviewport=1))

The only remaining task is to place calls to the loginLog macro in various other macros. In this case, one might call loginLog('Login'):\$res from the bootup macro and loginLog('Logout'):\$res from the exit macro. If one wanted to monitor "operator" logins, one could and additional keywords such as operatorlogin and Operatorlogout to the above file and then call loginLog('Operatorlogin'):\$res from the operatorlogin macro and call loginLog('Operatorlogout'):\$res from the

macro and call loginLog('Operatorlogout'):\$res from the
operatorlogout macro.

sqmode Study queue mode (P)

Description A global parameter that specifies the study queue mode. It is used to determine if the study queue acquisition is chained or not. See also NMR Spectroscopy User Guide, VnmrJ Walkup Related startq Start a chained study queue (M) xmnext Find next prescan or next experiment in study queue (M) xmwexp Processing macro for end of acquisition in study queue (M)

sqname Study queue parameter template (P)

Description Stores a string in the global tree that determines where a study is stored. It is set from the *Save data setup* dialog in the *Utilities* menu. Dollar signs (\$) are used to delimit a string to search for a parameter to be used in the study file name. Percent signs (%) are used to delimit a numeric extension, e.g. %Rn%, or time specifications. Strings from the sampleinfo file are not used, since studies are created in foreground, not automation. Text not delimited by dollar signs or percent signs is copied from sqname without any changes.

If sqname does not start with a slash mark (/), the study is stored in the path given by autodir or globalauto; otherwise the name is used as is. A revision number is automatically appended. Values: If sqname is a null string, it defaults to %R2%, and the resulting study id is a two-digit revision number. The resulting path and file name must be accessible (with read-write permission) by that user.

Examples sqname='s_%DATE%_%R3%' studyid='s_20040501_001'
sqname='s_\$loc\$_' studyid='s_7_01'
sqname='r\$vrack\$z\$vzone\$/well\$loc\$%R0%'
studyid='r1z3/well16'

See also NMR Spectroscopy User Guide, VnmrJ Walkup

autodir	Automation directory absolute path (P)
autoname	Prefix for automation data file (P)
globalauto	Automation directory name (P)
sqdir	Study queue directory (P)
sqname	Study queue parameter template (P)
studyid	Study identification (P)
Svfname	Create path for data storage (C)
	autodir autoname globalauto sqdir sqname studyid Svfname

sqpars

Create study queue parameters for imaging (M)

Applicability Imaging

Description	A macro to create study queue parameters for imaging. Usually called		
	by other macros, and not used from the command line.		
See also	VnmrJ Imaging User Guide		
Related	fixpar Correct parameter characteristics in experiment (M)		

sqprotocol Macro to create protocols (M)

Applicability	Imaging
Description	A macro to create protocols for imaging applications. Called by the
	Make protocols dialogs in the Utilities menu.

sgreset Reset study queue parameters for imaging (M)

Applicability	Imaging
Description	Reset study queue parameters for imaging. Usually called by other
	macros, and not used from the command line.

sqrt Return square root of a real number (0)

Description	A operator in a real numbe Operator is r	n MAGICAL programming that returns the square root of er. A negative argument to sqrt is evaluated to 0.0. not used from the command line.
Examples	a = sqrt(b)	
See also	User Programming	
Related	asin	Find arc sine of number (C)
	atan	Find arc tangent of a number (C)
	cos	Find cosine value of an angle (C)
	exp	Find exponential value (C)
	ln	Find natural logarithm of a number (C)
	tan	Find tangent value of an angle (C)
	trunc	Truncates real numbers (0)
	typeof	Return identifier for argument type (O)

sqsavestudy Macro to save study parameters for imaging (M)

Applicability	Imaging
Description	A macro to save study parameters in the imaging study queue. Usually
	called by other macros, and not used from the command line.

See also	VnmrJ Imaging	User Guide
Related	acquire	Acquire data (M)
	sqend	End a study queue (M)
	studypar	Study parameters (P)

sqsinebel1 Set up unshifted sinebell-squared window function (M)

Syntax	sqsinebell<(<t1_inc><,t2_inc>)></t1_inc>		
Description	Sets up an unshifted sinebell-squared window function in 1, 2, or 3 dimensions. The macro checks whether the data is 1D, 2D, and 3D.		
Arguments	t1_inc is the number of t1 increments. The default is ni.		
	t2_inc is	the number of t2 increments. The default is ni2.	
See also	NMR Spectroscopy User Guide		
Related	gaussian	Set up unshifted Gaussian window function (M	
	ni	Number of increments in 1st indirectly detected dimension (P)	
	ni2	Number of increments in 2nd indirectly detected dimension (P)	
	pi3ssbsq	Set up pi/3 shifted sinebell-squared window function (M)	
	pi4ssbsq	Set up pi/4 shifted sinebell-squared window function (M)	
	sqcosine	Set up unshifted cosine-squared window function (M)	

srate Spinning rate for magic angle spinning (P)

Systems with solids module.		
Set to the spinning speed for magic angle spinning (MAS). srate must be correct for the pulse sequence set up by xpolar1 to run TOSS or dipolar dephasing correctly. If hsrotor='y', the measured spinning speed is reported in srate for systems that have rotor synchronization.		
0 to 10 ⁷ , in Hz.		
NMR Spectroscopy User Guide		
hsrotor xpolar1	Display rotor speed for solids operation (P) Set up parameters for XPOLAR1 pulse sequence (M)	
	Systems with Set to the spi be correct fo dipolar deph speed is repo- synchronization 0 to 10 ⁷ , in <i>NMR Spectro</i> hsrotor xpolar1	

sread Read converted data into VnmrJ (C)

Syntax sread(file<,template>)
Description	Reads 32-bit data files into VnmrJ. For Bruker data files in the AMX and AM formats, each file must first be converted using the convertbru command before sread can read the data in the file into VnmrJ.	
Arguments	file is the name of a file containing data converted using convertbru.	
	template is the full path of a parameter template file, but without appending the .par extension on the file name. The default is bruker.par. If no parameter template is specified and bruker.par cannot be found in the user or system parlib directory, sread aborts with an error message.	
Examples	<pre>sread('brudata.cv','/vnmr/parlib/bruker')</pre>	
See also	NMR Spectroscopy User Guide	
Related	convertbru Convert Bruker data (M,U)	

srof2 Calculate exact rof2 value for Cold Probes (M)

Applicability	Systems with Agilent, Inc. Cold Probes		
Description	Calculates the exact value needed for rof2 to result in a lp=0		
	condition for the given sw. Works with either $dsp='r'$ and $fsq='y'$		
	or with dsp='i'. Not compatible with gcomp.		
Related	dsp Type of DSP for data acquisition (P)		

rof2 Receiver gating time following pulse (P)

ss Steady-state transients (P)

Description	Sets the number of complete executions of the pulse sequence not			
	accompanied by data collection prior to the acquisition of the real data			
(sometimes known as <i>dummy scans</i>). If ss is positive, ss ste				
	transients are applied on the first increment only, and if ss is negative,			
	-ss steady-state transients are applied at the start of each increment.			
Values	'n', -32768 to 32767			
See also	NMR Spectroscopy User Guide; User Programming			

ssecho Set up solid-state echo pulse sequence (M)

Applicability	Systems with a solids module.
Syntax	ssecho
Description	Converts a standard two-pulse experiment to a ready-to-run solid-state NMR echo (SSECHO) pulse sequence.

ssecho1 Set up parameters for SSECH01 pulse sequence (M)

Applicability	System with a wideline solids module.
Description	Sets up a parameter set for the quadrupole echo pulse sequence SSECHO1.
See also	NMR Spectroscopy User Guide

Full bandwidth of digital filter to yield a filtered FID (P) ssfilter

Description	Specifies the full bandwidth of the digital filter applied to the original FID to yield a filtered FID for solvent subtraction. If ssfilter does not exist in the current experiment, enter addpar('ss') to add it. The command addpar('ss') creates additional time-domain solvent subtraction parameters ssfilter, sslsfrg, ssntaps, and ssorder.			
Values	<pre>'n', 10 to sw/2, in steps of 0.1 Hz. The default is 100 Hz. If ssfilter is set to a value and ssorder is set to some value, the zfs (zero-frequency) option of solvent subtraction is selected.</pre>			
	If ssfilter is set to 'n', ("Not Used"), both the lfs (low-freque suppression) and zfs options are turned off.			
See also	NMR Spectroscopy User Guide			
Related	addpar ft	Add selected parameters to the current experiment (M) Fourier transform 1D data (C)		
	parfidss	Create parameters for time-domain solvent subtraction (M)		
	ssntaps	Number of coefficients in the digital filter (P)		
	sslsfrq	Center of solvent-subtracted region of spectrum (P)		
	ssorder	Order of polynomial to fit digitally filtered FID (P)		
	SW	Spectral width in directly detected dimension (P)		
	wft	Weight and Fourier transform 1D data (C)		

Center of solvent-suppressed region of spectrum (P) sslsfrq

Description Specifies the location of the center of the solvent-suppressed region of the spectrum. If sslsfrq does not exist in the current experiment, enter addpar('ss') to add it. addpar('ss') also creates time-domain solvent subtraction parameters ssfilter, ssntaps, and ssorder.

Values 'n' (or 0) specifies solvent suppresses a region centered about the transmitter frequency. This is the default

Non-zero value shifts the solvent-suppressed region by sslsfrq Hz. Multiple regions may be suppressed by arraying the value of sslsfrq. Up to 4 values are allowed.

- See also NMR Spectroscopy User Guide
 - Related addpar Add selected parameters to the current experiment (M) parfidss Create parameters for time-domain solvent subtraction (M) ssfilter Full bandwidth of digital filter to yield a filtered FID (P) ssntaps Number of coefficients in the digital filter (P) ssorder Order of polynomial to fit digitally filtered FID (P)

ssntaps Number of coefficients in digital filter (P)

- Description Specifies the number of taps (coefficients) to be used in the digital filter for solvent subtraction. If ssntaps does not exist in the current experiment, enter addpar('ss') to add it. addpar('ss') also creates time-domain solvent subtraction parameters ssfilter, sslsfrg, and ssorder.
 - Values Integer from 1 to np/4. The default is 121. An odd number is usually best.

The more taps in a filter, the flatter the passband response and the steeper the transition from passband to stopband, giving a more rectangular filter.

For the lfs (low-frequency suppression) option, the default is suitable.

For the zfs (zero-frequency suppression) option, a value between 3 and 21 usually works better.

See also NMR Spectroscopy User Guide

Related	addpar	Add selected parameters to the current experiment (M)
	ft	Fourier transform 1D data (C)
	ni	Number of increments in 1st indirectly detected
		dimension (P)
	np	Number of points (P)
	parfidss	Create parameters for time-domain solvent subtraction
		(M)
	ssfilter	Full bandwidth of digital filter to yield a filtered FID (P)
	sslsfrq	Center of solvent-suppressed region of spectrum (P)
	ssorder	Order of polynomial to fit digitally filtered FID (P)
	wft	Weight and Fourier transform 1D data (C)

ssorder Order of polynomial to fit digitally filtered FID (P)

Description Specifies the order of the polynomial to fit the digitally filtered FID if the zfs (zero-frequency suppression) option is selected for solvent

subtraction. ssorder is not used if the lfs (low-frequency suppression) option is selected. If ssorder does not exist in the current experiment, enter addpar('ss') to add it. addpar('ss') also creates time-domain solvent subtraction parameters ssfilter, sslsfrq, and ssntaps.

The solvent subtraction option (zfs or lfs) is selected as follows:

- If ssorder and ssfilter are both set to values, zfs is selected.
- If ssorder='n' and ssfilter is set to a value, lfs is selected.
- If ssorder='n' and ssfilter='n', zfs and lfs are both turned off.

'n', integer from 1 to 20. The default is 'n'. Values

See also

NMR Spectroscopy User Guide Related addpar Add selected parameters to the current experiment (M) Create parameters for time-domain solvent subtraction parfidss (M) ssfilter Full bandwidth of digital filter to yield a filtered FID (P) sslsfrq Center of solvent-suppressed region of spectrum (P) ssntaps Number of coefficients in the digital filter (P) wft Weight and Fourier transform 1D data (C)

Stacking mode for processing and plotting arrayed spectra stack (M)

Syntax stack(mode)

- Description When processing and plotting arrayed 1D spectra, VnmrJ automatically determines if the *stacking mode* is horizontal, vertical or diagonal from the number of traces and the number of lines in the spectrum. If you do not want this automatic function (or it makes an undesirable decision), you can override it by placing the stack macro in the experiment startup macro or by calling stack before processing (or reprocessing) a spectrum. The macro autostack switches back to automatic determination of the stack mode by destroying the parameter stackmode.
- Arguments mode is one of the stacking modes 'horizontal', 'vertical', or 'diagonal'.
 - NMR Spectroscopy User Guide See also

Related autostack Automatic stacking for processing and plotting arrays (M) procarray Process arrayed 1D spectra (M) Plot arrayed 1D spectra (M) plarray stackmode Stacking control for processing (P)

stackmode Stacking control for processing arrayed 1D spectra (P)

- Description Controls whether stacking for processing arrayed 1D spectra is automatic or nonautomatic. The *automatic stacking mode* can be overridden by creating and setting stackmode in the startup macro or before calling procplot or procarray. The autostack macro switches back to automatic determination of the stack mode by destroying this parameter.
 - Values 'horizontal', 'vertical', or 'diagonal'.
 - See also NMR Spectroscopy User Guide
 - Related autostack Automatic stacking for processing and plotting arrays (M) procarray Process arrayed 1D spectra (M) procplot Automatically process FIDs (M) stack Fix stacking mode for processing and plotting arrayed spectra (M)

startq Start a chained study queue (M)

Description Start a chained acquisition for a study queue.

Related	sqmode	Study queue mode (P)
	xmnext	Find next prescan or next experiment in study
		queue (M)

status Display status of sample changer (C,U)

Applicability	Systems with an automatic sample changer.			
Syntax	<pre>status<(directory<,config_file>)> (From UNIX) status directory <config_file></config_file></pre>			
Description	Displays a status window with a summary of all experiments and a scrollable list of individual experiments. Individual experiments are selected by clicking anywhere on the experiment of interest. status updates as the state of an automation run changes. If an experiment finishes or a new experiment is added, the status display is updated			
Arguments	directory is the path to the directory where the done queue (doneQ) is stored. In the UNIX shell, a directory path is required. In VnmrJ, a directory path is optional. The default is the automation mode directory.			
	config_file is the name of a user-supplied file that customizes status for local use. Refer to the manual <i>User Programming</i> for details.			

Examples	(From VnmrJ) status			
	(From VnmrJ) status('/home/vnmr1/AutoRun_621')			
	(From UNIX) status /home/vnmr1/AutoRun_621 mystatus			
See also	VnmrJ Walkup; User Programming			
Related	autodir	Automation directory absolute path (P)		
	autoname	Prefix for automation data file (P)		
	enter	Enter sample information for automation run (C,U)		

std1d Apptype macro for Standard 1D experiments (M)

Applicability	Liquids		
Description	Perform the actions for Standard 1D protocols to set up, process, and plot experiments.		
See also	NMR Spectroscopy	User Guide, VnmrJ	Walkup
Related	apptype	Application type (P)	
	execpars	Set up the exec	parameters (M)

stdshm Interactively create a method string for autoshimming (M)

Syntax	stdshm
--------	--------

Description	Creates a method string to be used in adjusting the spinning controls
	z1, z2, z3, and z4 when a sample is changed. If non-spin controls also
	need adjusting, further shimming operations are required.

The method string is constructed in answer to questions about the sample length, the time available for shimming, and the solvent T_1 or, in FID shimming, the T_1 of the sample. In asking about sample height, stdshm assumes that z3 and z4 need adjusting only with short samples; therefore, select "sample height will vary" if z3 and z4 shimming is definitely wanted.

Try lock shimming first to see if it produces a satisfactory result. Lock shimming requires a much shorter shimming time than FID shimming and usually adjusts z1 and z2 just as well. If lock shimming is unsatisfactory, try FID shimming. Again, when z3 and z4 adjustment is required, lock shimming is faster, but FID shimming is more effective. stdshm displays the estimated shimming time, permitting revision when the time is too long.

To shim after running stdshm, enter method='std' (for lock shimming) or method='fidstd' (for FID shimming). Then enter shim or set the wshim parameter to shim before the start of acquisition.

Note that the command newshm is much like stdshm but that newshm provides more flexibility in making method strings

See also	NMR Spectroscopy User Guide	
Related	dshim Display a shim method string (M)	
	method	Autoshim method (P)
	newshm	Interactively create a shim method with options (M)
	shim	Submit an Autoshim experiment to acquisition (C)
	wshim	Conditions when shimming is performed (P)

sth Minimum intensity threshold (P)

Description	Intensity threshold above which transitions are printed and included in the simulated spectrum. Transitions whose intensity falls below this threshold are omitted from the simulation.		
Values	0 to 1.00. A typical value is 0.05.		
See also	NMR Spectroscopy User Guide		
Related	spins Perform spin simulation calculation (C)		
	spsm	Enter spin system (M)	
	th	Threshold (P)	

string Create a string variable (C)

Syntax	string(variable)
Description	Creates a string variable without a value.
Arguments	variable is the string variable to be created.
Examples	<pre>string('strvar1')</pre>
See also	User Programming

string2arrayFormats a String Variable into an Array

Description	Converts a string variable into an array.
Syntax	string2array('parameter'):\$array
Examples	string2array():\$S1
Related	<pre>array2string, array2csv, array2strsv, srtsv2array</pre>

strstr Sets ret to the starting position of the first occurrence of string2 in string1

Syntax strstr(string1,string2):ret,s1,s2 - find position of one string in another strstr(string1,string2,'last'):ret,s1,s2 - find last

position of one string in another

Applicability VnmrJ 3.1

Description This command sets ret to the starting position of the first occurrence of string2 in string1. The first character position is 1. This command returns 1 if string2 is empty. It returns 0 if string2 does not occur in string 1. Two additional values can be returned. These correspond to the segments of string1 which precede and follow string2, respectively. If string2 does not exist is string1, the two returned segments are both set to a null string. This command can be used in a variety of ways. The examples below demonstrate determination of the file extension. Also, it can reproduce the UNIX basename and dirname commands.

Arguments

Examples n1='/export/home/vnmr1/vnmrsys/data/studies/s_2002-04-10_00 1/data/sems_01.fid'

 $n2='/s_{2002-04-10_001/data/'}$

strstr(n1,n2):\$ret,\$s1,\$s2

\$ret will be set to 40

\$s1 will be set to parent of the studies directory

'/export/home/vnmr1/vnmrsys/data/studies'

\$s2 will be set to the file name of a saved data set.

'sems_01.fid'

The combined \$s1+n2+\$s2 will be equal to n1.

If a third optional 'last' argument is given, then strstr will find the last occurrence of string2 in string1. The return arguments are the same. This might be used to find the extension of a file name. For example,

n1='/export/home/vnmr1/vnmrsys/data/old.studies/s_2002-04-10_001/ data/sems_01.fid'

strstr(n1,'.'):\$ret,\$s1,\$s2

\$ret will be set to 36

\$s1 will be set to

'/export/home/vnmr1/vnmrsys/data/old'

\$s2 will be set to

'studies/s_2002-04-10_001/data/sems_01.fid'

However,

strstr(n1, '.', 'last'):\$ret, \$s1, \$s2

\$ret will be set to 74

\$s1 will be set to

'/export/home/vnmr1/vnmrsys/data/old.studies/s_2002-04-10_001/data /sems_01'

\$s2 will be set to the extension.

'fid'

To find the directory and basename of a file path, the following can be used.

strstr(n1, '/', 'last'):\$ret, \$s1, \$s2

This will set \$s1 to the directory

'/export/home/vnmr1/vnmrsys/data/old.studies/s_2002-04-10_001/data' and \$s2 will be the basename 'sems 01.fid'

strsv2array Formats a String Separated Variable into an Array

Description	Converts a string separated variable into an array.
Syntax	strsv2array('parameter'):\$array
Examples	strsv2array():\$R1
Related	<pre>array2string, array2csv, array2stringview, string2array</pre>

strtext Starting point for LP data extension in np dimension (P)

Description Specifies inclusively the complex time-domain data point at which LP (linear prediction) data extension (alteration) is to begin in the np dimension. Enter addpar('lp') to create strtext and other np dimension LP parameters in the current experiment.

Values 1 to np/2

```
See also NMR Spectroscopy User Guide
```

- Related addpar Add selected parameters to the current experiment (M) lpalg LP algorithm in np dimension (P) np Number of data points (P)
 - strtlp Starting point for LP calculation in np dimension (P)

strtext1 Starting point for LP data extension in ni dimension (P)

Description Specifies inclusively the complex time-domain data point at which LP (linear prediction) data extension (alteration) is to begin in the ni

dimension. Enter addpar('lp',1) to create strtext1 and other ni dimension LP parameters in the current experiment.

Values 1 to ni/2

See also	NMR Spectroscopy User Guide		
Related	addpar Add selected parameters to the current experiment (M		
	lpalg1	LP algorithm in ni dimension (P)	
	ni	Number of increments in 1st indirectly detected	
		dimension (P)	
	strtlp1	Starting point for LP calculation in ni dimension (P)	

strtext2 Starting point for LP data extension in ni2 dimension (P)

Description	Specifies inclusively the complex time-domain data point at which LP (linear prediction) data extension (alteration) is to begin in the ni2 dimension. Enter $addpar('lp', 2)$ to create strtext2 and other ni2		
	dimension	LP parameters in the current experiment.	
Values	1 to ni2/2		
See also	NMR Spectroscopy User Guide		
Related	addpar	Add selected parameters to the current experiment (M)	
	lpalg2	LP algorithm in ni2 dimension (P)	
	ni2	Number of increments in 2nd indirectly detected	
		dimension (P)	
	strtlp2	Starting point for LP calculation in ni2 dimension (P)	

strtlp Starting point for LP calculation in np dimension (P)

Specifies the first complex, time-domain data point to be used in Description calculating the complex linear prediction (LP) coefficients in the np dimension. If lpopt='b', the strtlp-th complex time-domain data point and the ensuing (2*lpfilt-1) data points are used in this calculation. If lpopt='f', the strtlp-th complex time-domain data point and the preceding (2*lpfilt-1) data points are used in this calculation. Enter addpar('lp') to create strtlp and other np dimension LP parameters in the current experiment. See also NMR Spectroscopy User Guide Related addpar Add selected parameters to the current experiment (M) lpalg LP algorithm in np dimension (P) LP coefficients to calculate in np dimension (P) lpfilt LP number of data points in np dimension (P) lpnupts LP algorithm data extension in np dimension (P) lpopt strtext Starting point for LP data extension in np dimension (P)

strtlp1 Starting point for LP calculation in ni dimension (P)

Description Specifies the first complex, time-domain data point to be used in calculating the complex linear prediction (LP) coefficients in the ni dimension. It functions analogously to strlp. Enter addpar('lp',1) to create strlp1 and other ni dimension LP parameters in the current experiment. See also NMR Spectroscopy User Guide

RelatedaddparAdd selected parameters to the current experiment (M)lpalg1LP algorithm in ni dimension (P)lpfilt1LP coefficients to calculate in ni dimension (P)lpnupts1LP number of data points in ni dimension (P)lpopt1LP algorithm data extension in ni dimension (P)strtext1Starting point for LP data extension in ni dimension (P)

strt1p2 Starting point for LP calculation in ni2 dimension (P)

Description Specifies the first complex, time-domain data point to be used in calculating complex linear prediction (LP) coefficients in the ni2 dimension. strtlp2 functions analogously to strlp. Enter addpar('lp',2) to create strtlp2 and other ni2 dimension LP parameters in the current experiment. See also NMR Spectroscopy User Guide Related addpar Add selected parameters to the current experiment (M) lpalg2 LP algorithm in ni2 dimension (P) LP coefficients to calculate in ni2 dimension (P) lpfilt2 LP number of data points in ni2 dimension (P) lpnupts2 LP algorithm data extension in ni2 dimension (P) 1popt2 strtext2 Starting point for LP data extension in ni2 dimension (P)

studyid Study identification (P)

Applicability	Liquids			
Description	Specifies the relative directory where a study is stored. In Walkup, it is relative to autodir. In imaging it is relative to global auto: It is			
	set when a n	ew study is created.		
See also	NMR Spectroscopy User Guide, VnmrJ Walkup			
Related	autodir Automation directory absolute path (P)			
	globalauto	Automation directory name (P)		
	sqdir	Study queue directory (P)		
	sqname	Study queue parameter template (P)		

studypar Study parameters (P)

Applicability Liquids, Imaging

S

Description A global parameter that contains the list of parameters saved with a study. If the parameter does not exist, it is created by cqsavestudy for liquids or sqsavestudy for imaging when a study is saved.

See also NMR Spectroscopy User Guide, VnmrJ Automation, VnmrJ Walkup, VnmrJ Imaging User Guide

Related cqsavestudy Macro to save study queue parameters (M) sqsavestudy Macro to save study parameters for imaging (M)

studystatus Study status (P)

Applicability	VnmrJ Walkup		
Description	The status of a study for a sample. The status is set from the status of the experiments within the study by the macro $cqsavestudy$.		
See also	VnmrJ Walkup		
Related	cqsavestudy studytime	Macro to save study queue parameters (M) Study time (P)	

studytime Determine start and end times for studies (P)

Syntax studytime('next'):\$ret1,\$ret2 \$ret1- when will the dayQ for next (future) study start \$ret2 - when will the nightQ for the next (future) study start studytime(location):\$ret1,\$ret2,\$ret3,\$ret4 \$ret1 - when will the dayQ for the given location (arg1) begin \$ret2 - when will the nightQ for the given location (arg1) begin \$ret3 - when will the dayQ for the given location (arg1) finish \$ret4 - when will the nightQ for the given locaiton (arg1) finish studytime('', rack, zone, location):\$ret1, \$ret2, \$ret3, \$ret 4 Same as the 2nd example. This syntax is usually used in context with location selection in the tray display.

su Submit a setup experiment to acquisition (M)

Description Sets up the system hardware to match the current parameters but does not initiate data acquisition. Typical uses of su are to change the system frequency in preparation for probe tuning, to change the sample temperature in advance of beginning an experiment (or after a variable temperature experiment is run), and to turn the decoupler on or off. If load='y', su can be used to set shim values. su also sets lock parameters (lockpower, lockgain, lockphase) and the field offset parameter (z0).

su does *not* delete any existing data in the current experiment (only go, ga, and au do that). Everything that su does is also done by go, ga, and au.

Shim DAC values are automatically loaded when the acquisition system boots up; if the acquisition system has been recently rebooted, su must be entered before acqi or gtune can be run.

See also NMR Spectroscopy User Guide

Related	acqi	Interactive acquisition display process (C)
	au	Submit experiment to acquisition and process data (C)
	change	Submit a change sample experiment to acquisition (M)
	ga	Submit experiment to acquisition and FT the result (C)
	go	Submit experiment to acquisition (C)
	load	Load status of displayed shims (P)
	lock	Submit an Autolock experiment to acquisition (C)
	lockgain	Lock gain (P)
	lockphase	Lock phase (P)
	lockpower	Lock power (P)
	qtune	Tune probe using swept-tune graphical tool (C)
	sample	Submit change sample, autoshim experiment to
		acquisition (M)
	shim	Submit an Autoshim experiment to acquisition (C)
	spin	Submit a spin setup experiment to acquisition (C)
	z0	Z0 field position (P)

sub

Subtract current FID from add/subtract experiment (C)

- Syntax (1) sub<(multiplier<, 'new'>)>
 (2) sub('new')
 (3) sub('trace', index)
- Description Subtracts the last displayed or selected FID from the current contents of the add/subtract experiment (exp5). lsfid and phfid can be used to shift or phase rotate the selected FID before it is subtracted from the data in add/subtract experiment. A multi-FID add/subtract experiment can be created by using the 'new' keyword. Individual FIDs in a multi-FID add/subtract experiment can subsequently be

subtracted by using the 'trace' keyword followed by the index number of the FID.

The add and sub commands use the cexp command to create the add-subtract experiment. They take the same return values as the cexp command. These can be used to suppress messages. See "cexp Create aVnmr experiment (M)" on page 176 for a description of the return values.

Arguments multiplier is a value that the FID is to be multiplied by before being subtracted from the add/subtract experiment (exp5). The default is 1.0.

'new' is a keyword to create a new FID element in an add/subtract experiment.

'trace' is a keyword to use the next argument (index) as the number of the FID to subtract from in an add/subtract experiment. The default is to subtract from the first FID in a multi-FID add/subtract experiment.

index is the index number of the FID to be used as a target in a multi-FID add/subtract experiment.

Examples sub

sub(0.75)
sub('new')
sub('trace',2)

See also NMR Spectroscopy User Guide

Related	add	Add current FID to add/subtract experiment (C)
	clradd	Clear add/subtract experiment (C)
	lsfid	Number of complex points to left-shift ni interferogram
		(P)
	phfid	Zero-order phasing constant for np FID (P)
	select	Select a spectrum without displaying it (C)
	spsub	Subtract current spectra from add/subtract experiment (P)

substr Select a substring from a string (C)

Syntax	substr('string',word_number):\$n1<,\$n2<,\$n3>>
	substr('string',index,length<,'new_string'>): n1<,\$n2<,\$n3>>
	<pre>substr('string',word_number,'delimiter', 'delimiter_char'): n1<,\$n2<,\$n3>></pre>
Description	This command picks a substring or word out of a string. It can also be used to replace (or delete) a set of characters from a string. It can also count the number of words in a string. It can also treat a string

where string is the string or a string variable

word_number is the number of the word to be selected

In this context, a word is defined as any string of characters separated by spaces " " or tabs "\t". Alternate delimiters can be supplied, as described below.

stringvar is a string variable.

If additional return arguments are given, the position of the first character of the word is returned and the number of characters of the word are returned.

If a fourth return value is given, it is set to the input string with the requested word removed. If the requested word is the first word, then preceding and trailing whitespace is also removed. If the requested word is not the first word, then only preceding whitespace is removed.

If word_number is larger than the number of words in the 'string', nothing will be returned.

Examples

```
substr('There are 10 samples to be run',4):n2
string n2 = 'samples'
substr('There are 10 samples to be run',4):n2,$f,$num
sets n2 = 'samples' $f = 14 and $num = 7
substr('There are 10 samples to be
run',4):n2,$f,$num,$rem sets n2 = 'samples' $f =
14 $num = 7 and $rem = 'There are 10 to be run'
substr('There are 10 samples to be
run',11):n2,$f,$num leaves n2, $f and $num
unchanged from what they were prior to the call to
substr.
```

If four arguments are supplied, and the third argument is the keyword 'delimiter', then the fourth argument is used as the delimiter characters. These separate the words searched for, replacing the default delimiters of space and tab " t". Its usage is as follows.

substr('string',word_number,'delimiter',delimiter):s
 tringvar

where string is the string or a string variable

word_number is the number of the word to be selected

'delimiter' indicates the next argument is a delimiter

delimiter is a set of characters used to separate words

stringvar is a string variable.

Examples

```
substr('This is;a phrase',2):n2 n2 = 'is;a'
substr('This is;a phrase',2,'delimiter',';\
t'):n2,$f,$num n2 = 'is' $f = 6 and $num = 2
substr('This is;a phrase',2,'delimiter',';\
t'):n2,$f,$num,$rem
```

n2 = 'is' \$f = 6 \$num = 2 and \$rem = 'This; a phrase'

In this case, note that whitespace preceding the word 'is' is removed but the trailing whitespace (a semi-colon) is not removed.

Strings that represent "Comma Separated Values", or CSV strings have special rules defining separate words. In this case, commas separate words, unless the comma is enclosed in double quotes. Also, adjacent commas indicate additional words that are null strings and a comma at the end of the string indicates an additional null word. Use these parsing rules by using substr with 'csv' as the third argument. Its usage is as follows.

substr('string',word_number,'csv'):stringvar
where string is the string or a string variable
word_number is the number of the word to be selected
'csv' selects the "CSV" rules to parse the string.

stringvar is a string variable.

Examples

<pre>substr('sample,loc,,"my water",',1,'csv'):n2</pre>	<pre>sample, in string n2 = 'sample'</pre>
<pre>substr('sample,loc,,"my water",',2,'csv'):n2</pre>	<pre>sample, in string n2 = 'loc'</pre>
<pre>substr('sample,loc,,"my water",',3,'csv'):n2</pre>	<pre>sample, in string n2 = ''</pre>
<pre>substr('sample,loc,,"my water",',4,'csv'):n2 in water"'</pre>	<pre>sample, in string n2 = '"my sample,</pre>
<pre>substr('sample,loc,,"my water",',5,'csv'):n2</pre>	<pre>sample, in string n2 = ''</pre>
<pre>substr('sample,loc,,"my water",',6,'csv'):n2</pre>	sample, in error, no word 6

Case 2. Count words in a string.

substr('string','wc'):\$cnt

where string is the string or a string variable

'wc' is a keyword to give the word count of the string

In this context, a word is defined as any string of characters separated by spaces " " or tabs "t". Alternate delimiters can be supplied, as described below.

\$cnt is number of words found in the string

Examples

substr('There are 10 samples to be run', 'wc'):r1 r1 = 7

If four arguments are supplied, and the third argument is the keyword 'delimiter', then the fourth argument is used as the delimiter

characters. These separate the words searched for, replacing the default delimiters of space and tab " \t ". Its usage is as follows.

substr('string','wc','delimiter',delimiter):\$cnt

where string is the string or a string variable

 $\ensuremath{\mathsf{'wc'}}$ is the keyword to count the number of words in string.

'delimiter' indicates the next argument is a delimiter

delimiter is a set of characters used to separate words

\$cnt is a number of words found in the string.

Examples

```
substr('This is;a phrase','wc'):$cnt
$cnt=3
```

substr('This is;a phrase','wc','delimiter',';\t'):\$cnt
\$cnt=4

Strings that represent "Comma Separated Values", or CSV strings have special rules defining separate words. In this case, commas separate words, unless the comma is enclosed in double quotes. Also, adjacent commas indicate additional words that are null strings and a comma at the end of the string indicates an additional null word. Use these parsing rules by using substr with 'csv' as the third argument. Its usage is as follows.

substr('string','wc','csv'):\$cnt

where string is the string or a string variable

'wc' is the keyword to count the number of words in string.

'csv' selects the "CSV" rules to parse the string.

\$cnt is a number of words found in the string.

Examples

```
substr('sample, loc,, "my sample, in
    water",','wc','csv'):$cnt $cnt=5
substr('a,b','wc','csv'):$cnt
    $cnt=2
substr('a,b,','wc','csv'):$cnt
    $cnt=3
substr(',a,b,','wc','csv'):$cnt
    $cnt=4
substr(',a,b,,','wc','csv'):$cnt
    $cnt=5
```

Case 3. Get basename of a file path name

substr('string','basename'):\$base<,\$ext>

where string is the string or a string variable to be interpreted as a file path name.

'basename' is a keyword to give the base file name portion of the string. In this context, the base file name are all characters following the final '/' character in the string. If an optional second return value is requested, the basename is further separated into a name and extension. The extension is defined as all characters following a final '.' in the name.

substr('string','basename'<,'suffixes'>):\$base<,\$ext>

An optional third argument is a list of suffixes. If the basename has a dot (.) in it, the extension will only be removed if it matches one on the supplied suffixes.

The examples will illustrate the behavior.

Examples

```
substr('/home/vnmr1', 'basename'):$base
  $base='vnmr1'
substr('/home/vnmr1', 'basename'):$base,$ext
  $base='vnmr1' $ext=''
substr('s2pul', 'basename'):$base
  $base='s2pul'
substr('s2pul','basename'):$base,$ext
  $base='s2pul' $ext=''
substr('s2pul.c','basename'):$base
  $base='s2pul.c'
substr('s2pul.c','basename'):$base,$ext
  $base='s2pul' $ext='c'
substr('s2pul.', 'basename'):$base
  $base='s2pul.'
substr('s2pul.', 'basename'):$base,$ext
  $base='s2pul.' $ext=''
substr('/home/vnmr1/mydata.fid', 'basename'):$base,$ext
                                         $base='mydata'
  $ext='fid'
substr('/home/vnmr1/mydata.s2pul', 'basename'):$base,$e
  xt
                                         $base='mydata'
  $ext='s2pul'
  substr('/home/vnmr1/mydata.s2pul', 'basename', '.fid
  .REC .rec'):$base,$ext
$base='mydata.s2pul' $ext=''
```

Case 4. Get parent directory of a file path name

substr('string','dirname'):\$dir<,\$base<,\$ext>>

where string is the string or a string variable to be interpreted as a file path name.

'dirname' is a keyword to give the directory file name portion of the string. In this context, the directory file name are all characters before the final '/' character in the string. If the string contains no '/' character, a '.' is returned as the directory name. If two or three return values are requested, the second and third values are returned as in the basename case above.

The following magical expression will reconstruct the string from the \$dir,\$base, and \$ext components.

```
if ($ext = '') then
   $orig = $dir + '/' + $base
else
   $orig = $dir + '/' + $base + '.' + $ext
endif
```

The input string and \$orig may not be identical, but when interpreted as path names, they will describe the same file. That is, if the input string is 's2pul', \$orig will be './s2pul'

```
substr('string','dirname'<,'suffixes'>):$dir<,$base<,$e
xt>>
```

An optional third argument is a list of suffixes. If the basename has a dot (.) in it, the extension will only be removed if it matches one on the supplied suffixes.

```
Examples
substr('/home/vnmr1','dirname'):$dir
   $dir='/home'
substr('/home/vnmr1', 'dirname'):$dir,$base
   $dir='/home' $base='vnmr1'
                                             $dir='.'
substr('s2pul','dirname'):$dir
   $base='s2pul'
substr('s2pul','dirname'):$dir,$base
                                             $dir='.'
   $base='s2pul'
substr('','dirname'):$dir
                                                 $dir='.'
substr('/home/vnmr1/mydata.fid','dirname'):$dir,$base,$
   ext
   $dir='/home/vnmr1' $base='mydata' $ext='fid'
substr('/home/vnmr1/mydata.s2pul','dirname'):$dir,$base
   ,$ext
   $dir='/home/vnmr1' $base='mydata' $ext='s2pul'
substr('/home/vnmr1/mydata.s2pul','dirname','.fid .REC
   .rec'):$dir,$base,$ext
   $dir='/home/vnmr1' $base='mydata.s2pul' $ext=''
Case 5. Extract specific characters from a string
```

If the string and two indexes are supplied, its usage is as follows:

substr('string',index,length):stringvar

where string is the string or a string variable

index is what character to start from.

length is the length of substring.

stringvar is a string variable.

Example

```
substr('abcdefg',2,3):n2 string n2 = 'bcd'
```

If the string, two indexes, and a replacement string are supplied, its usage is as follows:

substr('string',index,length,'newstring'):stringvar

where string is the string or a string variable

index is what character to start from.

length is the length of substring.

newstring is the string to substitute for the range of characters specified by index and length. To delete characters from string, set newstring to an empty string. See the example below.

stringvar is a string variable.

Examples

```
substr('abcdefg',2,3):n2 string n2 = 'bcd'
substr('abcdefg',2,3,'1234'):n2 string n2 =
'a1234efg'
substr('abcdefg',2,3,''):n2 string n2 = 'aefg'
substr('aa bb;cc dd',2,'delimiter',' ;\t'):n2
sets string n2 = 'bb'
n1 = 'There are 10 samples to be run'
substr(n1,4):n2,$f,$num
substr(n1,$f,$num,'experiments'):n3
sets n3 = 'There are 10 experiments to be run'
```

Case 6.

```
substr('string','find','word'):$num,$index,$len,$newstr
ing
```

where string is the string or a string variable

'find' is a keyword to use the next argument as the word to search for in the string. In this context, a word is defined as any string of characters separated by spaces " " or tabs "\t". Alternate delimiters can be supplied, as described below.

\$num is returns as the the number of the word found in 'string'. A 0 is returned if 'word' is not found in 'string'.

If additional return arguments are given, the position of the first character of the word is returned and the number of characters of the word are returned. If a fourth return value is given, it is set to the input string with the requested word removed. If the requested word is the first word, then preceding and trailing whitespace is also removed. If the requested word is not the first word, then only preceding whitespace is removed.

If the 'word' exists multiple times in 'string', the first occurrence is returned.

Example

```
substr('This was a
    test','find','was'):$num,$index,$len,$newstring
$num = 2
$index = 6
$len = 3
$newstring = 'This a test'
```

If five arguments are supplied, and the fourth argument is a keyword 'delimiter', the fifth argument is used as a delimiter to separate the words searched for, replacing the default delimiters of space and tab " \t ". Its usage is as follows.

```
substr('string','find','word','delimiter',delimiter)
```

See also User Programming

Related	length	Determine length of a string (C)
	string	Create a string variable (C)

suselfrq Select peak, continue selective excitation experiment (M)

Syntax	suselfrq			
Description	Sets up selective frequency pulse, power, and shape and continue with the selective excitation experiment. Used by Noesy1d, and TOCSY1D.			
See also	NMR Spectroscopy User Guide, VnmrJ Walkup			
Related	Noesyld Change parameters for NOESY1D experiment (M)			
	setselinv	Set up selective inversion (M)		
	setselfrqc	Select selective frequency and width (M)		
	TOCSY1D	Change parameters for TOCSY1D experiment (M)		

svdat Save data (C)

Syntax	svdat(file<,'f' 'm' 'i' 'b'>)
Description	Outputs current data from the current experiment to a file. Integer data is scaled when it is written.
Arguments	file is the name of the data file. The file is created in the current directory VnmrJ is in unless a full directory path is given. If a file of the same name already exists, the user will queried to overwrite the

file. If a fully qualified filename is not given, the file will be created in VnmrJ's current directory.

'f'|'m'|'i'|'b' defines how the data is to be written out: 'f' is 32-bit floating point, 'm' or 'i' is 16-bit integer scaled to 12 bits, and 'b' is 8-bit byte integer. The default is 'f'.

Floating point data is not scaled when written.

Integer data is scaled when written. A data value x is scaled as ax+b where:

```
a = (vs*grays1*numgray)/64.0
b = numgray*(0.5-(grays1*grayctr/64.0))
```

where numgray (see below) has a default of 4096 for 'm'

and 'i' formats and a default of 256 for the 'b' format, graysl has a default of 1, and grayctr has a default of 32.0.

To scale 16-bit integer data other than 12-bits, the global parameter numgray can be created using create (numgray, real, global) and set to the value 2^n , where n is the number of bits desired. For example, to scale to 15-bits, set numgray=32768.

The display parameters graysl and grayctr are used to save data files for ImageBrowser.

Examples svdat(rathead, 'b')

See also VnmrJ Imaging NMR

RelatedcreateCreate new parameter in parameter tree (C)grayctrGray level window adjustment (P)grayslGray level slope (contrast) adjustment (P)

svf Save FIDs in current experiment (M)

Syntax svf<(file<, 'nolog'><, 'arch'><, 'force'><, 'nodb'>)>

- Description Saves parameters, text, and FID data in the current experiment to a file. No data is removed from the current experiment; svf merely saves a copy of the data in a different file. You can enter rt to retrieve the complete data set, or enter rtp to retrieve parameters only.
- Arguments file is the name of the file, with the suffix .fid added, to be created to save the data. The default is the system prompts for a file name. You are warned if you attempt to overwrite a file that already exists. In fact, if data has been acquired with the file parameter set, the data does not need to be saved. It is already stored in a named file.

'nolog' is a keyword to not save the log file with the data. The default is to save the log file.

'arch' is a keyword to assume that the data goes to a database and appends to the (or creates a) doneQ file with information that can be used by the command status.

If force is given, you are not warned and the older parameter set is removed.

nodb is a keyword to prevent svp from adding information to a database. This prevention is useful if temporary parameter files are saved that will soon be removed.

Examples	svf				
	<pre>svf('/home/vnmr1/mydatafile')</pre>				
See also	NMR Spectroscopy User Guide				
Related	file	File name (P)			
	rt	Retrieve FID (M)			
	rtp	Retrieve parameters (M)			
	status	Display status of all experiments (C)			

svfdf Save FID data in FDF format (M)

Syntax svfdf(directory)

Description Saves raw data from the FID file of the current experiment as an FDF (Flexible Data Format) file. Data is saved in multiple files, with one trace per file. The files are named fid0001.fdf, fid0002.fdf, etc. The procpar file from the current experiment is also saved in the same directory.

The FDF file format is described in the manual *User Programming*. Note that the data is complex (FDF type="complex"), and the FDF ordinate = {"intensity", "intensity"}, indicating that each point consists of a pair of intensities. The FDF headers also contain the following special fields:

- •nfile gives the sequential number of this file in the series.
- ct is the value of the ct parameter. The data should be divided by ct to give the average signal intensity for one scan.
- scale gives the power of two scaling factor for the data. The data should be multiplied by 2^{scale} to give the true values.

Arguments directory_name is the directory in which to store the files. The extension .dat is appended to the given name.

Examples svfdf(curexp+'/raw')

See also User Programming

Related ct Completed transients (P)

svfdir Directory for non-study data (P)

Description	Specifies the directory where data is saved when not using a study in VnmrJ.
See also	NMR Spectroscopy User Guide, VnmrJ Walkup
Related	fidsave Save data (M)

save	Save data (M)
svfname	Filename parameter template for non-study data
	((P)

svfj Save FID in JCAMP-DX format (M)

Syntax svfj<(filename<,opt>)>

Applicability VnmrJ 3.1

Description "svfj" saves the current 1D FID in JCAMP-DX format. "svfj" creates temporary files "/vnmr/tmp/jdxfid.real" and "/vnmr/tmp/jdxfid.imag"; it calls two external C programs "listparam" and "jdxfid". Only a single FID (the current trace in the case of an arrayed experiment) is saved. "svfj" does not work with nf>1. Arguments "filename" is the name of the target file. If no filename is supplied, the software checks if "file" differs from "exp" (i.e., you have used "rt" to load the dataset from disk). If "file<>'exp"" and if the FID file is writable, then the JCAMP-DX data are saved as

"{file}/dx_name.dx", where "{file}" has ".fid" added, if necessary, and "dx_name.dx" is the "basename" part of "file" (minus the ".fid" extension). If "file='exp" or if the FID directory is not writable, the user is prompted for the filename.

The resulting ASCII file by default has a ".dx" extension, unless the specified filename has an alternative extension. (the default can be altered in the header of the macro). NOTE: ALL files created - whether they are FIDs, spectra or line lists - have the same (default) extension (the distinction between FIDs and spectra is made within the JCAMP-DX format); it is up to the user to avoid overwriting files! Where multiple arguments are specified / allowed, "filename"MUST be the first argument.

"opt" is an optional argument that permits controlling the compression in the data part of the JCAMP-DX file. Possible options and their effects are:

	tbl	fix	pac	sqz	dup	dif	difdup	Effect	Description
X,Y	Х	-	-	-	-	-	-	X,Y	
								list	
FIX	-	Х	-	-	-	-	-	X(YY)	readable
PAC	-	-	Х	-	-	-	-	X(YY)	packed
SQZ	-	-	-	Х	Х	Х	Х	X(YY)	squeezed
DIF	-	-	-	-	-	Х	Х	SQZ	differences
DUP	-	-	-	-	Х	-	Х	SQZ	DUP mode

Table 3.Option

The default mode / option is "difdup" which usually gives the best compression. For human-readable data use the "'fix" option, for programs expecting output in X Y format (one ordinate value per line)

use the "'tbl'" option. All format options comply with the JCAMP-DX format and should be usable.

Examples svfj svfj('myfid') svfj('myfid.jdx','dup') select(3) svfj('myfid_3') Related listparam list parameters in simple format (UNIX) writetrace write ascii file from phasefile (f1 or f2) trace (M)

Svfname Create path for data storage (C)

Applicability	Automation			
Syntax	Svfname:\$path			
	<pre>Svfname(name_template):\$path</pre>			
	<pre>Svfname(name_template,suffix):\$path</pre>			
	<pre>Svfname(name_template,suffix, excluded_suffix'):\$path</pre>			
	Svfname(name_template,suffix<,'excluded_suffix', <'keepspaces' 'replacespaces'>):\$path			
Description	Determines the name used to store data. This command provides the functionality of the autoname parameter without being in automation mode.			
	Svfname default naming command with alternate suffixes is svfname and the default directory is svfdir. Svfname does not read a sample info file. A suffix is specified as the second argument. Use a suffix of " to access ordinary files and directories. Arguments used with Svfname are constructed the same way arguments are constructed for autoname.			
	The name is prefixed with using the value of the parameter autodir or userdir+'/data/' if name_template is a relative path.			
	The default suffix is .fid.			
Arguments	svfname is default naming parameter.			
	svfdir is default directory parameter.			
	<pre>name_template (no quotes) is string that contains keywords separated by substitution specifiers to represent the data storage path. Substitution specifiers in this template are either a percent sign (%) or a dollar sign (\$). The keywords are obtained using % substitution specifiers or VNMR parameters using \$ substitution specifiers.</pre>			
	Percent sign (%) substitution specifier is used to scan for the text specified by keyword between the first percent sign in the template string and the next percent sign. The text specified by the keyword between the % substitution specifiers is passed to \$path.			
	The following percent substitutions (% keywords) for time and date are obtained from the system clock, not from the sample info file:			

Keyword	Format	Description
%DATE%	YYYYMMDD	4-digit year, 2-digit month, 2-digit day
%TIME%	HHMMSS	2-digit hour, 2-digit minute, 2-digit second
%YR%	YYYY	4-digit year
%YR2%	YY	2-digit year
%MO%	MM	2-digit month
%DAY%	DD	2-digit day
%HR%	HH	2-digit hour
%MIN%	MM	2-digit month
%SEC%	SS	2-digit second

Dollar sign (\$) substitution specifier is used with the Svfname command to interpreted a VNMR parameter and substitute the value of this parameter a suffix.

Numeric parameters are truncated and represented as a string with the form: <optional string>parameter value<optional

string>. The name_template, pw=\$pw\$usec, with vnmr parameter pw having a value of 12.3 produces pw=12usec01 which is appended to .fid (or .img) and passed to \$path.

String parameters cannot not contain any of the following characters: ' ', '!', '''', '\$', '&', '\', '', '', '', '>', '?', '\\', '[', ']', '^', '`', '{', '}', '|', ', '\0'

A comma separated excluded suffix list appends a string based on the suffixes and excluded suffixes to the path. Using the keyword

'replacespaces' uses underscores (_) in place of spaces ' ' in the resulting path name. The keyword 'keepspaces' retains spaces in the resulting path name.

'keepspaces' | 'replacespaces' is an optional argument (includes quotes) that uses either of the following keywords: replacespaces or keepspaces. The argument is accepted if the third argument is a list of suffixes. The action is the same as described for the third argument

Version number is specified by Rn where n is an integer from 0 to 9 (default 2), as follows:

n= 0	Description no revision digits are appended (all names must be		
1 to 9	uniquely constructed without the revision number is padded with I an n-digit number. If more places	γ constructed without these revision digits). number is padded with leading zeroes to form jit number. If more places are needed than		
>9 (more than one digit) no %Rn%	specified, more zeroes are used. Rnn is still used as a search stri file. %Rn% must be specified at t name_template string. The r always appended except if %R0% default of %R2% is used	es are used. I search string in the sampleinfo pecified at the end of the string. The revision digits are cept if %R0% is used. sed		
See als	o NMR Spectroscopy User VnmrJ Walkup	• Guide, VnmrJ Automation User Guide,		
Relat	ed autoname Determine automatio autoname Temple de (P)	es path for data storage during an n run (C) etermining the path where is data stored		

sqname	Study queue parameter template (P)
svfname	Specifies the filename template (P)

svfname Filename parameter template for non-study data (P)

Description Specifies the filename template where data is saved when not using a study in VnmrJ. The template is constructed using the same keywords and delimiter, dollar sign (\$) and percent sign (%), as autoname.

Examples If svfdir=userdir+'/data', the result from fidsave is: svfname='\$pslabel\$_\$tn\$_' -> userdir+'/data/Proton_H1_01.fid' svfname='%DATE%/t%TIME%%R0%' -> userdir+'/data/20040501/t113005.fid'

See also NMR Spectroscopy User Guide, VnmrJ Walkup

	—	
Related	fidsave	Save data (M)
	Svfname	Create path for data storage (C)
	sqname	Study queue parameter template (P)
	save	Save data (M)
	svfname	Filename parameter template for non-study data $((P)$

svimg Generate and Save images as FDF files. (macro)

Syntax	<pre>svimg('directory_name'[,'outfmt'])</pre>
Applicability	VnmrJ 3.1
Description	The "svimg" command generates images from the current experiment and saves them into the specified directory as Flexible Data Format (fdf) files. It will save one image or a number of images in the case of multislice experiments. Currently the specified directory is made in the user's data directory, and will be appended with a ".dat". Image files will be created under this directory as "image0001.fdf", "image0002.fdf", and so on. A "procpar" file will also be saved into this directory.
Arguments	The 'outfmt' parameter is an optional character which defines the type of image data. It can take two character values:
	• 'f' - Outputs the data in floating point format.
	• 'm' - Outputs the data in 12 bit integer values in 16 bit words.
	The default is 'f' (floating point) and currently ImageBrowser only accepts data in floating point values. The macro only saves images with the new imaging parameters that support oblique imaging. Unlike "svsis" the macro does not care about the name of the sequence. It does however format the header according to the following parameters.
	•seqcon - Sequence loop control flag
	•nD - Data dimension assumed to be 2.

- sfrq,dfrq Spectrometer frequency (MHz)
- lro Size of FOV for read out axis (cm)
- lpe Size of FOV for phase encode axis (cm)
- pro Position of image center on the read out axis (cm)
- ppe Position of image center on 2D phase encode axis (cm)
- thk Slice thickness (mm)
- •pss Slice position (cm)
- psi, phi, theta Euler angles determining direction.

The macro uses a Vnmr command "svsdfd" to dump the transformed data out to the data file. After dumping the headers out a unix shell command "fdfgluer" is called to glue the headers to the data. The "svsdfd" command dumps the data in such a way that the (0,0) coordinates are the first data point in the file.

NOTES: Modifications to the macro should be made in the user's maclib. The output values of the direction cosines may not be correct.

See also svsis

svllj Save peak listing in JCAMP-DX X,Y or X,Y,M format (M)

Syntax	<pre>svllj<(filename<,'all'><,'noll'>)></pre>
Applicability	VnmrJ 3.1
Description	"svllj" saves a peak listing in X,Y format. If a file "dept.out" exists in the current experiment, peak multiplicities are added to the output as well (X,Y,M format).
Arguments	"all" ("svllj" only) causes solvent signals to be included in the peak listing (multiplicity marked as "U" = unassigned)
	"noll" ("svllj" only) causes "svllj" NOT to re-evaluate the line listing - the contents of the parameters "llfrq" and "llamp" are used instead.
Examples	<pre>svllj svllj('myspectrum') svllj('myspectrum','all') svllj('myspectrum','noll') svllj('myspectrum','noll','all') select(3) svllj('myspectrum_3')</pre>
See also	svfj
Related	list parameters in simple format (UNIX) writetrace write ascii file from phasefile (f1 or f2) trace (M)

svlsj Save large dynamic range spectrum in JCAMP-DX format (M)

Syntax svlsj<(filename<,opt>)>

Applicability VnmrJ 3.1

Description "svlsj" is the same as "svsj", except that the spectrum is saved with 8 extra bits of digital precision ("svsj" saves spectra with 16-bit precision), for spectra with very large dynamic range.

Arguments "filename" is the name of the target file. If no filename is supplied, the software checks if "file" differs from "exp" (i.e., you have used "rt" to load the dataset from disk). If "file<>'exp" and if the FID file is writable, then the JCAMP-DX data are saved as "{file}/dx_name.dx", where "{file}" has ".fid" added, if necessary, and "dx_name.dx" is the "basename" part of "file" (minus the ".fid" extension). If "file='exp!" or if the FID directory is not writable, the user is prompted for the filename.

The resulting ASCII file by default has a ".dx" extension, unless the specified filename has an alternative extension. (the default can be altered in the header of the macro). NOTE: ALL files created - whether they are FIDs, spectra or line lists - have the same (default) extension (the distinction between FIDs and spectra is made within the JCAMP-DX format); it is up to the user to avoid overwriting files! Where multiple arguments are specified / allowed, "filename"MUST be the first argument.

"opt" is an optional argument that permits controlling the compression in the data part of the JCAMP-DX file. Possible options and their effects are:

	tbl	fix	pac	sqz	dup	dif	difdup	Effect	Description
X,Y	Х	-	-	-	-	-	-	X,Y	
								list	
FIX	-	Х	-	-	-	-	-	X(YY)	readable
PAC	-	-	Х	-	-	-	-	X(YY)	packed
SQZ	-	-	-	Х	Х	Х	Х	X(YY)	squeezed
DIF	-	-	-	-	-	Х	X	SQZ	differences
DUP	-	-	-	-	X	-	X	SQZ	DUP mode

Table 4. Option

The default mode / option is "difdup!" which usually gives the best compression. For human-readable data use the "'fix'" option, for programs expecting output in X Y format (one ordinate value per line) use the "'tbl'" option. All format options comply with the JCAMP-DX format and should be usable.

Examples sv

```
svlsj
svlsj('myspectrum')
svlsj('myspectrum','tbl')
select(3) svlsj('myspectrum_3')
```

See also	svfj svsj	
Related	listparam	list parameters in simple format (UNIX)
	writetrace	write ascii file from phasefile (f1 or f2) trace (M)

svp Save parameters from current experiment (M)

Syntax svp(file) <(file<, 'force'><, 'nodb'>)>

- Description Saves parameters from current experiment to a file. The parameter set can be retrieved with the rtp and rt macros. svp reflects any changes made in parameters up to the moment of entering svp, including acquisition parameters (unlike macro svf).
- Arguments file is the name of the file, with the suffix .par added, to be created to save the parameters. The default is the system prompts for a file name. You are warned if you attempt to overwrite a parameter set that already exists.

If force is given, you are not warned and the older parameter set is removed.

nodb is a keyword to prevent svp from adding information to a database. This prevention is useful if temporary parameter files are saved that will soon be removed.

 Examples
 svp('/vnmr/stdpar/P31')

 svp('/usr/george/testdata')

 See also
 NMR Spectroscopy User Guide

 Related
 rt

 Retrieve FID (M)

 rtp
 Retrieve parameters (M)

	~					~
sví	Save	FIDs	in	current	experiment	(M)

svpdp Compare workspace parameters to parameter file

DescriptionCompares current workspace parameters to the parameter file. Any
current workspace parameter values that are different from the
parameter file are updated in the parameter file.Syntaxsvpdp<(parlib)>Argumentstarget parameter library

svr Save secured REC data for VnmrJ SE

Syntax svr - save changes in currently loaded REC data to a new datdir inside the REC

	<pre>svr_as(path) - save data in current exp as a REC data</pre>
Description	svr_as and svr are used to save secured REC data for VnmrJ SE.
	The argument path is a full path or a name, with or without the suffix .REC. If the argument is only a name, svfdir will be used to determine the full path. The suffix .REC will be added if t is missing.
Examples	A REC data contains the following files (for example)

acqfil/ acqfil/procpar acqfil/text acqfil/cmdHistory acqfil/log acqfil/curpar acqfil/fid acqfil/global acqfil/checksum acqfil/auditTrail datdir001/ datdir001/phasefile datdir001/procpar datdir001/text datdir001/cmdHistory datdir001/curpar datdir001/data datdir001/global datdir001/checksum datdir001/auditTrail

svs

Save shim coil settings (C)

Syntax svs(file)<:status>

Description S

on Saves all shim coil settings except Z0 to a file.

Arguments

file is the name of a file for saving the shim coil settings. If the file name is an absolute path, svs uses it with no modifications. Otherwise, svs saves the shim in the first application directory for which it has write permission.

The svs command reports where it stored the shims, unless it is requested to return the status.

status is a return variable with one of the following values after ${\tt svs}$ finishes:

- •0 indicates svs failed to store shim file.
- 1 indicates svs stored the shim file, either as an absolute path or in the shims directory of the first application directory.
- •>=2 indicates svs stored the file in shims directory of the second, third, or later application directory.

Examples	svs('acetone')
	svs('bb10mm'):r1
See also	NMR Spectroscopy User Guide
Related	rts Retrieve shim coil settings (C)

svs Spin simulation vertical scale (P)

Description	Vertical scale	for simulated spectrum.		
Values	0 to 1e10. A typical value is 200.			
See also	NMR Spectroscopy User Guide			
Related	spins	Perform spin simulation calculation (C)		
	spsm	Enter spin system (M)		

svsis Generate and Save images as FDF files. (macro)

Syntax	<pre>svsis('directory_name'[,'outfmt'])</pre>
Applicability	VnmrJ 3.1
Description	The "svsis" command generates images from the current experiment and saves them into the specified directory as Flexible Data Format (fdf) files. It will save one image or a number of images in the case of multislice experiments. Currently the specified directory is made in the user's data directory, and will be appended with a ".dat". Image files will be created under this directory as "image0001.fdf", "image0002.fdf", and so on. A "procpar" file will also be saved into this directory.
Arguments	The 'outfmt' parameter is an optional character which defines the type of image data. It can take two character values:
	•'f' - Outputs the data in floating point format.
	• 'm' - Outputs the data in 12 bit integer values in 16 bit words.
	The default is f' (floating point) and currently ImageBrowser only accepts floating point data.
	The macro only saves images from the standard SISCO imaging sequences: "image", "shorte", "stecho", "multiecho", "csi2D", and "ssfp". However, it can be easily modified to produce images from users own sequences provided the sequences use standard SISCO parameters, slice select pulse shapes, and generate data in the same manner as the standard SISCO sequences.
	To easily modify the macro to use a user's sequence the user need only add a line similar to the following in the "Valid Sequences" section:
	\$k=\$k+1 \$seqfil[\$k]='t1image' \$seq[\$k]='ncsnn' \$thk[\$k]='image'
	The new sequence name is 'tlimage'. Its reconstruction properties are given by \$seq whose values are similar to the parameter "seqcon". "seqcon"'s characters are defined as follows:

S

- First character: multiecho looping
- Second charcter: multislice looping
- Third charcter: 2D phase encode loop
- Fourth character: 3D phase encode loop
- Fifth character: 4D phase encode loop The values of each character are:
- 'n': null loop
- 's': standard loop
- 'c': compressed loop

In this case 'ncsnn' is a standard 2D image with compressed multislice. The \$thk value is the slice thickness type defined by the type of acquisition which in this case is the standard 'image' sequence.

More detailed modifications can be made to the macro but it is left to the user to make these adjustments. The macro uses a Vnmr command "svsdfd" to dump the transformed data out to the data file. After dumping the headers out a unix shell command "fdfgluer" is called to glue the headers to the data. The "svsdfd" command dumps the data in such a way that the (0,0) coordinates are the first data point in the file.

NOTE: Modifications to the macro should be made in the user's maclib. See also svimg

svsj Save spectrum in JCAMP-DX format (M)

Syntax svsj<(filename<,opt>)>

Applicability VnmrJ 3.1

Description "svsj" saves the current 1D spectrum in JCAMP-DX format. "svsj" creates a temporary file "/vnmr/tmp/jdxspec"; it calls two external C programs "listparam" and "jdxspec". Only a single 1D trace (the current trace in the case of an arrayed experiment) is saved; "svsj" does not work on 2D data after "wft1d" or "wft2d", but 2D data can be treated as arrayed 1D data sets using "wft" / "ft", which again permits saving traces.

Arguments "filename" is the name of the target file. If no filename is supplied, the software checks if "file" differs from "exp" (i.e., you have used "rt" to load the dataset from disk). If "file<>'exp" and if the FID file is writable, then the JCAMP-DX data are saved as "{file}/dx_name.dx", where "{file}" has ".fid" added, if necessary, and "dx_name.dx" is the "basename" part of "file" (minus the ".fid" extension). If "file='exp!" or if the FID directory is not writable, the user is prompted for the filename.

The resulting ASCII file by default has a ".dx" extension, unless the specified filename has an alternative extension. (the default can be altered in the header of the macro). NOTE: ALL files created - whether

they are FIDs, spectra or line lists - have the same (default) extension (the distinction between FIDs and spectra is made within the JCAMP-DX format); it is up to the user to avoid overwriting files! Where multiple arguments are specified / allowed, "filename"MUST be the first argument.

"opt" is an optional argument that permits controlling the compression in the data part of the JCAMP-DX file. Possible options and their effects are:

	tbl	fix	pac	sqz	dup	dif	difdup	Effect	Description
X,Y	Х	-	-	-	-	-	-	X,Y	
								list	
FIX	-	Х	-	-	-	-	-	X(YY)	readable
PAC	-	-	Х	-	-	-	-	X(YY)	packed
SQZ	-	-	-	Х	Х	Х	Х	X(YY)	squeezed
DIF	-	-	-	-	-	Х	Х	SQZ	differences
DUP	-	-	-	-	Χ	-	Χ	SQZ	DUP mode

Table 5. Option

The default mode / option is "'difdup'" which usually gives the best compression. For human-readable data use the "'fix'" option, for programs expecting output in X Y format (one ordinate value per line) use the "'tbl" option. All format options comply with the JCAMP-DX format and should be usable.

Examples svsj

svsj('myspectrum')
svsj('myspectrum','fix')
select(3) svsj('myspectrum_3')

Related listparam list parameters in simple format (UNIX) writetrace write ascii file from phasefile (f1 or f2) trace (M)

svtmp Move experiment data into experiment subfile (M)

Syntax svtmp<(file)>

Description Moves the experiment data (parameters, FID, and transformed spectrum) from current experiment into a subdirectory inside curexp+'/subexp'. Unlike the macro cptmp, the experiment data is no longer accessible in the current experiment; only a copy of the parameters is still present.
Arguments file is the name of the subfile that receives the experiment data. The default name is either the transmitter nucleus (if seqfil='s2pul') or the pulse sequence name.
Examples svtmp

```
svtmp('cosy')
```

See also NMR Spectroscopy User Guide

Related	cptmp	Copy experiment data into experiment subfile (M)
	curexp	Current experiment directory (P)
	rttmp	Retrieve experiment data from experiment subfile
	seqfil	Pulse sequence name (P)

svxyj Save spectrum in JCAMP-DX X,Y format (M)

Syntax svxyj<(filename)>

Applicability VnmrJ 3.1

Description "svxyj" is similar to "svsj", except that the spectrum is written out in X,Y (2-column) format, with referenced X values and Y values directly in mm (the other JCAMP-DX formats use a simple integer X and Y values, the scaling and referencing information is stored in header fields. NOTE: most JCAMP-DX import software expects "svsj" / "svlsj" output. "svxyj" output uses no compression - the resulting files are much bigger than with any of the output options of the other JCAMP-DX conversion macros for full spectra.

Arguments "filename" is the name of the target file. If no filename is supplied, the software checks if "file" differs from "exp" (i.e., you have used "rt" to load the dataset from disk). If "file<>'exp'" and if the FID file is writable, then the JCAMP-DX data are saved as "{file}/dx_name.dx", where "{file}" has ".fid" added, if

necessary, and "dx_name.dx" is the "basename" part of "file" (minus the ".fid" extension). If "file='exp'" or if the FID directory is not writable, the user is prompted for the filename.

The resulting ASCII file by default has a ".dx" extension, unless the specified filename has an alternative extension. (the default can be altered in the header of the macro). NOTE: ALL files created - whether they are FIDs, spectra or line lists - have the same (default) extension (the distinction between FIDs and spectra is made within the JCAMP-DX format); it is up to the user to avoid overwriting files! Where multiple arguments are specified / allowed, "filename"MUST be the first argument.

Examples	svxyj		
	svxyj('myspectrum')		
	<pre>select(3) svxyj('myspectrum_3')</pre>		
Related	listparam list parameters in simple format (UNIX)		
	writetrace write ascii file from phasefile (f1 or f2) trace (M		

Spectral width in directly detected dimension (P)

Description Sets the total width of the spectrum to be acquired, from one end to the other. All spectra are acquired using quadrature detection. The spectral width determines the sampling rate for data, which occurs at a rate of 2*sw points per second (actually sw pairs of complex points per second). Note that the sampling rate itself is not entered, either directly or as its inverse (known on some systems as the *dwell time*).

If a value of sw is entered whose inverse is not an even multiple of the time base listed above, sw is automatically adjusted to a slightly different value to give an acceptable sampling rate.

To enter a value in ppm, append the character p (e.g., sw=200p).

If a DSP facility is present in the system (i.e., dsp='i' or dsp='r') and oversampling in the experiment has not been turned off by setting oversamp='n', then the oversampling factor will be recalculated.

Values Number, in Hz. The range possible is based on the system:

100 Hz to 500 kHz.

solids systems: up to 5 MHz.

See also NMR Spectroscopy User Guide

Related	dp	Double precision (P)
	dsp	Type of DSP for data acquisition (P)
	oversamp	Oversampling factor for acquisition (P)
	setlp0	Set parameters for zero linear phase (M)
	sw1	Spectral width in 1st indirectly detected dimension (P)
	sw2	Spectral width in 2nd indirectly detected dimension (P)
	sw3	Spectral width in 3rd indirectly detected dimension (P)

sw1 Spectral width in 1st indirectly detected dimension (P)

Description Analogous to the sw parameter except that sw1 applies to the first indirectly detected dimension of a multidimensional data set. The increment of the variable evolution time d2 is automatically calculated from sw1. The number of increments for this dimension is set by ni. To create sw1 in the current experiment, as well as ni and phase, enter addpar('2d').

See also NMR Spectroscopy User Guide

RelatedaddparAdd selected parameters to the current experiment (M)d2Incremented delay in 1st indirectly detected dimension (P)niNumber of increments in 1st indirectly detected dimension (P)phasePhase selection (P)swSpectral width in directly detected dimension (P)sw2Spectral width in 2nd indirectly detected dimension (P)sw3Spectral width in 3rd indirectly detected dimension (P)

S

SW
Spectral width in 2nd indirectly detected dimension (P) sw2

Description Analogous to the sw parameter except that sw2 applies to the second indirectly detected dimension of a multidimensional data set. The increment of the variable evolution time d_3 is automatically calculated from sw2. The number of increments for this dimension is set by ni2. To create sw2 in the current experiment, as well as d3, ni2, and phase2, enter addpar('3d').

See also NMR Spectroscopy User Guide

Related addpar Add selected parameters to the current experiment (M) Incremented delay for 2nd indirectly detected dimension (P) Number of increments in 2nd indirectly detected dimension (P) Phase selection for 3D acquisition (P) phase2 SW Spectral width in directly detected dimension (P) Spectral width in 2nd indirectly detected dimension (P) sw1 sw3 Spectral width in 3rd indirectly detected dimension (P)

Spectral width in 3rd indirectly detected dimension (P) sw3

- Description Analogous to the sw parameter except that sw3 applies to the third indirectly detected dimension of a multidimensional data set. The increment of the variable evolution time d4 is automatically calculated from sw3. The number of increments for this dimension is set by ni3. To create sw3 in the current experiment, as well as d4, ni3, and phase3, enter addpar('4d').
 - NMR Spectroscopy User Guide See also
 - Related addpar Add selected parameters to the current experiment (M) d4 Incremented delay for 3rd indirectly detected dimension (P) ni3 Number of increments in 3rd indirectly detected dimension (P) par4d Create 4D acquisition parameters (C) Phase selection for 4D acquisition (P) Spectral width in directly detected dimension (P) SW sw1 Spectral width in 1st indirectly detected dimension (P) Spectral width in 2nd indirectly detected dimension (P) sw2

System gradient coil (P) sysgcoil

Description Specially reserved string parameter that specifies which physical gradient set is currently installed, and allows convenient updating of

939

important gradient characteristics when one gradient set is interchanged for another. The value to sysgcoil is assigned to the parameter gcoil when joining experiments or retrieving parameter sets.

This parameter is set in the Spectrometer Configuration window to the name of the gradient set in use. Once set, it is then available to all experiments and to all users.

See also VnmrJ Installation and Administration; VnmrJ Imaging NMR

RelatedconfigDisplay current configuration and possibly change it (M)gcoilCurrent gradient coil (P)gmaxMaximum gradient strength (P)setgcoilAssign sysgcoil configuration parameter (M)

system System type (P)

- Description A global parameter that sets the basic type of system: spectrometer or data station. The value is set using the System Type label in the Spectrometer Configuration window.
 - Values 'spectrometer' is a spectrometer system (Spectrometer choice in Spectrometer Configuration window).

'datastation' is a system used as a data station (Data Station choice in Spectrometer Configuration window). Acquisition is not allowed in this setting.

- See also VnmrJ Installation and Administration
 - Related config Display current configuration and possibly change it (M) Console System console type (P)

systemdir VnmrJ system directory (P)

Description Contains path to VnmrJ system directory, typically /vnmr. The UNIX environmental variable vnmrsystem initializes systemdir at bootup. See also NMR Spectroscopy User Guide



Agilent VnmrJ 4 Command and Parameter Reference Guide

t1	T_1 exponential analysis (M)
t1s	${\cal T}_1$ exponential analysis with short output table (M)
t2	T ₂ exponential analysis (M)
t2s	T_2 exponential analysis with short output table (M)
tabc	Convert data in table order to linear order (M)
tan	Find tangent value of an angle (C)
tape	Read tapes from VXR-style system (M,U)
tape	Control tape options of files program (P)
target_bval	Adjust ${\tt gdiff}$ to achieve target b-value (M)
tcapply	Apply Table Conversion Reformatting to Data (C)
tchan	RF channel number used for tuning (P)
tcl	Send Tcl script to Tcl version of dg window (C)
tcclose	Table Convert Close (C)
tcopen	Table Convert Open (C)
temp	Open the Temperature Control window (C)
temp	Sample temperature (P)
tempcal	Temperature calculation (C)
tempcalc	Measure approximate sample temperature in Cold Probes (M)
testacquire	Test acquire mode (P)
testct	Check ct for resuming signal-to-noise testing (M)
testsn	Test signal-to-noise of a spectrum (M)
teststr	Find which array matches a string M)
text	Display text or set new text for current experiment (C)
textis	Return the current text display status (C)



textvi	Edit text file of current experiment (M)
th	Threshold (P)
th2d	Threshold for integrating peaks in 2D spectra (P)
thadj	Adjust threshold for peak printout (M)
time	Display experiment time or recalculate number of transients (M)
tin	Temperature interlock (P)
tlt	First-order baseline correction (P)
tmove	Left-shift FID to time-domain cursor (M)
tmsref	Reference 1D proton or carbon spectrum to TMS (M)
tn	Nucleus for observe transmitter (P)
tncosyps	Set up parameters for TNCOSYPS pulse sequence (M)
tndqcosy	Set up parameters for TNDQCOSY pulse sequence (M)
tnmqcosy	Set up parameters for TNMQCOSY pulse sequence (M)
tnnoesy	Set up parameters for TNNOESY pulse sequence (M)
tnroesy	Set up parameters for TNROESY pulse sequence (M)
tntocsy	Set up parameters for TNTOCSY pulse sequence (M)
Tocsy	Convert the parameters to a TOCSY experiment (M)
Tocsyld	Convert the parameter set to a Tocsy1d experiment (M)
tocsyHT	Set up the tocsyHT experiment (M)
tof	Frequency offset for observe transmitter (P)
tpwr	Observe transmitter power level with linear amplifiers (P)
tpwrf	Observe transmitter fine power (P)
tpwrm	Observe transmitter linear modulator power (P)
trace	Mode for <i>n</i> -dimensional data display (P)
traymax	Sample changer tray slots (P)

trfunc	Translates screen co-ordinates to hertz or centimeters depending upon the axis parameter
trfuncd	Translates a screen distance into centimeters in a real image
troesy	Set up parameters for TROESY pulse sequence (M)
trtune	Allows the user to view multiple tuning traces apparently simultaneously
trunc	Truncate real numbers (0)
tshift	Adjust tau2 to current cursor position (M)
tugain	Amount of receiver gain used by qtune (P)
tune	Assign a frequency to a channel for probe tuning (C)
tunehf	Tune both H1 and F19 on an HFX probe (M)
tunematch	Default match target, in percent of optimum (P)
tunemethod	Method to use for tuning (P)
tuneResult	Message indicating how well the tuning succeeded (P)
tunerp	A pulse sequence for pulse tuning through the directional couplers in the VnmrJ display
tunesw	Width of the tuning sweep in Hz (P)
tupwr	Transmitter power used in tuning (P)
typeof	Return identifier for argument type (0)

t1 T₁ exponential analysis (M)

Description Processes data obtained using an array of values of the parameter d2 for a T_1 experiment. It runs expfit, which does an exponential curve fitting that determines the value of T_1 . The output is matched to the equation:

M(t) = (M(0) - M0) * exp(-t/T1) + M0

where M0 is the equilibrium Z magnetization and M(0) is the magnetization at time zero (e.g., immediately after the 180° pulse for an inversion recovery T_1 experiment). Notice that this equation will fit inversion recovery data (for which M(0) is approximately equal to -M0) or saturation recovery data (for which M(0) is 0).

The required input is the file fp.out from fp and the values of the arrayed parameter. The T_1 analysis is done for all the peaks listed in fp.out. Peaks are selected for analysis by entering

fp(index1, index2, ...) before running the analysis. The output file

is the analyze.list in the current experiment. The file analyze.out is used by expl to display the results. The output of the analysis program shows T_1 and its standard deviation, but does not explicitly show M(0), M0, or their standard deviations. The M(0) and M0 values can be found in "raw" form in analyze.out in the current experiment, but their standard deviations are not part of the program output.

- See also NMR Spectroscopy User Guide
 - Relatedd2Incremented delay in 1st indirectly detected dimension (P)expfitMake least squares fit to polynomial or exponential curve (C)fpFind peak heights (C)t1s T_1 exponential analysis with short output table (M)t2 T_2 exponential analysis (M)t2s T_2 exponential analysis with short output fable (M)

tls T_1 exponential analysis with short output table (M)

DescriptionPerforms the same analysis as t1 but produces a short output table
showing only a summary of the measured relaxation times.See alsoNMR Spectroscopy User Guide
Related
t1 T_1 exponential analysis (M)

t2 T₂ exponential analysis (M)

Description Processes data obtained using an array of values for the base time parameter bt for a T_2 experiment. It runs expfit, which does an exponential curve fitting that determines the value of T_2 . The output is matched to the equation:

 $M(t) = (M(0) - M(inf))^* exp(-t/T2) + M(inf)$

where M(0) is the magnetization at time zero (i.e., the full magnetization excited by the observe pulse) and M(inf) is the xy-magnetization at infinite time (zero unless the peak is sitting on an offset baseline).

The required input is the file fp.out from fp and the values of the arrayed parameter. The T_2 analysis is done for all the peaks listed in fp.out. Peaks are selected for analysis by entering

fp(index1, index2,...) before running the analysis. The output file is the file analyze.list in the current experiment. The file analyze.out is used by exp1 to display the results. The output of the analysis program shows T_2 and its standard deviation, but does not explicitly show M(0), M(inf), or their standard deviations. The M(0) and M(inf) values can be found in "raw" form in analyze.out in the current experiment, but their standard deviations are not part of the program output.

See also NMR Spectroscopy User Guide

Related expfit Make least squares fit to polynomial or exponential curve (C)

- fp Find peak heights (C)
- t1 T_1 exponential analysis (M)
- tls T_1 exponential analysis with short output table (M)
- t2s T_2 exponential analysis with short output fable (M)

t2s T_2 exponential analysis with short output table (M)

Description	Performs the same analysis as t_2 but produces a short output table showing only a summary of the measured relaxation times.
See also	NMR Spectroscopy User Guide
Related	T_2 T_2 exponential analysis (M)

tabc Convert data in table order to linear order (M)

Syntax tabc<(dimension)>

Description Converts arbitrarily ordered data obtained under control of an external AP table to linear monotonic order, suitable for processing in VnmrJ. The data must have been acquired according to a table in the tablib directory.

Imaging and other 2D experiments are normally acquired so that the order of the incremented acquisition parameter, such as the phase-encode gradient, is linear and monotonic. For a standard imaging experiment, this linear order means that the phase-encode gradient progresses from a starting negative value monotonically up through zero to a positive value (e.g., -64, -63, -62, ..., -1, 0, 1, ..., 62, 63). The ft2d program assumes this structure in its operation.

Data from table-driven 2D pulse sequences is used by entering tabc only once before normal 2D processing and/or parameter storage. In this situation, tabc takes no arguments and is executed by entering tabc in the command window. A simple check is done by tabc to prevent it from being executed more than once on the same data set.

2D data is expected to be in the standard VnmrJ format, but if the 2D data is in the compressed format, setting dimension to 1 converts the data. tabc supports all 2D data types recognized by VnmrJ: arrayed, compressed multislice, and arrayed compressed multislice,

3D data is expected to be in the compressed/standard format, in which there are ni standard 2D planes of data (the third dimension), each consisting of nf compressed FIDs (the second dimension). Setting dimension to 3 reorders 3D data acquired with an external table.

tabc reads the file fid in the acqfil subdirectory of the current
experiment. Before the data is reordered, this file is written to the file
fid.orig in the same acqfil directory. If for any reason tabc fails
or results in an unpredictable or undesired transformation, the original
raw data can be recovered by moving fid.orig back to fid. To gain
more disk space, you can delete fid.orig after you are satisfied that
conversion is successful.

Use tabc on saved data that has been loaded into an experiment or on data in an experiment that has just been acquired but not yet saved. In the first case, converted data must be resaved for the saved data set to reflect conversion.

tabc requires that data must have the same number of "traces" as the table elements. It does not support any of the advanced features of table expansion (e.g., the entire table must be explicitly listed in the table file), and expects to find only one table in a file; whether the table is t1 or t60 is unimportant.

Arguments dimension specifies the type of data to be converted: 1 for 2D compressed data, 2 for 2D standard data, or 3 for 3D compressed/standard data. The default is 2.

Examples tabc tabc(1) tabc(3)

See also VnmrJ Imaging NMR

Related flashc Convert compressed 2D data to standard 2D format (C) ft2d Fourier transform 2D data (C) ni Number of increments in 1st indirectly detected dimension (P) nf Number of FIDs (P)

tan Find tangent value of an angle (C)

Syntax	tan(angle)	<:n>
Description	Finds the tangent of an angle.	
Arguments	angle is an angle, in radians.	
	n is the retu display the t	rn value giving the tangent of angle. The default is to angent value in the status window.
Examples	<pre>tan(.5) tan(val):tan_val</pre>	
See also	User Programming	
Related	atan	Find arc tangent value of a number (C)
	COS	Find cosine value of an angle (C)
	exp	Find exponential value of a number (C)
	ln	Find natural logarithm of a number (C)
	sin	Find sine value of an angle (C)

tape Read tapes from VXR-style system (M,U)

Description Displays the contents of a VXR-style (Gemini, VXR-4000, or XL) 9-track tape for use with VnmrJ or reads one or several files from the tape into the current directory. Note that the *write* option is not supported (i.e., VnmrJ only *reads* tapes in a VXR-style format and does not write to a tape).

Arguments device is the tape drive device name. The default value is /dev/rst8. For AIX systems, device should be /dev/rmt0.If the default value is not set properly or another device name is wanted, be sure to type -d and a space before the device name you want to input.

type is the type of tape to be accessed. -q' or -s' select the 1/4-inch tape unit ("streaming" or cartridge tape); this is the default. -9', -h', or -n' select the 1/2- inch tape unit (open reel tape drive).

option is one of the following:

- 'help' is a keyword to display help on the use of the system.
- 'cat' is a keyword to display a catalog of files on tape.
- 'read' is a keyword to read one or more files. This option requires that the files be listed as the next argument.
- 'rewind' is a keyword to rewind tape (1/2-inch tape only).
- 'quit' is a keyword to release the tape drive (1/2-inch tape only).

file1, file2, ... are the names of one or more files to be read. Wildcard characters (* and ?) can be used.

- Examples tape('cat')
 tape('-h','read','mydata')
 tape -h read mydata
 tape -d /dev/rmt/01b read mydata
 - Related decomp Decompose a VXR-style directory (C) vxr_unix Convert VXR-style text files to UNIX format (M,U)

tape Control tape options of files program (P)

Description Defines device that files program accesses when it is instructed to read or write to a tape. The parameter tape is in the user's global parameter tree.

Values Name of a device. The default device is /dev/rst8. If tape does not exist or is set to the null string (two single quotes with no space between), files uses its default device value. Notice that different

computers define tape drives differently. For VnmrSGI, tape='/dev/tapens' is appropriate. For Solaris, tape='/dev/rmt/0mb'.

Related files Interactively handle files (C)

target bval Adjust gdiff to achieve target b-value (M)

Applicability Syntax	<u>Imaging Systems</u> target_bval(value)
Description	This macro iteratively adjusts gdiff and calls the sequence $(go('check'))$ to achieve the target b-value. The sequence is evoked because the contributions from the imaging gradients must be taken into account backwards calculation of b is not possible because the relationship between gdiff and b-value is not simple. The macro defaults to getting within 1 s/mm2 of the target or maximum of 20 iterations and exits if either condition is met.
Arguments	value, the target b-value in s/mm2.
Examples	target_bval(1000)
See also	VnmrJ Imaging User's Guide

Apply Table Conversion Reformatting to Data (C) tcapply

Syntax	tcapply	[<filename>]</filename>
--------	---------	--------------------------

Applicability VnmrJ 3.1

Description "tcapply" rearranges the spectra in a 2D dataset that reside in the current datafile. Using values from an AP table, it arranges the spectra corresponding to the value in the AP table from low value to high value. The values may have already been read in by the "tcopen" command or if the optional <filename> argument has been provided the values will be read in from \$vnmruser/tablib/<filename>.

> As mention before, this command uses spectra from the current datafile; which means that a "ftld" should have been done on the data before using this command. To give an example, for a standard imaging experiment the phase encode gradients will progress from a starting negative value monotonically up through zero to a positive value, e.g.:

 $-64, -63, -62, \dots, -1, 0, 1, \dots, 62, 63.$

It is possible to acquire the equivalent data in non-monotonic order, either by explicitly coding the desired progression into a pulse sequence, or by using an external AP table to control the order. In either case, "ft2d" will not be able to properly process the resulting data. "tcapply" and "tabc" are functions which reconstruct a properly ordered data set from any arbitrarily ordered data which has been acquired under control of an external AP table. The data must have

	been acquired different betw the first dime memory and	d according to a table in the "tablib" directory. The veen "tcapply" and "tabc" is that "tcapply" works on ension transformed spectra residing in Vnmr's data "tabc" works on and changes the raw data in the fid file.
Arguments	'filename' o resides in \$v	ptional argument specifying the AP table to be read which nmruser/tablib/ <filename>.</filename>
Examples	<pre>ftld(2) tcapply(pe ft2d(2)</pre>	table)
Related	tcclose tcopen tabc	Table Convert Close Table Convert Open

tchan RF channel number used for tuning (P)

Description	Set by the protune macro.	
See also	NMR Spectroscopy User Guide	
Related	protune	Macro to start ProTune (M)
	atune	ProTune Present (P)
	mtune	Tune probe using swept-tune graphical display (M)
	tugain	Receiver gain used in tuning (P)
	tunesw	Width of the tuning sweep in Hz (P
	tupwr	Transmitter power used in tuning (P)

tcl Send Tcl script to Tcl version of dg window (C)

Syntax	tcl(script)	
Description	Sends a Tcl (Tool Command Language) script to the Tcl version of the dg window. If this window is not active, this command does nothing	
Arguments	script is any legal Tcl script.	
See also	User Programming	
Related	dg Display group of acquisition/processing parameters (C)	

tcclose Table Convert Close (C)

Syntax	tcopen(<filename>)</filename>
	tcclose
Applicability	VnmrJ 3.1
Description	"tcopen" explicitly reads, sorts, and stores in memory a table convert
	file from <pre>\$vnmruser/tablib/<filename> which it will then use</filename></pre>

	when "tcapply" is called. Once the table has been read in "tcclose" command must be used to remove the table and free the memory used.
	"tcclose" removes the table and frees the memory used to store the sorted table indices read in with a "tcopen" command.
Arguments	'filename' argument specifying the file to be read which resides in \$vnmruser/tablib/ <filename>.</filename>
Examples	tcopen(petable) tcclose
Related	tcapply Apply Table Conversion Reformatting to Data

temp Open the Temperature Control window (C)

Applicability	Systems with a variable temperature (VT) controller.		
Description	Opens the capabilitie	e Temperature Control window, which has the following es:	
	• Turn temperature control off.		
	• Set temperature control on at a specified temperature in degrees C.		
	• Enable temperature control from within an experiment using th temp parameter and the su, go, ga, or au macros. This mode i default.		
	• Alternatively, turn off experiment control of the temperature and allow only the Temperature Control window (and sethw) to set the temperature. This mode has the advantage that, often times, temp different between experiments. Joining a different experiment and entering go can unexpectedly change the temperature. This mode prevents this problem.		
	 Resetting reconnect 	the temperature controller when the temperature cable is ed to a probe.	
See also	NMR Spee	ctroscopy User Guide	
Related	acqi au ga go readhw sethw su temp tin	Interactive acquisition display process (C) Submit experiment to acquisition and process data (M) Submit experiment to acquisition and FT the result (M) Submit experiment to acquisition (M) Read current values of acquisition hardware (C) Set values for hardware in acquisition system (C) Submit a setup experiment to acquisition (M) Sample temperature (P) Temperature interlock (P)	

temp

Sample temperature (P)

Applicability Systems with a variable temperature (VT) module.

Description	Sets the temperature of sample.	
Values	'n' or -150 to $+200$, in steps of 0.1° C. 'n' instructs the acquisition system not to change the VT controller and to ignore temperature regulation throughout the course of the experiment.	
See also	NMR Spectre	pscopy User Guide
Related	readhw	Read current values of acquisition hardware (C)
	temp	Open the Temperature Control window (C)
	tempcal	Temperature calculation (C)
	tin	Temperature interlock (P)
	vtc	Variable temperature cutoff point (P)

tempcal Temperature calculation (C)

Applicability	Systems with a variable temperature (VT) module.		
Syntax	<pre>tempcal(solvent)<:temperature></pre>		
Description	For exact determination of sample temperature when using the VT unit, a temperature calibration curve must be made for each probe used. All data, such as gas flow, must be noted. Use samples of ethylene glycol for high-temperature calibration, and use samples of methanol for low-temperature calibration. To make the calculation:		
	• Bring the sample to the desired temperature and allow sufficient time for equilibration, then obtain a spectrum.		
	•Next, align two cursors on the two resonances in the spectrum, then enter tempcal('e') for ethylene glycol, or enter tempcal('m') for methanol. The temperature is calculated based on the difference frequency between the cursors.		
Arguments	solvent is the sample solvent: 'glycol', 'e', or 'g' for ethylene glycol, or 'methanol' or 'm' for methanol.		
	temperature returns the calculated value of the sample temperature. The default is the system displays the value.		
Examples	tempcal('glycol') tempcal('m'):temp		
See also	NMR Spectroscopy User Guide		

tempcalc Measure approximate sample temperature in Cold Probes (M)

testacquire Test acquire mode (P)

Description	Allows test acquisitions to be done while a study queue is active, without using the study queue. When this mode is enabled, acquisition do not update the status of the currently loaded experiment in the study queue, and data is not saved in the study queue. This mode set from the Test mode check box in the Acquisition menu or from the command line.	
Syntax	testacquire=<'y' or 'n'>	
Values	'y' test acquire mode enabled	
	'n' test acquire mode disabled	
Related	acquireAcquire data (M)saveSave data (M)	

testct Check ct for resuming signal-to-noise testing (M)

Description	Used by the signal-to-nois	testsn macro to decide when to resume testing of e. See the description of testsn for details.
See also	NMR Spectro	scopy User Guide
Related	ct	Completed transients (P)
	testsn	Test signal-to-noise of a spectrum (M)

testsn Test signal-to-noise of a spectrum (M)

- Description Part of the automatic periodic signal-to-noise testing that occurs during various automated acquisitions, most notably c13. Transforms the data using fn=16000, and then baseline corrects, setting the left-most 10% of the spectrum and the right-most 2% as baseline. After the baseline correction, testsn uses getsn to calculate the signal-to-noise.
 - If signal-to-noise exceeds the desired goal in parameter sn (found in the standard carbon parameter set /vnmr/stdpar/c13), testsn aborts the experiment using the command halt, which initiates processing according to the wexp parameter.
 - If signal-to-noise is not reached, testsn estimates the signal-to-noise ratio at the end of the experiment. If signal-to-noise target will not be reached by then, it cancels subsequent signal-to-noise testing, but allows the experiment to proceed.

• If the signal-to-noise target will be reached before the end of the experiment, it saves the estimated number of transients required to reach the goal in the parameter r7 (using a conservative estimate), and then sets the processing at future blocks to be only testct, which simply tests if ct is greater than r7, and, if so, resumes testing of signal-to-noise with testsn.

See also NMR Spectroscopy User Guide

Related	c13	Automated carbon acquisition (M)
	fn	Fourier number in directly detected dimension (P)
	getsn	Get signal-to-noise estimate of a spectrum (M)
	halt	Abort acquisition with no error (C)
	r1-r7	Real parameter storage for macros (P)
	sn	Signal-to-noise ratio (P)
	testct	Check ct for resuming signal-to-noise testing (M)
	wexp	Specify action when experiment completes (C)

teststr Find which array matches a string M)

Syntax teststr(parameter,string <,tree>):\$ret

Description The teststr command requires at least two arguments. The first is the name of a string parameter. The first argument must generally be enclosed in single quotes. The teststr command needs the name of the parameter, not its values. The second is a string. The optional third argument is the parameter tree. The default is current.

Macro parameters can be used as the first argument. In this case, the third argument must be 'local'.

This command sets \$ret to the index of the array element that matches the second argument. If none of the array values of the parameter match the second argument, a zero is returned.

Examples n1='hello','labas','gidday','hola','bonjour','ciao'
 teststr('n1','labas'):r1

sets r1=2, since 'labas' matches element 2 of the n1 array.

The elements do not need to be single words. For example, n1='good night', 'labanaktis', 'bonne nuit', 'gute Nacht', 'boa noite', 'buonas noces'

teststr('n1', 'boa noite'):r1
sets r1=5. The strings must match exactly, including upper and lower
case

teststr('n1','gute nacht'):r1
sets r1=0, since the lower case n in nacht does not match the upper
case N in Nacht.

For local dollar variables, the 'local' argument must be used. Again, enclose the name of the local parameter in single quotes.

\$greet='hello','labas','gidday','hola', 'ciao'
teststr('\$greet','labas','local'):r1

text Display text or set new text for current experiment (C)

Syntax text<(text_string)><:string_variable>

- Description Associated with each experiment is a text file, consisting of a block of text, that can be used to describe the sample and experiment. text allows displaying the text file and changing the text file for the current experiment. A UNIX text editor, such as vi, or the macro textvi can also be used to edit the text file of the current experiment.
- Arguments text_string is a string of text that replaces the existing text file. The default is to display the text file in the current experiment. The characters \\ or \n can be used in the string to denote a new line, and the characters \t can be used to denote a tab (see example below).

string_variable returns the text in text_string as a string variable. Thus, for example, the text:n1 and text(n1+'cosy experiment') commands, where n1 is a string, can be used in a macro to add a "cosy experiment" to the text. An equivalent operation using the atext command would be atext('cosy experiment').

- Examples text('Sample 101\tCDCl3\\13 February')
- See also NMR Spectroscopy User Guide

Related	atext	Append string to the current experiment text (M)
	ctext	Clear the text of the current experiment (C)
	curexp	Current experiment directory (P)
	dtext	Display a text file in the graphics window (C)
	puttxt	Put text file into another file (C)
	textvi	Edit text file of current experiment (M)
	vnmrprint	Print text files (U)

textis Return the current text display status (C)

Syntax	(1) textis(command):\$yes_no(2) textis:\$display_command
Description	Determines if a command given by the user currently controls the text window (syntax 1) or returns the name of the command currently controlling the text window (syntax 2).
Arguments	command is the name of a command that potentially may be controlling the text window.
	<pre>\$yes_no returns 1 if command controls the text window, or 0 if it does not.</pre>
	\$display_command returns the name of the command currently controlling the text window.
Examples	textis:\$display if (\$display = 'dg') then endif
See also	User Programming
Related	graphis Return the current graphics display status (C)

textvi Edit text file of current experiment (M)

Description	Edits the text vi. textvi i	file of the current experiment using the UNIX text editor is equivalent to the command vi(curexp+'/text').
See also	NMR Spectroscopy User Guide	
Related	edit	Edit a file with user-selectable editor (M)
	text	Display text or set new text for current experiment
		(C)
	vi	Edit text file with vi editor (M)

th Threshold (P)

Description	Sets threshold for printout of peak frequencies so that peaks greater
	than th on the plot appear on any peak listings. th is always bipolar
	(i.e., negative peaks greater in magnitude than th also appear in peak
	listings).
Values	0 to 1e9, in mm.
See also	NMR Spectroscopy User Guide
Related	thadj Adjust threshold for peak printout (M)

th2d Threshold for integrating peaks in 2D spectra (P)

Description	Used by 112d when determining the bounds of a peak and calculating its volume. To create the 2D peak picking parameters th2d and xdiag in the current experiment, enter addpar('112d').	
Values	From 0.0 to 1.0. If th2d=1.0, 112d integrates all points in the peak that are above the current threshold for the spectrum (i.e., the portion of the peak that can be seen in a contour plot of the spectrum). A smaller value causes 112d to integrate a larger area when determining the volume of a peak. If th2d=0.5, for example, 112d integrates all points in a peak that are above 0.5 times the current threshold.	
See also	NMR Spectroscopy User Guide	
Related	addparAdd selected parameters to the current experiment (M)112dAutomatic and interactive 2D peak picking (C)xdiagThreshold for excluding diagonal peaks when peak picking (P)	

thadj Adjust threshold for peak printout (M)

Syntax thadj<(max_peaks<,noise_mult<,llarg1<,llarg2>>>)>

Description	Adjusts the threshold th so that no more than a specified maximum number of peaks are found in a subsequent line listing (see nll) and so that th is at least a specified noise multiplier times the root-mean-square noise level.		
Arguments	max_peaks is the maximum number of peaks in the displayed spectral range. The default is $wc/4$ (i.e., the threshold is adjusted such that ppf will produce a "reasonable" number of lines with any width of plot).		
	noise_mult is a noise multiplier used to calculate the minim for th from the size of the root-mean-square noise.		
	llarg1 is the noise_mult argument (the default is 3) to the nll command used inside this macro		
	llarg2 is the keyword argument ('pos', 'neg', 'all'; the defa is 'all'.) to the nll command used inside this macro.		
Examples	thadj thadj(50) thadj(200,4) thadj(200,4,2) thadj(200,4,2,'pos')		
See also	NMR Spectroscopy User Guide		
Related	nll ppf th	Find line frequencies and intensities (C) Plot teak frequencies over spectrum (M) Threshold (P)	
	vsadj vsadj2 vsadjc vsadjb	Automatic vertical scale adjustment (M) Automatic vertical scale adjustment by powers of two (M) Automatic vertical scale adjustment for ¹³ C spectra (M) Automatic vertical scale adjustment for ¹ H spectra (M)	
	WC	Width of chart (P)	

time Display experiment time or recalculate number of transients (M)

Syntax	<pre>time<(<hours,>minutes)></hours,></pre>
Description	Estimates the acquisition time or recalculates the number of transients so that the total acquisition time is approximately the requested time. The parameters looked at when calculating the time per transient are d1, d2, d3, at, ni, $sw1$, ni2, and $sw2$.
Arguments	hours and minutes are numbers making up a time to be used by the system to recalculate the parameter nt so that the total acquisition time is approximately the time requested; the default (no arguments) is for the system to estimate the acquisition time for a 1D, 2D, or 3D experiment using the parameters in the current experiment.
Examples	time time(2,45)

See also	NMR Spectroscopy User Guide		
Related	at	Acquisition time (P)	
	d1	First delay (P)	
	d2	Incremented delay in 1st indirectly detected dimension (P)	
	d3	Incremented delay in 2nd indirectly detected dimension (P)	
	exptime	Display experiment time (C)	
	ni	Number of increments in 1st indirectly detected dimension (P)	
	ni2	Number of increments in 2nd indirectly detected dimension (P)	
	nt	Number of transients (P)	
	sw1	Spectral width in 1st indirectly detected dimension (P)	
	sw2	Spectral width in 2nd indirectly detected dimension (P)	

tin Temperature interlock (P)

Description	Controls error handling based on temperature regulation. If
	temperature regulation is lost, tin can be used to select whether an
	error is generated and acquisition is halted or whether a warning is
	generated and acquisition continues. In both cases, the lost regulation
	will cause werr processing to occur, thus providing a user-selectable
	mechanism to respond to VT failure.
Values	'n' turns off the temperature interlock feature

w' indicates the variable temperature regulation light is monitored during the course of the experiment and, if it starts to flash (regulation lost), a warning is generated; however, acquisition is not stopped.

'Y' indicates the variable temperature regulation light is monitored during the course of the experiment and, if it starts to flash (regulation lost), the current data acquisition is stopped. The acquisition will not resume automatically if regulation is regained.

See also NMR Spectroscopy User Guide

Related in Lock and spin interlock (P) werr When error (P)

tlt First-order baseline correction (P)

Description When spectral display is active, the command dc turns on a linear drift correction (baseline correction). The result of this operation includes calculating a first-order baseline correction parameter tlt. The calculation is made by averaging of a small number of points at either end of the display and drawing a straight line baseline between them.

tmove Left-shift FID to time-domain cursor (M)

DescriptionProvides an alternative method of left shifting time-domain data. To
use this method, position the right time cursor at the place that should
be the start of the FID, then enter tmove. This adjusts lsfid to
left-shift the FID.See alsoNMR Spectroscopy User GuideRelatedlsfidNumber of complex points to left-shift np FID (P)

tmsref Reference 1D proton or carbon spectrum to TMS (M)

Syntax tmsref:tms_found

- Description Tries to locate a TMS line. If found, tmsref re-references the spectrum to the TMS line and returns a 1 to the calling macro; if not found, tmsref returns 0 and the referencing is left as it was. In the case of other signals (e.g., from silicon grease) immediately to the left of the TMS line (even if they are higher than the reference line), tmsref tries avoiding those by taking the rightmost line in that area, as long as it is at least 10% of the main Si-CH₃ signal. Large signals within 0.6 ppm for ¹H (or 6 ppm for ¹³C) to the right of TMS may lead to misreferencing.
 Arguments tms_found returns 1 if a TMS line was located or returns 0 if not. See also NMR Spectroscopy User Guide
 - Relatedc13Automated carbon acquisition (M)h1Automated proton acquisition (M)

tn Nucleus for observe transmitter (P)

- Description Changing the value of tn causes a macro (_tn) to be executed that extracts values for sfrq and tof from lookup tables. The tables, stored in the directory /vnmr/nuctables, are coded by atomic weights.
 - Values In the lookup tables, typically given by 'H1', 'C13', 'P31', etc. The value tn='lk' sets the deuterium frequency, and also holds the lock current and switches the relay in the automated deuterium gradient shimming module, if present, so that deuterium signal may be observed without disturbing lock. The frequency is the same as tn='H2'.

NMR Spectre	oscopy User Guide
dn	Nucleus for first decoupler (P)
dn2	Nucleus for second decoupler (P)
dn3	Nucleus for third decoupler (P)
sfrq	Transmitter frequency of observe nucleus (P)
tof	Frequency offset for observe transmitter (P)
	NMR Spectro dn dn2 dn3 sfrq tof

tncosyps Set up parameters for TNCOSYPS pulse sequence (M)

DescriptionSets up a homonuclear correlation experiment (phase-sensitive
version) with water suppression.See alsoNMR Spectroscopy User Guide

tndqcosy Set up parameters for TNDQCOSY pulse sequence (M)

Applicability Systems with a linear amplifier on the observe channel and a T/R switch.
 Description Sets up a 2D J-correlation experiment with water suppression.
 See also NMR Spectroscopy User Guide

tnmqcosy Set up parameters for TNMQCOSY pulse sequence (M)

- Applicability Systems with hardware digital phaseshifter for transmitting with direct- synthesis rf; otherwise, software small-angle phaseshifter for transmitting with the old-style rf is used.Description Sets up a multiple-quantum filtered COSY experiment with water suppression.
 - See also NMR Spectroscopy User Guide

tnnoesy Set up parameters for TNNOESY pulse sequence (M)

Applicability Systems with a linear amplifier on the observe channel and a T/R switch.
Description Sets up a 2D cross-relaxation experiment with water suppression.
See also NMR Spectroscopy User Guide

tnroesy Set up parameters for TNROESY pulse sequence (M)

DescriptionSets up a rotating-frame NOE experiment with water suppression.See alsoNMR Spectroscopy User Guide

tntocsy Set up parameters for TNTOCSY pulse sequence (M)

Applicability Systems with T/R switch, computer-controlled attenuators, and linear amplifiers on observe channel.
Description Sets up a total-correlation spectroscopy experiment (HOHAHA) with water suppression.
See also NMR Spectroscopy User Guide

TOCSY Convert the parameters to a TOCSY experiment (M)

Description	Convert parameters to a TOCSY experiment.		
See also	NMR Spectro	scopy User Guide	
Related	ft1dac	Combined arrayed 2D FID matrices (M)	
	ft2dac	Combined arrayed 2D FID matrices (M)	
	wft1dac	Combined arrayed 2D FID matrices (M)	
	wft2dac	Combined arrayed 2D FID matrices (M)	

TOCSY1d Convert the parameter set to a Tocsy1d experiment (M)

Description	Convert th	e parameter set to a Tocsy1d experiment.
See also	NMR Spect	roscopy User Guide
Related	Proton	Set up parameters for ¹ H experiment (M).
	sel1d	Selective 1D protocols to set up (M).

tocsyHT Set up the tocsyHT experiment (M)

Description	Sets up parameters for a Hadamard-encoded tocsy experiment.		
See also	NMR Spectroscopy User Guide		
Related	htofs1	Hadamard offset in ni (P)	
	fn1	Fourier number in 1st indirectly detected dimension (P)	
	ni	Number of increments in 1st indirectly detected	
		dimension (P)	

Fourier transform 2D data (C)
Set Hadamard frequency list from a line list (M)
Set up parameters for a TOCSY pulse sequence (M)
Hadamard frequency list in ni (P)

tof Frequency offset for observe transmitter (P)

Description Controls the exact positioning of the transmitter. As the value assigned to tof increases, the transmitter moves to a higher frequency (toward the left side of the spectrum). The minimum step size of tof is determined by the type of rf hardware in the spectrometer. The limit is specified using the Step Size label in the Spectrometer Configuration window. Systems with broadband style rf (rftype='b') generally have 100-Hz resolution; all other systems have 0.1 Hz resolution. Values Approximate, depends on frequency-100000 to 100000, in Hz. See also NMR Spectroscopy User Guide Related config Determine current configuration and possibly change it (M) dof Frequency offset for first decoupler (P) Frequency offset for second decoupler (P) dof2 dof3 Frequency offset for third decoupler (P) Type of rf generation (P) rftype

tpwr Observe transmitter power level with linear amplifiers (P)

Applicability Systems with a linear amplifier on the observe channel.

Description Controls transmitter power. The value of the attenuator upper safety limit is set using the Upper Limit label in the Spectrometer Configuration window. Depending on hardware adjustments, the system may saturate at a given value of tpwr (i.e., values above a certain value may give equal output).

Values On systems with 63-dB attenuator installed: 0 to 63 (63 is maximum power), in units of dB. About 55 to 60 is normal. Lower values (e.g., 49) might be used for water suppression experiments like 1-3-3-1.

On systems with 79-dB attenuator installed: -16 to 63 (63 is maximum power), in units of dB.

CAUTION

Continuous power greater than 2 watts in a switchable probe will damage the probe. Always carefully calibrate power to avoid exceeding 2 watts. The maximum value for tpwr on a 200-MHz, 300-MHz, or 400-MHz system with a linear amplifier on the decoupler channel has been set to 49, corresponding to about 2 watts of power. Before using tpwr=49 for continuous decoupling, ensure safe operation by measuring the output power. This should be done during system installation and checked periodically by the user.

See also NMR Spectroscopy User Guide

Related	cattn	Coarse attenuator (P)
	config	Determine current configuration and possibly change it (M)
	dpwr	Power level for first decoupler with linear amplifiers (P)
	dpwr2	Power level for second decoupler (P)
	dpwr3	Power level for third decoupler (P)
	dpwrf	First decoupler fine power (P)
	fattn	Fine attenuator (P)
	tpwrf	Observe transmitter fine power (P)

tpwrf Observe transmitter fine power (P)

Systems with a fine attenuator on the observe transmitter channel.		
Controls the transmitter fine attenuator. Systems with this attenuator are designated using the Fine Attenuator label in the Spectrometer Configuration window. The fine attenuator is linear and spans 60 dB or 6 dB. If tpwrf is not present, enter create('tpwrf', 'integer') setlimit('tpwrf', 4095, 0, 1) to create it.		
0 to 4095, where 4095 is maximum power. If tpwrf does not exist in the parameter table, a value of 4095 is assumed.		
NMR Spe	ctroscopy User Guide	
config dpwr dpwrf fattn tpwr tpwrm	Determine current configuration and possibly change it (M) Power level for first decoupler with linear amplifiers (P) First decoupler fine power (P) Fine attenuator (P) Observe transmitter power level with linear amplifier (P) Observe transmitter linear modulator power (P)	
	Systems of Controls are desig Configura or 6 dB. I setlimi 0 to 4095 the parar <i>NMR Spe</i> config dpwr dpwrf fattn tpwr tpwrm	

tpwrm Observe transmitter linear modulator power (P)

Description Controls the power level on the observe transmitter linear modulator. The fine power control is linear and spans 0 to tpwr.

Values	0 to 4095, where 4095 is maximum power. If tpwrm does not exist in the parameter table, a value of 4095 is assumed.		
See also	NMR Spectroscopy User Guide		
Related	config dpwrf	onfig Determine current configuration and possibly change it (M) pwrf First decoupler fine power (P)	
	fattn	Fine attenuator (P)	

trace Mode for *n*-dimensional data display (P)

Description	Sets the multidimensional data display mode.			
Values	$^{\prime}{\tt fl}^{\prime}$ displays the ${\tt f}_1$ axis horizontally and allows ${\tt f}_1$ traces to be displayed.			
	$^{\prime}$ f2 $^{\prime}$ displays the f_2 axis horizontally and allows f2 traces to be displayed.			
	$^{\prime}$ f3 $^{\prime}$ displays the f3 axis horizontally and allows f_3 traces to be displayed if the data set is 3D.			
See also	NMR Spectroscopy User Guide			

traymax Sample changer tray slots (P)

Applicability	Systems with an automatic sample changer.			
Description	Specifies the type of sample changer. It also can be used to disable the sample changer. The value is set using the Sample Changer label in the Spectrometer Configuration window.			
Values	0 is setting for no sample changer present or, if a sample changer is attached, to disable the changer (None choice in the Spectrometer Configuration window).			
	9, 50, 100, 96, 48 are traymax values that indicate the number of sample slots for the corresponding sample changer (9 is for Carousel, 50 is for SMS/ASM 50 Sample, 100 is for SMS/ASM 100 Sample, 96 is for VAST, and 48 is for NMS, 768 for 768AS).			
See also	VnmrJ Installation and Administration; VnmrJ Walkup			
Related	config Display current configuration and possibly change it (M)			

trfunc Translates screen co-ordinates

Syntax trfunc(\$x,\$y):\$xincm,\$yincm Applicability VnmrJ 3.1

Description	trfunc translates screen co-ordinates to hertz or centimeters
	depending upon the axis parameter.
Examples	call trfunc(\$x,\$y):\$xincm,\$yincm

trfuncd Translates a screen distance

Syntax	trfuncd		
Applicability	VnmrJ 3.1		
Description	trfuncd translates a screen distance into centimeters in a real ima It is only useful in axis='cc' (aspect ratio constrained) images.		
Examples	trfuncd(\$screenlength):\$imagelength		

troesy Set up parameters for TROESY pulse sequence (M)

Description	Sets up parameters for the transverse cross-relaxation experiment in
	a rotating frame.
See also	NMR Spectroscopy User Guide

trunc Truncate real numbers (0)

Description	In MAGICAL programming, an operator that truncates real numbers.		
Examples	\$3 = trunc(3.6)		
See also	User Programming		
Related	acos	Find arc cosine of number (C)	
	asin	Find arc sine of number (C)	
	atan	Find arc tangent of a number (C)	
	COS	Find cosine value of an angle (C)	
	exp	Find exponential value (C)	
	ln	Find natural logarithm of a number (C)	
	tan	Find tangent value of an angle (C)	
	sqrt	Return square root of a real number (0)	
	typeof	Return identifier for argument type (O)	

trtuneAllows the user to view multiple tuning traces apparently
simultaneously

Syntax trtune Applicability VnmrJ 3.1 Description "trtune" allows the user to view multiple tuning traces apparently simultaneously. A tune sweep executes on tn nucleus (typically H1), then the dn nucleus, the dn2, and so on. A color key is displayed to the right and above the axis on the display. The # traces selection (the nf parameter) controls how many traces are performed, the maximum number of traces is the number of rf channels present. If probeConnect is present, it is used. If not, the channel order is '12345' if tn is highband, and '21345' otherwise.

> There is only one vertical scale control. The traces may be adjusted by independent gain control (gain, gaind, gaind2, gaind3 etc.) which are defined in the prarameter set. The power may be adjusted independently as well (tupwr, tupwrd, tupwrd2 etc.). It is preferable to keep power levels low, and adjust gain. Adjusting the display is easiest setting number of traces to 1 and autoscale. Trtune does not support shared RF channel nor does it support quadrature tuning.

tshift Adjust tau2 to current cursor position (M)

Applicability Systems with a solids module.

Description Adjusts tau2 to make the current time cursor position the start of acquisition. As the time-domain cursor can move between points, this macro allows the accurate adjustment of tau2 so as to start another acquisition exactly at the top of an echo.

See also User Guide: Solid-State NMR

tugain Receiver gain used in tuning (P)

Description	Used internally by the protune macro to set the receiver gain. NMR Spectroscopy User Guide		
See also			
Related	protune	Macro to start ProTune (M)	
	atune	ProTune Present (P)	
	mtune	Tune probe using swept-tune graphical display (M)	
	tchan	RF channel number used for tuning (P)	
	tunematch	Default match target, in percent of optimum (P)	
	tunesw	Width of the tuning sweep in Hz (P	
	tupwr	Transmitter power used in tuning (P)	

tune Assign a frequency to a channel for probe tuning (C)

Syntax (1) tune(freq1,<freq2,freq3,freq4>)
(2) tune(chan1,freq1,<chan2,freq2,...>)

Description Assigns a frequency to a channel when tuning the probe. The frequency assignment remains in effect (as a tune frequency) until the next su or go command is executed. Although only the first synthesizer is connected to the tuning system, the console is programmed to set this synthesizer to the desired frequency based on the channel shown on the CHAN readout on the TUNE INTERFACE unit.

The tune program has two formats. If syntax 1 is used, frequencies are assigned to channels based on the order of the arguments. The first argument is interpreted and assigned to the first (observe) channel, the second argument is assigned to the second (decoupler) channel. A third or fourth argument would be interpreted and assigned in a similar manner.

If syntax 2 is used, the arguments are entered in pairs, with the first argument specifying the rf channel and the next argument specifying the frequency.

tune selects the format based on the first argument. If the first argument is a name for an rf channel, syntax 2 is assumed; otherwise, syntax 1 is used.

Arguments freq1, freq2, freq3, and freq4 specify the frequency of the rf channel as a value in MHz (e.g., 200 or 300) or indirectly using the nucleus for tuning the probe (e.g., 'H1' or 'C13'). If a nucleus is entered, it must be found in the nucleus table. The frequency of any channel without an argument is unaffected. For example, tune('H1','C13','N15') sets the first channel to tune at the ¹H, the second channel at ¹³C, and the third channel at ¹⁵N. If a fourth channel is present, it is not affected. Entering tune('H1','C13',200) assigns the same frequencies for the first and second channels but the third channel tunes to 200 MHz, regardless of the proton frequency.

chan1, chan2, chan3, and chan4 specify the channel directly:

- 'todev' or 'ch1' specify channel 1 (observe transmitter).
- 'dodev' or 'ch2' specify channel 2 (first decoupler).
- 'do2dev' or 'ch3' specify channel 3 (second decoupler).
- 'do3dev' or 'ch4' specify channel 4 (third decoupler).

Only one of these keywords is used per channel (do not enter the channel using just its number). If a channel does not have a keyword entered as an argument, that channel is not affected (e.g., tune('ch4','P31') selects the frequency corresponding to ³¹P on the fourth channel, but leaves the first three channels unaffected).

Examples tune('H1','C13','N15') tune('H1','C13',200) tune('ch4','P31')

See also NMR Spectroscopy User Guide

Related	dfrq	Transmitter frequency of first decoupler (P)
	dfrq2	Transmitter frequency of second decoupler (P)
	dfrq3	Transmitter frequency of third decoupler (P)
	go	Submit experiment to acquisition (C)

mtune	Tune probe using swept-tune graphical display (M)
qtune	Tune probe using swept-tune graphical tool (C)
sfrq	Transmitter frequency of observe nucleus (P)
spcfrq	Display frequencies of rf channels (M)
su	Submit a setup experiment to acquisition (C)
tune	Assign frequencies (C)

tunehf Tune both H1 and F19 on an HFX probe (M)

Syntax	tunehf<('x')>		
Description	Tune both H1 and F19 on an HFX probe. Including the optional argument, $tunehf('x')$ also tunes the low band channel to dn (dfrq).		
Arguments	'x'- low band channel to dn (dfrq)		
See also	NMR Spectroscopy User Guide		
Related	protune Macro to start ProTune (M)		

tunematch Default match target, in percent of optimum (P)

Description	The default match target, in percent of optimum. This local real parameter must be created. It is used as the match criterion in call of the form protune(599.96)		
See also	NMR Spectroscopy User Guide and VnmrJ Walkup		
Related	protune	Macro to start ProTune (M)	
	create	Create new parameter in a parameter tree (C)	
	atune	ProTune Present (P)	
	mtune	Tune probe using swept-tune graphical display (M)	
	tchan	RF channel number used for tuning (P)	
	tugain	Receiver gain used in tuning (P)	
	tunesw	Width of the tuning sweep in Hz (P	
	tupwr	Transmitter power used in tuning (P)	

tunemethod Method to use for tuning (P)

Applicability Liquids, VnmrJ Walkup, Automation
Description Specify probe tuning method. Methods are located in:
 \$home/vnmrsys/tune/methods for local user or
 /vnmr/tune/methods for access by all users.
 The method determines the nucleus to tune and how coarse or fine
 the probe is tuned as a percentage of the optimal pw.

Values	'lohi' -tune low band to medium criterion then tune high band medium criterion		
	<pre>' - user defined method.</pre>		
See also	NMR Spectro	scopy User Guide and VnmrJ Walkup	
Related	atune protune wtune	ProTune Present (P) Macro to start ProTune (M) Specify when to tune (P)	

tuneResult Message indicating how well the tuning succeeded (P)

- Description Message indicating how well the tuning succeeded. This local string parameter is created by ProTune and set to a string describing the result of the tuning. The first word of the message will be "ok" if tuning is successful, "failed" if it fails, and "Warning:" if tuning was not done but the experiment should proceed.
 See also NMR Spectroscopy User Guide and VnmrJ Walkup
 - Related protune Macro to start ProTune (M)

tunerp A pulse sequence for pulse tuning through the directional couplers in the VNMRJ display

Syntax	tunerp
Applicability	VnmrJ 3.1
Description	A pulse sequence for pulse tuning through the directional couplers in the VNMRJ display. Tunerp is used for high-power pulsed tuning and for characterization of phase transient. It provides a phase-detected output of the pulse that can be displayed in phased or absolute value mode. Launch Tunerp with the Fidscan button on the Shims page in Setup tab.
To use Tunerp it is preferable to be able to measure the reformed to reflected power. The standard directional coupler wired to measure reflected power only. The arrow on the scoupler should point back toward the Front End to measure power. To measure forward power reverse the coupler so the arrow points toward the probe.	
	A second optional bidirectional coupler is available on some systems. With this coupler reverse the direction of the arrow by turning the knob on the top.
	Setup
	Load a calibrated data set or load Settancpx into a workspace that will not be used to acquire data. Convert the data set with Tunerp. Set Tunerp to obs the desired channel with the procedure below. It is

helpful to set up Tunerp for each of 1-4 channels in the first 1-4 workspaces and join each of them when tuning is needed.

To tune a particular channel, enter its number (1-4) in Channel entry box on the Sequence page. Also set the particular channel as observe on the channels page and choose the nucleus.

For two-channel experiments, where channels 1 and 2 are used as obs and dec, it is simply necessary to enter the desired nucleus in the observe nucleus entry box. The correct channel will be selected automatically and the channel number will be displayed.

For three-channel experiments or any time channels 3 and 4 are involved, it is necessary to configure probeConnect and preAmpConfig before tuning. See the instructions below for configuration of these parameters. Enter one of the nuclei designated in probeConnect in the observe transmitter-nucleus entry box. The correct channel will be selected automatically and the channel number will be displayed

Set aTune and tpwr to appropriate values. The amplitude of the tuning pulse is determined by aTune (not aX90) and tpwr. One should tune routinely with about 25 to 50 Watts of power or less. Sometimes it is necessary to retune with the precise amplitude to be used in the experiment.

Pulse Tuning

Before pulse-tuning always rough-tune the probe with the mtune function.

Press the Tune button to set pwTune at 300 us and select a full FID display. Note that Tune sets a 5.0 MHz spectral width and the appropriate acquisition time. Enter Fidscan on the Shims page of the Setup tab and select magnitude mode only. Adjust the repetition rate with d1 as desired. Alternatively type av and collect one-scan displays with acquire.

Switch to forward power and measure the pulse shape. It may be necessary to reduce reciever gain to avoid receiver overload. For high-power tuning it may be necessary to put attenuation between the coupler and the Front End.

Switch to reflected power and tune the probe to minimize the central component of the pulse. Characterize the forward/reflected ratio by recording two traces with the same value of vertical fid scale vf. Good tuning is a ratio of > 30/1.

Phase Transient

Minimization of phase transient on the proton channel is needed for multiple-pulse proton experiments such as Hetcorlgcp2d. Minimization of phase transient on the X channel is needed for Pisema2d and is desired for multiple-pulse X experiments such as C7inad2d.

Be sure the probe is tuned before measuring phase transient.

To characterize phase transient press the button labeled Transient to set a 10 us pulse whose rise and fall are clearly visible. Note that the Transient button sets a 5.0 MHz spectral width and the appropriate acquisition time. Enter Fidscan on the Shims page of the Setup tab and select real and imaginary modes. Adjust the repetition rate with d1 as desired. Alternatively type av and collect one-scan displays with acquire.

Set the coupler for forward power. Collect a trace and phase it so that the real channel is 90 degrees out of phase and the imaginary channel is in phase. In this mode one will see zero amplitude with two transients of opposite phase at the beginning and ends of the pulse.

These transients are the phase transient and represent pulse amplitudes during the rise and fall times that are 90 degrees out of phase from the pulse.

Adjust probe tuning or cable lengths so as to minimize the amplitude of the two transients. Note that removal of phase transient with the probe tuning alone will detune the probe and increase reflected power. Generally one cannot achieve both good tuning and no transient by changing only the probe.

To remove phase transient by probe tuning adjust the Tune knob on the probe to move the tuning dip either up or down in frequency. Rephase the display and note whether the transients have gained or lost amplitude. Choose a value that minimizes the transient.

Phase transient can be removed permanently by adjusting the cable length between the probe and the directional coupler. Phase transient is a minimum for cable lengths that are multiples of 1/2 wavelength plus a constant. To find the correct length it is helpful to have a set of short cables and connectors and experiment with different lengths. The high-band channel can be adjusted with a set of elbow connectors. Once the correct length is found it is desirable to have a single permanent length made. Be sure that the probe remains tuned during this process. Note that one must have a different cable length for each different nucleus.

Three and Four Channel Experiments

To tune on channels 3 and 4 one must set probeConnect and preAmpConfig. These two parameters are "Global" strings than must be created manually by the system manager or user. As global parameters these strings apply to all workspaces in a user and do not affect other users. Note that these parameters are NOT created in the "Update User" function of the VNMRJ administrator interface or by the "makeuser" function.

probeConnect is a global string whose entries are the nuclei to be assigned to each channel. Create it with the command create 'probeConnect','string','global'). Type

display('probeConnect', 'global') to verify its exisitence. Type display('probeConnect') alone to verify that a "current" version of probeConnect does not exist. The result should be negative.

Set [rpbeConnect equal to the nuclei for channels 1 to the number of channels, numrfch, in order, separated by spaces. For example:

probeConnect = 'H1 C13 F19 N15'

т

sets up a four channel spectrometer with an HFXY probe tuned to the indicated nuclei. Note that the first entry is always highband and the second always low band. On three-channel spectrometers the third entry must match the band of channel-three amp. On four channel spectrometers a second highband amp is always placed on channel 3 if it is present.

preAmpConfig is a global string whose entries indicate the receiver function attached to each channel. Create it with the command create('preAmpConfig','string','global'). Type

display('preAmpConfig','global') to verify its exisitence. Type display('preAmpConfig') alone to verify that a "current" version of preAmpConfig does not exist. The result should be negative.

The characters of preAmpConfig can be "H" for highband, "L" for lowband and "X" for no preamp. The band of the preamp on a channel must match the band of the amplifier. A channel must have a preamp to be selected as the observe function.

An example for preAmpConfig is:

preAmpConfig = 'HLHL'

for the four-channel machine above.

probeConnect and preAmpConfig are present on the Channels page of all sequences. An output of "---" means that parameter does not exist. An Output of " " means that the parameter exists but has null value.

Parameter Groups

tune: Module: no

Sequence: tunerp.c

Description: Implements a directional-coupler pulse on a selected hardware channel for pulse tuning.

Parameters: Sequence Page

Arguments atune: the amplitude of the tune pulse.

chtune: the hardware channel to be tuned.

pwtune: the length of the tune pulse.

tunesw Width of the tuning sweep in Hz (P)

Description	Sets the widt	h of the tuning sweep in Hz and is set by the protune	
~ .	macro.		
See also	NMR Spectroscopy User Guide and VnmrJ Walkup		
Related	protune	Macro to start ProTune (M)	
	atune	ProTune Present (P)	
	mtune	Tune probe using swept-tune graphical display (M)	
	tchan	RF channel number used for tuning (P)	
	tugain	Receiver gain used in tuning (P)	

tunematchDefault match target, in percent of optimum (P)tupwrTransmitter power used in tuning (P)

tupwr Transmitter power used in tuning (P)

Description	The transmitter power used in tuning. The aptune pulse sequence uses this to set the transmitter power. Set by the protune macro.	
See also	NMR Spectroscopy User Guide and VnmrJ Walkup	
Related	protune	Macro to start ProTune (M)
	atune	ProTune Present (P)
	mtune	Tune probe using swept-tune graphical display (M)
	tchan	RF channel number used for tuning (P)
	tugain	Receiver gain used in tuning (P)
	tunematch	Default match target, in percent of optimum (P)
	tunesw	Width of the tuning sweep in Hz (P

typeof Return identifier for argument type (0)

Syntax	typeof	
Description	In MAGICAL programming, an operator that returns an identifier (0 or 1) for the type (real or string) of an argument.	
Examples	if typeof('\$1') then \$arg=1 else \$arg=\$1 endif	
See also	User Programming	
Related	isreal	Utility macro to determine a parameter type (M)
	isstring	Utility macro to determine a parameter type (M)
	on	Make a parameter active or test its state (C)
	size	Return number of elements in an arrayed parameter (0)

Т



Agilent VnmrJ 4 Command and Parameter Reference Guide

ultra8	Selects the Ultra 8 shim configuration (M)	
ultra18	Selects the Ultra 18 shim configuration (M)	
undospins	Restore spin system as before last iterative run (M)	
undosy	Restore original 1D NMR data from sub experiment (M)	
undosy3D	Restores 2D DOSY data stored by the dosy macro in 3D DOSY(M)	
unit	Define conversion units (C)	
unixtime	Return marker for current time to a Magical variable	
unlock	Remove inactive lock and join experiment (C)	
updatepars	Update all parameter sets saved in a directory (M)	
updateprobe	Update probe file (M)	
updaterev	Update after installing new VnmrJ version (M)	
updtgcoil	Update gradient coil (M)	
updtparam	Update specified acquisition parameters (C)	
usemark	Use "mark" output as deconvolution starting point (M)	
userdir	VnmrJ user directory (P)	
usergo	Experiment setup macro called by go, ga, and au (M)	
userfixpar	Macro called by fixpar (M)	

ultra8

selects the Ultra 8 shim configuration (M)

Syntax ultra8

Description The ultra8 macro selects the Ultra 8 shim configuration and selects an appropriate template for the dgs command and manual shim panel. Administrator privilege is required to change the shim configuration. The shims are: zlc z2c x1 y1 xz yz xy x2y2.

Related ultra18 selects the Ultra 18 shim configuration (M)



Select 18 shim configuration for Ultra 18 shim power ultra18 supply (M)

Syntax ultra18

Description Selects the 18 shim configuration for the Ultra 18 shim power supply and selects an appropriate template for the dgs command and manual shim panel. Administrator privilege is required to change the shim configuration.

> The shims are: z1 z1c z2 z2c z3c z4c x1 y1 xz yz xy x2y2 x3 y3 xz2 yz2 zxy zx2y2

selects the Ultra 8 shim configuration (M) Related ultra8

Restore spin system as before last iterative run (M) undospins

- Description Returns the values of the line assignments and the chemical shifts and coupling constants existing before the last iterative adjustment with spins('iterate'), and then runs spins. The parameters are returned from the file spini.inpar and the transitions from the file spini.savela in the current experiment. See also NMR Spectroscopy User Guide
 - Perform spin simulation calculation (C) Related spins

Restore original 1D NMR data from sub experiment (M) undosy

- Restores the 1D DOSY data stored by the dosy macro (if data exists) Description by recalling the data stored in the file subexp/dosy2Ddisplay in the current experiment. undosy and redosy enable easy switching between the 1D DOSY data (spectra as a function of gzlvl1) and the 2D DOSY display (signal as a function of frequency and diffusion coefficient). NMR Spectroscopy User Guide See also

Related	dosy	Process DOSY experiments (M)
	redosy	Restore 2D DOSY display from subexperiment (M)

undosy3D

Syntax	undosy3D
Applicability	VnmrJ 3.1
See also dosy

unit Define conversion units (C)

Syntax unit<(suffix,label,m<,tree><,'mult'|'div'> \
 ,b<,tree><,'add'|'sub'>)>

Description Defines a linear relationship that can be used to enter parameters with units. The unit is applied as a suffix to the numerical value (e.g., 10k, 100p). The definition of the linear relations follows the traditional y=mx+b equation, where x is the input value and y is the converted result.

Entering the unit command with no arguments displays all currently defined units. To remove a unit, define the unit with a 0 for the slope.

A convenient place to put unit commands for all users is in the bootup macro. Put private unit commands in a user's login macro.

Arguments suffix is a string identifying the name for the unit. The length of the string is limited to 12 characters.

label is a string for the name to be displayed when the axis parameter is set to the value of the suffix (if the suffix is only a single character). The length of the string is limited to 12 characters.

m is the slope of the linear relationship, defined either as a numerical value or as the name of a parameter. If a parameter name is used, it may be optionally followed with the parameter tree to use (argument tree) and by another optional keyword that specifies whether the parameter value should be a multiplier (keyword 'mult') or divisor (keyword 'div').

tree is the parameter tree to use (i.e.,'current', 'processed',
'global', or 'systemglobal'). The default tree is 'current'.

'mult' is a keyword that specifies that a parameter value used for the slope should be a multiplier. This is the default for the slope.

 ${}^{\prime}\text{div}{}^{\prime}$ is a keyword that specifies that a parameter value used for the slope should be a divisor.

b is the intercept of the linear relationship, defined either as a numerical value or as the name of a parameter. If a parameter name is used, it may be optionally followed with the parameter tree to use (argument tree) and by another optional keyword that specifies whether the parameter value should be added (keyword 'add') or subtracted (keyword 'sub').

'add' is a keyword that specifies that a parameter value used for the intercept should be a added. This is the default for the intercept.

'sub' is a keyword that specifies that a parameter value used for the intercept should be a subtract.

Examples unit Displays all currently defined units unit('k','kHz',1000) r1=10k will set r1 to 10000 unit('p','ppm','reffrq','processed') r1=10p will set r1 to 10*reffrq, where reffrq from processed tree unit('p','',0) r1=10p will set r1 to 10 and give an error "unknown unit p" unit('F','degF',5/9,-32*5/9) r1=212F will set r1 to 100 (degrees C) unit('C','degC',9/5,32) r1=100C will set r1 to 212 (degrees F) NMR Spectroscopy User Guide, User Programming See also Related axis Axis label for displays and plots (P) bootup Macro executed automatically when VnmrJ is activated (M)

unixtime Return marker for current time to a Magical variable

Syntax unixtime:r1,r2- Return marker for current time to a Magical variable systemtime:r1,r2- synonym for unixtime

Applicability VnmrJ 3.1

Description unixtime and systemtime are two names for the same function. They determine the current date and time as a system-dependent integer. The return value is in seconds. This value is usually defined as the elapsed time from an "epoch", which is often 1970. A second return value will give a microsecond value, for higher resolution.

The unixtime command helps time the execution of commands. It returns a marker representing the current time, in seconds. Call unixtime at the start and the end of a sequence of operation and then subtract the starting from the ending time to get the elapsed time.

unixtime accesses only the wall clock time, not the CPU time or any other statistic connected with the current process. The units for values returned are seconds and values should be accurate to within a few milliseconds.

Be aware that unixtime cannot time operations that run in background, for example, the ft3d command or go and its aliases.

The following Magical code fragment illustrates how you time something:

\$t1 = 0
\$t2 = 0
\$t3 = 0
unixtime:\$t1

ft2d unixtime:\$t2 \$t3=\$t2-\$t1 write('line3','elapsed time for ft2d is %f secs',\$t3 For more information, consult the UNIX manual entries time and get time of day.

unlock Remove inactive lock and join experiment (C)

Syntax unlock(exp_number,'force')

Description In attempting to join another experiment, the jexp command may abort claiming the experiment is locked. This feature prevents two users from processing the same experimental data at the same time, which could corrupt the data (a "user" can also be a background operation invoked by the same user, such as in wexp processing). This lock can be left behind if the program or the computer crashes.

> The unlock command removes the lock if it is inactive and joins the unlocked experiment. The command will fail if the lock is still active (i.e., the process that made the lock is still executing) or if the lock was placed on the experiment by a remote host. The latter situation can only occur when one or more nodes are sharing the same file system (and experimental data).

Arguments exp_number is the number of the experiment from 1 to 9 to be unlocked. force unlocks an experiment under all circumstances and joins the unlocked experiment.

Examples unlock(3) See also NMR Spectroscopy User Guide Related jexp Join existing experiment (C)

updatepars Update all parameter sets saved in a directory (M)

Syntax	updatepars(directory)
Description	Corrects saved parameter sets. Starting with VNMR version 4.2, all parameters, upper limit, lower limit, and step sizes have been tightened. Further additions were made in VNMR 4.3. updatepars searches a directory for parameter and FID files and corrects the procpar files found. This macro overwrites parameters in the current experiment. The corrections applied to the parameter sets are defined by the parfix macro. Because updatepars uses the current experiment to process the parameter sets, the experiment chosen for running updatepars should not contain a valuable data set.
Arguments	directory is the name of the directory to be searched.

Examples	updatepars	('myparlib')
	updatepars	('mydata')
See also	NMR Spectro	scopy User Guide
Related	parfix	Update parameter sets (M)
	parversion	Version of parameter set (P)

updateprobe Update probe file (M)

Syntax	updateprobe(<probe 'tmplt'><,'system'>)</probe 'tmplt'>		
Description	Updates the current existing probe file or probe template.		
Arguments	probe is the probe parameter to update. The default is the current probe parameter value.		
	'tmplt' is a keyword to update the local probe template. The default is the current probe file.		
	'system' is a keyword to update the system template or probe file, providing you have write permission to the file. The default is to update the local template or probe file.		
Examples	updateprobe updateprobe('autosw') updateprobe('autosw','system') updateprobe('tmplt')		
See also	NMR Spectroscopy User Guide		
Related	addparamsAdd parameter to current probe file (M)getparamReceive parameter from probe file (M)setparamsWrite parameter to current probe file (M)		

updaterev Update after installing new VnmrJ version (M)

DescriptionUpdates experiment parameters and the global file following
installation of a new VNMR software version. updaterev is called by
the makeuser command during the installation process.See alsoVnmrJ Installation and Administration

updtgcoil Update gradient coil (M)

ApplicabilitySystems with three-axis gradients.DescriptionCreates the gcoil parameter, if it does not exist, and sets it to the
current value of the system gradient coil sysgcoil. updtgcoil only
executes if gradients are configured in the system.

	The updtgcoil	macro is called when a new experiment is joined or
	new parameters	are read into an experiment; however, it is only called
	at these times if	the gcoil parameter exists. If sysgcoil is set to a
	gradient table na	ame and if the values of sysgcoil and gcoil are
	different, a mess	sage is displayed in the Status window to let the user
	know that the g	radient coil parameters have been updated.
	updtgcoil can parameter set w	be called directly if the user wants to update the ith the gcoil and gradient table parameters.
See also	NMR Spectrosco NMR	py User Guide; User Programming; VnmrJ Imaging
Related	gcoil	Read data from gradient calibration tables (P)
	sysgcoil	System gradient coil (P)

updtparam Update specified acquisition parameters (C)

Description	Enables interactive	updating of specified acquisition parameters.
See also	SpinCAD	
Related	psgupdateoff	Prevent update of acquisition parameters (C)
	psgupdateon	Enable update of acquisition parameters (C)

usemark Use "mark" output as deconvolution starting point (M)

Description In some cases it is not possible to produce a line list that is a suitable starting point for a deconvolution (e.g., lines may overlap so severely that a line list does not find them). In this case, or in any case, the results of a "mark" operation during a previous spectral display (ds) may be used to provide a starting point. If the "mark" has been made with a single cursor, the information in the file mark1d.out contains only a frequency and intensity, and the starting linewidth is taken from the parameter slw.

If the "mark" is made with two cursors, placed symmetrically about the center of each line at the half-height point, mark1d.out contains two frequencies and an intensity. In this case, the starting frequency is taken as the average of the two cursor positions; the starting linewidth is taken as their difference (thus allowing different starting linewidths for each line).

See also NMR	Spectroscopy	User	Guide	
--------------	--------------	------	-------	--

Related ds Display a spectrum (C) slw Spin simulation linewidth (P)

- Description Stores the full UNIX path of the directory that contains a user's private VnmrJ files. These include a user's private maclib, menulib, shims, psglib, experiments, etc. This parameter is initialized at bootup by the UNIX environmental variable vnmruser.
 - Values Typical value is /home/vnmr2/vnmrsys
 - See also NMR Spectroscopy User Guide
 - Related curexp Current experiment directory (P) systemdir VnmrJ system directory (P)

usergo Experiment setup macro called by go, ga, and au (M)

Description Called by macros go, ga, or au before starting an experiment. The user typically creates usergo as a means to set up general experiment conditions.

See also NMR Spectroscopy User Guide

Related	au	Submit experiment to acquisition and process data (M)
	ga	Submit experiment to ac acquisition and FT the result (M)
	go	Submit experiment to acquisition (M)
	go_	Pulse sequence setup macro called by go, ga, and au (M)

userfixpar Macro called by fixpar (M)

DescriptionCalled by the macro fixpar to provide an easy mechanism to
customize parameter sets.See alsoNMR Spectroscopy User GuideRelatedfixparCorrect parameter characteristics in experiment (M)



Agilent VnmrJ 4 Command and Parameter Reference Guide

vastgetSelects and displays VAST spectra (M)vastglueAssemble 1D datasets into a 2D (or pseudo-2D) datasets (M)vastglue2Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M)vastgoTurn off LC stop flow automation, start VAST automation (M)vbgRun VNMR processing in background (U)vfVertical scale of FID (P)viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vastglueAssemble 1D datasets into a 2D (or pseudo-2D) datasets (M)vastglue2Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M)vastgoTurn off LC stop flow automation, start VAST automation (M)vbgRun VNMR processing in background (U)vfVertical scale of FID (P)viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vastglue2Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M)vastgoTurn off LC stop flow automation, start VAST automation (M)vbgRun VNMR processing in background (U)vfVertical scale of FID (P)viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vastgoTurn off LC stop flow automation, start VAST automation (M)vbgRun VNMR processing in background (U)vfVertical scale of FID (P)viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vbgRun VNMR processing in background (U)vfVertical scale of FID (P)viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vfVertical scale of FID (P)viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
viEdit text file with vi text editor (M)vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vibraddDisplay relative amplitudes of Cold Probe vibrations (M)vjhelpDisplay VnmrJ help (U)	
vjhelp Display VnmrJ help (U)	
vn Start VNMR directly (U)	
Vnmr Start VNMR in current windowing system (U)	
vnmr_accounting Open Accounting window (U)	
vnmremail Utility to Send Files via Email	
vnmrexit Exit from the VNMR system (C)	
vnmrj Start VnmrJ (U)	
vnmrjcmd() Commands to invoke the GUI popup(C)	
vnmrjOptions Installer for passworded VnmrJ options (C)	
vnmrplot Plot files (U)	
vnmrprint Print text files (U)	
vo Vertical offset (P)	
VP Vertical position of spectrum (P)	
vpaction Set initial state for multiple viewports (M)	



vpf	Current vertical position of FID (P)
vpfi	Current vertical position of imaginary FID (P)
vpset3def	Set the viewport state to three default viewports (M)
vpsetup	Set new viewports (M)
vs	Vertical scale (P)
vs2d	Vertical scale for 2D displays (P)
vsadj	Automatic vertical scale adjustment (M)
vsadj2	Automatic vertical scale adjustment by powers of 2 (M)
vsadjc	Automatic vertical scale adjustment for ¹³ C spectra (M)
vsadjh	Automatic vertical scale adjustment for ¹ H spectra (M)
vsproj	Vertical scale for projections and traces (P)
vtairflow	Variable Temperature Air Flow (P)
vtairlimits	Variable Temperature Air Flow Limits (P)
vtc	Variable temperature cutoff point (P)
vtcomplvl	Variable temperature compensation for gradient shimming (P)
vttype	Variable temperature controller present (P)
vtwait	Variable temperature wait time (P)
vxr_unix	Convert VXR-style text files to UNIX format (M,U)

vast1d Set up initial parameters for VAST experiments (M)

Applicability	Systems with VAST accessory.
Description	Sets up initial VAST parameters from the /vnmr/stdpar directory or from the user's stdpar directory if the appropriate file exists there. Any changes made to the files in these directories are reflected in the setup. The file
	/vnmr/stdpar/vast1d.par contains the "default" parameters for VAST spectra and should be modified as needed to produce spectra under desirable conditions. After running vast1d, the solvent parameter can be set by choosing it from the list of solvents listed in /vnmr/solvents.
See also	NMR Spectroscopy User Guide

vastget Selects and displays VAST spectra (M)

Applicability	Systems with VAST accessory.
Syntax	<pre>vastget(<well>, <well>,)></well></well></pre>
Description	Selects and displays the spectra from any arbitrary well or wells using the well label(s) as arguments. the spectra are displayed in a dss stacked plot.
Arguments	well is the well label from which you want to select and display spectra. The wells are labeled [A->H][1-8].
Examples	vastget('B6','B7','C11','G3')
See also	NMR Spectroscopy User Guide

vastglue Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M)

Applicability	Systems with the VAST accessory.		
Syntax	<pre>vastglue(<rack,<zone>)</rack,<zone></pre>		
	<pre>vastglue(<glue order="">, <plate>)</plate></glue></pre>		
Description	Used to artificially reconstruct a 2D datasets from a series of 1D data sets having similar filenames. It is crucial to ensure that the format of the file names of each of the 1D data sets is identical. vastglue reads in each 1D file, in succession, and adds it to the previous data, but in a 2D format. It assumes that file names are of the format obtained when using the default setting of autoname (autoname=''). If autoname has been redefined, use a macro like vastglue2. Save the resulting reconstructed 2D datasets in the normal manner using svf.		
Arguments	rack is the rack number; the default is 1. If you enter a rack number, you must also enter a zone number.		
	zone is the zone number; the default is 1. If you want to specify a zone number, you must enter a rack number.		
	glue order is the specific glue order to be defined based on the order defined in a plate_glue file. If glue order is specified, you can provide a plate number as the second argument and used with the glue order argument.		
See also	NMR Spectroscopy User Guide		
Related	autonamePrefix for automation data file (P)vastglue2Assemble related 1D datasets into a 2D (or pseudo-2D) datasets (M)		

vastglue2 Assemble 1D datasets into a 2D (or pseudo-2D) datasets (M)

Applicability Systems with the VAST accessory Syntax vastglue2<(number)> Description Used to artificially reconstruct a 2D data set from a series of 1D datasets having similar filenames. It is crucial to ensure that the format of the file names of each of the 1D datasets is identical. vastqlue2 reads in each 1D file, in succession, and adds it to the previous data, but in a 2D format. It assumes that file names are of the format obtained using a nondefault setting of autoname (autoname='filename_R%RACK:%_Z%ZONE:%_S%SAMPLE#:%_'). This definition must be hard coded into the macro by the user. If autoname has not been redefined, use a macro like vastqlue. Save the resulting reconstructed 2D data set in the normal manner using svf. Arguments number is used to specify that only spectra from 1 through number are to be glued. The default is to glue all the spectra stored in the

Arguments number is used to specify that only spectra from 1 through number are to be glued. The default is to glue all the spectra stored in the current directory that have the proper file name format (from 1 through arraydim).

See also NMR Spectroscopy User Guide

Related autoname Prerix for automation data file (P) vastglue Assemble related 1D datasets into a 2D (or pseudo-2D) data set (M)

vastgo Turn off LC stop flow automation, start VAST automation (M)

ApplicabilitySystems with the LC-NMR and VAST accessoryDescriptionTurns off LC stopped flow use of automation and starts VAST
automation run.

vbg Run VNMR processing in background (U)

Syntax(From UNIX) vbg exp_number command_string <prefix>DescriptionEnables user to perform VNMR tasks in the background. vbg (for
"VNMR background processing") must be run from within a UNIX shell,
and *no* foreground or other background processes can be active in the
designated experiment (e.g., if you are working in exp2 in VNMR (in
the foreground), you cannot execute background processing in exp2 as
well).

Foreground processing causes a lock file to be placed in the appropriate experiment. The file has a format such as f.1268, where

	1268 indicates the process number in the process table (accessed in UNIX by entering the command $ps -e$). Background processing causes a lock file to be in the appropriate experiment as well. This file has a format such as b.4356, where 4356 indicates the process number. By displaying the files within an experiment, the user can readily determine whether any foreground or background processes are active in that experiment.
Arguments	exp_number is the number of the experiment, from 1 to 9, in the user's directory in which the background processing is to take place.
	command_string is the command string to be executed by VNMR in the background. Double quotes enclosing the string are mandatory (e.g., "fn=4096 fn1=2048 wft2da").
	prefix is a prefix to be added to the name of the log file, making the name prefix_bgf.log. The default name is exp_number_bgf.log, where exp_number is the experiment number. The log file is placed in the experiment in which the background processing takes place.
Examples	(From UNIX) vbg 1 "wft2da bc('f1')"
	(From UNIX) vbg 3 "vsadj pl pscale pap page" plotlog
See also	User Programming

Vertical scale of FID (P)

Description	In normalized intensity (nm) mode, vf is the height of the largest FID In absolute intensity (ai) mode, vf is a multiplier that is adjusted to produce a desired vertical scale, using the appearance on the display screen as a guide (full scale on the screen gives full scale on the plotter).	
	vf can be entered in the usual way or interactively controlled by clicking the middle mouse button in the graphics window during a FID display (click above the FID to increase vf or below the FID to decrease it).	
Values	1e-6 to 1e9, in mm (in nm mode) or as a multiplier (in ai mode).	
See also	NMR Spectroscopy User Guide	
Related	ai Select absolute intensity mode (C)	
	df Display a single FID (C)	
	nm Select normalized intensity mode (C)	
	sf Start of FID (P)	
	wf Width of FID (P)	

vi

vf

Edit text file with vi text editor (M)

Syntax vi(file)

Description Invokes the UNIX text editor vi for editing the file name given. On the Sun workstation, a popup screen contains the editing window. On the GraphOn terminal, the main screen becomes the editing window. vi is a powerful text editor, but its user interface is limited: the mouse is not used, menus are not available, and status information is virtually nonexistent.

vi operates in three modes: the *command mode* (for moving the cursor and editing text), the *insert mode* (for inserting text into the file), and the *last line mode* (for special operations). Each mode is described below.

Command mode

vi starts up in the command mode. In this mode, user commands consist mostly of a single character, sometimes in combination with another character, or a number, or both. A number preceding a command typically defines how many times a command should be executed (e.g., 3dd means delete three lines). The commands available include the following:

G	go to the start of the last line in the file
3G	go to the start of line 3
0	(zero) go to the start of the current line
\$	go to the end of the current line
Return or +	go to start of next line
-	(hyphen) go to start of previous line
Ctrl-d	scroll down (forward) half a screen
Ctrl-f	scroll forward by a full screen
Ctrl-u	scroll up (back) half a screen
Ctrl-b	scroll back by a full screen
/expression	find next expression and jump to its first character
?expression	find previous expression, jump to its first character
n	find next expression (from the last search)
N	find previous expression (from the last search)
dd	delete one line and put it into the buffer
3dd	delete three lines and put them into the buffer
dw	delete word
x	erase one character forward (under cursor)
Х	erase one character backwards (before cursor)
3x	erase three characters forward
rcharacter	erase character and replace with character
ZZ	write if necessary and quit vi
	(period) repeat the last command
u	undo the last command

J	join the next line to the current line
уу ог ч	yank one line and put into a buffer (called yank buffer)
р	put contents of yank buffer after the cursor
Р	put contents of yank buffer before the cursor
"aY	yank line into buffer a (buffers b to z also available)
"ap	put contents of buffer a below current line
"aP	put contents of buffer a above current line

Because there is no command line, these commands do not show up on the screen but are *executed immediately* (without pressing the Return key).

Insert mode

In the insert mode, characters typed on the keyboard (except for the Esc key) show up in the text. The insert mode is entered by typing one of the following commands from the command mode:

a text Esc	append text after the current cursor position
A text Esc	append text to the end of current line
i text Esc	insert text before current cursor position
cw word Esc	change word from current cursor position to end
2cw words Esc	change two words from current cursor position to end
o text Esc	open line below current line and append text
0 text Esc	open line above current line and append text

The only way to exit the insert mode is by pressing the Esc key, which leads back to the command mode. Unfortunately, there is no indication on the screen whether vi is in the command mode or in the insert mode. Inexperienced users often press the Esc key to make sure they are still in the command mode. The Esc key can also be used to avoid execution of commands that have been typed partially (e.g., the number has been typed, but not the last character).

You can insert special (normally nondisplayable) characters into the text if they are preceded by a Ctrl-v (e.g., entering Ctrl-v Ctrl-q is displayed in the text as Q).

Changing selected occurrences

The following actions find one or more occurrences of a particular word and change it to another word:

- First, type /word and press Return, where / is a forward slash and word is word you want to change.
- Next, press n as necessary until you reach the occurrence of the word you want to change.
- Finally, type cw newword and press Esc, where newword is replacement word.

• To repeat for another occurrence of word, press n as necessary to scan forward, and then type . (a period) to repeat cw newword (or whatever was the last change)

Changing selected occurrences of an expression (one or more words) is similar. To change two words, for example, take the same actions as above but use the command 2cw (or c2w) instead.

Last line mode

The last line mode is initiated with a colon; thereafter, commands such as the following can be used (press Return to execute these commands):

r filename:	read file named filename (insert in currently open file)
:w	write (save) file
:w filename	write under a new file named filename
:e filename	edit a different file named filename
:d	quit vi (only possible if file has been written back)
:wq	write back file (save changes) and quit vi
:q!	quit vi without saving changes

Exiting from vi is accomplished by using the ZZ command in the command mode, or with the :q, :wq, or :q! commands in the last line mode.

This description lists only a selection of the most important commands. For more information on vi, refer to UNIX books and manuals.

```
Examples vi(userdir+'/psglib/apt.c')
vi(curexp+'/text')
```

See also User Programming

Related	edit	Edit a file with user-selectable editor (M)
	paramvi	Edit a parameter and its attributes with vi text editor (\ensuremath{M})
	macrovi	Edit a user macro with the vi text editor (C)
	menuvi	Edit a menu with the vi text editor (M)
	textvi	Edit text file of current experiment (M

vibradd Display relative amplitudes of Cold Probe vibrations (M)

Applicability	Systems with Agilent, Inc. Cold Probes
Description	Display the relative amplitudes of the vibrations reaching the probe.
	Requires a doped HOD sample.

vjhelp Display VnmrJ help (U)

Syntax vjhelp file:///vnmr/jhelp/jhelp.html Description Displays the VnmrJ help in a Web browser.

vn Start VNMR directly (U)

Syntax	(From UNIX) vn <-display Xserver> <-fn font> &	
Description	Starts the VNMR application directly without checking the operating system and attempting to run the window manager.	
Arguments	-display Xserver specifies X server display (e.g., hostname:0.0) The default is the environment set by the DISPLAY variable.	
	-fn font specifies the size of the font displayed (e.g., $9x15$, $8x13$, or $7x13$). The default is the font set in the .Xdefaults file. Note that the size of the font affects the size of the VNMR window.	
Examples	vn & vn -display hostname:0.0 & vn -font 8x13 &	
See also	NMR Spectroscopy User Guide	
Related	vnmr Start VNMR (U)	

vnmr Starts VnmrJ (U)

Applicability	VnmrJ
Syntax	vnmr
Description	Starts the VnmrJ application
See also	NMR Spectroscopy User Guide
Related	vnmrj Start VnmrJ (U)

vnmr_accountingOpen Accounting window (U)

Description Opens a window for creating and maintaining cost accounting data for groups of users on a spectrometer system. The program accommodates multiple rate schedules for spectrometer usage. A calendar tool can be used to define holidays for holiday rates. There is no limit on the number of rates that can be defined. Multiple printers can be selected. Any user can view the accounting information (enter cd /vnmr/bin followed by ./vnmr_accounting), but to update information, the user must have root privileges.

See also System Installation and Administration Related operator Operator name (P) operatorlogin Sets work space and parameters for the operator (M)

vnmremail Utility to Send Files via Email

Description	Sends a file to an email address. Files are sent after uuencode.
	Directories are converted into tar files or zip files and sent.
Syntax	<pre>vnmremail(<'-m'>,filename,address)</pre>
Examples	<pre>vnmremail('myfile','nmr@agilent.com')</pre>
Arguments	The -m option is used to concatenate the specified file to the body of
	the email.

vnmrexit Exit from the VNMR system (C)

Description Exits from the VNMR system in a graceful manner by writing parameters and data to the disk, removing lock files, and restoring the terminal (if on a GraphOn). To provide flexibility when exiting VNMR, the macro exit calls vnmrexit to exit from VNMR.

CAUTION When you exit from the VNMR user interface on your X display system, whether you are using an X terminal or a Sun computer, and whether you are using OpenWindows, CDE, or Motif, you must first exit from any copy of VNMR running on your system. Failure to do this can cause current parameter values and even current data to be lost.

vnmrj Start VnmrJ (U)

Applicability	VnmrJ
Syntax	vnmrj
Description	Starts the VnmrJ application
See also	NMR Spectroscopy User Guide; VnmrJ Walkup
Related	vnmr Starts VnmrJ (U)

vnmrjcmd() Commands to invoke the GUI popup(C)

Syntax	<pre>vnmrjcmd('command1','command2',, parametername)</pre>	
	vnmrjcmd('command1','command2',<, callback>)	
Description	1 The vnmrjcmd() commands are needed in order to invoke the GU popup in which the user enters the parameters.	
	Note that vnmrbg and VnmrJ cannot be easily synchronized. When a macro invokes VnmrJ via vnmrjcmd, the VnmrJ thread runs	
	independently and the macro continues on and takes action without	

otherwise having knowledge of VnmrJ. In order to have events associated with required parameters occur in the proper order, a callback strategy was devised. In simple terms, the vnmrj commands can have a callback string such that when the required parameters are established in VnmrJ, vnmrbg can be re- invoked - the foremost example of this is re-entering the 'go' macro after the parameters are established in VnmrJ.

Examples Sends parameters one at a time to VnmrJ to be eventually displayed in an entry popup:

vnmrjcmd('reqpar','warngui','set', 'real',
parametername)
vnmrjcmd('reqpar','warngui','set', 'string',

parametername)

Display a GUI panel listing required parameters sent from vnmrbg in the previous 'set' option above:

vnmrjcmd('reqpar','warngui','show')

vnmrjcmd('reqpar','warngui','show', callback)

The callback is a command string to be sent back to vnmrbg, if needed. See the reqpartest macro source code for examples of how to use callback.

See also VnmrJ User Programing

Related go Submit experiment to acquisition (M) reqpartest Tests whether required parameters are set (M)

vnmrjOptionsInstaller for passworded VnmrJ options (C)

Applicability	VnmrJ 3.2
Description	VnmrJ passworded options can be installed after the VnmrJ software
	in installed from the distribution media. This tool provides the
	mechanism to specify the passwords and install the options. If you run
	this tool and do not have permission to write to the /vnmr system
	directory, it will show you what options are currently loaded.

vnmrplot Plot files (U)

Syntax	(From UNIX) vnmrplot <file></file>		
Description	A UNIX command that plots files from inside VNMR commands. To plot a file, you should use the page command, which uses vnmrplot internally.		
Arguments	file is the name of the file to be plotted.		
See also	NMR Spectroscopy User Guide		
Related	vnmrprint Print text files (U)		

vnmrprint Print text files (U)

Syntax	(From UNIX) vnmrprint printfile <printcap> <printer_type <clear="" file="" ="">></printer_type></printcap>		
Description	A UNIX command installed as part of the VNMR system to print text files. The printon and printoff commands use vnmrprint to print files. vnmrprint can also be used to delete a print file or save a print file to a different name.		
Arguments	printfile is the name of the text file to be printed.		
	printcap is a UNIX printcap entry (e.g. LaserJet_300) for the printer to print the text file. The default is the printer selected by the -p option of the UNIX 1p command.		
	printer_type is the type of printer from the list of VNMR printers (e.g., LaserJet_300). printer_type is required as an argument when it is desired to clear the printer file or save the printer file to another name.		
	clear is a keyword to delete the current print file. Deleting this file also requires that the printfile, printcap, and printer_type arguments be entered so that clear is the fourth argument.		
	file is the name of the file to use in saving the printfile. If a file with the name specified already exists, it is overwritten. Saving the file also requires that the printfile, printcap, and printer_type arguments be entered so that file is the fourth argument.		
Examples	vnmrprint /vnmr/psglib/tocsy.c LaserJet_300 vnmrprint myfile LaserJet_300 LaserJet_300 clear vnmrprint myfile ps PS_AR yourfile		
See also	NMR Spectroscopy User Guide		
Related	printoffStop sending text to printer and start print operation (C)printonDirect text output to printer (C)vnmrplotPlot files (U)		

Vertical offset (P)

vo

Description Sets the vertical offset, for 1D data sets, of the each spectrum in a stacked display with respect to the previous spectrum. The parameter ho sets the horizontal offset. For a "left-to-right" presentation, ho is typically negative; for a "bottom-to-top" presentation, vo is positive. For 2D data sets, the parameter wc2 sets the distance between the first and last trace and the vo parameter is inactive.
Values Number, in mm.
See also NMR Spectroscopy User Guide
Related ho Horizontal offset (P) wc2 Width of chart in second direction (P)

Vp Vertical position of spectrum (P)

Description	Contains vertical position of spectrum with respect to the bottom of		
	the display or plotter.		
Values	-200 to +200, in mm.		
See also	NMR Spectroscopy User Guide		
Related	vpf Current vertical position of FID (P)		
	vpfi Current vertical position of imaginary FID (P)		

vpaction Set initial state for multiple viewports (M)

Applicability	VnmrJ Walkup		
Description	Sets the initial state for multiple viewports. Used by the viewport editor dialog under Edit -> Viewports .		
See also	User Programming	,	
Related	jcurwin	Work space numbers of all viewports (P)	
	jviewportlabel jviewports	Work space labels for all viewport buttons (P) Viewport layout (P)	

vpf Current vertical position of FID (P)

Description Contains the current vertical position of an FID. To create this parameter and the other FID display parameters axisf, crf, deltaf, dotflag, and vpfi (if the parameter set is older and lacks these parameters), enter addpar('fid').

Values	Number, in mm. If vpf=0, the FID is positioned in the middle of the screen.		
See also	NMR Spectroscopy User Guide		
Related	addpar	Add selected parameters to the current experiment (M)	
	axisf	Axis label for FID displays and plots (P)	
	crf	Current time-domain cursor position (P)	
	deltaf	Difference of two time-domain cursors (P)	
	dotflag	Display FID as connected dots (P)	
	vp	Vertical position of spectrum (P)	
	vpfi	Current vertical position of imaginary FID (P)	

vpfi Current vertical position of imaginary FID (P)

Description	Contains the current vertical position of the imaginary part of an FID. To create this parameter and the other FID display parameters axisf, crf, deltaf, dotflag, and vpf (if the parameter set is older and lacks these parameters), enter addpar('fid').	
Values	Number, in mm. In vpfi=0, the imaginary part is positioned in the middle of the screen.	
See also	NMR Spectroscopy User Guide	
Related	addpar Add selected parameters to the current experiment (M) axisf Axis label for FID displays and plots (P) crf Current time-domain cursor position (P) deltaf Difference of two time-domain cursors (P dotflag Display FID as connected dots (P) vp Vertical position of spectrum (P) vpf Current vertical position of FID (P)	

vpset3def Set the viewport state to three default viewports (M)

Description	Sets the number of viewports to three, and resets the viewport button labels.		
See also	User Programming		
Related	jcurwin jviewportlabel jviewports	Work space numbers of all viewports (P) Work space labels for all viewport buttons (P) Viewport layout (P)	

Set new viewports (M) vpsetup

Description Sets the viewports from the selections made in the viewport editor dialog. For each viewport, it checks the work space number to join, then joins the appropriate work space.

See also User Programming

Related jcurwin Work space numbers of all viewports (P) jviewportlabel Work space labels for all viewport buttons (P) Viewport layout (P) jviewports

Vertical scale (P) vs

Description	In normalized (nm) mode, vs is the height of the largest peak in the
	spectrum. In absolute intensity (ai) mode, vs is a multiplier that is
	adjusted to produce a desired vertical scale, using the appearance on
	the display screen as a guide (full scale on the screen gives full scale
	on the plotter). vs can be entered in the usual way or interactively
	controlled by clicking the middle mouse button.

Values 1e-6 to 1e9, in mm (in nm mode) or as a multiplier (in ai mode).

NMR Spectroscopy User Guide See also

Related	ai	Select absolute intensity mode (C)
	isadj	Adjust integral scale (M)
	nm	Select normalized intensity mode (C)
	thadj	Adjust threshold for peak printout (M)
	vsadj	Automatic vertical scale adjustment (M)
	vsadj2	Automatic vertical scale adjustment by powers of two (M)
	vsadjc	Automatic vertical scale adjustment for ¹³ C spectra (M)
	vsadjh	Automatic vertical scale adjustment for ¹ H spectra (M)

vs2d Vertical scale for 2D displays (P)

Sets a multiplier for 2D spectra and images that is adjusted to produce Description a desired vertical scale for display or plotting. vs2d takes the place of vs for 2D data display and can be adjusted by explicitly setting it to a value or by clicking the middle mouse button when pointing to a point on a 2D display. If vs2d does not exist, it can be created by running par2d.

- NMR Spectroscopy User Guide See also
 - Related par2d Create 2D acquisition, processing, and display parameters (M)
 - Select vertical scale (C) vs
 - vsproj Adjust vertical scale for projections and traces (M)

Syntax	vsadj<(height)>	
Description	Automatically sets the vertical scale vs in the absolute intensity (ai) mode so that the largest peak is at the requested height.	
Arguments	height is the desired height, in mm, of the largest signal in the displayed portion of the spectrum. The default is $0.9*(wc2max-vp-sc2)$.	
Examples	vsadj vsadj (100)	
See also	NMR Spectroscopy User Guide	
Related	aiSelect absolute intensity mode (C)isadjAdjust integral scale (M)thadjAdjust threshold for peak printout (M)vsVertical scale (P)vsadj2Automatic vertical scale adjustment by powers of two (M)vsadjcAutomatic vertical scale adjustment for ¹³ C spectra (M)vsadjhAutomatic vertical scale adjustment for ¹ H spectra (M)wc2maxMaximum width of chart in second direction (P)	

vsadj2 Automatic vertical scale adjustment by powers of 2 (M)

Syntax	vsadj2<(height)>:scaling_factor	
Description	Adjusts the vertical scale by powers of two as required for expansion plots (see aexppl for more information).	
Arguments	height is desired height of largest (or largest relevant) signal in displayed portion of the spectrum. The default is $0.9*(wc2max-vp-sc2)$.	
	scaling_factor returns to the calling macro the ratio of the new compared to the old value of vs .	
Examples	vsadj2 vsadj2(50):r1	
See also	NMR Spectroscopy User Guide	
Related	aexpplAutomatic expansions plot (M)isadjAdjust integral scale (M)sc2Start of chart in second direction (P)thadjAdjust threshold for peak printout (M)vpVertical position of spectrum (P)vsVertical Scale (P)vsadjAutomatic vertical scale adjustment (M)vsadjcAutomatic vertical scale adjustment for ¹³ C spectra (M)vsadjhAutomatic vertical scale adjustment for H1 spectra (M)	
	 thadj Adjust threshold for peak printout (M) vp Vertical position of spectrum (P) vs Vertical Scale (P) vsadj Automatic vertical scale adjustment (M) vsadjc Automatic vertical scale adjustment for ¹³C spectra (M) vsadjh Automatic vertical scale adjustment for H1 spectra (M) wc2max Maximum width of chart in second direction (P) 	

Syntax	vsadjc<(height)>	
Description	Functionally the same as the macro $vsadj$, except excludes solvent and TMS signals from the carbon spectra for the adjustment of vs .	
Arguments	height is desired height of largest (or largest relevant) signal in displayed portion of the spectrum. The default is $0.9*(wc2max-vp-sc2)$.	
Examples	vsadjc vsadjc(wc2max-sc2-wc2-5)	
See also	NMR Spectroscopy User Guide	
Related	isadjAdjust integral scale (M)thadjAdjust threshold for peak printout (M)vsVertical Scale (P)vsadjAutomatic vertical scale adjustment (M)vsadj2Automatic vertical scale adjustment by powers of two (M)vsadjhAutomatic vertical scale adjustment for H1 spectra (M)	

vsadjh Automatic vertical scale adjustment for ¹H spectra (M)

Syntax	vsadjh<(height<,do_not_ignore_solvent>)>	
Description	Works as the same as the macro vsadj, except disregards solvent and TMS signals from proton spectra and, if from the remaining spectrum the highest line is more than three times as high as the second highest line, the spectrum is scaled to this second highest signal (otherwise the highest signal is taken as relevant).	
Arguments	height is desired height of largest (or largest relevant) signal in displayed portion of the spectrum. If height is 0 or a negative value, it defaults to $0.9*(wc2max-vp-sc2)$, which is also the default with no arguments.	
	do_not_ignore_solvent is any second argument. If present, it signals vsadjh to not ignore the solvent line and regard the solven line as normal signal (i.e, only exclude the TMS line). This argumen was added for the situation where frequently there are high "real" signals at the position of the solvent line. Such signals could otherwi be regarded as solvent line and would then be ignored. This could the lead to overscaling in the result.	
Examples	vsadjh vsadjh(0.7*wc2max)	
See also	NMR Spectroscopy User Guide	
Related	isadjAdjust integral scale (M)sc2Start of chart in second direction (P)thadjAdjust threshold for peak printout (M)vsVertical scale (P)	

V

vsadj	Automatic vertical scale adjustment (M)
vsadj2	Automatic vertical scale adjustment by powers of two (M)
vsadjc	Automatic vertical scale adjustment for ¹³ C spectra (M)

vsproj Vertical scale for projections and traces (P)

- Description Sets a multiplier that is adjusted to produce a desired vertical scale for projections or traces of 2D data sets. vsproj can be explicitly adjusted by setting it to a value or by clicking the middle mouse button when pointing at the projection or trace. When interactively adjusting the scale with the mouse, the higher the pointer is in the trace display, the larger the vertical scale. If the parameter does not exist, it can be created by running the par2d macro.
 - See also NMR Spectroscopy User Guide
 - Related par2d Create 2D acquisition, processing, and display parameters (M) vs Select vertical scale (C)
 - S Select vertical scale (0)
 - vs2d Adjust vertical scale for 2D displays (M)

vtairflow Variable Temperature Air Flow (P)

Description	This global parameter sets the VT air flow, in l/min. The adjustment is coarse, +/- 1 l/min. If there is not enough air flow available it may not reach the requested value.	
Values	0 - 25	
Related	pin vtairlimit s	Pneumatics router interlock (P) Variable temperature air flow limits (P)

vtairlimits Variable Temperature Air Flow Limits (P)

Description This global parameter determines the range of safe VT air flow, as indicated by the LEDs on the flow meter. It sets the LEDs on the air flow meter, upper and lower LEDs are orange, in between are green. As long as the ball in the air flow meter is next to a green LED the air flow is considered safe. If the air flow drops or increases such that the ball is next to an orange LED, the pneumatics box will turn the VT Controller off and notify the experiment, provided the switch is in the 'run' position. A bit value of 1 sets an unsafe orange state, a bit value of 0 sets a safe green state. To create the parameter:

	create('vt setlimit('	airlimits','integer','global') vtairlimits',1023,0,1,'global')
Examples	a value of 775 or 0x307 will set the two lower and the three upper LEDs (orange) and clear the remaining 5 in between (green). Note tha the upper bits determine the lower LEDs. If the parameter does not exist the value defaults to 0x307 for liquids; 0x200 for solids.	
Values	0 - 1023	
Related	pin	Pneumatics router interlock (P)
	tin	Temperature interlock (P)
	vtairflow	Variable temperature air flow (P)

vtc Variable temperature cutoff point (P)

Applicability	Systems with a variable temperature (VT) module.	
Description	Sets a VT cutoff point. Above this temperature, VT air flows straight into the probe, past the heater, then past the sample. Below this temperature, air goes first through the heat exchange bucket, for cooling by the heat exchange fluid, and then into the probe and past the heater.	
Values	0 to 50, in degrees celsius. vtc is typically set 5°C higher than the supply gas used for VT regulation.	
See also	NMR Spectroscopy User Guide	
Related	tempSample temperature (P)tinTemperature interlock (P)	

vtcomplvl Variable temperature compensation for gradient shimming (P)

Description Specifies the level of VT compensation used by gradient shimming. Values 0, disable VT compensation.

- 1, enable VT compensation
- 2, enable VT compensation with extra gradient dephasing.
- Related gmapz Get parameters and files for gmapz pulse sequence (M) gmapsys Run gradient autoshimming, set parameters, map shims (M) gzsize Number of z-axis shims used by gradient shimming (P) temp Sample temperature (P) vttype Variable temperature controller present (P)

V

vttype Variable temperature controller present (P)

Description In the Spectrometer Configuration window, this parameter specifies whether a variable temperature (VT) controller is present or not on the system. The value is set using the VT Controller label in the Spectrometer Configuration window.

When entered from command line in VNMR, control of the variable temperature (VT) controller from the current experiment is either engaged (*vttype=2*) or disengaged (*vttype=0*). The current state of the variable temperature (VT) controller is not changed when *vttype* is set in the command window.

The variable temperature (VT) controller setting in Spectrometer Configuration is not affected by entering *vttype* on the command line.

Values 2 is setting for VT controller (Present choice in Spectrometer Configuration window).

0 is setting for no VT controller (Not Present choice in Spectrometer Configuration window).

- Examples If temp='some temperature' while vttype=2 and vttype is then changed to vttype=0 on the command line, the variable temperature (VT) controller will continue regulate the sample at the value set by temp. While vttype=0 changes to temp will have no effect.
- See also VnmrJ Installation and Administration; NMR Spectroscopy User Guide
 - Related config Display current configuration and possibly change values (M)
 - masvt Type of variable temperature system (P)

vtwait Variable temperature wait time (P)

- Applicability Systems with a variable temperature (VT) module.
 Description Sets a time for establishing temperature regulation. If temperature interlock tin is set and regulation is not established after the time set by vtwait, VNMR displays the message "VT FAILURE" and aborts the experiment.
 Values Number, in seconds, A typical value is 180 seconds.
 See also NMR Spectroscopy User Guide
 Related pad Preacquisition delay (P)
 - tin Temperature interlock (P)

vxr_unix Convert VXR-style text files to UNIX format (M, U)

Syntax	<pre>(From VNMR) vxr_unix(VXR_file<,UNIX_file>) (From UNIX) vxr_unix VXR_file UNIX_file</pre>	
Description	Converts a VXR-style text file (from a Gemini, VXR, or XL system) to the UNIX format.	
Arguments	VXR_file is the name of the input file, which must be a text file.	
	UNIX_file is the name of the output file after conversion. The names of the input and output files must be different.	
Examples	<pre>(From VNMR) vxr_unix('oldtextfile','newtextfile') (From UNIX) vxr_unix oldtextfile newtextfile</pre>	
See also	NMR Spectroscopy User Guide	
Related	convertConvert data set from a VXR-style system (C,U)decompDecompose a VXR-style directory (C)	



Agilent VnmrJ 4 Command and Parameter Reference Guide

W	Who is using system (C)
walkup	Walkup automation (M)
walkupQ_runtime	Macro to Control Study Queue
waltz	WALTZ decoupling present (P)
warmprobe	Tells the system a warm probe is present
wbs	Specify action when bs transients accumulate (C)
wbs	When block size (P)
WC	Counts Words in a String
wc2	Width of chart in second direction (P)
wcmax	Maximum width of chart (P)
wc2max	Maximum width of chart in second direction (P)
wdone	Specify action when experiment is done (C)
wdone	Specify action when experiment is done (P)
wds	
werr	Specify action when error occurs (C)
werr	When error (P)
wet	Flag to turn on or off wet solvent suppression ((P)
Wetld	Set up parameters for wet ¹ H experiment (M)
wetdqcosy	Set up parameters for a WETDQCOSY pulse sequence (M)
wetgcosy	Set up parameters for a WETGCOSY pulse sequence (M)
wetghmqcps	Set up parameters for a WETGHMQCPS pulse sequence (M)
wetghsqc	Set up parameters for a WETGHSQC pulse sequence (M)



wetgmqcosy	Set up parameters for a WETGHSQC pulse sequence (M)
wetit	Set up and create pulse shapes for Wet1d experiment (M)
wetnoesy	Set up parameters for a WETNOESY pulse sequence (M)
wetpeaks	Number of peaks for wet solvent suppression (P)
wetpwxcal	Set up parameters for a WETPWXCAL pulse sequence (M)
wettntocsy	Set up parameters for a WETTNTOCSY pulse sequence (M)
wetshape	Shape for pwwet pulses (P)
wexp	Specify action when experiment completes (C)
wexp	When experiment completes (P)
wf	Width of FID (P)
wf1	Width of interferogram in 1st indirectly detected dimension (P)
wf2	Width of interferogram in 2nd indirectly detected dimension (P)
wfgtest	Waveform generator test (M)
wft	Weight and Fourier transform 1D data (C)
wftld	Weight and Fourier transform ${\rm f_2}$ for 2D data (C)
wft1da	Weight and Fourier transform phase-sensitive data (M)
wft1dac	Combine arrayed 2D FID matrices (M)
wft2d	Weight and Fourier transform 2D data (C)
wft2da	Weight and Fourier transform phase-sensitive data (M)
wft2dac	Combine arrayed 2D FID matrices (M)
wftt3	Process f ₃ dimension during 3D acquisition (M)
which	Display which command or macro is used (M)
wnt	Specify action when nt transients accumulate (C)
wnt	When number of transients (P)
wp	Width of plot in directly detected dimension (P)
wp1	Width of plot in 1st indirectly detected dimension (P)

wp2	Width of plot in 2nd indirectly detected dimension (P)
write	Write formatted text to a device (C)
writefid	Write numeric text file using a FID element (C)
writejxy	Create x,y ascii file from phasefile for JCAMP-DX conversion (M)
writeparam	Write one of more parameters to a file (C)
writespectrum	Write a spectrum to a binary file (C)
writetrace	Create ascii file from phasefile (f1 or f2) trace (M)
writexy	Create x,y ascii file from phasefile (f1 or f2) trace (M)
wrtp	Command string executed after rtp command (P)
wsram	Send hardware configuration to acquisition console (C)
wstart	
wshim	Conditions when shimming is performed (P)
wtfile	User-defined weighting in directly detected dimension (P)
wtfile1	User-defined weighting in 1st indirectly detected dimension (P)
wtfile2	User-defined weighting in 2nd indirectly detected dimension (P)
wtgen	Compile user-written weighting functions (M,U)
wti	Interactive weighting (C)
wtia	Interactive weighting for 2D absorptive data (M)
wtune	Specify when to tune (P)
wtunedone	What to do after ProTune tuning is done (P)
wysiwyg	Set plot display or full display (P)

W

Who is using system (C)

Displays information about users currently on the system. It functions Description like the UNIX command of the same name.

See also User Programming

walkup Walkup automation (M)

- Description Enables using sample changers for continuous "walk-up" operation. Click on Utilities -> New automation run to run this macro from the VnmrJ Walkup interface. The macro creates a new automation directory each day with the name auto_yyyy.mm.dd, where yyyy is the year, dd is the day of the month, and mm is the month (e.g., auto_20040601). The automation directory is saved in a directory specified by the global parameter globalauto. walkup creates the directory globalauto and the parameter globalauto, and then sets the globalauto parameter. See also VnmrJ Walkup
 - Related enter Enter sample information for automation run (M,U) globalauto Automation directory name (P)

walkupQ_runtime Macro to Control Study Queue

```
Syntax
       """ walkupQ_runtime - this is typically used at runtime
       by ""
                        "" CMD protocols to modify the queue as
                        appropriate ""
                        "" Usage:
                        walkupQ_runtime(keyword1, experiment/nod
                        e,keyword2,..) ""
                        . .
                             keyword1 = add / delete /customize
                        . .
                        п п
                              arg2 = '' is interpreted as ALL
                        experiments in the queue ""
                        "" arg2='' and keyword1='add' is invalid
                        . .
                        п п
                             keyword2 = next / last / all / night
                        / '' / node
                                        "" keyword2='node' interprets arg2 as
                        nodename ""
                        п п
                             else arg2 is experimentname ""
                        "" keyword2='node' and arg2='' is
                        invalid combination ""
                        "" keyword2='all' and keyword1='add' is
                        invalid
                                 .....
                        "" keyword2='' is same as
                        keyword2='next'
                        "" keyword2='night' and arg2='' include
                        an implicit 'all' ""
```

```
. .
                                        4th argument is required for
                                 customize option
Description this is typically used at runtime by ""
                                 "" CMD protocols to modify the queue as appropriate
                                              пп
Arguments
             .....
                 Number of Arguments: 3
                                 "" walkupQ runtime('add','gHSQCAD','next') ""
                                 "" Adds gHSQCAD as next experiment in the queue ""
                                 "" walkupQ runtime('add','gHSQCAD','gCOSY 02')
                                 ••••
                                 "" Add gHSQCAD after gCOSY 02 node ""
                                 "" walkupQ runtime('add','gHSQCAD','last') ""
                                 "" Add gHSQCAD as last experiment in the queue ""
                                 "" walkupQ runtime('add','gHSQCAD','night') ""
                                 "" Add gHSQCAD to the night queue ""
                                 ******
                                 "" Number of Arguments: 3
                                                            ....
                                 "" walkupQ runtime('delete',",'next')
                                                                   ....
                                 "" Delete the next experiment in the queue
                                                                       ""
                                 "" walkupQ runtime('delete',",'last') ""
                                 "" Delete the last experiment in the queue ""
                                 "" walkupQ runtime('delete',",'all') ""
                                 "" Delete all pending experiments in the queue ""
                                 "" walkupQ runtime('delete','HSQCAD','next or last or
                                 all') ""
                                 "" Delete next(last or all) HSQCAD experiments ""
                                 "" walkupQ runtime('delete','gHSQCAD 02','node')
                                 ....
                                 "" Delete gHSQCAD 02 in the queue ""
                                     *****
                                                                 *****
                                 *******
                                                            ""
                                 "" Number of Arguments: 4
                                 "" walkupQ runtime('customize',",'keyword','nt=32')
                                 ....
                                 ....
                                                                             ....
                                      keyword='next' or 'last' or 'all'
                                 ....
                                      Set nt=32 for the next/last/all experiments
                                                                                ....
                                 ....
                                 walkupQ runtime('customize','HSQCAD','keyword','nt
                                 =32') ""
                                 "" keyword='next' or 'last' or 'all' ""
```

W

"" Set nt=32 for the next/last/all HSQCAD experiments

walkupQ_runtime('customize','gHSQCAD_02','node','n
t=32') ""

"" Set nt=32 for the gHSQCAD_02 experiment in the queue ""

waltz WALTZ decoupling present (P)

Description	Sets whether system is equipped for WALTZ decoupling. The value is changed by normal parameter entry rather than using the Spectrometer Configuration window
Values	'n' sets WALTZ decoupling not present.

'y' sets WALTZ decoupling present.

"

See also VnmrJ Installation and Administration

warmprobe Tells the system a warm probe is present

Applicability	VnmrJ 3.1
Description	If a C13 observe coldprobe is being used, the value of rof2 should not be less than 350 user. The coldprobe macro tells the system that
	a coldprobe is present so that the rof2 rule is enforced.
	The warmprobe macro tells the system that a warm probe is present so that the rof2 rule is not enforced.

Related coldprobe Tells the system a coldprobe is present

wbs Specify action when bs transients accumulate (C)

Syntax	wbs(string)
Description	Specifies what action to take when bs transients accumulate. The <i>command</i> wbs sets the corresponding <i>parameter</i> wbs. Using the command, rather than setting the parameter value explicitly, notifies the acquisition process that the associated parameter value has changed. Thus, the desired operation can be effected even if the experiment has already started.
Arguments	string is a string argument containing the command or macro to be executed when this event happens. The string must be enclosed in single quotes. If single quotes are required <i>within</i> the text string, place a backslash character before each of the interior single quotes $(\)$.

	Maximum length of the string is 256 characters. To turn off wbs processing, enter wbs(''), where the argument is two single quotes with no space between.	
Syntax	wbs('dg wf wbs('mf(3) wbs('')	t') ')
See also	NMR Spectro	scopy User Guide
Related	bs	Block size (P)
	makefid	Make a FID element using numeric text input (C)
	phfid	Zero-order phasing constant for np FID (P)
	wbs	When block size (P)
	werr	Specify action when error occurs (C)
	wexp	Specify action when experiment completes (C)
	wnt	Specify action when nt transients accumulate (C)

wbs When block size (P)

Description	Invokes an action to occur automatically after each bs block of transients is completed. For example, wbs='wft' results in an automatic weighting and Fourier transformation after each bs transients. To specify no wbs processing, set wbs to the null string. If the acquisition has already started, the wbs <i>command</i> must be used to change this parameter.	
Values	Command, macro, or null string (wbs='', where the value is given by two single quotes with no space between them).	
See also	NMR Spectroscopy User Guide	
Related	bsBlock size (P)wbsSpecify action when bs transients accumulate (C)	

WC

Counts Words in a String

Syntax	wc(string)
Description	Utility to emulate the "wc -w" command in Unix. Called on a string
	variable, it returns the number of words in the string.
Examples	<pre>wc('textfile'):r1</pre>

wc2 Width of chart in second direction (P)

Description Specifies width of chart (plotting or printing area) along the second axis (or y axis) of a 2D contour plot or 2D "stacked display." For plots

made in the cutoff mode, wc2 specifies the width of the plotted area along the y-axis.

Values	Width, in mm.	
See also	NMR Spectroscopy User Guide	
Related	cutoff	Data truncation limit (P)
	ho	Horizontal offset (P)
	sc2	Start of chart in second direction (P)
	wcmax	Maximum width of chart (P)
	wc2max	Maximum width of chart in second direction (P)

wcmax Maximum width of chart (P)

Description	Specifies the maximum width of a chart (plotting or printing area). Set when plotter or printer is installed.	
Values	Width, in mm.	
See also	NMR Spectroscopy User Guide	
Related	wc Width of chart (P)	
	wc2 Width of chart in second direction (P)	

wc2max Maximum width of chart in second direction (P)

Description	Specifies the maximum width of a chart (plotting or printing area) in the second direction $(y$ -axis). Set when the plotter or printer is installed.	
Values	Width, in mm.	
See also	NMR Spectro	scopy User Guide
Related	wc2	Width of chart in second direction (P)
	wcmax	Maximum width of chart (P)

wdone Specify action when experiment is done (C)

Syntax	wdone(string)
Description	Specifies the action to take when the experiment is done, after wexp has been executed. The wdone command sets the corresponding parameter wdone. Using the command, rather than setting the parameter value explicitly, notifies the acquisition process that the associated parameter value has changed and the desired operation is affected area if the ameriment has alwardy started
	enected even if the experiment has aready started.
Arguments	The string argument contains the command or macro to be executed when the experiment is done. The string must be enclosed in single
quotes. If single quotes are required within the text string, place a backslash character before each of the interior single quotes ('). Maximum length of the string is 256 characters.

'' (null string) turns off wdone processing.

Related wexp Specify action when experiment completes (C)

wdone Specify action when experiment is done (P)

Syntax wdone'<command, macro, or null string >'

Description Invokes a single action to occur just after wexp is executed. As with wexp, it is executed automatically after the experiment is finished, which can occur at the end of a single FID or after the last fid in a multi-FID experiment. To specify no wdone processing, set wdone to the null string. If the acquisition has already started, the wdone command must be used to change the wdone parameter. For wdone to execute after an experiment finishes and after wexp has executed, start the experiment with the au command.

If the wexp action sets the wdone parameter, the new value of the wdone parameter will be executed and the old value will be ignored.

werr Specify action when error occurs (C)

Syntax we:	rr(string)
------------	------------

Description Specifies what action to take if an error occurs during acquisition. The *command* werr sets the corresponding *parameter* werr. Using the command, rather than setting the parameter value explicitly, notifies the acquisition process that the associated parameter value has changed. Thus, the desired operation can be effected even if the experiment has already started.

Arguments string is a string argument containing the command or macro to be executed when this event happens. The string must be enclosed in single quotes. If single quotes are required *within* the text string, place a backslash character before each of the interior single quotes (\'). Maximum length of the string is 256 characters. To turn off werr processing, enter werr(''), where the argument is two single quotes with no space between them.

Examples	werr('react werr('')	t')
See also	NMR Spectro	scopy User Guide
Related	wbs	Specify action when bs transients accumulate (C)
	werr	When error (P)
	wexp	Specify action when experiment completes (C)
	wnt	Specify action when nt transients accumulate (C)

werr When error (P)

Description	Specifies a r action when processing, s been started <i>parameter</i> . A werr in acq refer to the o directory /v	macro (e.g., werr='react') that will take appropriate an error occurs during acquisition. To specify no werr set werr to the null string. If the acquisition has already , the werr <i>command</i> must be used to change the werr Arrayed parameter acqstatus provides the error code to status[1] and acqstatus[2]. For a list of error codes, description of acqstatus or view the file acq_errors in nmr/manual.
Values	Macro or nu quotes with	ll string (werr='', where the value is given by two single no space between them).
See also	NMR Spectre	oscopy User Guide
Related	acqstatus react werr	Acquisition status (P) Recover from error conditions during werr processing (M) Specify action when error occurs (C)

wet Flag to turn on or off wet solvent suppression ((P)

Description Specifies if wet solvent suppression is turned on or off. It is now a standard option in many liquids pulse sequences, including Wet1d and sequences of apptype hetero2d and homo2d.

apptype	Application type (P)
hetero2d	Execute protocol actions of apptype hetero2d (M)
homo2d	Execute protocol actions of apptype homo2d (M)
std1d	Execute protocol actions of apptype std1d (M)
Wet1d	Set up parameters for a WET1D pulse sequence (M)
	apptype hetero2d homo2d std1d Wet1d

Wet1d Set up parameters for wet ¹H experiment (M)

Description Set up parameters for wet ¹H experiment.

wetdqcosy Set up parameters for a WETDQCOSY pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Description	Sets up for a WETDQCOSY LC-NMR experiment.
See also	NMR Spectroscopy User Guide

wetgcosy Set up parameters for a WETGCOSY pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Description	Sets up for a WETGCOSY LC-NMR experiment.
See also	NMR Spectroscopy User Guide

wetghmqcps Set up parameters for a WETGHMQCPS pulse sequence (M)

ApplicabilitySystems with LC-NMR accessory.DescriptionSets up for a WETHMQCPS LC-NMR experiment.See alsoNMR Spectroscopy User Guide

wetghsqc Set up parameters for a WETGHSQC pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Syntax	wetghsqc('nucleus')
Description	Sets up for a WETGHSQC LC-NMR experiment.
See also	NMR Spectroscopy User Guide

wetgmqcosy Set up parameters for a WETGHSQC pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Description	Sets up for a WETGMQCOSY LC-NMR experiment.
See also	NMR Spectroscopy User Guide

wetit Set up and create pulse shapes for Wet1d experiment (M)

Applicability	VnmrJ Walkup
Description	A macro to set up and create pulse shapes for a Wetld experiment.
	It is based on suppressing the largest N peaks found in a spectrum.
Related	wetpeaks (P)

1013

wetnoesy Set up parameters for a WETNOESY pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Description	Sets up for a WETNOESY LC-NMR experiment.
See also	NMR Spectroscopy User Guide.

wetpeaks Number of peaks for wet solvent suppression (P)

Applicability	Walkup
Description	Sets the number of peaks to be suppressed by wet solvent suppression for the Wetld protocol. The wetit macro suppresses the N tallest peaks found in the scout spectrum, where N is specified by wetpeaks. The parameter is set by the <i>Number of peaks to suppress menu</i> on the Prescan page.
Values	1 to 7 for DirectDrive or UnityInova systems; 3 for Mercury systems are the default values.
Related	Wetld Set up parameters for wet 1H experiment (M) wetit Set up and create pulse shapes for Wetld experiment (M)

wetpwxcal Set up parameters for a WETPWXCAL pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Description	Sets up for a WETPWXCAL LC-NMR pulse width calibration.
See also	NMR Spectroscopy User Guide

wettntocsy Set up parameters for a WETTNTOCSY pulse sequence (M)

Applicability	Systems with LC-NMR accessory.
Description	Sets up for a WETTNTOCSY LC-NMR experiment.
See also	NMR Spectroscopy User Guide

wetshape Shape for pwwet pulses (P)

Applicability	Systems with LC-NMR accessory.
Description	Sets the name of the shape used for pwwet pulses (e.g.,
	wetshape='wet').
See also	NMR Spectroscopy User Guide

Syntax wexp(string)

Description	Specifies what action to take when the experiment completes. The
	wexp command sets the corresponding parameter wexp. Using the
	command, rather than setting the parameter value explicitly, notifies
	the acquisition process that the associated parameter value has
	changed. Thus, the desired operation can be effected even if the
	experiment has already started.

Arguments string is a string argument containing the command or macro to be executed when the experiment completes. The string must be enclosed in single quotes. If single quotes are required *within* the text string, place a backslash character before each of the interior single quotes ('). Maximum length of the string is 256 characters. To turn off wexp processing, enter wexp(''), where argument is two single quotes with no space between them.

Examples	wexp('wft(wexp('')	\'all\') calcT1')
See also	NMR Spectro	oscopy User Guide
Related	wbs	Specify action when bs transients accumulate (C)
	werr	Specify action when error occurs (C)
	wexp	When experiment completes (P)
	wnt	Specify action when nt transients accumulate (C)

wexp When experiment completes (P)

Description Invokes a single action to occur automatically after the experiment is finished, which can occur after a single FID or after a number of FIDs in a multi-FID experiment. To specify no wexp processing, set wexp to the null string. If the acquisition has already started, the wexp command must be used to change the wexp parameter. For wexp to execute after an experiment finishes, start the experiment with the au command.

> wexp processing occurs after wnt processing in a single FID experiment, and both can be used. wexp also occurs after wnt during the last FID of a multi-FID experiment. Thus, wnt='wft(\'all\')' wexp='calcT1' and wexp='wft(\'all\') calcT1' transforms each FID in a T_1 experiment as it is performed, and when each of the FIDs has been collected, performs the calculation of the T_1 using a hypothetical macro command calcT1. Notice the use of the backslash to include a single quotation mark inside the string.

Values Command, macro, or null string (wexp='', where the value is given by two single quotes with no space between them). If the command or macro uses a file name as an argument, specifying an absolute path is best. Be sure the path is valid and you have the appropriate write permission.

W

See also	NMR Spectroscopy User Guide		
Related	wexp	Specify action when experiment completes (C)	
	wnt	When number of transients (P)	
	au	Submit experiment to acquisition and process data (C)	

wf Width of FID (P)

Description Width of the FID display. This parameter can be entered in the usual way or interactively controlled by selecting the sf wf button during a FID display.

Values 0 to the value of at, in seconds.

See also NMR Spectroscopy User Guide

Related	at	Acquisition	time	(P)	
					-

- dcon Display noninteractive color intensities map (C)
- dconi Interactive 2D data display (C)
- df Display a single FID (C)
- sf Start of FID (P)
- vf Vertical scale of FID (P)
- wfl Width of interferogram in 1st indirectly detected dimension (P)
- wf2 Width of interferogram in 2nd indirectly detected dimension (P)

wf1 Width of interferogram in 1st indirectly detected dimension (P)

- Description Sets the width of the interferogram display in the first indirectly detected dimension.
 - Values 0 to $(2 \times ni)/sw1$, in seconds.
 - See also NMR Spectroscopy User Guide
 - Related ni Number of increments in 1st indirectly detected dimension (P)
 - sf1 Start of interferogram in 1st indirectly detected dimension (P)
 - sw1 Spectral width in 1st indirectly detected dimension (P)
 - wf Width of FID (P)

w£2 Width of interferogram in 2nd indirectly detected dimension (P)

Description	Sets the width of the interferogram display in the second indirectly detected dimension.			
Values	0 to (2	0 to $(2 \times ni2)/sw^2$, in seconds.		
See also	NMR S ₁	NMR Spectroscopy User Guide		
Related	ni2	Number of increments in 2nd indirectly detected dimension (P)		
	sf2	Start of interferogram in 2nd indirectly detected dimension (P)		
	sw2	Spectral width in 2nd indirectly detected dimension (P)		
	wf	Width of FID (P)		

wfgtest Waveform generator test (M)

Applicability	Systems with a waveform generator.
Description	Retrieves a parameter set and pulse sequence, and compiles the
	sequence, in order to set up an experiment to test the waveform generators.
See also	Waveform Generator Kit Installation

wft Weight and Fourier transform 1D data (C)

Syntax	<pre>(1) wft<(<opt (2) wft('inve</opt </pre>	tions,><'nf'><,start><,finish><,step>)> erse',exp_number,expansion_factor)
Description	Performs a Fourier transform on one or more 1D FIDs with weighting applied to the FID. The command executes a left-shift, zero-order phase rotation, and a frequency shift according to the parameters lsfid, phfid, and lsfrq, respectively, on the time-domain data prior to the weighting and Fourier transformation. The type of Fourier transformation to be performed is determined by proc. wft uses the same arguments as the command ft, and except for weighting, it functions the same as the ft command.	
See also	NMR Spectrose	copy User Guide
Related	ft lsfid lsfrq phfid proc	Fourier transform 1D data (C) Number of points to left-shift np FID (P) Frequency shift of the fn spectrum in Hz (P) Zero-order phasing constant for np FID (P) Type of processing on np FID (P)

Syntax	(1) wft1d(ele	ement_number)	
	(2) wft1d<(<	options,> <coefficients>)></coefficients>	
Description	Performs the first Fourier transformation along the dimension defined by sw , with weighting and matrix transposition. This allows the display of t_1 interferograms with the dcon and dconi commands.		
	Except for weig See the descrip	wftld functions the same as the ftld command. ption of ftld for further information.	
Arguments	Same as the arguments to ftld. See the ftld command for details.		
See also	NMR Spectroscopy User Guide		
Related	dcon	Display noninteractive color intensity map (C)	
	dconi	Interactive 2D data display (C)	
	ft1d	Fourier transform along f_2 dimension (C)	
	SW	Spectral width in directly detected dimension (P)	

wftlda Weight and Fourier transform phase-sensitive data (M)

wftlda<(options)>		
Processes 2D FID data as well as 2D planes at particular t_1 or t_2 times from a 3D data set for a pure absorptive display.		
wftlda d data is pe of ftlda	iffers from ftlda only in that weighting of the time-domain erformed prior to the Fourier transform. See the description for further information.	
Same as arguments to ft2da. See the ft2da command for details.		
NMR Spectroscopy User Guide		
ft1da ft2da wft2da	Fourier transform phase-sensitive data (M) Fourier transform phase-sensitive data (M) Weight and Fourier transform phase-sensitive data (M)	
	wftlda<0 Processes from a 3E wftlda d data is pe of ftlda Same as a <i>NMR Spec</i> ftlda ft2da wft2da	

wftldac Combine arrayed 2D FID matrices (M)

Syntax	wft1dac<(<mult1>,<mult2>, ,<multn>)></multn></mult2></mult1>
Description	Allows the ready combination of 2D FID matrices within the framework of the 2D Fourier transform program. Weighting is performed. This command requires that the data be acquired either without f_1 quadrature or with f_1 quadrature using the TPPI method. wftldac is used with TOCSY (with multiple mixing times).
Arguments	mult1,mult2,,multn are multiplicative coefficients. The n th argument is a real number and specifies the multiplicative coefficient for the n th 2D FID matrix.

See also	NMR Spectroscopy User Guide	
Related	ft1dac	Combine arrayed 2D FID matrices (M)
	Tocsy	Set up parameters for TOCSY pulse sequence (M)
	wft2dac	Combine arrayed 2D FID matrices (M)

wft2d Weight and Fourier transform 2D data (C)

Syntax wft2d<(<options,>coefficients)> Description Performs a complete 2D transformation with weighting after 2D data has been acquired. If the first Fourier transformation has already been done using ftld, wftld, ftlda, or wftlda, then the wft2d command performs only the second transform. For arrayed 2D experiments, a single array element can be transformed and weighted using the array element number as an argument. Interferograms can be constructed explicitly using the following coefficient table: wft2d(rr1, ir1, rr2, ir2, ... ri1, ii1, ri2, ii2, ...). wft2d('ptype',...) transforms P-type spectra, and wft2d('ntype',...) transforms N-type spectra. The default is N-type. wft2d also completes a 2D transform that has been started with wftld (or related commands such as wftlda). The first transform will not be done again if it has already been performed. For phase-sensitive 2D experiments, the coefficients must be applied as part of the first transform (e.g., with wftlda) since the interferograms are formed at that stage. These coefficients need not be repeated when invoking the subsequent transform: a simple wft2d or ft2d can suffice. See the ft2d command description for further information. Same as the arguments to ft2d. See the ft2d command for details. Arguments Examples wft2d(1,0,0,0) wft2d(2) wft2d(1,0,1,0,0,1,0,1) wft2d(.67,0,.33,0,0,.67,0,.33) NMR Spectroscopy User Guide See also Related dconi Interactive 2D data display (C) Fourier transform along f₂ dimension (C) ft1d Fourier transform "halfway" for pure absorption 2D data ft1da (M) ft2d Fourier transform 2D data (C) wft1d Weight and Fourier transform f₂ for 2D data (C) wft1da Weight and FT "halfway" for pure absorption 2D data (M) wft2da Weight and transform for pure absorption 2D data (M)

wft2da Weight and Fourier transform phase-sensitive data (M)

Syntax	wft2da<(options)>		
Description	Processes 2D FID data, as well as 2D planes at particular $t_1 \mbox{ or } t_2$ times, from a 3D data set for a pure absorptive display.		
	wft2da differs from ft2da only in that weighting of the time-domain data is performed prior to the Fourier transform. See the description of ft2da for further information.		
Arguments	Same as used with ft2da. See the ft2da command for details.		
See also	NMR Spectroscopy User Guide		
Related	ftlda Fourier transform phase-sensitive data (M)		
	ft2da Fourier transform phase-sensitive data (M)		
	wftlda Weight and Fourier transform phase-sensitive data (M)		

wft2dac Combine arrayed 2D FID matrices (M)

Syntax wft2dac<(<mult1><,mult2>,...<,multn>)>

Allows the ready combination of 2D FID matrices within the framework Description of the 2D Fourier transform program. Weighting is performed. This command requires that the data be acquired either without f_1 quadrature or with f_1 quadrature using the TPPI method. wft2dac is used with TOCSY (with multiple mixing times). mult1, mult2, ..., multn are multiplicative coefficients. The nth Arguments argument is a real number and specifies the multiplicative coefficient for the *n*th 2D FID matrix. See also NMR Spectroscopy User Guide Related ftldac Combine arrayed 2D FID matrices (M) Combine arrayed 2D FID matrices (M) ft2dac Set up parameters for TOCSY pulse sequence (M) Tocsy

Combine arrayed 2D FID matrices (M)

wftt3 Process f₃ dimension during 3D acquisition (M)

wft1dac

Description Allows f_3 processing of 3D data to be performed concurrently with data acquisition. To invoke this function, set wnt='wftt3' and use au to start the acquisition of the 3D data. When wftt3 detects that all the FIDs comprising a (t1,t2) block have been acquired, it starts up the ft3d program in background to process that block of FIDs in f_3 . The 3D processing information file, created by entering set3dproc within VnmrJ, does not need to contain valid f_1 and f_2 processing information but only valid f_3 processing information. Once the f_3 processing is complete, a new 3D information file can be created for the f_1 - f_2 processing stages that contains valid f_1 and f_2 processing information.

The non-standard string parameter path3d can be used to specify the directory into which the f₃ processed 3D data is to be stored. Normally, path3d is absent in the parameter set. If this is the case or if path3d='', the f₃-processed 3D data is stored in the directory curexp/datadir. path3d can be created by entering create('path3d','string') setgroup('path3d','display').

See also NMR Spectroscopy User Guide

Related	au	Submit experiment to acquisition and process data (C)
	create	Create new parameter in a parameter tree (C)
	ft3d	Perform a 3D Fourier transform (M,U)
	getplane	Extract planes from a 3D spectral data set (M)
	path3d	Path to currently displayed 2D planes from a 3D data
		set (P)
	select	Select a spectrum or 2D plane without displaying it (C)
	set3dproc	Set 3D processing (C)
	setgroup	Set group of a parameter in a tree (C)
	wnt	When number of transients (P)

which Display which command or macro is used (M)

Syntax	which(name)	
--------	-------------	--

Description Searches VnmrJ libraries and then displays on line 3 which VnmrJ command or macro with the given name will be executed. For macros, which displays the type of macro (user, local, application, or Agilent) and the path to the library.

- Arguments name is the name of a command or macro.
- Examples which('wft')
- See also User Programming
 - Related exists Determine if a parameter, file, or macro exists (C) hidecommand Execute macro instead of command with same name (M)

Specify action when nt transients accumulate (C) wnt

Syntax wnt(string)

Description Specifies what action to take when nt transients accumulate. The wnt command sets the corresponding parameter wnt. Using the command, rather than setting the parameter value explicitly, notifies the acquisition process that the associated parameter value has changed. Thus, the desired operation can be effected even if the experiment has already started.

Arguments	 string is a string argument containing the command or macro to be executed when this event happens. The string must be enclosed in single quotes. If single quotes are required within the text string, place a backslash character before each of the interior single quotes (\'). Maximum length of the string is 256 characters. To turn off wnt processing, enter wnt(''), where the argument is two single quotes with no space between them. 	
Examples	<pre>wnt('wft(\'all\')') wnt('')</pre>	
See also	NMR Spectroscopy User Guide	
Related	nt	Number of transients (P)
	wbs	Specify action when bs transients accumulate (C)
	werr	Specify action when error occurs (C)
	wexp	When experiment completes (P)
	wnt	When number of transients (P)

wnt When number of transients (P)

Description	Invokes a single action to occur automatically after the FID is finished	
	(ct=nt) or after each FID in a multi-FID experiment involving an	
	arrayed parameter. The most common processing to occur after an FID	
	is an automatic weighting and Fourier transformation (i.e.,	
	wnt='wft'); however, this is normally not needed because the	
	command ga is the exact equivalent of wnt='wft(\'acq\')' au (i.e.,	
	ga sets the wnt action automatically). To specify no wnt processing,	
	set wnt to the null string. If the acquisition has already been started,	
	the wnt command must be used to change this parameter.	
Values	Command, macro, or null string (wnt='', where the value is given by two single quotes with no space between them).	
See also	NMR Spectroscopy User Guide	
Related	nt Number of transients (P)	
	wnt Specify action when nt transients accumulate (C)	

wp Width of plot in directly detected dimension (P)

Description	Sets the width of the displayed or plotted region of the spectrum.	
Values	Always stored in Hz, but can be entered in ppm by using the p suffix (e.g., $wp=6p$ sets the width of plot to 6 ppm).	
See also	NMR Spectroscopy User Guide	
Related	wp1 Width of plot in 1st indirectly detected dimension (P)	
	Wp2 Width of plot in 2nd indirectly detected dimension (P)	

wp1 Width of plot in 1st indirectly detected dimension (P)

Description	Analogous to the wp parameter except that wp1 applies to the first indirectly detected dimension of a multidimensional data set.
See also	NMR Spectroscopy User Guide
Related	wpWidth of plot in directly detected dimension (P)wp2Width of plot in 2nd indirectly detected dimension (P)

Wp2 Width of plot in 2nd indirectly detected dimension (P)

Description	Analogous to the wp parameter except that wp2 applies to the second indirectly detected dimension of a multidimensional data set.
See also	NMR Spectroscopy User Guide
Related	WP Width of plot in directly detected dimension (P)
	wp1 Width of plot in 1st indirectly detected dimension (P)

write Write formatted text to a device (C)

Syntax	<pre>(1) write('keywords'><,color pen></pre>		
	<,'reverse'>,x,y<,template>) <:height>		
	(2) write('alpha' 'printer' 'line3' 'error',template)		
	<pre>(3) write('reset' 'file' 'fileline',file<,template>)</pre>		
	<pre>(4) write('net',host,port, template)`</pre>		

- Description Writes text to a graphics screen or plotter in a given format (syntax 1), writes formatted text to another device (syntax 2), clears a file (syntax 3), or writes to a file (syntax 3). The input to the command comes from arguments in template, which can be parameters such as n1 or pw.
- Arguments 'keywords' identify the output device ('graphics'|plotter') and the drawing mode ('xor'|'normal'|'newovly'|'ovly'| 'ovlyC').
 - 'graphics'| 'plotter' is a keyword selecting the output device. The default is 'plotter'. The output selected is passed to subsequent pen, move, or draw commands and remains active until a different mode is specified.
 - ''xor', 'normal' is a keyword for the drawing mode when using the 'graphics' output device. The default is 'normal'. In the 'xor' mode, if a line is drawn such that one or more points of the line are in common with a previous 'xor' line, the common points are erased. In the normal mode, the common points remain. The mode selected is passed to subsequent pen, move, and draw commands and remains active until a different mode is specified.

• 'newovly', 'ovly', and 'ovlyC' are keywords that specify an interactive drawing capability that is slightly slower than the 'xor' mode but more consistent in color. 'newovly' clears any previous draws, boxes, and writes made with the 'ovly' modes and draws the figure. 'ovly' draws without clearing so that multi-segment figures can be created. 'ovlyC' clears without drawing.

color is the color of the text on a color display: 'red', 'yellow', 'green', 'cyan', 'blue', 'magenta', and 'white'. The default is 'yellow'.

pen is the plotter pen: 'pen1', 'pen2', etc.

'reverse' is a keyword specifying a sideways orientation of the output.

x and y are coordinates on the screen or plotter, in mm.

template is a string of formatting characters along with arguments to those characters. The format is the same as used with the UNIX printf command (for details, see any basic UNIX manual or enter man printf in UNIX). For example, 'pw = %12.5f' is a template to format the parameter pw as fixed point with a field width of 12 spaces and 5 decimal places. The following format characters are implemented:

character	%C
integer	%d
hexadecimal	%h
exponential:	%e
fixed point	%f
exponential/fixed point	%g
octal	80
string	°∕S
write a % character	usewrite('%s','%'

height returns the height of the characters on the screen or plotter. This is useful for positioning multiple-line displays. See the source code of the macro dtext in the maclib directory for an example of usage.

)

'alpha' is a keyword to write text to the alphanumeric screen.

'printer' is a keyword to print text on the printer

'line3' is a keyword to write text as a message on line 3.

'error' is a keyword to write text as an error on line 3 and sound a beep.

'reset' is a keyword to clear the file specified.

'file' is a keyword to append data to the file specified. Existing data in the file is not overwritten. By writing repeated 'file' calls, a formatted data file can be created (see the fifth example below). Each write command automatically appends a carriage return (line feed) to the end of the string defined by the template argument. To append data without the automatic line feed, use the 'fileline' keyword instead of 'file'. Also, two backslashes () are interpreted as a new line.

'fileline' is a keyword to append data to the file specified, the same as using the 'file' keyword, but without automatically appending a carriage return (line feed) to the end of the data. Any line feeds desired must be explicitly defined (using $\n)$ by the template argument (see the sixth example below). Furthermore, two backslashes ($\n)$ output a single backslash into the file.

file is the name of the file used with the 'reset', 'file', and 'fileline' keywords.

'net' is a keyword for writing to a network program. The host name and port number must be supplied. The host name may also be an IP address, such as 10.190.x.y. The hostname of the local computer is stored in the instrument parameter. The command serverport may be used to get the port number for the currently executing VnmrJ program.

```
Examples write('graphics',100,100):$ys
write('plotter',20,180, 'pw = %12.5f',pw)
write('line3', 'Too many arguments')
write('reset','temp1')
write('file','temp1','%10f %10.1f',n1,pw)
write('fileline','temp1','\nEnd of data\n\n')
serverport:$port
write('net',instrument,$port,'banner(`hello`)')
See also User Programming
```

RelateddtextDisplay a text file in the graphics window (M)serverportReturns the value of the VnmrJ network listening
port (C)

writefid Write numeric text file using a FID element (C)

Syntax writefid(file<,element_number>)

- Description Writes a text file using data from the selected FID element. The program writes two values per line—the first is the value from the X (or real) channel and the second is the value from the Y (or imaginary) channel. writefid writes the raw FID data (i.e., FID data processing based on the parameters phfid, lsfid, and lsfrq does not occur).
- Argumentsfile is the name of a text file to store the data.element_number is an integer larger than 0 for the number of a FID
element. The default is 1.See alsoNMR Spectroscopy User Guide, User ProgrammingRelatedlsfidNumber of complex points to left-shift np FID
(P)lsfrgFrequency shift of fn spectrum in Hz (P)
 - lsfrqFrequency shift of fn spectrum in Hz (P)makefidMake a FID element using numeric text input
(C)

phfidZero-order phasing constant for np FID (P)writespectrumWrite a spectrum to a binary file (C)

writejxy Create x,y ascii file from phasefile for JCAMP-DX conversion (M)

Syntax	<pre>writejxy<(traceno)></pre>
Applicability	VnmrJ 3.1
Description	"writejxy" does almost the same as "writexy", but in a mode that
	is adjusted for calls by the "svxyj" macro (JCAMP-DX X,Y data
	conversion).

writeparam Write one of more parameters to a file (C)

Syntax	writeparam	<pre>file,parlist[,tree]['add'</pre>	<pre>'replace')</pre>
--------	------------	--------------------------------------	-----------------------

Description The writeparam command will write one or more parameters to a specified file.

Arguments The first argument is the name of the file. The second argument is a list of the names of the parameters to be written or it is the name of an arrayed temporary \$ variable. If it is a list, it is a string parameter and the names can be separated either by a space or a comma. If it is an arrayed temporary \$ varaible, each array element is a single parameter name. The optional third argument is the tree from which the parameters are copied. The variable trees are 'current', 'global'. 'processed' and 'systemglobal'. An optional final argument is the keyword 'add' or 'replace'. The add keyword will cause the parameters to be appended to the specified file. If they already exists in the file, their values will be updated. The replace keyword will replace the values in the file with the current values from the tree. The parameters must exist in both the file and the tree. A special case for the replace option occurs when the parameter list is an empty string. In this case, all the parameters in the file will be updated with the current values in the tree. If the parameter does not exist in the tree, no change will be made for that parameter.

This command may be used to store temporary values. For example, you may want to save wexp, wbs, wnt, etc. in order to run a setup acquisition. When it is done, you want to reset the original values. The fread command can to used to read the parameters back into an appropriate parameter tree.

Examples

writeparam(curexp+'/mypar','in') writes the parameter in into the file mypar in the current experiment directory.

writeparam(curexp+'/mypar','tin il','add') appends the
parameters tin and il to the file

writeparam(curexp+'/mypar','','replace') replace all of the parameters in mypar with the corresponding values from the current tree.

writeparam(curexp+'/mypar', 'sw ct np', 'processed') writes the parameters sw, ct, and np from the processed tree into the file mypar in the current experiement directory.

writeparam(curexp+'/mypar', 'at', 'processed', 'add')
appends the at parameter to the file.

writeparam(curexp+'/mypar', 'np sw d1', 'processed', 'replace') replace np and sw in mypar with the corresponding values from the processed tree. Since d1 did not exist in mypar, it is not added.

\$list='np sw d1'
writeparam(curexp+'/mypar',\$list,'processed','replace')
This is the same as the previous example.

\$arraylist='np','sw','d1'
writeparam(curexp+'/mypar','\$arraylist','processed','re
place') This is also the same as above example, however the variable
names are passed as an arrayed temporary \$ variable \$arraylist.
Note the single quotes around the second argument to writeparam. The
name of the local temporary \$variable is passed to the command, not
its value. This format is useful if the list of parameters to write
becomes large.

writespectrum write a spectrum to a binary file (C)

Description Writes out the current spectrum as a binary file. The file has no header information and is written in the native format (little-endian on Linux; big-endian on Solaris). writespectrum scales the data by vs, determines the mode selected, ph, av, or pwr, and writes whatever is displayed by ds. The file is written in the current experiment as specN, where N is the element number. Examples Write files spec1, spec2, spec3 ... spec{arraydim} in the current experiment directory: wft \$i=0 while (\$i < arraydim) do \$i = \$i + 1 select(\$i) writespectrum endwhile Write the real and imaginary components if phase mode is selected. wft

```
wit
ph
$i=0
$index=''
while ($i < arraydim) do
    $i = $i + 1
    format($i,0,0):$index
    select($i)</pre>
```

```
writespectrum
mv(curexp+'/spec'+$index, curexp+'/
spec'+$index+'.re')
rp = rp + 90
writespectrum
mv(curexp+'/spec'+$index, curexp+'/
spec'+$index+'.im')
rp = rp - 90
endwhile
```

```
Related writefid Write numeric text file using a FID element (C)
```

writetrace Create ascii file from phasefile (f1 or f2) trace (M)

Syntax	writetrace<(traceno)>		
Applicability	VnmrJ 3.1		
Description	ption "writetrace" creates an ASCII file from a phasefile trace in the current experiment. The argument indicates the number of the trace that is to be "asciified". The trace orientation depends on the orientation of the current data set (trace parameter). "writetrace works on fids (1D, arrayed, 2D), interferograms and 1D/2D spectr Trace counting starts at 1. The default trace is the current one. Th output will be written into a file in the current experiment, using th trace number as filename extension:		
	• curexp+'/trace.1': 1D spectrum (can be 1st of an array)		
	• curexp+'/trace.8': 8th trace from arrayed 1D data set		
	• curexp+'/f2trace.13': 13th f2 trace from 2D data set		
	• curexp+'/f1trace.1024': 1024th f1 trace from 2D data set		
	NOTE: the data MUST have been displayed using the "ds" (1D) or "dcon" or related (2D) commands, otherwise the phased spectrum is not even generated, and "writetrace" can't work. For 2D data, also traces that are currently not on display must have been displayed in the current orientation once before, otherwise they may not exist in phasefile!		
Examples	<pre>writetrace writetrace(13) writetrace(1024)</pre>		

writexy Create x,y ascii file from phasefile (f1 or f2) trace (M)

Syntax	writexy<(traceno)>
Applicability	VnmrJ 3.1
Description	"writexy" does the same thing as "writetrace", except that it creates
	an output file with x and y pairs (one pair per line, x values in

referenced Hz). Also here, the output will be written into a file in the current experiment, using the trace number as filename extension:

- curexp+'/xytrace.1': 1D spectrum (can be 1st of array)
- $\bullet curexp+'/xytrace.8':$ 8th trace from arrayed 1D data set
- curexp+'/f2xytrace.13': 13th f2 trace from 2D data set
- curexp+'/flxytrace.1024': 1024th f1 trace from 2D data set

Examples writexy writexy(13)

wrtp Command string executed after rtp command (P)

Description Holds the command string that is executed after an rtp command finishes. It is mostly used to set frequency-dependent parameter values, such as sw, so that one parameter set can be used on all spectrometers.

Examples wrtp='setsw(13p,-2p)'

wsram Send hardware configuration to acquisition console (C)

Syntax	wsram<:\$success>	
Description	Sends new hardware configuration information to the acquisition console when config is used (e.g., to set lockfreq). wsram (write to static RAM) is not normally entered directly by the user.	
Arguments	success returns 1 if wsram is successful, or 0 otherwise.	
See also	VnmrJ Installation and Administration.	
Related	configDisplay current configuration and possibly change it (M)lockfrLock frequency (P)	
	eq	

wshim Conditions when shimming is performed (P)

Description	Specifies when automatic shimming is to be used, according to the method specified by the parameter method.
Values	'n' sets that no automatic shimming is performed. Even with wshim set to this value, the shimming procedure specified by the parameter method can be activated by using the shim command.
	'e' or 'exp' sets that automatic shimming is done before data acquisition.

's' or 'samp' sets that automatic shimming is done only at the beginning of the first experiment, following the change of a sample using the automatic sample changer.

'g' sets that automatic shimming using gradient shimming is done only at the beginning of the first experiment, following the change of a sample using the automatic sample changer. The parameter method is ignored. This option is only available in automation and is not used with the go, ga, or au commands.

'f' or 'fid' set automatic shimming is done prior to the data collection of each new array member in a multi-FID experiment.

'fn', where *n* is an integer, sets shimming is done prior to data collection of every *n*th FID (e.g., wshim='f16' shims prior to acquiring FIDs 1, 17, 33, etc.). This method is only relevant to arrayed or 2D experiments.

See also NMR Spectroscopy User Guide

Related gf Prepare parameters for FID/spectrum display in acqi (M) method Autoshim method (P)

wtfile User-defined weighting in directly detected dimension (P)

Description Set to name of the file containing the user-written weighting function along the directly detected dimension. This dimension is referred to as the f_2 dimension in 2D data sets, the f_3 dimension in 3D data sets, etc. The shellscript wtgen is used to compile the user-written weighting module into an executable program. The source file is stored in the directory vnmruser+'/wtlib' with a .c file extension. The executable file is in the same directory and has the same name as the source file but has no file extension.

Values file is the name of the executable weighting function or the name of the weighting function text file.

'' (two single quotes with no space in between) indicates wtfile is inactive and VnmrJ should not look for a user-written weighting function.

- See also NMR Spectroscopy User Guide; User Programming
 - Related wtfile1 User-defined weighting in 1st indirectly detected dimension (P)
 - wtfile2 User-defined weighting in 2nd indirectly detected dimension (P)
 - wtgen Compile user-written weighting functions (C,U)

- - See also NMR Spectroscopy User Guide; User Programming
 - Related wtfile User-defined weighting in directly detected dimension (P) wtfile2 User-defined weighting in 2nd indirectly detected dimension (P)

wtfile2 User-defined weighting in 2nd indirectly detected dimension (P)

Description Set to the name of the file containing the user-written weighting function along the second indirectly detected dimension. This dimension is often referred to as the f₂ dimension of a multidimensional data set. wtfile2 can be set with wti on the 2D interferogram data. Otherwise, wtfile2 is analogous to wtfile.
See also NMR Spectroscopy User Guide; User Programming
Related wtfile User-defined weighting in directly detected dimension (P) wtfile1 User-defined weighting in 1st indirectly detected dimension (P)
wti Interactive weighting (C)

wtgen Compile user-written weighting functions (M,U)

Syntax (From VnmrJ) wtgen(file<.c>)
 (From UNIX) wtgen file<.c>
Description Allows compilation of a user-written weighting function that
 subsequently can be executed from within VnmrJ. wtgen performs the
 following functions:
 Checks for the existence of the /vnmr/bin directory and aborts if
 the directory is not found.
 Checks for files usrwt.o and weight.h in the /vnmr/bin directory
 and aborts if either of these two files cannot be found there.
 Checks for the existence of the user's directory and creates this
 directory if it does not already exist.
 Establishes in the wtlib directory soft links to usrwt.o and
 weight.h in the /vnmr/bin directory.
 }
}

- Compiles the user-written weighting function, which is stored in the wtlib directory, link loads it with usrwt.o, and places the executable program in the same directory; any compilation and/or link loading errors are placed in the file errmsg in wtlib.
- Removes the soft links to usrwt.o and weight.h in the /vnmr/bin directory.

The name of the executable program is the same as that for the source file without a file extension (e.g., testwt.c is the source file for the executable file testwt).

Examples (From VnmrJ) wtgen('testwt') (From UNIX) wtgen testwt.c

See also User Programming

wti Interactive weighting (C)

Syntax wti<(element_number)>

Description Allows weighting parameters to be set interactively for both t2 FID's and t1 interferograms. The optional argument "fidnum" specifies which FID element or interferogram trace is to be used in adjusting the weighting parameters. The default value is the currently active element or trace. These commands respond appropriately to "phfid" and "lsfid" for t2 FID's and to "phfid1" and "lsfid1" for t1 interferograms.

Parameters

1b, 1b1 - line broadening factor in Hz; a positive value gives sensitivity enhancement; a negative value gives resolution enhancement.

sb, sb1 - sine bell time period in sec; a negative value give a sine squared bell.

sbs, sbs1 - sine bell shift in sec; shifts the origin of the sine bell; active only if sb (or sb1) is active.

gf, gf1 - gaussian apodization constant in sec.

gfs, gfs1 - gaussian function shift in sec; shifts the origin of the gaussian function; active only if gf (or gf1) is active.

sa – sampling window in data points. Range is 8 to np/2. All points in the FID greater than sa will be set to zero. The FID points start from point 1 at the beginning of the FID and np/2 at the end of the FID.

sas - sampling window shift in data points. All points in the FID less than or equal to sas will be set to zero. The FID points start from point 1 at the beginning of the FID and np/2 at the end of the FID. The minimum value of sas is 0. The maximum value is np/2 - sa.

awc, awc1 - additive weighting constant; it is added in to the weighting function after the lb and sb (sbs) contributions but before the gf (gfs) contributions

These parameters can be typed in or changed with the left mouse button in the weighting function field. "vs" and "vf" can be changed with the center button in the proper field. The right mouse button allows to turn off the spectrum for a faster response to changes in the weighting function.

See also wti

wti(3) NMR Spectroscopy User Guide

RelatedlsfidNumber of complex points to left-shift np FID (P)lsfid1Number of complex points to left-shift ni interferogram (P)phfidZero-order phasing constant for np FID (P)phfid1Zero-order phasing constant for ni interferogram (P)wtiaInteractive weighting for 2D absorptive data (C)

wtia Interactive weighting for 2D absorptive data (M)

Syntax wtia<(element_number)>

Description	Allows weighting parameters to be set interactively for both t_2 FIDs and t_1 interferograms in 2D absorptive data. Refer to the description of the wti command for further information.	
Arguments	element_number specifies which FID element or interferogram trace is to be used in adjusting the weighting parameters. The default is the currently active trace.	
See also	NMR Spectroscopy User Guide	
Related	lsfid	Number of complex points to left-shift np FID (P)
	lsfid1	Number of complex points to left-shift ni interferogram (P)
	phfid	Zero-order phasing constant for np FID (P)
	wti	Interactive weighting (C)

wtune Specify when to tune (P)

Applicability	Liquids, VnmrJ Walkup, Automation	
Description	Specify when automatic probe tuning will happen.	
Syntax	wtune ='value1 <value2>'</value2>	
Values	s 's' – when a new sample is inserted	
	'e' – before each experiment	
	'o' – change of operator	
	'v' – change of solvent	
	't' – change of temperature	

	'1' - change of high band frequency (tn or dn)	
	'2' - change of low band frequency (dn or tn)	
	'n' - do not tune, if 'n' is included in argument list, no tuning will	
	occur.	
Examples	wtune ='st12'	
	The system will tune when a new sample is inserted (s) or the temperature changes for the current or new sample (t) or there is a change in the high band frequency (tn or dn) (1) or there is a change of low band frequency (dn or tn) (2).	
See also	NMR Spectroscopy User Guide and VnmrJ Walkup	
Related	tunemethod Method to use for tuning (P)	
	protune Macro to start ProTune (M)	
	wtunedone What to do after ProTune tuning is done (P)	

wtunedone What to do after ProTune tuning is done (P)

Description	Specific what to do after ProTune tuning is done. This is a local string parameter that does not exist by default and must be created to specify a command to be executed after tuning is finished.	
See also	NMR Spectroscopy	User Guide and VnmrJ Walkup
Related	protune	Macro to start ProTune (M)
	create	Create new parameter in a parameter tree
		(C)
	wtune	Specify when to tune (P)

wysiwyg Set plot display or full display (P)

Description	Sets whether the window display is the same as the plot ("what you see is what you get," or WYSIWYG) or is expanded to fill the window. This allows the user to scale the image to the full window, making it easier to view. This parameter is in the user's global parameter file.
Values	'y' makes the window picture size depend on the current plotter setting. Scaling the window does not change the ratio of the picture. This value is the default display condition.
See also	'n' makes the window display expand, giving a full display. NMR Spectroscopy User Guide



Agilent VnmrJ 4 Command and Parameter Reference Guide

x0	X-zero position of HP pen plotter or Postscript device (P)
x1	X1 shim gradient (P)
x2y2	X2Y2 shim gradient (P)
x3	X3 shim gradient (P)
x4	X4 shim gradient (P)
xdiag	Threshold for excluding diagonal peaks when peak picking (P)
xgate	Load time counter (M)
xm1	Utility macro for study queue experiment manager (M)
xmaction	Perform study queue action (M)
xmactionw	Perform study queue action for walkup (M)
xmaddreq	Add a required protocol before the main protocol (M)
xmcheckreq	Check required protocol name (M)
xmconvert	Convert a temporarily stored study into a submitted study (M)
xmcopy	Copy protocols in a study queue (M)
xmdelete	Delete nodes in a study queue (M)
xmenablepanel	Enable or disable a parameter panel (M)
xmendq	End a chained study queue (M)
xmgetatts	Get study queue attributes (M)
xmHprescan	Set up and process Proton prescans (M)
xminit	Initialize an imaging study queue (M)
xmlockup	Move a study queue node up and lock it (M)
xmmakenode	Make a new study queue node (M)



xmnext	Find next prescan or next experiment in study queue (M)
xmprescan	Run prescans in study queue (M)
xmreact	Recover from error conditions during automation study (M)
xmreadnode	Read attributes from a study queue node (M)
xmrtpar	Retrieve parameters from a study queue node (M)
xmsample	Write enterQ entry for a sample for study queue – automation (M)
xmsara	Write sample enterQ entry for study queue— imaging (M)
xmsatfrq	Processing for Presat experiment (M)
xmselect	Action when study queue node is selected (M)
xmsetattr	Set an attribute for a study queue node (M)
xmsetatts	Set an attribute for a study queue node (M)
xmshowdata	Show data from a study queue node (M)
xmstartnightq	Start the night queue (M)
xmsubmit	Submit sample(s) to the study queue (M)
xmtime	Update the study queue time (M)
xmtune	Check tune parameter during automation (M)
xmwerr	Recover from acquisition error in study queue (M)
xmwexp	Processing macro for end of acquisition in study queue (M)
xmwritenode	Write study queue node attributes (M)
xmwritesq	Write study queue node order (M)
xpol	Cross-polarization (P)
xpolar1	Set up parameters for XPOLAR1 pulse sequence (M)
ху	XY shim gradient (P)
XZ	XZ shim gradient (P)
xz2	XZ2 shim gradient (P)

x0 X-zero position of HP pen plotter or Postscript device (P)

Applicability Systems with a Hewlett-Packard pen plotter or a Postscript output device. Adjusts the x-zero position on the chart. Use hpa to adjust x0 (and Description y_0) to place the numbers in a pleasing position when filled in on the blank lines. x0 is part of vnmrsys/global and hence common to all experiments. Number, in mm. Values See also NMR Spectroscopy User Guide Related hpa Plot parameters on special preprinted chart paper (C) y0 Y-zero position of HP plotter or Postscript device (P)

x1 X1 shim gradient (P)

Description	Holds current setting of the X1 radial shim gradient.			
Values	If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no curren			
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.			
See also	NMR Spectroscopy User Guide			
Related	shimset Type of shim set (P)			

x2y2 X2Y2 shim gradient (P)

Description Holds current setting of the X2Y2 radial shim gradient.
Values If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide Related shimset Type of shim set (P)

x3 X3 shim gradient (P)

Description	Holds current setting of the X3 radial shim gradient.		
Values	If shimset is 1, 2, 10: -2048 to +2047, steps of 1, 0 is no current.		
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.		
See also	NMR Spectroscopy User Guide		
Related	shimset Type of shim set (P)		

x4 X4 shim gradient (P)

Х

Description	Holds current setting of the X4 radial shim gradient.
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

xdiag Threshold for excluding diagonal peaks when peak picking (P)

Description Used by the 112d program to exclude diagonal peaks when peak picking.
To create the 2D peak picking parameters xdiag and th2d in the current experiment, enter addpar('112d').
Values Peaks within xdiag Hz of the diagonal will not be picked by 112d. Setting xdiag to 0.0 will cause 112d to pick all peaks, including diagonal peaks.
See also NMR Spectroscopy User Guide
Related addpar Add selected parameters to the current experiment (M) 112d Automatic and interactive 2D peak picking (C)

th2d Threshold for integrating peaks in 2D spectra (P)

xgate Load time counter (M)

Applicability Systems with a solids module.

Syntax xgate(counts)

Description Loads the (12-bit) time counter on the pulse programmer with the specified number of counts and switches the counter to the external time base (the external trigger). On each trigger, the counter counts one unit down, and the next pulse sequence event starts when the count reaches zero. Often that time count will be just 1 (1.0, as the argument must be a floating point number). If the final pulse is to be performed after a longer delay, two options are available:

• Perform a normal delay, followed by the xgate(1.0) call.

• Calculate how many rotor cycles that delay would be (calculation is typically done based on a parameter srate) and then perform xgate with that calculated number of rotor triggers. Be aware that the only number of rotor cycles that can be counted this way is 4096, because the pulse programmer uses a 12-bit counter). At typical rotor speeds of 5 to 10 kHz, the "counted" delay is limited to 0.8 to 0.4 seconds.

Arguments counts is the number of counts to load into the time counter. The value must be a floating point number.

Examples	xgate(5.0)
See also	User Guide: Solid-State NMR; VNMR Pulse Sequences
Related	srate Spinning rate for magic angle spinning (P)

xm1 Utility macro for study queue experiment manager (M)

Description A utility macro for setting study queue attributes and other study queue operations. Usually called from other macros, and not from the command line.

xmaction Perform study queue action (M)

Applicability	VnmrJ Walkup, Imaging		
Description	Perform an action on an experiment node in the study queue. Usually		
	called from study queue actions, and not from the command line.		

xmactionw Perform study queue action for walkup (M)

Applicability	VnmrJ Walkup
Description	Perform an action on an experiment node in the study queue. Usually
	called from other macros, and not from the command line.

xmaddreq Add a required protocol before the main protocol (M)

Applicability	VnmrJ Walkup, Imaging	
Description	Add a required protocol before the main protocol, when adding a protocol to the study queue. Usually called from other macros, and not from the command line.	
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide	
Related	xmmakenode Make a new study queue node (M)	

xmcheckreq Check required protocol name (M)

Applicability VnmrJ Walkup, Imaging

Description	Check if a required protocol exists in the study queue, and return		
	full path filename to data, if data has been acquired. Usually called		
	from plotting macros, and not from the command line.		
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide		
Related	complet Macro to perform generic 2D plot (M)		
	plot2D Plot 2D spectra (M)		

xmconvert Convert a temporarily stored study into a submitted study (M)

Applicability	VnmrJ Walkup, Imaging		
Description	Convert a temporarily stored study into a submitted study. Usually		
	only called from	other macros.	
See also	VnmrJ Walkup,	VnmrJ Imaging User's Guide	
Related	xmsubmit	Submit sample(s) to the study queue (M)	

xmcopy Copy protocols in a study queue (M)

Applicability	VnmrJ Walkup, Imaging
Description	Copy protocols within a study queue. Usually only called from other macros.
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide
Related	xmaction Perform study queue action (M)
	xmactionw Perform study queue action for walkup (M)

xmdelete Delete nodes in a study queue (M)

Applicability	VnmrJ Walkup, Imaging	
Description	Delete nodes within a study queue. Usually only called from other macros.	
See also	VnmrJ Walkup,	VnmrJ Imaging User's Guide
Related	sqfilemenu	Study queue file menu commands (M)
	xmaction	Perform study queue action (M)
	xmactionw	Perform study queue action for walkup (M)

xmenablepanel Enable or disable a parameter panel (M)

Description Enable or disable a parameter panel. Usually used to disable the Acquire panel for Imaging applications. Usually called only from a panel.

xmendq End a chained study queue (M)

Applicability	VnmrJ Walkup		
Description	End a chained study queue in the Walkup interface. Usually called by other macros.		
See also	VnmrJ Walkup		
Related	\times mnext Find next prescan or next experiment in study queue (M)		

xmgetatts Get study queue attributes (M)

Applicability	VnmrJ Walkup, Ima	aging
Description	Get study queue att	ributes.
See also	VnmrJ Walkup, Vn	mrJ Imaging User's Guide
Related	xmaction	Perform study queue action (M)

xmHprescan Set up and process Proton prescans (M)

Applicability	VnmrJ Walkup	
Description	A macro to set up and process prescans for Proton-type experiments (Proton, Presat, or Wet1d protocols). Usually called from other macros, and not from the command line.	
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide	
Related	HprescanProton prescan (P)stdldApptype macro for Standard 1D experiments (M)	

xminit Initialize an imaging study queue (M)

Applicability	Imaging
Description	Initialize an imaging study queue. Usually called from other macros, and not from the command line.

See also	VnmrJ Imaging	User's Guide	
Related	sqfilemenu	Study queue file menu commands (M)

xmlockup Move a study queue node up and lock it (M)

Applicability	VnmrJ Walkup, Imaging
Description	A macro to move a study queue node up above other completed nodes in the study queue, and lock it so it cannot be moved. This is usually done just prior to acquisition. Usually called from other macros, and not from the command line.
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide
Related	acquire Acquire data (M)

xmmakenode Make a new study queue node (M)

VnmrJ Walkup, Imaging		
Create a new node in the study queue. Usually only called by other		
macros.		
VnmrJ Walkup, VnmrJ Imaging User's Guide		
locactionLocator action (M)xmaddreqAdd a required protocol before the main protocol (M)		

xmnext Find next prescan or next experiment in study queue (M)

Applicability	VnmrJ Walkup		
Description	Find the next prescan or next experiment in a study queue. It is used for chaining prescans and experiments. Usually only called by other macros.		
See also	VnmrJ Walkup		
Related	acquire	Acquire data (M)	
	startq	Start a chained study queue (M)	
	xmprescan Run prescans in study queue (M)		
	xmwexp	Processing macro for end of acquisition in study queue	
		(M)	

xmprescan Run prescans in study queue (M)

Applicability VnmrJ Walkup

Description	Run prescans in a study queue. Usually only called by other macros				
See also	VnmrJ Walkup				
Related	cqfindz0	cqfindz0 Run an experiment to find the value of z0 (M)			
	gmapshim Start gradient autoshimming (M)				
	prescan	Study queue prescan (P)			
	xmnext	Find next prescan or next experiment in study queue			
		(M)			

xmreact Recover from error conditions during automation study (M)

Applicability	VnmrJ Walkup		
Description	A macro to recover from error conditions during a study queue automated acquisition. Usually only called by other macros.		
See also	VnmrJ Walkup		
Related	acquire Acquire data (M) react Recover from error conditions during werr processing (M)		

xmreadnode Read attributes from a study queue node (M)

Applicability	VnmrJ Walkup, Imaging		
Description	Read attributes from a study queue node. Usually only called by other macros		
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide.		
Related	xmactionPerform study queue action (M)xmactionwPerform study queue action for walkup (M)reactRecover from error conditions during werr processing		
		(M)	

xmrtpar Retrieve parameters from a study queue node (M)

Applicability	Imaging		
Description	Retrieve parameters from a study queue node after its parameters have been customized. Usually only called by other macros.		
See also	VnmrJ Imaging User's Guide		
Related	xmmakenode xmselect	Make a new study queue node (M) Action when study queue node is selected (M)	

xmsample Write enterQ entry for a sample for study queue - liquids (M)

Applicability	<i>VnmrJ Walkup</i> , syste	ems with automation such as sample changer or
Description	Write the information required for a sample in the study queue when the sample is submitted. Usually only called by other macros.	
See also	VnmrJ Walkup	
Related	loc	Location of sample in tray (P)
	xmsubmit	Submit sample(s) to the study queue (M)

xmsara Write enterQ entry for a sample for study queue – imaging (M)

Applicability	Imaging
Description	Halt or resume acquisition in the study queue, especially when using
	multiple viewports. Usually only called from interface panels.

xmsatfrq Processing for Presat experiment (M)

Applicability	VnmrJ Walkup	
Description	A macro to handle processing steps for the Presat experiment. It is optimized for use with water. Usually only called from other macros.	
See also	VnmrJ Walkup	
Related	xmHprescan	Set up and process Proton prescans (M)

xmselect Action when study queue node is selected (M)

Applicability	VnmrJ Walkup		
Description	A macro to specify the action taken when a study queue node is selected by double-clicking on it. The action depends on the node status, which is Ready for acquisition, Executing, Completed, etc. The macro also runs the macros associated with selecting a study queue node, and saves the parameters of the current node before retrieving parameters of the selected node.		
See also	VnmrJ Walku		
Related	xmaction Perform study queue action (M)		
	xmactionw Perform study queue action for walkup (M)		
	xmrtpar Retrieve parameters from a study queue node (M)		

xmsetatts Set an attribute for a study queue node (M)

Applicability	VnmrJ Wal	kup, Imaging	
Description	Set an attribute for a study queue node.		
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide		
Related	xmaction	Load colors for graphics window and plotters (M)	
	xmaction	Location of sample in tray (P)	
	W		

xmsetattr Set an attribute for a study queue node (M)

Applicability	VnmrJ Walk	up, Imaging
Description	Set an attrib	ute for a study queue node.
See also	VnmrJ Walk	up, VnmrJ Imaging User's Guide
Related	xmaction Load colors for graphics window and plotters (M)	
	xmactionw	Location of sample in tray (P)

xmshowdata Show data from a study queue node (M)

Applicability	VnmrJ Walkup, Imaging		
Description	A macro that retrieves data from a completed study queue node. In the Walkup liquids interface, data is also processed if <i>Process data on</i> <i>drag-and-drop</i> from locator is selected in the <i>System settings</i> dialog in the Utilities menu.		
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide		
Related	xmselect Action when study queue node is selected (M)		

xmstartnightq Start the night queue (M)

Applicability	VnmrJ Walkup
Description	Start the night queue. It also is used to initialize the night queue settings in the Utilities menu.
Examples	xmstartnightq start the night queue xmstartnightq('at') initialize the night queue settings.
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide
Related	walkup Walkup automation (M)

X

xmsubmit Submit sample(s) to the study queue (M)

Applicability	<i>VnmrJ Walkup</i> , systems with automation such as sample changer or LC-NMR.	
Description	Submit the sample or samples selected in the study queue tray. If the Submit DayQ button below the study queue area is selected, samples are submitted to the DayQ. If the Submit NightQ button is selected, samples are submitted to the NightQ.	
See also	VnmrJ Walkup	
Related	xmsample	Write enterQ entry for a sample for study queue – automation (M)

xmtime Update the study queue time (M)

Applicability	VnmrJ Walkup, syste	ems with automation such as sample changer or
Description	Update the study que called from panels or	eue time for both DayQ and NightQ. Usually only c other macros.
See also	VnmrJ Walkup	
Related	sqfilemenu	Study queue file menu commands (M)
	startq	Start a chained study queue (M)
	studytime	Study time (P)
	xmsubmit	Submit sample(s) to the study queue (M)

xmtune Check tune parameter during automation (M)

Applicability	Automation	
Syntax	xmtune	
Description	Check tune parameters in the study queue during automation and determine if tuning will occur. Macro is usually called from within automation and not from the command line.	
See also	NMR Spectroscopy User Guide and VnmrJ Walkup	
Related	protuneMacro to start ProTune (M)tunemethodMethod to use for tuning (P)wtuneSpecify when to tune (P)	

xmwerr

Recover from acquisition error in study queue (M)

Applicability VnmrJ Walkup, Imaging
Description	Recover from an acquisition error in a study queue when not runnin		
	automation. Usually only called from other macros.		
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide		
Related	acquire Acquire data (M)		
	xmreact Recover from error conditions during automation study (M)		

EXAMPLE 2 Processing macro for end of acquisition in study queue (M)

Applicability	VnmrJ Wa	lkup, Imaging
Description	A processing macro; runs at the end of acquisition in the study queue and keeps track of study queue parameters and settings. Usually only called from other macros.	
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide	
Related	acquire xmreact	Acquire data (M) Recover from error conditions during automation study (M)

xmwritenode Write study queue node attributes (M)

Applicability	VnmrJ Wall	kup, Imaging
Description	Write study macros.	queue node attributes. Usually only called from other
See also	VnmrJ Wall	kup, VnmrJ Imaging User's Guide
Related	xmaction	Load colors for graphics window and plotters (M)
	xmactionw	Location of sample in tray (P)
	xmsetattr	Set an attribute for a study queue node (M)

xmwritesq Write study queue node order (M)

Applicability	VnmrJ Walkup, Imaging
Description	Write the study queue node order. Usually only called from other macros.
See also	VnmrJ Walkup, VnmrJ Imaging User's Guide
Related	xmactionLoad colors for graphics window and plotters (M)xmactionwLocation of sample in tray (P)

xpol Cross-polarization (P)

Applicability	Systems with	n a solids module.
Description	Selects cross- experiments	-polarization or direct polarization in solid-state NMR such asXPOLAR1.
Values	'n' sets the 'y' sets the	experiment for direct polarization. experiment for cross-polarization.
See also	User Guide:	Solid-State NMR
Related	xpolar1	Set up parameters for XPOLAR1 pulse sequence (M)

xpolar1 Set up parameters for XPOLAR1 pulse sequence (M)

Applicability	Systems with	n solids modules.
Description	Sets up the susing the particular functionality	solid-state NMR cross-polarization experiment XPOLAR rameters. Otherwise, xpolar1 contains the same as xpolar.
See also	User Guide:	Solid-State NMR
Related	hsrotor	Display rotor speed for solids operation (P)
	rotorsync	Rotor synchronization (P)

xy XY shim gradient (P)

Description	Holds current setting of the XY radial shim gradient.
Values	If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

xz XZ shim gradient (P)

Description	Holds current setting of the XZ radial shim gradient.
Values	If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

xz2 XZ2 shim gradient (P)

Description Holds current setting of XZ2 radial shim gradient.
Values If shimset is 2, 8: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide

Related shimset Type of shim set (P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

У0	Y-zero position of HP pen plotter or Postscript device (P)
y1	Y1 shim gradient (P)
у3	Y3 shim gradient (P)
у4	Y4 shim gradient (P)
Уz	YZ shim gradient (P)
yz2	YZ2 shim gradient (P)

YO Y-zero position of HP pen plotter or Postscript device (P)

Applicability	Systems with device.	a Hewlett-Packard pen plotter or a Postscript output
Description	Adjusts the y-zero position on the chart. Use hpa to adjust $y0$ (and $x0$) to place numbers in a pleasing position when filled in on the bland lines. $y0$ is part of vnmrsys/global; therefore, it is common to all experiments.	
Values	Number, in n	nm.
See also	NMR Spectroscopy User Guide	
Related	hpa	Plot parameters on special preprinted chart paper (C)
	x 0	X-zero position of HP plotter or Postscript device (P)

y1

Y1 shim gradient (P)

Description Holds current setting of the Y1 radial shim gradient.



Values If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide
Related shimset Type of shim set (P)

Y3 Y3 shim gradient (P)

Description Holds current setting of the Y3 radial shim gradient.
Values If shimset is 1, 2, 10: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide Related shimset Type of shim set (P)

y4 Y4 shim gradient (P)

Description	Holds current setting of the Y4 radial shim gradient
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

yz YZ shim gradient (P)

Description Holds current setting of the YZ radial shim gradient.
Values If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide Related shimset Type of shim set (P)

yz2 YZ2 shim gradient (P)

Description	Holds current setting of the YZ2 radial shim gradient.
Values	If shimset is 2, 8: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)



Agilent VnmrJ 4 Command and Parameter Reference Guide

Z	Add integral reset point at cursor position (C)
zO	Z0 field position (P)
z1	Z1 shim gradient (P)
z1c	Z1C shim gradient (P)
z2	Z2 shim gradient (P)
z2c	Z2C shim gradient (P)
z2x2y2	Z2X2Y2 shim gradient (P)
z2x3	Z2X3 shim gradient (P)
z2xy	Z2XY shim gradient (P)
z2y3	Z2Y3 shim gradient (P)
z3	Z3 shim gradient (P)
z3c	Z3C shim gradient (P)
z3x	Z3X shim gradient (P)
z3x2y2	Z3X2Y2 shim gradient (P)
z3x3	Z3X3 shim gradient (P)
z3xy	Z3XY shim gradient (P)
z3y	Z3Y shim gradient (P)
z3y3	Z3Y3 shim gradient (P)
z4	Z4 shim gradient (P)
z4c	Z4C shim gradient (P)
z4x	Z4X shim gradient (P)
z4x2y2	Z4X2Y2 shim gradient (P)
z4xy	Z4XY shim gradient (P)
z4y	Z4Y shim gradient (P)
z5	Z5 shim gradient (P)



z5x	Z5X shim gradient (P)
z5y	Z5Y shim gradient (P)
z6	Z6 shim gradient (P)
z7	Z7 shim gradient (P)
z8	Z8 shim gradient (P)
zeroneg	Set all negative intensities of 2D spectra to zero (C)
zoom	Adjust display to given width (M)
zx2y2	ZX2Y2 shim gradient (P)
zx3	ZX3 shim gradient (P)
zxy	ZXY shim gradient (P)
zy3	ZY3 shim gradient (P)

Add integral reset point at cursor position (C)

Syntax z<(reset1,reset2,...)>

Description	Resets the int The command generally be (resets). The is changed.	tegral to zero at the point marked by the displayed cursor. It cz removes all such integral resets and it should used before starting to enter a series of integral zeros resets are stored as frequencies and do not change if fn
Arguments	reset1, res The default is order.	set2, are reset points entered, in either Hz or ppm. the cursor position). Reset points can be entered in any
Examples	Z	
	z(7.5*sfrq	,5*sfrq,2.5*sfrq,0.1*sfrq)
See also	NMR Spectro	scopy User Guide
Related	CZ	Clear integral reset points (C)
	dlni	Display list of normalized integrals (C)
	ds	Display a spectrum (C)
	fn	Fourier number in directly detected dimension (P)
	nli	Find integral values (C)

z0 Z0 field position (P)

Description Holds current setting of the Z0 setting. The value of z0 can be set by su. lockfreq can be used to find the lock signal or resonance. To use the lock frequency, deactivate z0 by typing the statement z0='n'. To activate z0, enter z0='y'.

 \mathbf{z}

Values	If shimset i	s 1, 2, 8, 10: –2048 to +2047, steps of 1, 0 is no current
	If shimset i	s 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current
See also	NMR Spectro	scopy User Guide
Related	lockfreq	Lock frequency (P)
	su	Submit a setup experiment to acquisition (M)

z1 Z1 shim gradient (P)

Description Holds current setting of the Z1 axial shim gradient.
Values If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide Related shimset Type of shim set (P)

z1c Z1C shim gradient (P)

Description	Holds current setting of the Z1C axial shim gradient.
Values	If shimset is 1, 2, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 5 or 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

z2 Z2 shim gradient (P)

Description	Holds current setting of the Z2 axial shim gradient.
Values	If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

z2c Z2C shim gradient (P)

Description	Holds current setting of the Z2C axial shim gradient.
Values	If shimset is 1, 2, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 5 or 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

z2x2y2 Z2X2Y2 shim gradient (P)

Description Holds current setting of the Z2X2Y2 radial shim gradient. Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z2x3 Z2X3 shim gradient (P)

DescriptionHolds current setting of the Z2X3 radial shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z2xy Z2XY shim gradient (P)

Description	Holds current setting of the Z2XY radial shim gradient.
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide

z2y3 Z2Y3 shim gradient (P)

DescriptionHolds current setting of the Z2Y3 radial shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z3 Z3 shim gradient (P)

Description	Holds current setting of the Z3 axial shim gradient.
Values	If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to $+32767$, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

z3c Z3C shim gradient (P)

Description Holds current setting of the Z3C radial shim gradient.

Ζ

Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z3x Z3X shim gradient (P)

Description Holds current setting of the Z3X radial shim gradient. Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z3x2y2 Z3X2Y2 shim gradient (P)

Description	Holds current setting of the Z3X2Y2 radial shim gradient
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide

z3x3 Z3X3 shim gradient (P)

DescriptionHolds current setting of the Z2X3 radial shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z3xy Z3XY shim gradient (P)

DescriptionHolds current setting of the Z3XY radial shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z3y Z3Y shim gradient (P)

Description Holds current setting of the Z3Y radial shim gradient. Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z3y3 Z3Y3 shim gradient (P)

Ζ

Description Holds current setting of the Z3Y3 radial shim gradient. Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z4 Z4 shim gradient (P)

Description Holds current setting of the Z4 shim gradient.
Values If shimset is 1, 2, 8, 10: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide Related shimset Type of shim set (P)

z4c Z4C shim gradient (P)

DescriptionHolds current setting of the Z4C shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z4x Z4X shim gradient (P)

Description Holds current setting of the Z4X shim gradient. Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z4x2y2 Z4X2Y2 shim gradient (P)

DescriptionHolds current setting of the Z4X2Y2 radial shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z4xy Z4XY shim gradient (P)

Description Holds current setting of the Z4XY radial shim gradient.

Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

z4y Z4Y shim gradient (P)

DescriptionHolds current setting of the Z4Y shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z5 Z5 shim gradient (P)

Description	Holds current setting of the Z5 axial shim gradient.
Values	If shimset is 2, 10: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

z5x Z5X shim gradient (P)

Description	Holds current setting of the Z5X radial shim gradient
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide

z5y Z5Y shim gradient (P)

Description	Holds current setting of the Z5Y radial shim gradient
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide

z6 Z6 shim gradient (P)

Description	Holds current setting of the Z6 axial shim gradient.
Values	-32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide

z7 Z7 shim gradient (P)

DescriptionHolds current setting of the Z7 axial shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

z8 Z8 shim gradient (P)

DescriptionHolds current setting of the Z8 shim gradient.Values-32768 to +32767, steps of 1, 0 is no current.See alsoNMR Spectroscopy User Guide

zeroneg Set all negative intensities of 2D spectra to zero (C)

Description	Sets to zero all negative intensities of 2D-J spectra.		
See also	NMR Spectroscopy User Guide		
Related	foldj	Fold J-resolved 2D spectrum about $f_1=0$ axis (C)	
	rotate	Rotate 2D data (C)	

zoom Adjust display to given width (M)

Syntax	zoom(width)
Description	Adjusts the display limits. It is useful in the display of powder patterns after split has been used. zoom both zooms in and out from the current display.
Arguments	width is the total display width, in Hz. Display limits are set to \pm width/2.
See also	NMR Spectroscopy User Guide
Related	split Split the difference between two cursors (M)

zx2y2 ZX2Y2 shim gradient (P)

Description	Holds current setting of the ZX2Y2 shim gradient.
Values	If shimset is 2, 8: -2048 to +2047, steps of 1, 0 is no current.
	If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also	NMR Spectroscopy User Guide
Related	shimset Type of shim set (P)

Description Holds current setting of the ZX3 shim gradient. Values -32768 to +32767, steps of 1, 0 is no current. See also NMR Spectroscopy User Guide

ZXY ZXY shim gradient (P)

Description Holds current setting of the ZXY shim gradient.
Values If shimset is 2, 8: -2048 to +2047, steps of 1, 0 is no current. If shimset is 3 to 7, 9: -32768 to +32767, steps of 1, 0 is no current.
See also NMR Spectroscopy User Guide Related shimset Type of shim set (P)

zy3 ZY3 shim gradient (P)

DescriptionHolds current setting of the ZY3 shim gradient.Values-32768 to +32767, steps of 1, 0 as no current.See alsoNMR Spectroscopy User Guide



© Agilent Technologies, Inc. Printed in USA, Rev. A, March 2013 G7446-90519

